

# Gone fishing: Adélie penguin site-specific foraging tactics and breeding performance

SILVIA OLMASTRONI <sup>1,2</sup>, NICCOLÒ FATTORINI <sup>3</sup>, FRANCESCO PEZZO<sup>4</sup> and SILVANO FOCARDI<sup>1</sup>

<sup>1</sup>*Dipartimento di Scienze Fisiche, della Terra e dell'Ambiente, Università di Siena, Via Mattioli 4, 53100 Siena, Italy*

<sup>2</sup>*Museo Nazionale dell'Antartide 'F. Ippolito', Via Laterina 8, 53100 Siena, Italy*

<sup>3</sup>*Dipartimento di Scienze della Vita, Università di Siena, Via Mattioli 4, 53100 Siena, Italy*

<sup>4</sup>*Istituto Superiore per la Protezione e la Ricerca Ambientale (ISPRA), via Ca Fornacetta 9, 40064 Ozzano Emilia (BO), Italy*  
[silvia.olmastroni@unisi.it](mailto:silvia.olmastroni@unisi.it)

**Abstract:** The ecological drivers underlying breeding performance are expected to differ across the geographical range of seabird species, but few studies have compared trade-offs between colonies with different local conditions. During chick-rearing (2000–01), we compared the foraging trips, diet and breeding parameters of two Adélie penguin colonies in the Ross Sea, at Edmonson Point (EdPo; ~2000 breeding pairs) and Inexpressible Island (InIs; ~24 000 breeding pairs). Penguins from InIs travelled farther and performed longer feeding trips. The quantity of food brought to the nest was the same for the two colonies, but penguins from InIs brought more fish and less krill. Eggs hatched earlier at EdPo. Breeding success did not differ, but chick weight during hatching–fledging was greater at InIs. Despite worse weather conditions at InIs, the larger proportion of high-energy food brought by penguins from InIs (i.e. fish) may explain their offspring's better performance. In addition, the persistence of fast ice at EdPo may have led to greater energy expenditure of breeding individuals, possibly reducing chick growth. The greater intraspecific competition expected at InIs may have been reduced by longer foraging trips and/or counteracted by the more nutritious diet. Our findings reveal complex trade-offs between foraging effort and environmental constraints in determining the breeding performance of Adélie penguins.

Received 5 July 2019, accepted 10 January 2020

**Key words:** Adélie penguin, breeding success, diet, foraging trip, polynya, Ross Sea

## Introduction

Seabird breeding performance is expected to vary regionally as a consequence of ecological drivers affecting species across their range. Antarctic seabirds have to cope with extreme environmental stressors to survive and to reproduce successfully: sea-ice conditions, weather and food availability are reported to be crucial determinants shaping the distributions and life histories of these species (Wilson *et al.* 2001, Ainley *et al.* 2005, Jenouvrier *et al.* 2005, 2009). Seabirds inhabiting polar regions consequently show high geographical variability in foraging strategies and breeding performance (Inchausti *et al.* 2003, Tremblay & Cherel 2003). Sea ice, in particular, is a dynamic structural element of the polar environment. Antarctic seabirds have evolved a number of different ecological adaptations to cope in this highly variable habitat that shapes their breeding and feeding activity (e.g. Ainley *et al.* 1984, Smith *et al.* 1999, Le Guen *et al.* 2018). Continuous and rapid changes in sea ice during the breeding season make it necessary for

birds such as penguins to deal with highly variable situations between the nesting site and foraging areas (Forcada & Trathan 2009).

The Adélie penguin, *Pygoscelis adeliae* (Hombron & Jacquinot 1841), is one of two penguin species that only breed on the Antarctic continent and is relatively abundant along the Antarctic coasts (the other is the Emperor penguin, *Aptenodytes forsteri*, Gray 1844). *Pygoscelis adeliae* has developed extreme adaptations in feeding behaviour, such as diving. It breeds in colonies in ice-free areas during the summer. It needs land to breed and food to be available within a limited foraging range and at critical times during the breeding season. Physical factors related to environmental variability, including extent and quality of sea ice (e.g. Watanuki *et al.* 2002, Ropert-Coudert *et al.* 2015, Widmann *et al.* 2015), as well as oceanographic conditions (Forcada & Trathan 2009), affect the availability (i.e. abundance and distribution) of their principal prey (euphausiaceans and fish). Local meso-scale pack-ice cover is also known to affect primary productivity and therefore to affect prey availability

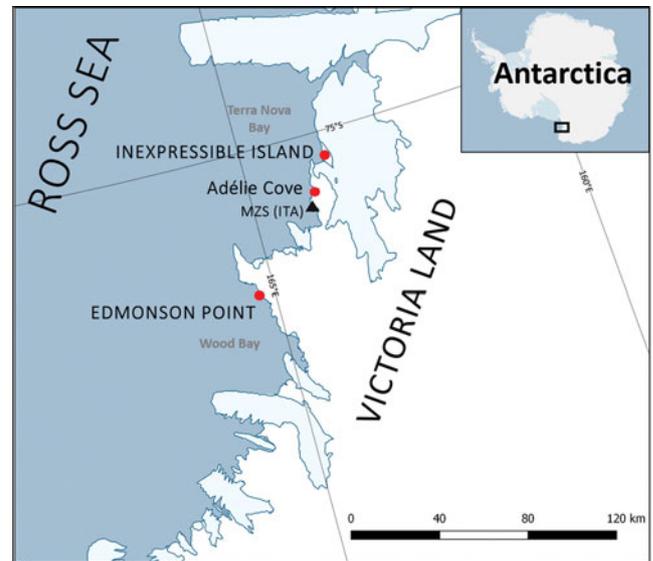
(Arrigo *et al.* 2002). Individual traits such as sex, age and physiological status are also known to influence foraging behaviour (Ballard *et al.* 2010). Access to prey can be influenced by population size due to intraspecific competition for food resources (Ainley *et al.* 2004, Ballance *et al.* 2009).

In the Ross Sea region, the diet of Adélie penguins during the reproductive period consists mainly of crystal krill, *Euphausia crystallophias*, and Antarctic silverfish, *Pleuragramma antarctica* (Clarke *et al.* 1998, Olmastroni *et al.* 2000, Ainley *et al.* 2003, 2018). These two species are at intermediate levels in the pelagic food web. Antarctic krill, *Euphausia superba*, is a dietary item of penguins of northern colonies, but it is relatively rare in the stomach contents of penguins nesting further south in Victoria Land (Ainley 2002). The distribution of krill species has various latitudinal and longitudinal gradients, with crystal krill living closer to coasts and the pelagic Antarctic krill preferring the continental slope (Azzali *et al.* 2006). This prey is influenced by sea-ice distribution (Davis *et al.* 2017), which in turn affects the aggregation and foraging trips of top predators.

Previous studies on the Adélie penguin have documented various foraging tactics in colonies affected by sea ice on various spatiotemporal scales (Ainley *et al.* 1998, Clarke *et al.* 1998, Watanuki *et al.* 2002, 2004, Widmann *et al.* 2015, Le Guen *et al.* 2018), although studies comparing site-specific foraging strategies, diet and breeding performance are still needed in the Victoria Land sector. Comparing colonies where penguins may adopt various foraging strategies and/or yield various levels of breeding performance is useful to predict variations in population-specific responses to environmental changes.

In this study, we observed and compared differences in foraging trips, diet, breeding success and chick growth rate of Adélie penguins in two colonies in central Victoria Land (Ross Sea) during summer 2000–01. The colonies had different population sizes and were exposed to specific local environmental conditions.

