



Neurodegeneration with Brain Iron Accumulation and a Brief Report of the Disease in Iran

Reza Hajati, Maziar Emamikhah , Fardad Danaee Fard, Mohammad Rohani, Afagh Alavi 

ABSTRACT: Neurodegeneration with brain iron accumulation (NBIA) is a term used for a group of hereditary neurological disorders with abnormal accumulation of iron in basal ganglia. It is clinically and genetically heterogeneous with symptoms such as dystonia, dysarthria, Parkinsonism, intellectual disability, and spasticity. The age at onset and rate of progression are variable among individuals. Current therapies are exclusively symptomatic and unable to hinder the disease progression. Approximately 16 genes have been identified and affiliated to such condition with different functions such as iron metabolism (only two genes: Ferritin Light Chain (*FTL*) Ceruloplasmin (*CP*)), lipid metabolism, lysosomal functions, and autophagy process, but some functions have remained unknown so far. Subgroups of NBIA are categorized based on the mutant genes. Although in the last 10 years, the development of whole-exome sequencing (WES) technology has promoted the identification of disease-causing genes, there seem to be some unknown genes and our knowledge about the molecular aspects and pathogenesis of NBIA is not complete yet. There is currently no comprehensive study about the NBIA in Iran; however, one of the latest discovered NBIA genes, GTP-binding protein 2 (*GTPBP2*), has been identified in an Iranian family, and there are some patients who have genetically remained unknown.

RÉSUMÉ : La neurodégénérescence avec surcharge cérébrale en fer et bref rapport sur la maladie en Iran. La neurodégénérescence avec surcharge cérébrale en fer (NBIA) est un terme désignant un groupe de troubles neurologiques héréditaires, accompagnés d'une accumulation anormale de fer dans les noyaux gris centraux. Ces troubles diffèrent grandement sur les plans génétique et clinique, et se manifestent par des symptômes tels que la dystonie, la dysarthrie, le parkinsonisme, une déficience intellectuelle et la spasticité. De plus, l'âge d'apparition de la maladie et sa vitesse d'évolution varient d'une personne à l'autre. Par ailleurs, les traitements actuels ne traitent que les symptômes et ne permettent pas de ralentir l'évolution de la maladie. Seize gènes ont été identifiés jusqu'à maintenant et touchent différentes fonctions, par exemple le métabolisme du fer (deux gènes seulement : *FTL* et *CP*), le métabolisme des lipides, les fonctions lysosomiales et l'autophagie, mais il reste d'autres fonctions à découvrir. Une catégorisation des sous-groupes de la NBIA a été établie en fonction des gènes mutants. Bien que, au cours des dix dernières années, la mise au point de la technique de séquençage complet de l'exome ait facilité l'identification de gènes pathogènes, il semble exister d'autres gènes inconnus, et l'on ne connaît pas tout encore des aspects moléculaires et de la pathogénèse de la NBIA. Enfin, aucune étude exhaustive ne porte à ce jour sur la NBIA en Iran; toutefois, l'un des derniers gènes liés à la NBIA, le *GTPBP2*, a été identifié dans une famille iranienne, et il y a certains patients dont on ne connaît pas encore le patrimoine génétique.

Keywords: Neurodegeneration with brain iron accumulation (NBIA), Basal ganglia, Iron metabolism, Genetic heterogeneity

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INTRODUCTION

Neurodegeneration with brain iron accumulation (NBIA) belongs to neurodegenerative disorders. Its main feature is the accumulation of iron in several parts of the brain, especially in Globus Pallidus (GP).¹ The clinical features of NBIA include dystonia, parkinsonism, spasticity, cognitive impairment, and intellectual disability.^{2,3} The age at onset varies from early childhood to late adulthood.¹ There is a lack of trustworthy

information about the prevalence of NBIA in different geographical areas. The overall frequency of each subtype is shown in Figure 1. In fact, some areas such as the Dominican Republic are reported to have more cases of some NBIA subtypes. However, estimations are about 1/500,000 in some populations. There are different subgroups of NBIA of which pantothenate kinase-associated neurodegeneration (PKAN), phospholipase A2-associated neurodegeneration (PLAN), mitochondrial protein-associated

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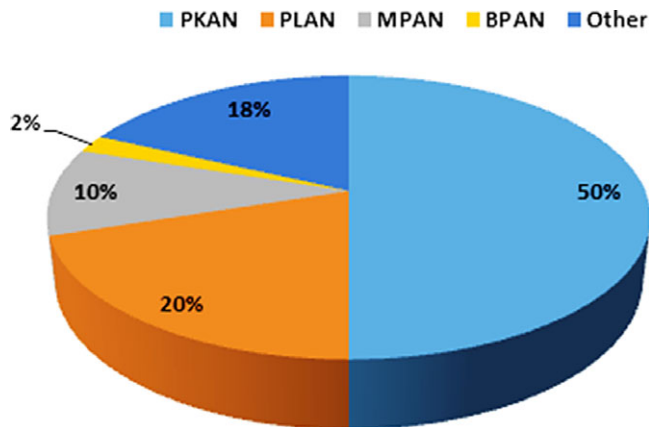


Figure 1: The frequency of neurodegeneration with brain iron accumulation (NBIA) subtypes in the literature.

neurodegeneration (MPAN), and Beta propeller-associated neurodegeneration (BPAN) are the most common forms. Mutations in pantothenate kinase 2 (*PANK2*), Phospholipase A2 Group VI (*PLA2G6*), Chromosome 19 Open Reading Frame 12 (*C19orf12*), and WD Repeat Domain 45 (*WDR45*) genes cause the aforementioned subgroups, respectively.⁴ Proteins encoded by these genes mediate different cellular functions, including lipid metabolism, autophagy, coenzyme-A (CoA) biosynthesis, and mitochondrial and lysosomal function regulation.^{1,2} NBIA inheritance patterns are autosomal recessive (AR), autosomal dominant, and X-linked.⁵ Diagnosis depends on clinical and magnetic resonance imaging (MRI) findings. In addition to color changes in MRI resulting from iron accumulation, some types of NBIA show a unique pattern in the MRI. The “eye of tiger” sign is the most well-known one in many cases of PKAN.⁶ Since a significant number of patients remain without a proper diagnosis, genetic testing based on clinical features and MRI findings is beneficial.⁵ NBIA is classified based on disease-causing genes, which will be demonstrated later on.

NBIA Classifications

Pantothenate Kinase-Associated Neurodegeneration; PKAN-NBIA1 (*PANK2*; Online Mendelian Inheritance in Man (OMIM)*606157)

PKAN was first described by two German physicians, Hallervorden and Spatz, so it was named Hallervorden–Spatz disease. However, due to their cooperation with the Nazi party today it is known as PKAN. It is the most common form of NBIA, which includes almost half of the cases (Figure 1). The exact prevalence of PKAN is yet unknown, but it is estimated to be 1–3/1,000,000.^{2,4} It can occur from early childhood to middle age. There is an association between the age at onset and the disease course. The early-onset form of PKAN has a higher rate of progression than the late-onset form of the disease.⁴

Although this NBIA is reported in many ethnicities and the dominant clinical picture is similar, there have been some variations in different populations. Lee et al. have found that cranial dystonias were less common in Asians than in Caucasians, and Asians tended to have segmental dystonia, whereas Caucasians were more likely to present generalized dystonia. In classic PKAN, dysarthria was more frequent in Asians, but parkinsonism

in Caucasians. Pyramidal signs and mental impairment were less prevalent in Asians.⁷

Classic Form of PKAN

This form is characterized by an early start and rapid progression. It occurs due to complete loss of function mutations in the *PANK2* gene and often develops before the age of 6 years (average age at onset is 3 years old with a median survival of 9.5 years after onset). The main presentation of PKAN is considered to be a gait disturbance. Most of the affected children to the classic form of PKAN show a history of mild growth and developmental delay as well as dyspraxia before dystonia and spasticity. Pigmented retinopathy is a common feature in classical PKAN, but seizures are not. Although PKAN is known as neuroacanthocytosis disorder, the acanthocytosis, abnormal red blood cell morphology, is found in only some patients; since the high sensitivity genetic tests are available in many countries, this feature of erythrocytes is not a common diagnostic criterion.^{4,8}

Atypical Form of PKAN

In this form of PKAN, the clinical features are different from the classic form and start at later age (with a median age at onset of 18 years and 2% mortality in the 10 years follow up after onset).⁸ It is often presented with speech abnormalities. Progression happens at a slower pace, and it is the result of an incomplete loss of function mutations in the *PANK2* gene. Other features of this form include dystonia and dysarthria. Among neuropsychiatric impairments in patients with the atypical form of PKAN, impulsivity and obsessive-compulsive disorder are the most common manifestations.⁴ Pigmentary retinopathy is rare in this form, though extraocular movement abnormalities are common.⁹

Genetics of PKAN

The pattern of inheritance is AR and the disease results from mutations in the *PANK2* gene encoding for pantothenate kinase 2 within the 20p13 chromosomal position.^{2,10} Currently, according to Human Gene Mutation Database (HGMD) professional 2019.4, 188 mutations in *PANK2* have been reported. Chang et al. have proposed that the proportion of the patients carrying two null alleles is significantly higher in early-onset (classic) patients compared to late-onset (atypical) cases (27.5% vs. 0.8%). The null allele was presumed as “loss-of-function” allele.⁸

Pathophysiology of PKAN

PANK2 enzyme is a CoA sensitive sensor in the mitochondrial inner membrane.¹¹ *PANK2* mutations lead to CoA metabolism impairments as a result of defects in the pantothenate kinase enzyme, leading to the accumulation of some of the CoA pathway substrates such as cysteine and compounds containing cysteine in basal ganglia. The result of these events is the iron accumulation in GP under the influence of rapid, toxic, and spontaneous cysteine oxidation, which leads to free radical production. Since basal ganglia and retina are sensitive to oxidative stress, the involvements of these sites are more prominent in PKAN patients.^{12,13} A specific pattern of iron accumulation is detected in many PKAN cases, which is called the “eye of tiger” sign (Figure 2A and B).⁶

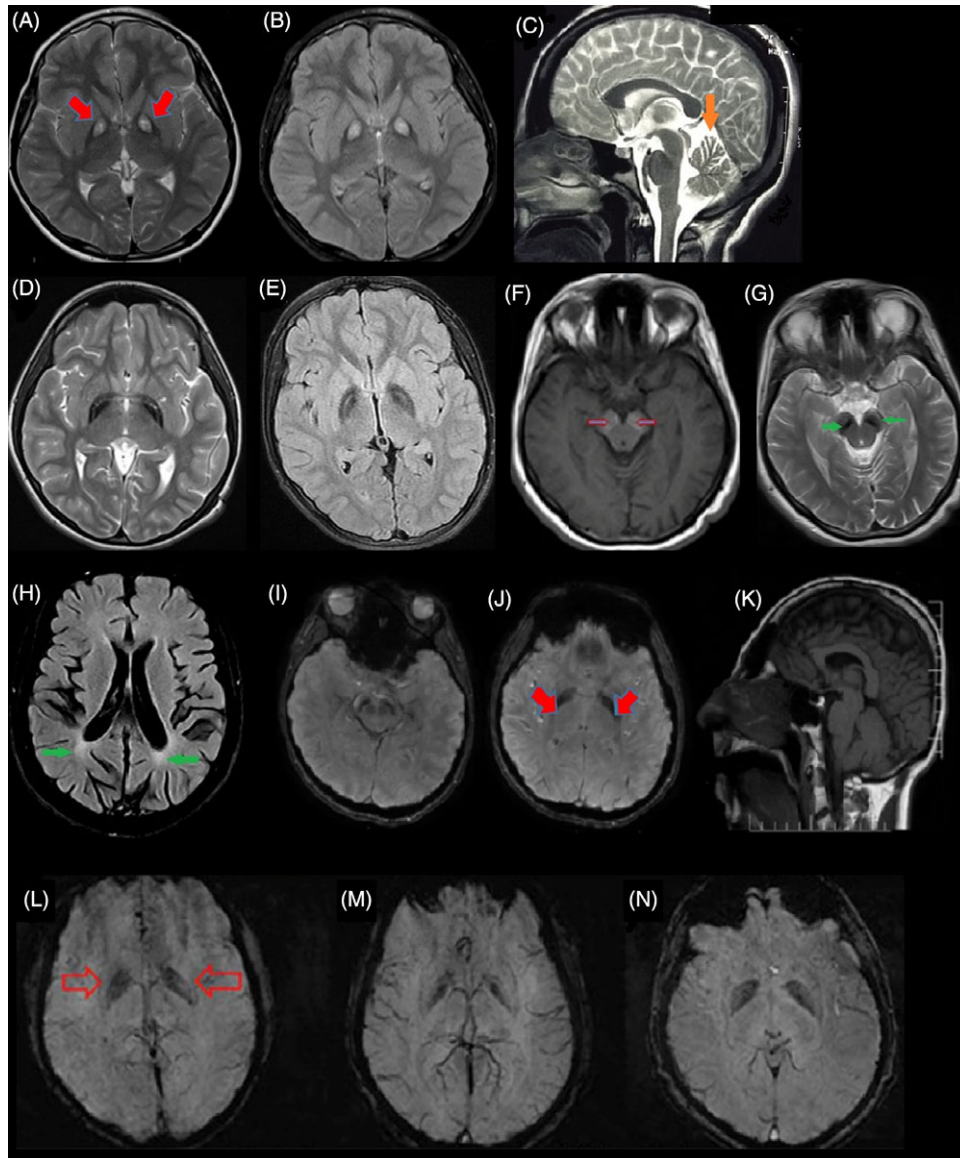


Figure 2: Brain magnetic resonance imaging (MRI) of different subtypes of NBIA from Iran. (A & B) Eye of tiger sign pattern (red arrow) in MRI of pantothenate kinase-associated neurodegeneration (PKAN) patients. (C) cerebellar atrophy (orange arrow) in phospholipase A2-associated neurodegeneration (PLAN). (D & E) iron deposition in globus pallidus (GP) with preserved medial medullary lamina in MPAN. (F & G) Halo sign in cerebral peduncles (substantia nigra [SN]) on T1-weighted sequences (empty red arrows) and iron deposition in the corresponding T2-weighted sequences in SN (green arrow) in a Beta propeller-associated neurodegeneration (BPAN) patients. (H) deep white-matter changes suggesting leukodystrophy (green arrows) in a fatty acid hydroxylase-associated neurodegeneration (FAHN) patient. (I–K) T2-weighted hypointensity signal in GP (red arrows) and SN and small pituitary gland (K) in the Woodhouse–Sakati syndrome (WSS) patients. (L–N) Susceptibility weighted imaging (SWI) in three affected to JABELS showing mild hypointensity in GP (empty red arrows) compatible with iron deposition.

Phospholipase A2-Associated Neurodegeneration; PLAN-NBIA2 (*PLA2G6*; OMIM*603604)

PLAN-NBIA2 comprises about 20% of NBIA cases (Figure 1), manifesting a broad range of symptoms, from psychological disorders in childhood to parkinsonism–dystonia in adulthood. Pathological abnormalities in the central nervous system (CNS) and peripheral nervous system (PNS) have been detected.^{2,4} This type of disease is categorized into three phenotypic forms with clinical and radiological overlap.