It can be expected that birds breeding at colonies relatively close to each other but exposed to different local sea-ice conditions have site-specific foraging patterns. Our two study colonies are part of a clustered group of three in central Victoria Land (see clusters 7–9 in Ainley *et al.* 1995, p. 174) and differ as much as tenfold in breeding population size. This difference in numbers should lead to differences in intraspecific competition over food resources, which may in turn have cascading effects on foraging strategies, diet and breeding performance (Birt *et al.* 1987, Ainley *et al.* 1995). The two colonies also differ in local sea-ice dynamics, leading to potential differences in prey distribution and foraging costs for penguins. Until late January, the smaller colony is generally separated from the sea by several kilometres of fast ice. Conversely, the larger colony is located on



**Fig. 1.** Locations of our study colonies (Edmonson Point and Inexpressible Island) in central Victoria Land (Ross Sea). Locations of a third colony in the same area (Adélie Cove) and the Italian Research Station (Mario Zucchelli Station; MZS) are also shown.

Terra Nova Bay close to a wind-generated coastal polynya (Vacchi *et al.* 2012), which may play an important role in shaping foraging trips and diet (e.g. by facilitating access to foraging areas and reducing travel costs throughout the breeding season).

## Methods

### Study areas

Our study was conducted from 30 October 2000 to 10 February 2001 in two penguin colonies: Edmonson Point (EdPo; 74°20'S, 165°08'E) and Inexpressible Island (InIs; 74°54'S, 163°39'E). These areas are located in Wood Bay and Terra Nova Bay, ~75 km apart, in the Ross Sea (Antarctica) (Fig. 1). Breeding populations, measured by ground count (SC-CAMLR 2004) on 29 November and 6 December 2000, were 1987 and 24 142 breeding pairs, respectively. During the study period, fast ice at EdPo stretched for 20–24 km, while the wind-driven polynya off InIs was 20–40 km from the shore of the island. Weather at the two colonies was determined from data of the weather stations InIs 'Manuela' (<http://amrc.ssec.wisc.edu/aws/api/form.html>) and EdPo 'Penguin' (<http://www.climantartide.it>). Meteorological data recorded every 10 min were averaged daily. Differences in daily weather at the two colonies in the period from November 2000 to January 2001 ( $n=92$  days) were assessed using two-tailed paired tests (relative humidity: paired Wilcoxon test; mean daily temperature: paired  $t$ -test, after checking for normal distribution of data by the

Shapiro–Wilk test,  $W=0.985$ ,  $P=0.354$ ). Strong katabatic winds are common around InIs (Bromwich 1985), although we could not formally test differences in wind chill between colonies because anemometer data were not available for InIs. At InIs, unpredictable and extreme weather events seem to play a major role in breeding outcomes for Adélie penguins (Olmastroni, unpublished data 2000). For example, during a visit to InIs in mid-November 2000, we found that many nests had been buried in snow after a strong blizzard. Many of them failed in subsequent days (Olmastroni, personal observation 2000).

#### *Penguin capture and handling*

Whenever our data collection involved capturing penguins, either to study their foraging trips or to assess their diet, we observed the following Convention for the Conservation of Antarctic Marine Living Resources (CCAMLR) procedures. We captured penguins using a hand net. We determined the sex of adults on the basis of papillae by cloacal inspection (Sladen 1978). We marked adult penguins with passive transponders (TIRIS™, Texas Instruments Registration and Identification System) and externally with temporary marking dye before releasing them. The latter allowed us to avoid recapturing the same individual and facilitated observations to check that penguin breeding was not impaired by handling. Chicks weighed to plot their growth curve were initially marked with dye or an adjustable numbered Velcro™ band on the left flipper. Velcro flipper bands were adjusted according to chick size and were all removed at the end of the season. Handling time was less than 20 min per individual, and all penguins returned safely to their nests or crèche group.

#### *Foraging trips*

In the period from late December 2000 to January 2001, we tracked seven adult breeding penguins at EdPo and five at InIs using ST-10 Platform Terminal Transmitters (PTTs; Telonics, USA, packaged by Sirtrack, New Zealand). The devices were attached to the middle-lower back of penguins to reduce drag (Bannasch *et al.* 1994) using fast-setting cyanoacrylic glue (Loctite™ 401) and cable ties (Clarke *et al.* 2006). Birds were allowed to perform approximately two to four trips before the instruments were removed. Satellite position data were downloaded from ARGOS, and processed by SO through the Australian Antarctic Division data centre. Each foraging trip was analysed using the following criteria: colony position start/end of foraging trip; delta time between fixes 0.25 h, maximum allowed speed 7 kph; all data were filtered for minimum quality accuracy 1 (within 1 km) (Clarke *et al.* 2006). Whenever possible, consistency of penguin departure/arrival time was also checked by direct observation at the nest. Data

were analysed using *ArcView* 3.2 (ESRI, Inc.) with the *Animal Movement* extension (USGS\_BRD, Alaska Biological Science Center). We calculated the duration (decimal hours), overall distance travelled (km) and maximum linear distance travelled from the colony (km) of each foraging trip.

#### *Diet sampling and analysis*

We collected stomach contents of penguins by method A8 of CCAMLR Standard Methods (SC-CCAMLR 2004): during chick rearing, and at least 10 days after the hatching peak, adults returning from foraging trips were stomach flushed by the water-offloading technique (Wilson 1984), which was repeated in order to obtain complete stomach contents (i.e. until clear water emerged). In the period from 2 to 30 January 2001, we obtained complete stomach contents from nine individuals (three females, six males) at EdPo and ten individuals (five females, six males) at InIs. In order to avoid temporal inconsistencies, we attempted to balance diet sampling in the two study colonies (~2–3 day intervals). As penguins were individually marked, stomach contents can be regarded as independent samples (i.e. belonging to different individuals). Stomach samples were preserved in ethanol and were analysed according to SC-CCAMLR (2004) guidelines. The mass of each sample (g fresh weight) was determined after draining through a sieve. We subtracted the weight of rocks (range 0–15.36 g) from total drained stomach content weight to obtain the total weight of food brought back to the nest by penguins. Contents were divided into four main categories (krill, amphipods, fish, squid), each of which was then weighed and measured. The proportion of each prey category was calculated based on the total amount of prey ingested for each individual.

We distinguished krill species (Antarctic and crystal krill) and amphipod families (Gammariidae and Hyperiididae) according to Food and Agriculture Organization (FAO) species identification sheets (FAO 1985). As digestion reduced the number of krill specimens with an intact carapace, up to ten integral specimens of *E. superba* and *E. crystallophias* were randomly selected from each sample, and their body lengths (mm) were measured with a calliper to the nearest 1 mm ( $n=149$  *E. superba*,  $n=150$  *E. crystallophias*).

A total of 324 fish otoliths were analysed to identify fish species (preferably) or family using a 10x stereomicroscope. Recognition of otoliths to the closest taxon and measurements of otolith length were carried out following Williams & McEldowney (1990) and with the aid of an Aforo Shape Analysis of Fish Otoliths (<http://aforo.cmima.csic.es/index.jsp>). We determined the number of individuals per species and per sample by calculating the minimum number of individuals, according

to the method described by Emslie *et al.* (1998). Maximum otolith length was measured with a micrometric eyepiece (10x) to an accuracy of 0.1 mm. The lengths of 58 intact otoliths (i.e. not showing signs of erosion) of *P. antarctica* were measured in order to obtain the standard lengths of fish ingested according to the regression formula developed by Williams & McEldowney (1990, p. 109). In order to assess the variety of fish species preyed on, we calculated the Shannon–Weaver diversity ( $H'$ ) and Pielou evenness ( $J$ ) indices (Peet 1975).

### Breeding parameters

Whenever possible, hatching date was recorded (EdPo: 106 nests; InIs: 72 nests). Breeding success was assessed on 224 nests with eggs at EdPo and 241 nests with eggs at InIs. We considered the ratio of the observed number of chicks successfully in crèche to the number of nests with eggs. To determine the growth rate of offspring in each colony, chicks from at least 20 nests per colony were weighed regularly from hatching to the end of the crèche period (see Table SI in the supplementary material for breeding chronology). We also weighed 46 chicks at EdPo and 30 at InIs after fledging (2–6 February).