Infantile-Onset PLAN or Infantile Neuro-Axonal Dystrophy (INAD)

Early-onset form of disease emerges between the age of 6 months to 3 years (around 2 years) with growth retardation, hypotonia, and spastic tetraparesis.^{14–16} Many of the affected children can never walk. Seizure and ophthalmological impairment (strabismus, nystagmus, and optic atrophy) are prominent in these patients and death occur in infancy and early childhood.¹⁶

Childhood-Onset PLAN or Atypical Neuro-Axonal Dystrophy

Atypical neuro-axonal dystrophy shows more phenotypic variations in comparison with INAD. The age at onset can vary from early childhood (usually after 3 years) to the end of the second decade of life. These children show ataxia, dystonia, spasticity, delay in speaking, and autistic symptoms with degrees of intellectual disability. Strabismus, nystagmus, optic atrophy, and neuropsychological disorders are also common. These patients have slower progression and longer survival times compared to INAD cases.^{4,16}

Adult-Onset PLAN or Dystonia–Parkinsonism

Patients affected with this form present their symptoms at various ages, but most patients present symptoms early in adulthood (20–40 years). Patients manifest dystonia and Parkinsonism from late adolescence to early twenties and it might be accompanied by mild mental impairment. Survival for several decades is not unusual.^{4,16,17}

Genetics of PLAN

Mutation in *PLA2G6*, encoding for phospholipase A2, within 22q13.1 chromosomal position results in PLAN. The pattern of inheritance is considered AR.¹⁸ Most of the patients have missense mutations that do not disrupt the catalytic activity of the protein.¹⁹ Currently, according to the HGMD professional 2019.4, 191 mutations in *PLA2G6* have been reported.

Pathophysiology of PLAN

PLA2G6 encodes for a specific type of phospholipase, which catalyzes the release of fatty acid from phospholipid. It has been proposed that this gene plays a crucial role in phospholipid homeostasis, and the impairment of this enzyme increases the level of phospholipid and acetyl-CoA. Impairment in phospholipid homeostasis may change the lipid components of membrane, vesicles, or endosomes, resulting in membrane permeability, fluidity, iron homeostasis disturbance, and disruption of the autophagy process.^{20,21} Alpha-synuclein and neurofibrillary tangles in the pathologic study of PLAN have suggested a link to Parkinson's disease. Although some studies have implicated that *PLA2G6* mutation is not a major risk factor for Parkinson's disease in Asian populations.¹⁶

Mitochondrial Protein-Associated Neurodegeneration; MPAN-NBIA4 (*C19orf12*; OMIM*614297)

Historically, this type of NBIA was first discovered in a group of patients in eastern Europe.²² Global prevalence is estimated to be fewer than 1/1,000,000 and until now less than 100 patients are confirmed to be affected to MPAN. The clinical features include dystonia, Parkinsonism, spasticity, and psychological problems such as anxiety, depression, inattention, hallucination, and hyperactivity. Onset is usually in the first or second decade (median onset of 9 years), but adult-onset cases are also reported.²³ The late-onset form of disease starts in the third or fourth decade of life with disorders like psychosis and dementia.^{4,22,24} The disease severity and progression of MPAN are thought to be generally similar or milder than PKAN but data

on this are still lacking due to the rarity of this NBIA and the absence of large cohorts. There are reports of cases with preserved ambulation after two decades of symptom onset.^{22–25}

Genetics of MPAN

MPAN is caused by mutations in *C19orf12* on 19q12, a gene encoding a mitochondrial membrane protein.²² The pattern of inheritance is AR, but there are reports of autosomal dominant in some cases.²⁴ Currently, according to the HGMD professional 2019.4, 51 mutations in *C19orf12* have been reported. The most frequent type of mutations affecting this gene are p.Gly69Argfs*10 and p.Thr11Met.³ Landouere and colleagues reported c.187G>C; p.Ala63Pro mutation in two sisters affected to hereditary spastic paraplegia type-43 (HSP-SPG43) without iron accumulation in their brain MRI. This mutation was also reported in at least two other families affected by MPAN, thus the determination of genotype–phenotype correlation is challenging.²⁶

MPAN Pathophysiology

C19orf12 encodes for a small transmembrane protein localized in the mitochondrial membrane. The precise functionality remains to be unraveled, but based on several studies, fatty acid biosynthesis and destruction of branched amino acids (valine, leucine, and isoleucine) are two prominently involved pathways.²² Studies on postmortem affected samples showed increased alpha-synuclein in stained tissue and Lewy body formation in the brain. Also, it seems that spheroidal eosinophilic structures in GP are byproducts of neuronal degeneration. More investigations are needed to understand the exact pathophysiology of mutations in this gene.²⁴

Beta Propeller-Associated Neurodegeneration; BPAN-NBIA5 (*WDR45*; OMIM*300526)

The initial phenotype includes neural developmental delay in early childhood, intellectual disability (autism or a Rett-like syndrome), seizures, and sleep problems.^{27,28} Subsequently, speech skills become impaired. Ataxia, dystonia, and Parkinsonism are later features that add to the clinical picture in adolescence or early adulthood (at a mean age of 27 years).^{4,29} The initial non-motor symptoms may be mild or moderate and the clinical picture discloses itself by later motor symptoms and typical MRI features, which may be nonspecific or insignificant in early childhood imaging.^{30,31}

Genetics of BPAN

BPAN is the only form of NBIA with an X-linked dominant pattern of inheritance, resulting from mutations in the *WDR45* gene on Xp11.23. This gene encodes an autophagy protein, beta-propeller.²⁷ Up to now, most of the BPAN cases were as a result of de novo mutations, and some affected men were mosaic; probably because of the post-zygotic mutations. By the development of next generation sequencing and whole exome sequencing (WES), many cases were identified and the spectrum of clinical features has been widening.⁴ Currently, according to the HGMD professional 2019.4, 102 mutations in *WDR45* have been reported.

Pathophysiology of BPAN

In this form of the disease, there are numerous findings indicating autophagy pathway impairments. WDR45 protein is a member of a protein family that facilitates the formation of the protein complex and plays an essential role in many biological processes such as signaling, apoptosis, and gene expression regulation. WDR45 protein can bind to Autophagy Related 2A (ATG2A) and Autophagy Related 2B (ATG2B) that are two other autophagy-related proteins, indicating the affiliation of this protein in the autophagy process.^{32,33} Lymphoblastic cell line in patients affected to BPAN, with severe decrease of WDR45 protein and lower autophagy function rate leads to abnormal accumulation of initial autophagy structures.³⁴ Neuropathological investigations of BPAN affected patients showed severe neurodegeneration, axonal spheroids, siderophages, and reactive astrocytes. Abnormality in the autophagy process leads to the accumulation of degraded components in the cell, toxic conditions, and cell stress resulting in neuronal degeneration.³⁵ Iron accumulation in BPAN patients in substantia nigra (SN) resembles a halo sign pattern on T1-weighted sequences of MRI³⁶ (Figure 2F).

Coasy Protein-Associated Neurodegeneration; CoPAN-NBIA6 (Coenzyme A synthetase (COASY); OMIM*609855)

CoPAN-NBIA is a rare form of NBIA reported in 2014 in two Italian patients.³⁷ Clinical features manifested in the first decade with mild cognitive impairment and gait disturbance. Then through the second decade, oromandibular dystonia, dysarthria, and progressive spasticity, and axonal neuropathy were pursued. The complete severe phenotype in the third decade included exacerbation of prior symptoms as well as the emergence of parkinsonism, depression, and obsessive-compulsive disorder.^{1,4,9,37-39}

Genetics of CoPAN

Mutations in *COASY*, showing an AR pattern of inheritance, are the genetic causes of CoPAN. *COASY* is located on 17q21, which encodes the CoA-synthase enzyme.³⁷ Currently, according to the HGMD professional 2019.4, five mutations in *COASY* have been reported.

Pathophysiology of CoPAN

COASY is a critical enzyme in CoA biosynthesis. Defects in this gene lead to a reduction of CoA and acetyl-CoA levels. Acetyl-CoA is involved in the regulation of the autophagy process. Investigation of CoPAN patients shows strong links between lipid metabolism, CoA synthesis, and autophagy in NBIA pathology. In patients' fibroblasts culture, it has been shown that *COASY* protein and acetyl-CoA levels are lower than usual significantly.^{33,37,40,41}

Fatty Acid Hydroxylase-associated Neurodegeneration; FAHN (FA2H; OMIM*611026)

Clinical hallmarks of this disease include ataxic spastic gait, starting in the first decade (median age of 4 years) followed by a decline in intellectual function, dystonia, seizure, divergent

strabismus, and exacerbated cerebellar and pyramidal signs in the second decade. With the disease progression, as a matter of time (median 7 years), dystonia and spasticity limit the ability of movement; thus, patients require a wheelchair in the adolescence or early twenties.^{1,4,42-44}

Genetics of FAHN

Defects in this gene were described in HSP (HSP-SPG35) and since then, mutations in this gene were introduced in movement disorders with brain iron accumulation.⁴² Nowadays, HSP-SPG35 is regarded as a type of FAHN disease because of the pattern of iron accumulation in basal ganglia in many reported cases.⁴ FAHN is due to AR mutations in *FA2H* on 16q23.1,² also there are some reports of uniparental disomy.⁴⁵ Currently, according to the HGMD professional 2019.4, 65 mutations in *FA2H* have been reported.

Pathophysiology of FAHN

FA2H enzyme is a 2-hydroxylase enzyme expressed in Schwann cells and oligodendrocytes.⁴⁶ 2-hydroxy sphingolipids produced by *FA2H* are the most abundant lipids in the myelin sheath. Mutations in this gene lead to a decreased level of enzyme activity and, subsequently, myelin sheath degeneration.^{33,47} *FA2H* is also a lipid raft movement regulator. Lipid rafts are cholesterol and sphingolipid structures of membrane involved in membrane transporting and signaling. Guo and colleagues showed that *FA2H*-dependent hydroxylation affects membrane protein transport due to lysosomal degradation. Based on these findings, defects in *FA2H* lead to changes in the 2-hydroxy sphingolipid profile that can affect membrane lipids.^{33,48}

Aceruloplasminemia (CP; OMIM*117700)

Aceruloplasminemia is one type of NBIA with direct relation to iron homeostasis and is reported globally, but it seems to be more common in the Japanese population (1/2,000,000). Accumulation of iron in the brain and other organs leads to neurodegeneration, retinopathy, and diabetes mellitus. Neurologic symptoms mainly include movement disorders including ataxia, chorea, dystonia, and tremor with cognitive and psychological changes. The age at onset is mainly in early to middle adulthood (median age of neurologic symptom onset, 51 years).^{4,5,49} Diabetes is reported in almost 70% of affected individuals with microcytic or normocytic anemia because of iron accumulation in the pancreas. Since diabetes mellitus and microcytic anemia often occur one or two decades earlier than neurologic signs (median age, 35 years), it is an important clue in the early diagnosis of patients.⁴⁹⁻⁵¹ A mortality of 30% is reported in patients under chelation therapy after a median of 7 years of neurological symptoms onset (mean age at death, 60 years).⁴⁹

Genetics of Aceruloplasminemia

Loss of function mutations in the *CP* gene results in the AR form of NBIA, called aceruloplasminemia. *CP* is located on 3q21-24 and encodes a plasma membrane glycoprotein that plays a role in iron transport and processing in a way that helps the binding of iron to transferrin. Transferrin transports the iron to the red blood cells to carry oxygen. Mutations in *CP* lead to unstable and nonfunctional protein, which does not transport the iron

out of cells and its accumulation will damage the cells and tissues. Currently, according to the HGMD professional 2019.4, 65 mutations in this gene have been reported.^{52,53}

Pathophysiology of Aceruloplasminemia

Ceruloplasmin is the only ferroxidase, which is expressed by astrocytes. Defects in this protein lead to a disability of astrocytes in oxidizing the iron that is directly being transferred to CNS.⁵⁴ There are two forms of ceruloplasmin protein; the secretory form that is mainly expressed in the liver and the GPI-linked form that is mainly expressed in the brain.⁵⁵ Based on in vivo and in vitro studies, several factors contribute to the pathogenesis of disease such as: (1) ceruloplasmin dysfunction that leads to iron accumulation; (2) oxidative stress and the reactive oxygen species (ROS) production; (3) increased level of lipid peroxidation; and (4) change in lipid environment that leads to mitochondrial dysfunction and neuronal degeneration.⁵⁶

Neuroferritinopathy-NBIA3 (*FTL*; OMIM*134790)

Neurological signs such as chorea, dystonia, cerebellar symptoms, psychosis, depression, cognitive impairment, Parkinsonism, dysarthria, insomnia, sleep apnea, areflexia, a positive Romberg test, and dysautonomia are the clinical features of this type of NBIA.^{57–59} The onset is between the second to fifth decades (mean age, 40 years) depending on the type of mutation. Neuroferritinopathy progresses gradually but in those with c.458dupA mutation a rapid progression of Parkinsonism, ataxia, and neuropsychiatric symptoms has been reported. Death is mostly due to aspiration pneumonia.⁵⁹

Genetics of Neuroferritinopathy

Neuroferritinopathy is due to mutations in the *FTL* gene, which encodes the light chain of ferritin. Ferritin is regarded as the main iron storage protein. *FTL* is located on 19q13.33. Mutations in this gene are transmitted by an autosomal-dominant pattern of inheritance with complete penetrance. Most of these mutations cluster in the fourth exon. Cataract hyperferritinemia syndrome is due to a mutation in the 5'-untranslated region (5'UTR) in *FTL* with no neurological symptoms.^{2,4} Currently, according to the HGMD professional 2019.4, 66 mutations in *FTL* have been reported, and interestingly, 36 mutations are in regulatory regions of the gene.

Pathophysiology of Neuroferritinopathy

The affected individuals' fibroblast culture revealed the accumulation of ferritin in the cytoplasm and nucleus, the lower expression of transferrin receptor, and the higher expression of divalent metal transporter 1 and ROS formation. Defect in ferritin leads to a higher level of unstable iron in cells, therefore increases the ROS production and oxidized protein, decreasing the proteasome proficiency.⁶⁰ Pathologic investigations showed ferritin inclusion body in the cytoplasm and nucleus of glia and CNS neurons and also in other organs related to iron.

Kufor-Rakeb Syndrome; KRS (ATPase Cation Transporting 13A2 (*ATP13A2*); OMIM*610513)

It is a rare type of atypical young onset Parkinsonian syndrome, which occurs before the age of 20 years (Parkinson disease 9 (PARK9)). "Kufor-Rakeb" is the name of a region in

Jordan, where it was first reported.⁶¹ Clinical features like visual hallucinations, dementia, perioral myokymia (facial-facial-finger mini myoclonus), supranuclear ophthalmoplegia, and spasticity have been reported. Since, in some cases, the iron accumulation in basal ganglia is detectable on brain MRI, it is suggested to be considered as NBIA syndrome. The Parkinsonism responds to levodopa but is complicated by the early development of motor fluctuations.^{4,5,9,62}

Genetics of Kufor-Rakeb Syndrome

It is caused by mutations in *ATP13A2* located on 1p36.13, encoding a p-type ATPase. Affected individuals show juvenile Parkinsonism, and in some cases, there are reports of iron accumulation in basal ganglia.^{63,64} The pattern of inheritance is AR and currently, according to the HGMD professional 2019.4, 53 mutations in *ATP13A2* have been reported. Mutations in this gene were also reported in autosomal recessive-hereditary spastic paraplegia (AR-HSP), spastic paraplegia 78 (SPG78). The disease emerges in adulthood, presenting mainly with spasticity and rarely Parkinsonism.⁶⁵