### Statistical analysis

We assessed differences in foraging trips between colonies using generalized linear mixed models. For each foraging trip, we evaluated three response variables separately: trip

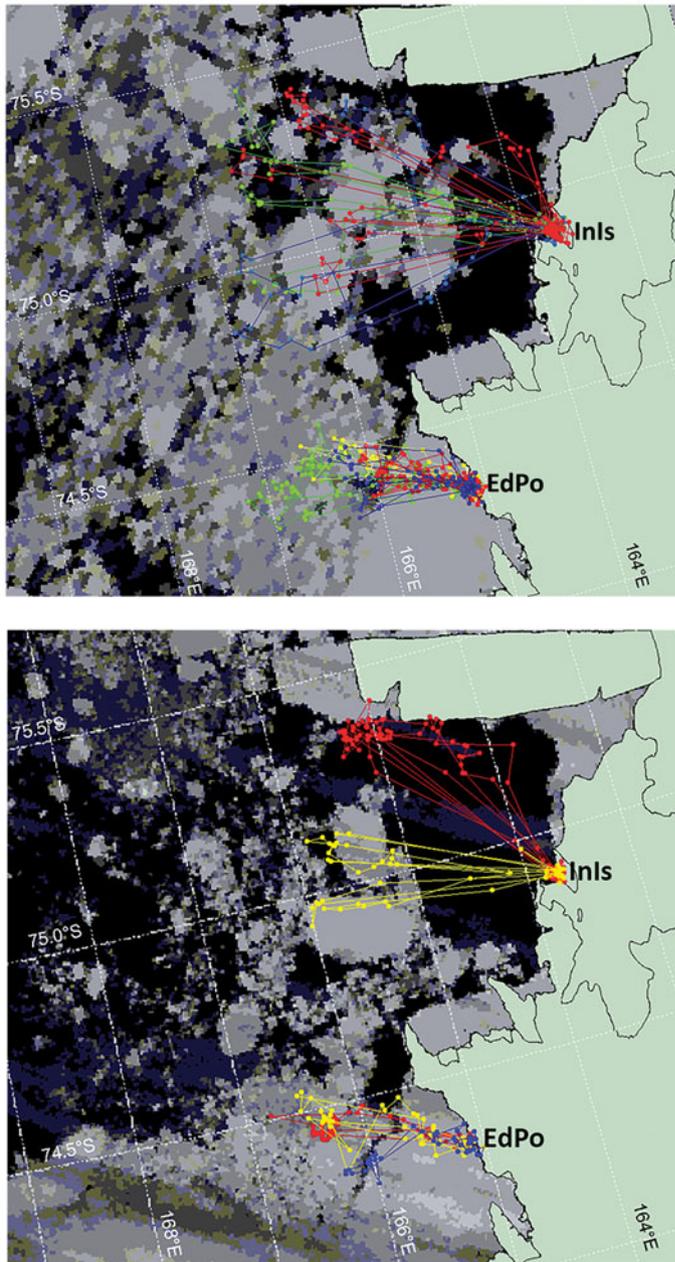
duration, overall distance travelled and maximum distance travelled from the colony. We modelled them using Gamma errors (link function: log) because response variables were  $0^+$  values and because the same analysis conducted with a Gaussian error distribution did not support normality of residuals. We included the colony as the categorical predictor (reference level: EdPo). We considered breeding period (guard *vs* crèche) and individual differences by including individual identity as a random intercept nested in the period. The data were weighted according to the number of fixes recorded in each foraging trip, and the significance of the predictor was tested by assessing whether 95% confidence intervals contained zero.

We assessed individual differences in meal quantity between colonies using the *t*-test, after validating normality and homoscedasticity assumptions (Shapiro–Wilk test, EdPo:  $W = 0.875$ ,  $P = 0.139$ , InIs:  $W = 0.898$ ,  $P = 0.208$ ; *F*-test:  $F = 1.52$ ,  $P = 0.561$ ). We looked for an overall difference in meal composition (quantities of fish, krill, amphipods and other items) between the two colonies by comparing the total amount of each food category using a  $\chi^2$  test based on 99 999 Monte Carlo permutations. Due to the non-normal distribution of the percentages of food categories per individual, we used the Mann–Whitney test to assess individual differences in prey items between the colonies, considering the four prey categories, and also considering the krill and fish species or amphipod families identified. We used the *t*-test to compare size

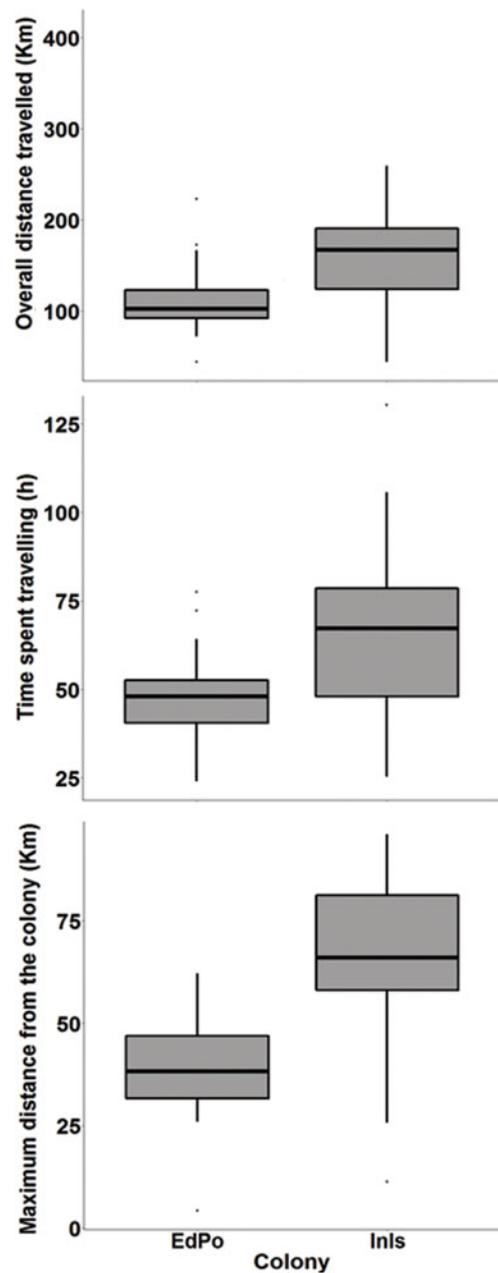
**Table I.** Results of univariate statistical tests assessing differences between the two study colonies in weather, diet, prey size and breeding parameters. For details on sample size, validation of assumptions and descriptive statistics, see text.

Variable	Test statistic	<i>P</i> -value
Weather at the colony		
Relative humidity (%)	Paired Wilcoxon test; $W = 2398$ , $Z = 1.008$	0.313
Mean daily temperature (°C)	Paired <i>t</i> -test; $t = 7.082$	< 0.001
Diet		
Quantity of food brought back to nest (g)	<i>t</i> -test; $t = 1.01$	0.327
Krill in diet (%)	Mann–Whitney test; $U = 11.0$ , $Z = -2.53$	0.011
Fish in diet (%)	Mann–Whitney test; $U = 19.0$ , $Z = -2.08$	0.037
Amphipods in diet (%)	Mann–Whitney test; $U = 27.5$ , $Z = -1.39$	0.163
Squid in diet (%)	Mann–Whitney test; $U = 36.0$ , $Z = -0.97$	0.330
Antarctic krill in diet (%)	Mann–Whitney test; $U = 36.5$ , $Z = -0.66$	0.511
Crystal krill in diet (%)	Mann–Whitney test; $U = 36.5$ , $Z = -0.66$	0.511
Gammariid amphipods in diet (%)	Mann–Whitney test; $U = 20.0$ , $Z = -0.78$	0.435
Hyperiid amphipods in diet (%)	Mann–Whitney test; $U = 20.0$ , $Z = -0.78$	0.435
Prey size		
Antarctic krill size (mm)	<i>t</i> -test; $t = 0.89$	0.370
Crystal krill size (mm)	Mann–Whitney test; $U = 2039.5$ , $Z = -2.94$	0.003
Antarctic silverfish size (mm)	Mann–Whitney test; $U = 188.5$ , $Z = -3.33$	< 0.001
Breeding parameters		
Hatching date (day in December)	Mann–Whitney test; $U = 8476.5$ , $Z = -3.79$	< 0.001
Breeding success (no. chicks at fully crèche stage/no. eggs laid)	$\chi^2$ test; $\chi^2 = 0.025$	0.895
Chick mass at fledging (g)	<i>t</i> -test; $t = -3.02$	0.003

a.



b.



**Fig. 2. a.** Foraging trips of Adélie penguins from Edmonson Point (EdPo) and Inexpressible Island (InIs) during guard (top; satellite image: 26 December 2000) and crèche stages (bottom; satellite image: 21 January 2001). Satellite images were georeferenced and the data were projected on the map by *Envi 3.4* (Research Systems, Inc.). The map is from the SCAR Antarctic Digital Database version 3 (sheet no. SS58-60). **b.** Differences in overall distance travelled, time spent travelling and maximum distance from colony during foraging trips by members of the two colonies (the differences are significant irrespective of individual and breeding stage: see text). Boxplots show medians (thick lines), interquartile ranges (boxes), ranges (whiskers) and outliers (dots). For details on statistical analysis, see text.

(length) of Antarctic krill between colonies (Shapiro–Wilk test, EdPo:  $W=0.98$   $P=0.11$ , InIs:  $W=0.97$ ,  $P=0.24$ ;  $F$ -test:  $F=1.11$ ,  $P=0.64$ ), while we used the Mann–Whitney test to assess differences in length of crystal krill and Antarctic silverfish between colonies.

We compared the hatching date (as a day in December) between the colonies by the Mann–Whitney test. We used the  $\chi^2$  test based on 99 999 Monte Carlo permutations to assess differences in breeding success between the colonies by comparing the number of chicks successfully in crèche

**Table II.** Coefficients (*B*) with relevant 95% confidence intervals (CIs) estimated for generalized linear mixed models and a linear mixed model testing differences in foraging trips and chick growth between the two colonies. The reference category for colony is Edmonson Point. Variance of random factors (*var*) and model fit ( $R^2$ ) are also shown for each model.

Response variable	Predictor	<i>B</i>	95% CI
Distance travelled at sea (km) (Penguin ID:breeding stage) var = 0.17 $R^2 = 0.99$	Intercept	4.75	4.58–4.91 <sup>a</sup>
	Colony (InIs)	0.46	0.20–0.71 <sup>a</sup>
Time spent travelling at sea (h) (Penguin ID:breeding stage) var = 0.09 $R^2 = 0.99$	Intercept	3.85	3.74–3.97 <sup>a</sup>
	Colony (InIs)	0.44	0.26–0.62 <sup>a</sup>
Maximum distance from the colony (km) (Penguin ID:breeding stage) var = 0.14 $R^2 = 0.85$	Intercept	3.62	3.43–3.80 <sup>a</sup>
	Colony (InIs)	0.59	0.29–0.88 <sup>a</sup>
Chick mass (g) (Sampling period) var = 1 291 741 $R^2 = 0.93$	Intercept	1551	650.25–2452.37 <sup>a</sup>
	Colony (InIs)	141	72.61–210.07 <sup>a</sup>

<sup>a</sup> Confidence intervals that do not include zero.

InIs: Inexpressible Island.

to the expected number of chicks in sampled nests with eggs. We ran a linear mixed model to compare chick mass between EdPo and InIs over seven time periods, established arbitrarily between the hatching date and when chicks entered the crèche (SC-CCAMLR 2004): 0–5 days from hatching date, 6–10 days, 11–15 days, 16–20 days, 21–25 days, 26–30 days and crèche stage. We set colony as the categorical predictor (reference level: EdPo) and time period as a random intercept. We tested predictor significance by assessing whether 95% confidence intervals contained zero. We used the *t*-test (after checking for normality and homoscedasticity; Shapiro–Wilk test, EdPo:  $W = 0.979$ ,  $P = 0.581$ , InIs:  $W = 0.954$ ,  $P = 0.226$ ; *F*-test:  $F = 1.332$ ,  $P = 0.379$ ) to compare chick weight at fledging between colonies, as these data were not recorded on the same nests as those of chick weight from hatching to crèche.

Statistical analysis was performed using *Past* and *R Stats* packages ([www.R-project.org](http://www.R-project.org)) and *lme4* (Bates *et al.* 2015). All tests were two-tailed and the significance level was set at  $\alpha = 0.05$ . All values are reported as mean  $\pm$  standard error (for data analysed by two-sample tests) or as 95% confidence intervals of effect-size estimates (for model predictions), unless otherwise specified.

## Results

### *Weather at the colonies*

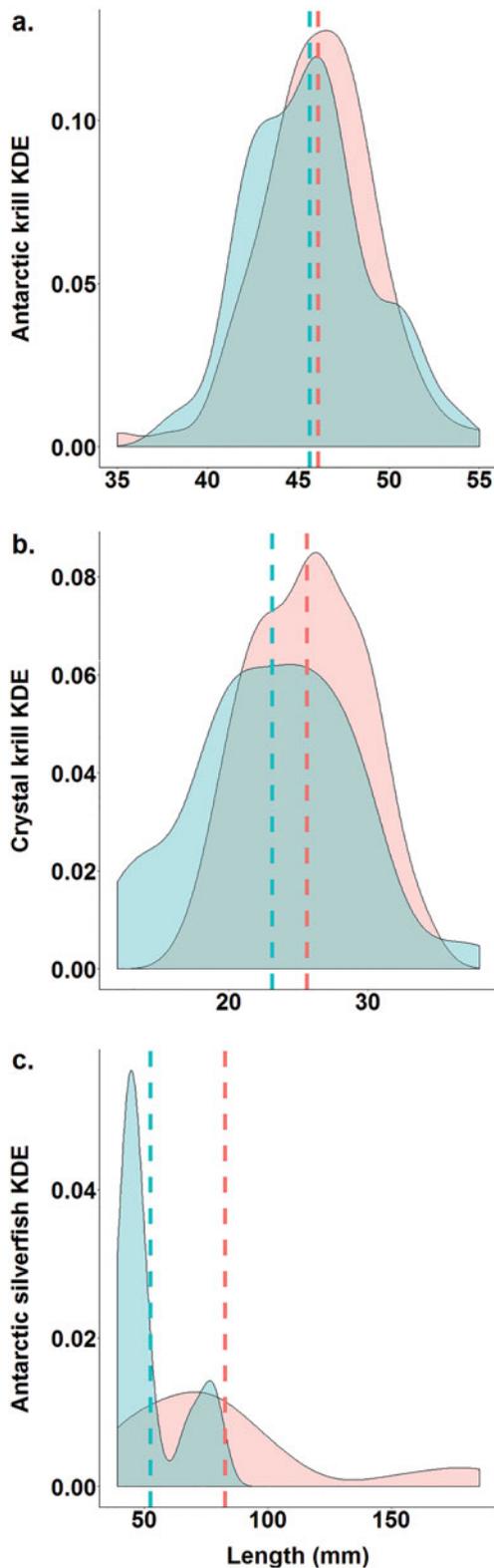
In the period from November 2000 to January 2001, the weather was harsher at InIs. Despite similar relative humidity (EdPo:  $55.8 \pm 1.03\%$ , InIs:  $54.2 \pm 1.05\%$ ; Table I), a colder mean daily temperature was recorded at InIs (EdPo:  $-5.09 \pm 0.33^\circ\text{C}$ , InIs:  $-6.37 \pm 0.35^\circ\text{C}$ ; Table I).

### *Foraging trips*

During the study period, birds from the two colonies travelled to distinct foraging areas (Fig. 2a). Considering breeding period and penguin identity, the models predicted that penguins from InIs performed foraging trips that were on average 21.75–142.14 km longer than those by penguins at EdPo (EdPo: 98.22–136.17 km; Fig. 2b & Table II) and spent on average 12.22–46.03 more hours travelling than penguins from EdPo (EdPo: 41.94–52.97 h; Fig. 2b). Penguins from InIs also travelled on average 10.41–63.44 km farther from the colony than did penguins from EdPo (EdPo: 30.85–44.86 km; Fig. 2b & Table II).