Pathophysiology of Kufor-Rakeb Syndrome

In vitro studies show that mutant *ATP13A2* protein remains in the endoplasmic reticulum (ER), and then is degraded by the proteasome. The mutant protein accumulation can lead to ER stress induction.^{66,67} Investigation of affected individual cultured fibroblast shows that lack of *ATP13A2* leads to lysosomal dysfunction because of vesicle structures (lysosomes and autophagy vacuoles) accumulation and proteolytic dysfunction of the lysosomal enzymes, and disability to digest lysosomal substrates.^{68,69} Deficiencies in the mitochondrial respiratory chain lead to a decrease in ATP production. In Kufor-Rakeb patients, mitochondrial DNA is fragmented, and its integrity is lost.⁷⁰

Woodhouse-Sakati Syndrome; WSS (DDB1 And CUL4 Associated Factor 17 (*DCAF17*); OMIM*612515)

This form of NBIA is a rare AR disorder, which occurs in adolescence. WSS consists of mixed neurologic and endocrine features including deafness, alopecia, progressive dystonia, dysarthria, dysphagia, cognitive impairment, intellectual disability, hypogonadism, and diabetes mellitus. Some patients show iron accumulation in GP and SN (Figure 2I–K). WSS occurs due to mutations in the *DCAF17* gene on 2q31.1 that encodes a transmembrane nuclear protein with unknown function.^{71,72} Currently, according to the HGMD professional 2019.4, 21 mutations in *DCAF17* have been reported mainly in the Middle East. Two clinical subtypes based on the severity and progressive disabilities (mainly due to neurological symptoms) are proposed. The severe subtype manifests earlier (mean age, 12.6) with progressive generalized dystonia leading to disability in mean 7.4 years whereas the second type starts later (mean age, 18.1) with focal limb or cervical dystonia and patients remain ambulatory for at least two decades.⁷³

Jaberi-Elahi Syndrome; JABELS SYNDROME (*GTPBP2*; OMIM*607434)

This form is also known as neurodevelopmental retardation with a delay in growth and intellectual disability. It starts in childhood, and the severity of symptoms is variable. Some

patients have social interactions, with ataxic gait, tremor, or dystonia, while others might not achieve any movement control and are not able to speak. Optic anomaly, microcephaly, abnormality in hand and foot structure, kyphoscoliosis, dysmorphic face, and seizures are their other features. MRI usually shows cerebellar and corpus callosum hypoplasia and atrophy. Recently, Jaber and colleagues reported an Iranian family with consanguineous marriage in which four affected children with neurodevelopmental impairment and age at onset in childhood were born. All affected children showed psychomotor retardation including mild delay in motor milestones, speech delay, intellectual disability, and behavior abnormalities. MRI findings indicated cerebellum atrophy, hypointensity in GP, and SN indicating abnormal iron deposition (Figure 2L–N). Homozygosity mapping and WES analysis in this family revealed a homozygous splice-site variant, in the *GTPBP2* gene.⁷⁴ *GTPBP2* is located on 6p21.1 and encodes a GTP-binding protein, which participates in the reporter mRNA stability but the precise function is still unknown.⁷⁵ GTP-binding protein 1 (GTPBP1) and GTPBP2 are members of the GTPase superfamily, which participate in cell proliferation, differentiation, and protein synthesis.^{76,77}

Leukoencephalopathy with Dystonia and Motor Neuropathy; LKDMN (Sterol Carrier Protein 2 (SCP2); OMIM*184755)

LKDMN is an AR disorder, reported in 2006 for the first time in an adult person with dystonic head tremor, spasmodic torticollis, azoospermia, hypogonadotropic hypogonadism, mild cerebellar ataxia, gait impairment, and hyposmia. Plasma metabolic analysis revealed pristanic acid accumulation and abnormal excretion of bile alcohol glucuronides in the urine. Brain MRI revealed bilateral hyperintensity signal on thalami, Pons, and occipital lobes.⁷⁸ Molecular analysis revealed a homozygous missense mutation in *SCP2*. Recently, the second affected individual, a 51 years old man with a compound heterozygous missense mutation in this gene, was identified. He had movement impairment and ataxia with NBIA features in MRI but no leukoencephalopathy.⁷⁹ The *SCP2* gene is located on 1p32.3 and encodes two different proteins; sterol carrier protein X (SCPx) and SCP2 that are result of transcription initiation with two unrelated regulatory promoters.⁸⁰ Both these products are peroxisomal enzymes with thiolase activity utilized for branched-chain fatty acids breakage. Pathologic hallmarks in this disorder are probably due to the branched chain fatty acids accumulation.⁸¹ Affected individuals with mutations in *SCP2* may have an abnormality in fatty acid acetyl-CoA metabolism, which is a common mechanism in NBIA pathogenicity. This fact indicates that iron accumulation in the brain can be a secondary phenomenon due to metabolic defects. By this hypothesis, iron chelation-based therapeutics remain uncertain.⁷⁹

New Subtypes of NBIA

NBIA7 (Rala Binding Protein 1 (RALBP1) Associated Eps Domain Containing 1 (REPS1); OMIM*614825)

Drecourt and colleagues reported two sisters with unrelated healthy parents who originated from France. The clinical features in the first sister were speech and motor delay, hypotonia, progressive cerebellar ataxia, losing the ability to walk at the

age of nine, and nystagmus. Dysarthria, dysmetria, spasticity of the lower limbs, and pes cavus were also reported. Cerebellar and cerebral atrophy and iron accumulation in the GP and cerebral peduncles showed on the brain MRI. She eventually died at the age of 20 years. The second sister had the same but milder features. She can walk yet but with help at the age of 14 years. WES identified a compound heterozygous variant, c.232G>C and c.338C>A, in the *REPS1* gene. Both variants were in an epsin-15 homology domain, a conserved region that interacts with RALBP1 to form the endosome recycling compartment. *REPS1* is located on 6q24.1 and encodes RALBP1-associated Eps domain-containing protein-1. This protein is affiliated to autophagy because of its involvement in endocytosis, vesicle transport, localization in the cytoplasm, and endosome. This type of disease is known as NBIA7.^{82,83}

NBIA8 (Carnitine O-Acetyltransferase (CRAT); OMIM*600184)

Drecourt et al. also reported another subtype of NBIA in a Turkish girl with first cousin parents. Clinical manifestations include hypotonia, brisk deep tendon reflexes, sensory neuropathy, tremor, dysmetria with evidence of cerebellar atrophy, posterior leukoencephalopathy, hyperintensity of basal ganglia, hypointensity of GP, and SN which is suggestive for iron accumulation. Single-nucleotide polymorphism (SNP) genotyping and WES revealed a homozygous variant c.962G>A in the *CRAT* gene. This gene is located on 9q34.11 and encodes carnitine O-acetyltransferase. The encoded protein is localized in mitochondria and catalyzes the reversible transfer of acyl groups from carnitine to CoA and regulates the acyl-CoA/CoA ratio. It also plays a crucial role in the transport of fatty acids for beta-oxidation. This type of disease is known as NBIA8.^{82,83}

Hereditary Spastic Paraplegia-50; HSP-SPG50 (Adaptor Related Protein Complex 4 Subunit Mu 1 (AP4M1); OMIM*602296) or AP4M1-Associated NBIA

This gene encodes a subunit of the heterotetrameric AP-4 complex. AP4 is composed of four chains: beta-4 and epsilon-4 (AP4B1 and AP4E1 large chains), mu-4 (AP4M1 medium-chain), and sigma-4 (AP4S1 small chain). *AP4M1* gene is located on 7q22.1 and encodes the mu subunit of the adaptor protein complex-4. This gene is ubiquitously expressed in the CNS and plays a role in vesicle formation, post-Golgi protein trafficking, and sorting processes.^{83,84} Although mutations in genes encoding the different subunits of adaptor protein complex-4 have been reported in patients affected to different types of HSP, Roubertie and colleagues in 2018 reported the clinical features of three patients originating from a large consanguineous Moroccan family who exhibited early-onset developmental delay, juvenile motor function deterioration, spasticity, and pyramidal tract signs and intellectual deficiency in which brain MRI findings were compatible with disorders in the NBIA spectrum. Their MRI showed the isointense pattern of the GP and hypointensity of the GP on T1 and T2 sequences, respectively. WES was performed and revealed a homozygous mutation c.916C>T;p.Arg306* in the *AP4M1* gene. These data show the overlap between NBIA and hereditary spastic paraplegia.⁸⁵ Previously, Najmabadi and colleagues reported a mutation in *AP4M1* in a patient with microcephaly and paraplegia.⁸⁶