### *Diet*

In January 2001, the quantity of food brought back to the colony per individual did not differ between EdPo and InIs (EdPo:  $419.06 \pm 57.59$  g,  $n = 9$ , InIs:  $328.46 \pm 67.56$  g,  $n = 10$ ; Table I), yet its overall composition differed between colonies ( $\chi^2 = 107.69$ ,  $df = 1$ ,  $P < 0.0001$ ). At the individual level, the percentage of krill per penguin was greater at EdPo (EdPo:  $94.32 \pm 2.34\%$ , InIs:  $72.83 \pm 8.59\%$ ; Table I), whereas that of fish was greater at InIs (EdPo:  $3.89 \pm 1.28\%$ , InIs:  $16.01 \pm 6.88\%$ ; Table I). The percentages of amphipods (EdPo:  $1.77 \pm 1.11\%$ , InIs:  $7.58 \pm 4.01\%$ ; Table I) and squid per penguin (EdPo:  $0.02 \pm 0.01\%$ , InIs:  $3.58 \pm 3.58\%$ ; Table I) did not differ between colonies. Percentages of Antarctic krill (EdPo:  $43.95 \pm 11.33\%$ , InIs:  $57.03 \pm 14.06\%$ ; Table I) and crystal krill (EdPo:  $56.05 \pm 11.33\%$ , InIs:  $42.96 \pm 14.06\%$ ; Table I) also did not differ between colonies. Antarctic silverfish was the predominant fish species in the diets of both colonies (EdPo: 82.35%,  $n = 34$  fish, InIs: 98.62%,  $n = 145$  fish). Other fish species were *Trematomus bernacchii*



**Fig. 3.** Differences in kernel density estimates (KDEs) of the lengths of prey species (**a.** Antarctic krill, *Euphausia superba*; **b.** crystal krill, *Euphausia crystallorophias*; **c.** Antarctic silverfish, *Pleuragramma antarctica*) between colonies (red: Edmonson Point; blue: Inexpressible Island). Dashed lines show mean values.

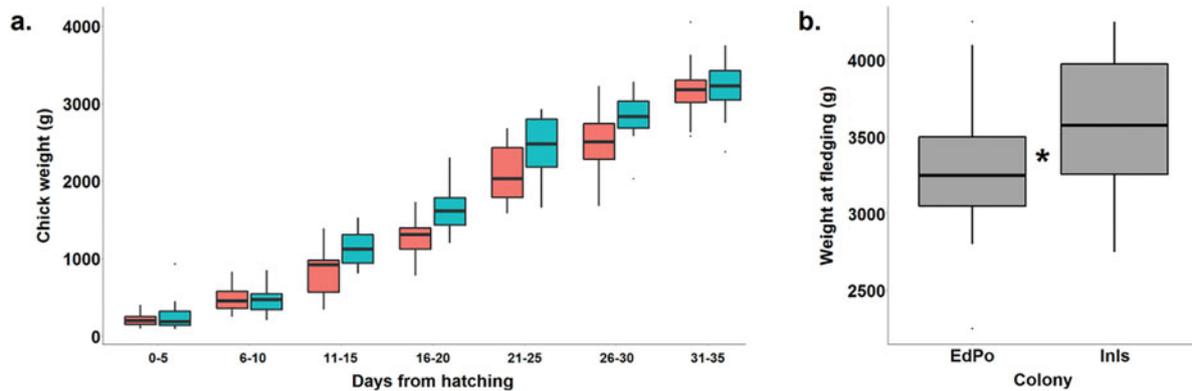
(EdPo: 8.82%,  $n = 34$  fish, InIs: 0.69%,  $n = 145$  fish), *Chaenodraco wilsoni* (EdPo: 5.88%,  $n = 34$  fish, InIs: 0.69%,  $n = 145$  fish) and *Pagothenia borchgrevinki* (EdPo: 2.94%,  $n = 34$  fish, InIs: 0%,  $n = 145$  fish). Percentages of Gammarid (EdPo:  $73.91 \pm 15.26\%$ , InIs:  $63.26 \pm 12.29\%$ ; Table I) and Hyperiid amphipods (EdPo:  $26.87 \pm 15.26\%$ , InIs:  $36.63 \pm 12.29\%$ ; Table I) did not differ between colonies. The sizes of Antarctic krill (EdPo:  $46.09 \pm 0.34$  mm, InIs:  $45.06 \pm 0.43$  mm; Fig. 3a & Table I) did not differ between colonies, but sizes of crystal krill (EdPo:  $25.62 \pm 0.43$  mm, InIs:  $23.12 \pm 0.70$  mm; Fig. 3b & Table I) and Antarctic silverfish (EdPo:  $82.59 \pm 8.83$  mm, InIs:  $52.29 \pm 2.22$  mm; Fig. 3c & Table I) were smaller at InIs. The Shannon and Pielou indices were higher for EdPo ( $H_{\text{EdPo}} = 0.64$ ,  $H_{\text{InIs}} = 0.08$ ,  $J_{\text{EdPo}} = 0.46$ ,  $J_{\text{InIs}} = 0.07$ ).

#### Breeding chronology, breeding success and chick growth

The chronology of the breeding cycle in the 2000–01 summer is summarized in Table SI. Hatching occurred  $\sim 2$  days earlier at EdPo (EdPo:  $18.82 \pm 0.34$  December,  $n = 170$  eggs, InIs:  $20.63 \pm 0.37$  December,  $n = 135$  eggs; Table I). Breeding success did not differ between colonies (EdPo: 0.91 chicks/nest, InIs: 0.90 chicks/nest; Table I). Chicks sampled regularly from study nests were significantly heavier at InIs throughout the breeding season (i.e. regardless of sampling period): the model predicted chicks from InIs to be 72.61–210.07 g heavier than those from EdPo (EdPo: 650.26–2452.37 g; Fig. 4a & Table II). Chicks from InIs achieved greater body mass at fledging (EdPo:  $3291.80 \pm 56.68$  g,  $n = 46$  chicks, InIs:  $3581.20 \pm 81.01$  g,  $n = 30$  chicks; Fig. 4b & Table I).

#### Discussion

We showed that two colonies with various local ecological conditions displayed differences in foraging distance, diet and breeding performance. In group-living foragers, intraspecific competition for food may influence social and spatiotemporal behaviour: the larger the number of individuals in the group, the greater the foraging interference (e.g. Birt *et al.* 1987). Accordingly, foraging competition should be greater in large colonies of seabirds, and availability of resources per individual should decrease in proportion to colony size (e.g. Cairns 1989, Ainley *et al.* 2018). In turn, all other factors being equal, individuals from large colonies should forage over a larger area to avoid competitors and to achieve adequate food intake (e.g. Ballance *et al.* 2009 for Adélie penguins). Our findings on the spatial range of foraging trips may support this hypothesis: the penguin population at InIs was ten times larger than the one at EdPo, and birds from InIs did indeed travel further and for longer than those at EdPo. There is a third Adélie



**Fig. 4. a.** Differences in chick weight between colonies in the hatching-to-crèche stage (red: Edmonson Point, EdPo; blue: Inexpressible Island, InIs; differences are significant irrespective of sampling period: see text). **b.** Difference in chick weight between colonies at fledging (\* significant). Boxplots show medians (thick lines), interquartile ranges (boxes), ranges (whiskers) and outliers (dots).

penguin colony, Adélie Cove, ~17 km north of InIs (Fig. 1). On 12 December 2000, we estimated a population of 7735 breeding pairs by ground count at Adélie Cove. These extra competitors may have contributed to prey depletion in the Terra Nova Bay area, forcing penguins nesting at InIs to travel longer distances from the coast and to perform longer-lasting trips than those at EdPo in order to avoid foraging areas used by conspecifics. However, as no synoptic foraging data for the Adélie Cove colony or information on prey abundance are available for this season, we could not test whether extra competitors from Adélie Cove and/or food were limiting factors. Alternative explanations, such as varying prey distribution/availability, could also have influenced their foraging tactics (e.g. Watanuki *et al.* 1993). Overall, we found that Adélie penguins nesting in central Victoria Land fed mainly on krill, fish and, to a lesser extent, amphipods. This is in line with our previous findings for the EdPo colony in various years (Olmastroni *et al.* 2000) and with those reported by Ainley *et al.* (2003) for colonies on Ross Island (95% crystal krill and silverfish, 5% amphipods).

The quantitative data for krill species of our study indicate that *E. superba* was on the whole the predominant species in the diet of penguins in January 2001 (30–50% of stomach contents in both colonies). The frequency distribution of body length of krill specimens in our samples also indicates that juvenile and adult stages of *E. crystallophias* and *E. superba* (according to the age classification reported by Sala *et al.* 2002 and Ainley *et al.* 2003) were both present in the Terra Nova Bay area in summer 2000–01. The stomach contents of penguins from InIs contained greater quantities of *E. superba* than *E. crystallophias*, which, conversely, was more abundant in samples from EdPo, although differences were not significant. Penguins from InIs travelled longer times and distances

than those at EdPo. This could explain the higher percentage of *E. superba* found in samples from penguins of InIs and the predominance of *E. crystallophias* in the stomach contents of penguins from EdPo. In fact, oceanographic surveys in the western Ross Sea, described in Sala *et al.* (2002) and Azzali *et al.* (2006), report only a small overlap in the distributions of the two krill species, with a spatiotemporal distribution linked to sea-ice conditions. *Euphausia superba* has a prevalently more pelagic distribution than *E. crystallophias*. Antarctic krill is the principal penguin prey throughout Antarctica, including northern Victoria Land. Conversely, it is reported to be absent or rare in the diets of penguins breeding at southern colonies (table 2.1 in Ainley 2002, Kristan *et al.* 2019). Our study shows that *E. superba* may be available to penguins nesting in central Victoria Land.

Studies on the diet of Adélie penguins of southern Ross Sea colonies (> 74°S) over three breeding seasons (Ainley *et al.* 1998) found crystal krill and Antarctic silverfish to be the main prey. *Pleuragramma antarctica* was the only fish present, constituting up to 50–90% of the stomach contents of penguins foraging in open-water habitats adjacent to fast ice. In January 2001, penguins from the two study colonies fed mainly on silverfish (95% of the species consumed), confirming that this species is a major food item when energy demand is high for raising chicks (Ainley *et al.* 2003). The low percentages (1–2%) of other fish species may indicate opportunistic catches in trophic niches shared by a number of fish or varying predatory behaviour by individual Adélie penguins in the study area (Clarke *et al.* 1998, Nesti *et al.* 2010).

The sizes of *P. antarctica* ingested by InIs penguins were smaller than those ingested by penguins from EdPo. According to La Mesa *et al.* (2010), silverfish in the ranges 28–33 and 53–74 mm are post-larval specimens and 1 year and over (i.e. juveniles) respectively, living in

the water column at depths of 0–200 m. Post-larval and juvenile stages constitute 95–99% of the ichthyoplankton on the Ross Sea continental shelf (Granata *et al.* 2002), where they tend to occupy extensive areas near submarine banks often in association with krill (Sala *et al.* 2002). During the study period, penguins used areas off their colonies for a mean linear distance of 40–60 km, closer to the coast in the case of EdPo and beyond the external edge of the Terra Nova Bay polynya in the case of InIs. The regions where the two groups foraged may have underwater topographic features (basins and banks; see fig. 1 in La Mesa *et al.* 2010) that aid upwelling and concentration of prey. The fact that more fish were caught off InIs may also be due to favourable conditions for rapid growth of *P. antarctica* larvae near polynya, in line with autumnal phytoplankton blooms (Vacchi *et al.* 2012). Indeed, Terra Nova Bay seems to be particularly rich in silverfish at various growth stages (Davis *et al.* 2017, O'Driscoll *et al.* 2018).

Concerning the slight difference in breeding chronology between the two colonies, the colder weather and blizzard we recorded at InIs during early incubation could have delayed hatching. One might expect greater breeding success and chick growth at EdPo because offspring enjoyed a more sheltered (and therefore less energy-consuming) situation compared to InIs. Surprisingly, our findings showed that penguins at InIs reared heavier chicks than penguins nesting at EdPo, although there were no differences in breeding success, and despite the harsher weather, greater colony size and longer-lasting foraging trips were also associated with the InIs colony. We suggest that two factors may explain the varying early growth patterns of chicks in the two colonies, although other hypotheses remain to be tested. 1) Adults from EdPo had to walk several kilometres on fast ice to reach the foraging grounds and return to feed chicks, whereas penguins from InIs could swim all of the way. For penguins, locomotion on ice and snow is known to require greater energy expenditure than swimming (Pinshow *et al.* 1976, Wilson *et al.* 1991). This difference in foraging trips to the sea may have affected parents from EdPo, causing them to consume a greater proportion of the food ingested in order to cover the extra exertion of walking (than in the case of penguins from InIs), thus reducing the food delivered to their young (*sensu* Ainley *et al.* 1998, Emmerson & Southwell 2008). If so, less of the similar food loads foraged by adults of the two colonies would have been delivered to chicks at EdPo. 2) As the quantity of food ingested by adults did not differ between the two colonies, chicks at InIs may have benefitted from the larger proportion of fish foraged by their parents. Fish has been shown to have a higher caloric content than krill (Ainley *et al.* 2003, 2018, Whitehead *et al.* 2015, Jennings *et al.* 2016). Chicks at InIs, fed a larger quantity of fish, may

therefore have built up more fat reserves than chicks at EdPo, achieving faster growth and higher body weight at fledging. However, more data are needed to distinguish the effects of confounding variables in order to verify either hypothesis.

Clustered colonies are reported to have similar diets, with birds breeding in smaller colonies reducing their foraging distances (Ainley *et al.* 2003). However, differences in population size, access to foraging grounds (in relation to fast ice and/or polynya), sea ice, sea currents, intraspecific competition and prey distribution/abundance can all be determinants for neighbouring colonies (Emmerson & Southwell 2008). The role of the Terra Nova Bay polynya has not been widely investigated for the top predators living in this area. However, this productive hotspot (Davis *et al.* 2017) may improve foraging efficiency and prey quality (i.e. higher energy intake), ultimately supporting the larger penguin population at InIs and the neighbouring colony at Adélie Cove with respect to the Wood Bay colony (EdPo).

In describing two Adélie penguin colonies in central Victoria Land, including breeding phenology, reproductive output and (during chick rearing) foraging areas and diet, we observed that penguins breeding in neighbouring colonies with varying habitat conditions and population sizes successfully tackled both situations by implementing various foraging strategies in relation to the physical characteristics of their nesting and feeding sites. However, our analysis would have benefitted from a greater sample size and repeated data from various years. In particular, interannual comparisons of foraging trips and breeding success at the cluster of colonies in mid-Victoria Land could complete the complex scenario of the Ross Sea Adélie penguin population, which accounts for 38% of the global population of this species (Ainley 2002). To this end, our local baseline data may help future research in this area to clarify potential concerns regarding regional changes in sea-ice dynamics and the biological responses of Antarctic seabirds (Massom & Stammerjohn 2010), as is required by the monitoring and management research plan for the Ross Sea Marine Protected Area.

#### Acknowledgements

We are grateful to K.R. Kerry for conceiving and supervising the research and for commenting on a draft of the manuscript. We are indebted to I. Bisogno and B. Sprugnoli for help with diet analysis and to V. Volpi for assistance with GIS analysis. The authors appreciated the support of the Australian Antarctic Division, the PNRA Meteo-Climatological Observatory at MZS and Victoria Land ([www.climantartide.it](http://www.climantartide.it)), the dataset, data display and information provided by the University of Wisconsin-Madison's Automatic Weather Station programme, grant number ANT-1543305 from the National Science Foundation and the map in Fig. 1

from the Quantarctica's Norwegian Polar Institute. We thank the anonymous reviewers for their insightful comments and suggestions.