• PKAN	Generalized dystonia including oromandibular dystonia, retinitis pigmentosa, eye of the tiger sign in MRI
• MPAN	Spastic paraplegia, optic atrophy, dystonia, MRI shows iron in GP with preserved medial medullary lamina
• PLAN	Neuropathy, optic atrophy, Imaging shows cerebellar atrophy
• Aceruloplasminemia	Diabetes mellitus, diffuse iron deposition in brain including thalami in MRI, low ceruloplasmin
• Neuroferritinopathy	AD inheritance, chorea, dementia, MRI: diffuse iron deposition, putamenal necrosis and cavitation
• BPAN	Static encephalopathy of childhood, autism, Rette like syndrome, Parkinsonism and dystonia in adolescence, MRI: Halo sign in SN
• WSS	Hypogonadism, alopecia, deafness diabetes mellitus, MRI shows white matter hyperintensities
• FAHN	Spastic paraplegia, MRI reveals evidences of leukodystrophy
• KRS	Parkinsonism, supranuclear vertical gaze palsy, cognitive impairment
• CoasyPAN	Pyramidal and extrapyramidal signs, swollen putamen and caudate GP calcification in imaging

Figure 3: Diagnostic clues for main subtypes of NBIA.

PARK15 (FBXO7; OMIM*605648) or FBXO7-Associated NBIA

The F-box only protein 7 (*FBXO7*) gene is located on 22q12.3 and encodes a protein involved in the ubiquitin–proteasome protein-degradation pathway and mitophagy. In 2008, Shojae and colleagues performed a linkage analysis in a large Iranian family with Parkinsonian-pyramidal syndrome and found a region on chromosome 22 linked to disease. Sequencing was done for candidate genes in that region which revealed a disease-associated variant in the *FBXO7* gene, PARK15. Recently Correa-Vela and colleagues reported a novel homozygous mutation, c.368C>G; p.S123*, in *FBXO7* in a child with spastic paraplegia, epilepsy, cognitive decline, cerebellar degeneration, levodopa nonresponsive Parkinsonism. Brain MRI revealed iron deposition in the GP and SN appeared as a hypointense signal on iron-sensitive sequences which is compatible with NBIA. All these findings suggest that *FBXO7*-associated NBIA can be a novel subtype of the NBIA and there are clinically and genetically overlap between NBIA and other neurodegenerative disorders.^{87,88}

Differential Diagnosis of NBIA

Clinical features of NBIA are often diverse and involved different systems like pyramidal, extrapyramidal, cortical, and cerebellar, so the differential diagnosis consists of a broad spectrum of disorders such as inborn errors of metabolism, lysosome storage diseases, hereditary spastic paraplegia, and other forms of parkinsonism–dystonia.⁷⁰ T2-weighted hypointensity in GP is also reported in cases with mutations in Lysine Methyltransferase 2B (*KMT2B*) (*KMT2B*-related dystonia), Solute Carrier Family 39 Member 14 (*SLC39A14*) (*SLC39A14*-related early-onset dystonia–parkinsonism) and Sequestosome 1 (*SQSTM1*) (neurodegeneration with ataxia, dystonia, and gaze palsy, childhood-onset [NADGP]) (<https://www.ncbi.nlm.nih.gov/>

[books/NBK121988/](#)). On the other hand, the lack of iron in the early stage of disease makes the diagnosis more challenging. It is noteworthy that the level of iron in the brain is deeply dependent on age in a way that there is no iron accumulation at birth but tends to increase during development.³ Diagnostic clues for the main subtypes of NBIA are summarized in Figure 3.

Molecular Mechanism of NBIA Proteins

Different cellular and molecular pathways are mediated by NBIA proteins such as CP and FTL, which are iron homeostasis regulator; PLA2G6, FA2H, SCP2, CRAT, and C19orf12, that participate in lipid metabolism; PANK2 and COASY are key enzymes in CoA biosynthesis; WDR45 and ATP13A2 that participate in autophagy process; FBXO7 involved in ubiquitin–proteasome protein-degradation pathway and mitophagy; REPS1 and AP4M1 are associated with vesicle trafficking; finally, DCAF17 and GTPBP2 with unclear function.^{4,83,87}

Treatment

Treatment of NBIA has been mostly symptomatic, supportive, and palliative for many decades. Drugs like tetrabenazine, baclofen, anticholinergics, antidopaminergic agents, and levodopa are partially useful in the treatment of major symptoms of NBIA.^{3,89} However, there has been a great struggle to implement disease-modifying drugs for NBIA patients. Chelation therapy for iron has been the major subject of research in this regard, but disease-specific treatment measures are being deployed such as (1) delivering alternative substrates downstream of the defective enzyme, (2) activating isoenzymes, (3) counteracting downstream cellular effects, (4) gene therapy, and (5) enzyme replacement therapy. Deferiprone, an oral iron chelator, which crosses the blood-brain easily is the most common chelating agent investigated in NBIA. Despite the numerous investigations conducted for PKAN the results regarding deferiprone efficacy in this NBIA is still controversial. There are few reports of chelation therapy in PLAN, BPAN, MPAN and neuroferritinopathy with mixed results and data in these subtypes are scarce. Regarding aceruloplasminemia, due to iron deposition in other organs alongside the brain, the number of studied cases and the results are much more promising. Beyond the iron chelation many studies in preclinical or clinical phases are investigating the effect of specific genetic or enzymatic interventions on different NBIA subtypes as alluded above.⁹⁰

NBIA in IRAN

There are more than 20 studies about NBIA in Iran (Table 1); however, there is a lack of exact information about the prevalence of NBIA among this population. Until the time of this paper, 39 families with 53 affected individuals are reported in Iran. Respectively, there were 23, 5, 3, 1, 3, 3, and one family with PKAN, MPAN, PLAN, JABELS, BPAN, Kufor-Rakeb and WSSs in Iran (Figure 4).^{74,91–109} Like the other countries, the most common form of NBIA was PKAN accounting for more than half of the cases (59%) (Figure 4). A high percentage of the PKAN may be due to the easy diagnostics procedures; the specific pattern on MRI, which is the “eye of tiger” sign. MPAN was the second most common form (13%) of NBIA among the Iranian population. While based on investigations in other countries, the PLAN seems to be more prevalent and accounts