### Author contributions

SO conceived the research and performed data collection and analysis. FP performed data collection. NF conceived and performed the statistical analysis. SF supervised the PNRA1999/1.3 project. SO and NF wrote the manuscript. All authors commented on the manuscript and approved the final draft.

### Financial support

The Programma Nazionale di Ricerche in Antartide (PNRA) provided logistical support. This research was funded by Ministry of Education, University and Research (MIUR) project PNRA1999/1.3.

### Supplemental material

A supplemental table and a supplemental figure will be found at <https://doi.org/10.1017/S0954102020000085>.

### References

- AINLEY, D.G. 2002. *The Adélie penguin: bellwether of climate change*. New York: Columbia University Press, 310 pp.
- AINLEY, D.G., NUR, N. & WOEHLE, E.J. 1995. Factors affecting the distribution and size of pygoscelid penguin colonies in the Antarctic. *Auk*, **112**, 10.2307/4088776.
- AINLEY, D.G., O'CONNOR, E.F. & BOEKELHEIDE, R.J. 1984. *The marine ecology of birds in the Ross Sea, Antarctica*. Washington, DC, American Ornithologists' Union, 97 pp.
- AINLEY, D.G., WILSON, P.R., BARTON, K.J., BALLARD, G., NUR, N. & KARL, B. 1998. Diet and foraging effort of Adélie penguins in relation to pack-ice conditions in the southern Ross Sea. *Polar Biology*, **20**, 10.1007/s003000050308.
- AINLEY, D.G., BALLARD, G., BARTON, K.R., KARL, B., RAU, G.H., RIBIC, C.A. & WILSON, P.R. 2003. Spatial and temporal variation of diet within a presumed metapopulation of Adélie penguins. *Condor*, **105**, 10.1093/condor/105.1.95.
- AINLEY, D.G., CLARKE, E.D., ARRIGO, K.R., FRASER, W.R., KATO, A., BARTON, K.J. & WILSON, P.R. 2005. Decadal-scale changes in the climate and biota of the Pacific sector of the Southern Ocean, 1950s to 1990s. *Antarctic Science*, **17**, 10.1017/S0954102005002567.
- AINLEY, D.G., DUGGER, K.M., LA MESA, M., BALLARD, G., BARTON, K.J., JENNINGS, S., *et al.* 2018. Post-fledging survival of Adélie penguins at multiple colonies: chicks raised on fish do well. *Marine Ecology Progress Series*, **601**, 10.3354/meps12687.
- AINLEY, D.G., RIBIC, C.A., BALLARD, G., HEATH, S., GAFFNEY, I., KARL, B.J., *et al.* 2004. Geographic structure of Adélie penguin populations: size, overlap and use of adjacent colony-specific foraging areas. *Ecological Monograph*, **74**, 10.1890/02-4073.
- ARRIGO, K.R., VAN DIJKEN, G.L., AINLEY, D.G., FAHNESTOCK, M.A. & MARKUS, T. 2002. Ecological impact of a large Antarctic iceberg. *Geophysical Research Letters*, **29**, 10.1029/2001GL014160.
- AZZALI, M., LEONORI, I., DE FELICE, A. & RUSSO, A. 2006. Spatial-temporal relationships between two euphausiid species in the Ross Sea. *Chemistry and Ecology*, **22**, 10.1080/02757540600670836.
- BALLANCE, L.T., AINLEY, D.G., BALLARD, G. & BARTON, K. 2009. An energetic correlate between colony size and foraging effort in seabirds, an example of the Adélie penguin *Pygoscelis adeliae*. *Journal of Avian Biology*, **40**, 10.1111/j.1600-048X.2008.04538.x.
- BALLARD, G., DUGGER, K.M., NUR, N. & AINLEY, D.G. 2010. Foraging strategies of Adélie penguins: adjusting body condition to cope with environmental variability. *Marine Ecology Progress Series*, **405**, 10.3354/meps08514.
- BANNASCH, R., WILSON, R.P. & CULIK, B. 1994. Hydrodynamic aspects of design and attachment of a back-mounted device in penguins. *Journal of Experimental Biology*, **194**, 83–96.
- BATES, D., MAECHLER, M., BOLKER, B. & WALKER, S. 2015. Fitting linear mixed-effects models using *lme4*. *Journal of Statistical Software*, **67**, doi:10.18637/jss.v067.i01.
- BIRT, V.L., BIRT, T.P., GOULET, D., CAIRNS, D.K. & MONTEVECCHI, W.A. 1987. Ashmole's halo: direct evidence for prey depletion by a seabird. *Marine Ecology Progress Series*, **40**, 10.3354/meps040205.
- BROMWICH, D.H. 1985. Katabatic wind interaction with Inexpressible Island, Terra Nova Bay. *Antarctic Journal of United States*, **20**, 196–198.
- CAIRNS, D.K. 1989. The regulation of seabird colony size: a hinterland model. *American Naturalists*, **134**, 10.1086/284970.
- CLARKE, J., EMMERSON, L.M. & OTAHAL, P. 2006. Environmental conditions and life history constraints determine foraging range in breeding Adélie penguins. *Marine Ecology Progress Series*, **310**, 10.3354/meps310247.
- CLARKE, J., MANLY, B., KERRY, K., GARDNER, H., FRANCHI, E., CORSOLINI, S. & FOCARDI, S. 1998. Sex differences in Adélie penguin foraging strategies. *Polar Biology*, **20**, 10.1007/s003000050301.
- DAVIS, L.B., HOFMANN, E.E., KLINCK, J.M., PIÑONES, A. & DINNIMAN, M.S. 2017. Distributions of krill and Antarctic silverfish and correlations with environmental variables in the western Ross Sea, Antarctica. *Marine Ecology Progress Series*, **584**, 10.3354/meps12347.
- EMMERSON, L. & SOUTHWELL, C. 2008. Sea ice cover and its influence on Adélie penguin reproductive performance. *Ecology*, **89**, 10.1890/08-0011.1.
- EMSLIE, S.D., FRASER, W., SMITH, R.C. & WALZER, W. 1998. Abandoned penguin colonies and environmental change in the Palmer Station area, Anvers Island, Antarctic Peninsula. *Antarctic Science*, **10**, 10.1017/S0954102098000352.
- FAO 1985. *FAO species identification sheets for fishery purposes Southern Ocean: Fishing Areas 48, 58 and 88 (CCAMLR Convention Area)*. FISCHER, W. & HUREAU, J.C. eds. Rome: Commission for the Conservation of Antarctic Marine Living Resources, Vol. 1, 232 pp.
- FORCADA, J. & TRATHAN, P.N. 2009. Penguin responses to climate change in the Southern Ocean. *Global Change Biology*, **15**, 10.1111/j.1365-2486.2009.01909.x.
- GRANATA, A., CUBATA, A., GUGLIELMO, L., SIDOTI, O., GRECO, S., VACCHI, M. & LA MESA, M. 2002. Ichthyoplankton abundance and distribution in the Ross Sea Region during 1987–1996. *Polar Biology*, **25**, 187–202.
- INCHAUSTI, P., GUINET, C., KOUDIL, M., DURBEC, J.P., BARBRAUD, C., WEIMERSKIRCH, H., *et al.* 2003. Inter-annual variability in the breeding performance of seabirds in relation to oceanographic anomalies that affect the Crozet and the Kerguelen sectors of the Southern Ocean. *Journal of Avian Biology*, **34**, 10.1034/j.1600-048X.2003.03031.x.
- JENNINGS, S., VARSANI, A., DUGGER, K.M., BALLARD, G. & AINLEY, D.G. 2016. Sex-based differences in Adélie penguin (*Pygoscelis adeliae*) chick growth rates and diet. *PLoS One*, **11**, 10.1371/journal.pone.0149090.
- JENOUVRIER, S., WEIMERSKIRCH, H., BARBRAUD, C., YOUNG-HYANG, P. & CAZELLES, B. 2005. Evidence of a shift in the cyclicity of Antarctic