Table 1: Iranian reported NBIA affected individuals

Disease/# family	Gene	Ref	Family	Consanguinity	Gender	Age at onset	Mutation		Zygoty
						(year)	cDNA	Protein	
PKAN/23	PANK2	79	1	+	M	6	c.1442G>C	p.Arg481Pro	Hom
			2	+	M	3	c.1594C>T	p.Arg532Trp	Hom
			3	+	M	3	c.1168A>T	p.Ile390Phe	Hom
			4	+	M	7	c.833G>T	p.Arg278Leu	Hom
			5	+	M	7	c.1069C>T	p.Arg357Trp	Hom
			6	+	F	12	c.1208A>T	p.Asp403Val	Hom
			7	+	M	17	c.700A>C	p.Thr234Pro	Hom
		76	8	NA	M	7	NA	NA	NA
			9	NA	F	14	NA	NA	NA
		78	10	+	M	2	c.1017-1020 del AGAT ins GCTTTGCAAAC	Premature Stop codon	Hom
		81	11	+	F	2	c.936T>A	p.Cys312*	Hom
					F	NA			Hom
		77	12	+	M	10	c.1069C>T	p.Arg357Trp	Hom
					F	10			Hom
					M	9			Hom
		93	13	NA	F	4	NA	NA	NA
		83	14	+	F	6	c.1208A>T	p.Asp403Val	Hom
		84	15	NA	F	12	c.1208A>T	p.Asp403Val	Hom
					M	24	c.G833T	p.Arg278Leu	Hom
					M	22	c.1583C>T	p.Thr528Met	Hom
		85	18	+	M	10	c.833G>T	p.Arg278Leu	Hom
		82	19	+	F	4	c.1426_1429delATGA	p.Met476fs	Hom
		87	20	+	F	16	c.706_708delGAA	p.Glu236del	Hom
M	18				c.706_708delGAA	p.Glu236del	Hom		
90	21	+	F	2	c.1168A>T	p.Ile390Phe	Hom		
			M	7	c.1168A>T	p.Ile390Phe	Hom		
			F	Early infancy	c.735-736insT	p.Lys246*	Hom		
MPAN/5	C19orf12	79	1	+	F	11	c.191insG	p.Ala67GlyfsX14	Hom
					F	11			Hom
			2	+	M	12	c.404insT	p.Met135IlefsX15	Hom
			3	+	M	21	c.32C>T	p.Thr11Met	Hom
			4	+	M	26	c.32C>T	p.Thr11Met	Hom
86	5	+	F	36	c.32C>T	p.Thr11Met	Hom		

PLAN/3	<i>PLA2G6</i>	82	1	+	M	1.5	c.3G>T	p.Met11le	Hom
		91	2	+	M	25	c.1894C>T	p.Arg632Trp	Hom
					M	22	c.1894C>T	p.Arg632Trp	Hom
					F	21	c.1894C>T	p.Arg632Trp	Hom
89	3	+	M	18	?	p.Ala681Cysfs*92	Hom		
JABELS/1	<i>GTPBP2</i>	59	1	+	F	NA	c.1237-1G>T	Mis-splicing	Hom
					M	NA			Hom
					M	NA			Hom
BPAN/3	<i>WDR45</i>	88	1	NA	F	9 months	c.439-2T>G	Mis-splicing	Het
			2	NA	F	Early infancy	c.411dupT (c.412 insT)	p.Glu138*	Het
			3	NA	F	Early infancy	c.729-2A>G	Mis-splicing	Het
KRS/3	<i>ATP13A2</i>	92	1	+	F	10	c.2455C>T	p.Arg819*	Hom
					F	10			Hom
					F	12			Hom
					M	11			Hom
					F	12			Hom
		80	2	+	F	14	c.2572C>T	p.Gln858*	Hom
					M	10			Hom
3	+	M	30	c.2572C>T	p.Gln858*	Hom			
WSS/1	<i>DCAF17</i>	75	1	-	M	24	c.1091+1G>A	Mis-splicing	Hom

NA: not available; M: male; F: female; Hom: homozygous; Het: heterozygous; KRS: Kufor-Rakeb syndrome; Ref: reference; WSS: Woodhouse-Sakati syndrome.

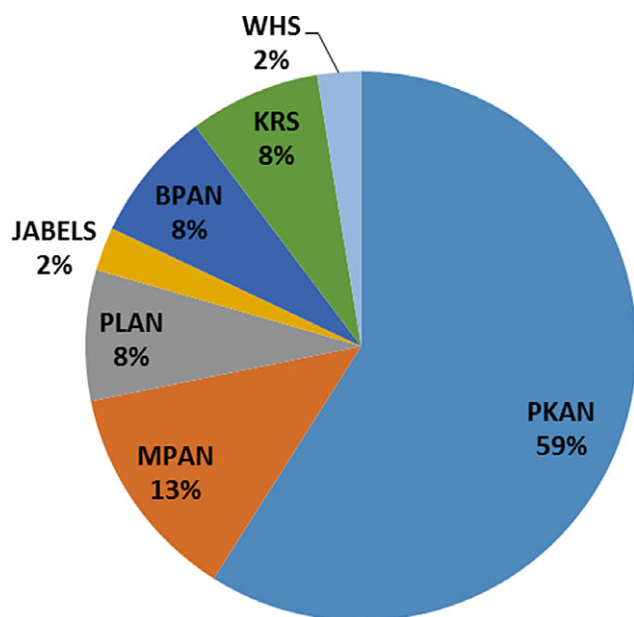


Figure 4: Prevalence of NBIA subtypes among the Iranian population.

for about 20% of NBIA cases, it was accounted for only 8% of the Iranian NBIA patients. The low frequency of PLAN may be explained by several reasons: (1) lower prevalence in Iran; (2) less diagnosis (milder phenotypes in atypical forms and overlaps with other neurodegenerative disorders); and (3) failure to report cases. In the authors' opinion, one main reason is the lack of referring these cases from neuropediatric clinics to movement disorders centers including ours.

BPAN and KRS had prevalences equal to PLAN cases. The reports of KRS from Iran (eight patients from three families) look very interesting since there are few case reports from all over the world. Interestingly, JABELS was reported from Iran for the first time, and subsequently, mutations in gene *GTPBP2* were reported in three unrelated Arab families from other countries, with a wide range of neurologic and non-neurologic features.¹¹⁰ The first only case of WSS in Iran has also been reported recently.⁹¹

Also, we found one case of FAHN and a family affected by aceruloplasminemia with two affected siblings who have not been reported yet. There are no reports of neuroferritinopathy and CoPAN in the Iranian population so far, meaning that these forms are very rare (Table 1).

CONCLUSION

Due to phenotype and genotype heterogeneity in NBIA, diagnosis is challenging and demanding. In many cases, due to lack of specific iron accumulation patterns on brain MRI, misdiagnosis is inevitable, while others have overlapping clinical symptoms. The aforementioned explanations plus unreported cases might result in lower prevalence compared to the real prevalence in some populations. In some cases, there are mutations in already-known genes related to NBIA without evidence of iron accumulation; on the contrary, some cases show sensible iron accumulation in the brain but no variation in already-known genes, making this disease far more complicated. Despite the undiscovered points about NBIA, the issue that there is no

relationship between many NBIA genes and iron deposition increases the complexity of NBIA.

This shows regarding the identification of molecular and pathophysiological aspects of NBIA, we are still at the beginning of the road to understand the real nature of this entity, and more investigations are needed. Since the pattern of inheritance in NBIA is mainly AR, in populations with consanguine marriage like Iran, it is likely to find novel NBIA genes and the consanguine marriages may explain the higher prevalence of NBIA in Iran in comparison with the worldwide prevalence. Recent findings of NBIA suggest that other HSPs and Parkinson's disease genes might be also involved in NBIA, so it indicates that some NBIA genes might not be reported yet.

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CONFLICTS OF INTEREST

The authors have no conflicts of interest to declare.

STATEMENT OF AUTHORSHIP

(1) Research project: A. Conception, B. Organization, C. Execution; (2) Statistical Analysis: A. Design, B. Execution, C. Review and Critique; and (3) Manuscript: A. Writing of the first draft, B. Review and Critique. Reza Hajati: 3B. Fardad Danaee Fard: 3B. Maziar Emamikhah: 2C, 3B. Mohammad Rohani: 1A, 1B and 3B. Afagh Alavi: 1A, 1B and 3B.