- seabird dynamics linked to climate. *Proceedings of the Royal Society B*, **272**, 10.1098/rspb.2004.2978.
- JENOUVRIER, S., CASWELL, H., BARBRAUD, C., HOLLAND, M., STROEVE, M.J. & WEIMERSKIRCH, H. 2009. Demographic models and IPCC climate projections predict the decline of an emperor penguin population. *Proceedings of the National Academy of Sciences of the United States of America*, **106**, 10.1073/pnas.0806638106.
- KRISTAN, A.K., EMSLIE, S.D. & PATTERSON W.P. 2019. Stable isotope analyses of ancient and modern Adélie penguin (*Pygoscelis adeliae*) mummies from the Ross Sea Region, Antarctica. *Polar Biology*, **42**, 10.1007/s00300-019-02513-4.
- LA MESA, M., CATALANO, B., RUSSO, A., GRECO, S., VACCHI, M. & AZZALI, M. 2010. Influence of environmental conditions of spatial distribution and abundance of early life stages of Antarctic silverfish, *Pleuragramma antarcticum* (Nototheniidae), in the Ross Sea. *Antarctic Science*, **22**, 10.1017/S0954102009990721.
- LE GUEN, C., KATO, A., RAYMOND, B., BARBRAUD, C., BEAULIEU, M., BOST, C.A., *et al.* 2018. Reproductive performance and diving behaviour share a common sea-ice concentration optimum in Adélie penguins (*Pygoscelis adeliae*). *Global Change Biology*, **24**, 5304–5317.
- MASSOM, R.A. & STAMMERJOHN, S.E. 2010. Antarctic sea ice change and variability – physical and ecological implications. *Polar Science*, **4**, 10.1016/j.polar.2010.05.001.
- NESTI, I., ROPERT-COUDERT, Y., KATO, A., BEAULIEU, M., FOCARDI, S. & OLMASTRONI, S. 2010. Diving behaviour of chick-rearing Adélie penguins at Edmonson Point, Ross Sea. *Polar Biology*, **10**, 10.1007/s00300-010-0775-2.
- O'DRISCOLL, D.L., LADROIT, Y., PARKER, S.J., VACCHI, M., CANESE S.P., GHIGLIOTTI L., *et al.* 2018. Acoustic deployments reveal Antarctic silverfish under ice in the Ross Sea. *Antarctic Science*, **30**, 10.1017/S0954102018000366.
- OLMASTRONI, S., CORSOLINI, S., PEZZO, F., FOCARDI, S. & KERRY, K. 2000. The first five years of Italian–Australian joint programme on the Adélie penguin: an overview. *Italian Journal of Zoology*, **67**(Suppl. 1), 10.1080/11250000009356369.
- PEET, R.K. 1975. Relative diversity indices. *Ecology*, **56**, 10.2307/1934984.
- PINSHOW, B., FEDAK, M.A., BATTLES, D.R. & SCHMIDT-NIELSEN, K. 1976. Energy expenditure for thermoregulation and locomotion in emperor penguins. *American Journal of Physiology*, **231**, 10.1152/ajplegacy.1976.231.3.903.
- ROPERT-COUDERT, Y., KATO, A., MEYER, X., PELLÉ, M., MACINTOSH, A.J.J., ANGELIER, F., *et al.* 2015. A complete breeding failure in an Adélie penguin colony correlates with unusual and extreme environmental events. *Ecography*, **38**, 10.1111/ecog.01182.
- SALA, A., AZZALI, M. & RUSSO, A. 2002. Krill of the Ross Sea: distribution, abundance and demography of *Euphausia superba* and *Euphausia crystallorophias* during the Italian Antarctic Expedition (January–February 2000). *Scientia Marina*, **66**, 10.3989/scimar.2002.66n2123.
- SC-CCAMLR 2004. *CCAMLR ecosystem monitoring program: standard methods for monitoring studies*. Hobart: CCAMLR, 233 pp.
- SLADEN, W.J. 1978. Sexing penguins by a cloacoscope. *International Zoo Yearbook*, **18**, 10.1111/j.1748-1090.1978.tb00224.x.
- SMITH, R.C., AINLEY, D., BAKER, K., DOMACK, E., EMSLIE, S., FRASER, B., *et al.* 1999. Marine ecosystem sensitivity to climate change: historical observations and paleoecological records reveal ecological transitions in the Antarctic Peninsula region. *BioScience*, **49**, 10.2307/1313632.
- TREMBLAY, Y. & CHEREL, Y. 2003. Geographic variation in the foraging behaviour, diet and chick growth of rockhopper penguins. *Marine Ecology Progress Series*, **251**, 10.3354/meps251279.
- VACCHI, M., DEVRIES, A.L., EVANS, C.W., BOTTARO, M., GHIGLIOTTI, L., CUTRONEO, L. & PISANO E. 2012. A nursery area for the Antarctic silverfish *Pleuragramma antarcticum* at Terra Nova Bay (Ross Sea): first estimate of distribution and abundance of eggs and larvae under the seasonal seaice. *Polar Biology*, **35**, 10.1007/s00300-012-1199-y.
- WATANUKI, Y., KATO, A., MORI, Y. & NAITO, Y. 1993. Diving performance of Adélie penguins in relation to food availability in fast sea-ice areas: comparison between years. *Journal of Animal Ecology*, **62**, 10.2307/5384.
- WATANUKI, Y., TAKAHASHI, A., KATO, A., SATO, K. & BOST, C.A. 2004. Inter-colony and sex differences in the effects of parental body condition and foraging effort on the brood growth of Adélie penguins. *Journal of Ethology*, **22**, 10.1007/s10164-003-0108-9.
- WATANUKI, Y., KATO, A., SATO, K., NIIZUMA, Y., BOST, C.A., LE MAHO, Y. & NAITO, Y. 2002. Parental mass change and food provisioning in Adélie penguins rearing chicks in colonies with contrasting sea-ice conditions. *Polar Biology*, **25**, 10.1007/s00300-002-0399-2.
- WHITEHEAD, A.L., LYVER, P.O.B., BALLARD, G., BARTON, K., KARL, B.J., DUGGER, K.M., *et al.* 2015. Factors driving Adélie penguin chick size, mass and condition at colonies of different sizes in the southern Ross Sea. *Marine Ecology Progress Series*, **523**, 10.3354/meps11130.
- WIDMANN, M., KATO, A., RAYMOND, B., ANGELIER, F., ARTHUR, B., CHASTEL, O., *et al.* 2015. Habitat use and sex-specific foraging behavior of Adélie penguins throughout the breeding season in Adélie Land, East Antarctica. *Movement Ecology*, **3**, 10.1186/s40462-015-0052-7.
- WILLIAMS, R. & McELDOWNY, T. 1990. A guide to the fish otoliths from waters off the Australian Antarctic Territory, Heard, and Macquarie Islands. *ANARE Research Notes*, No. 75, 173 pp.
- WILSON, P.R., AINLEY, D.G., NUR, N., JACOBS, S.S., BARTON, K.R., BALLARD, G. & COMISO, J.C. 2001. Adélie penguin population change in the Pacific Sector of Antarctica: relation to sea ice extent and the Antarctic circumpolar current. *Marine Ecology Progress Series*, **213**, 10.3354/meps213301.
- WILSON, R.P. 1984. An improved stomach pump for penguins and other seabirds. *Journal of Field Ornithology*, **55**, 109–112.
- WILSON, R.P., CULIK, B. & ADELUNG, D. 1991. To slide or stride: when should Adélie penguins (*Pygoscelis adeliae*) toboggan? *Canadian Journal of Zoology*, **69**, 10.1139/z91-033.