REFERENCES

- Schneider SA. Neurodegenerations with brain iron accumulation. *Parkinsonism Relat Disord.* 2016;22:S21–5.
- Di Meo I, Tiranti V. Classification and molecular pathogenesis of NBIA syndromes. *Eur J Paediatr Neurol.* 2018;22:272–84.
- Schneider SA. Neurodegeneration with brain iron accumulation. *Curr Neurol Neurosci Rep.* 2016;16:9.
- Hayflick SJ, Kurian MA, Hogarth P. Neurodegeneration with brain iron accumulation. *Handb Clin Neurol.* 2018;147:293–305.
- Wiethoff S, Houlden H. Neurodegeneration with brain iron accumulation. In: Kovacs GG, Alafuzoff I, editors. *Handbook of clinical neurology.* Amsterdam, the Netherlands: Elsevier; 2018, pp. 157–66.
- Sethi KD, Adams R, Loring D, El Gammal T. Hallervorden-Spatz syndrome: clinical and magnetic resonance imaging correlations. *Ann Neurol.* 1988;24:692–4.
- Lee C-H, Lu C-S, Chuang W-L, et al. Phenotypes and genotypes of patients with pantothenate kinase-associated neurodegeneration in Asian and Caucasian populations: 2 cases and literature review. *Sci World J.* 2013;2013:860539.
- Chang X, Zhang J, Jiang Y, Wang J, Wu Y. Natural history and genotype-phenotype correlation of pantothenate kinase-associated neurodegeneration. *CNS Neurosci Ther.* 2020;26:754–61.
- Hogarth P. Neurodegeneration with brain iron accumulation: diagnosis and management. *J Mov Disord.* 2015;8:1–13.
- Hartig MB, Hörtnagel K, Garavaglia B, et al. Genotypic and phenotypic spectrum of PANK2 mutations in patients with neurodegeneration with brain iron accumulation. *Ann Neurol.* 2006;59:248–56.
- Leonardi R, Rock CO, Jackowski S, Zhang Y-M. Activation of human mitochondrial pantothenate kinase 2 by palmitoylcarnitine. *Proc Natl Acad Sci.* 2007;104:1494–9.

12. Gregory A, Hayflick SJ. Neurodegeneration with brain iron accumulation. *Folia Neuropathol.* 2005;43:286–96.
13. Bokhari MR, Bokhari SRA. Hallervorden spatzi disease (Pantothenate kinase-associated neurodegeneration, PKAN). StatPearls Treasure Island (FL): StatPearls Publishing LLC; 2019.
14. Gregory A, Westaway S, Holm IE, et al. Neurodegeneration associated with genetic defects in phospholipase A2. *Neurology.* 2008;71:1402–9.
15. Kurian M, Morgan N, MacPherson L, et al. Phenotypic spectrum of neurodegeneration associated with mutations in the PLA2G6 gene (PLAN). *Neurology.* 2008;70:1623–9.
16. Guo Y-P, Tang B-S, Guo J-F. PLA2G6-associated neurodegeneration (PLAN): review of clinical phenotypes and genotypes. *Front Neurol.* 2018;9:1100.
17. Paisan-Ruiz C, Bhatia KP, Li A, et al. Characterization of PLA2G6 as a locus for dystonia-parkinsonism. *Ann Neurol.* 2009;65:19–23.
18. Morgan NV, Westaway SK, Morton JE, et al. Erratum: PLA2G6, encoding a phospholipase A2, is mutated in neurodegenerative disorders with high brain iron *Nature Gen.* 2006;38:957.
19. Engel LA, Jing Z, O'Brien DE, Sun M, Kotzbauer PT. Catalytic function of PLA2G6 is impaired by mutations associated with infantile neuroaxonal dystrophy but not dystonia-parkinsonism. *PLoS One.* 2010;5:e12897.
20. Malik I, Turk J, Mancuso DJ, et al. Disrupted membrane homeostasis and accumulation of ubiquitinated proteins in a mouse model of infantile neuroaxonal dystrophy caused by PLA2G6 mutations. *Am J Pathol.* 2008;172:406–16.
21. Shinzawa K, Sumi H, Ikawa M, et al. Neuroaxonal dystrophy caused by group VIA phospholipase A2 deficiency in mice: a model of human neurodegenerative disease. *J Neurosci.* 2008;28:2212–20.
22. Hartig MB, Iuso A, Haack T, et al. Absence of an orphan mitochondrial protein, c19orf12, causes a distinct clinical subtype of neurodegeneration with brain iron accumulation. *Am J Hum Genet.* 2011;89:543–50.
23. Sparber P, Krylova T, Repina S, et al. Retrospective analysis of 17 patients with mitochondrial membrane protein-associated neurodegeneration diagnosed in Russia. *Parkinsonism Relat Disord.* 2021;84:98–104.
24. Hogarth P, Gregory A, Kruer MC, et al. New NBIA subtype: genetic, clinical, pathologic, and radiographic features of MPAN. *Neurology.* 2013;80:268–75.
25. Schulte EC, Claussen MC, Jochim A, et al. Mitochondrial membrane protein associated neurodegeneration: a novel variant of neurodegeneration with brain iron accumulation. *Mov Disord.* 2013;28:224–7.
26. Gregory A, Hartig M, Prokisch H, Kmiec T, Hogarth P, Hayflick SJ. Mitochondrial membrane protein-associated neurodegeneration. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. *GeneReviews*® [Internet]. Seattle (WA): University of Washington; 2014.
27. Haack TB, Hogarth P, Kruer MC, et al. Exome sequencing reveals de novo WDR45 mutations causing a phenotypically distinct, X-linked dominant form of NBIA. *Am J Hum Genet.* 2012;91:1144–9.
28. Hayflick SJ, Kruer MC, Gregory A, et al. Beta-propeller protein-associated neurodegeneration: a new X-linked dominant disorder with brain iron accumulation. *Brain.* 2013;136:1708–17.
29. Stige KE, Gjerde IO, Houge G, Knappskog PM, Tzoulis C. Beta-propeller protein-associated neurodegeneration: a case report and review of the literature. *Clin Case Rep.* 2018;6:353–62.
30. Chard M, Appendino JP, Bello-Espinosa LE, et al. Single-center experience with Beta-propeller protein-associated neurodegeneration (BPAN); expanding the phenotypic spectrum. *Mol Gen Metab Rep.* 2019;20:100483.
31. Kimura Y, Sato N, Ishiyama A, et al. Serial MRI alterations of pediatric patients with beta-propeller protein associated neurodegeneration (BPAN). *J Neuroradiol.* 2021;48:88–93.
32. Behrends C, Sowa ME, Gygi SP, Harper JW. Network organization of the human autophagy system. *Nature.* 2010;466:68.
33. Meyer E, Kurian MA, Hayflick SJ. Neurodegeneration with brain iron accumulation: genetic diversity and pathophysiological mechanisms. *Annu Rev Genomics Hum Genet.* 2015;16:257–79.
34. Saito H, Nishimura T, Muramatsu K, et al. De novo mutations in the autophagy gene WDR45 cause static encephalopathy of childhood with neurodegeneration in adulthood. *Nature Genet.* 2013;45:445.
35. Hayflick SJ. Defective pantothenate metabolism and neurodegeneration. *Biochem Soc Trans.* 2014;42:1063–8.
36. Kruer MC, Hiken M, Gregory A, et al. Novel histopathologic findings in molecularly-confirmed pantothenate kinase-associated neurodegeneration. *Brain.* 2011;134:947–58.
37. Dusi S, Valletta L, Haack TB, et al. Exome sequence reveals mutations in CoA synthase as a cause of neurodegeneration with brain iron accumulation. *Am J Hum Genet.* 2014;94:11–22.
38. Hinarejos I, Machuca-Arellano C, Sancho P, Espinós C. Mitochondrial dysfunction, oxidative stress and neuroinflammation in neurodegeneration with brain iron accumulation (NBIA). *Antioxidants.* 2020;9:1020.
39. Annesi G, Gagliardi M, Iannello G, Quattrone A, Iannello G, Quattrone A. Mutational analysis of COASY in an Italian patient with NBIA. *Parkinsonism Relat Disord.* 2016;28:150–1.
40. Martinez DL, Tsuchiya Y, Gout I. Coenzyme A biosynthetic machinery in mammalian cells. *Biochem Soc Trans.* 2014;42:1112–7.
41. Mariño G, Pietrocola F, Eisenberg T, et al. Regulation of autophagy by cytosolic acetyl-coenzyme A. *Mol Cell.* 2014;53:710–25.
42. Kruer MC, Paisán-Ruiz C, Boddaert N, et al. Defective FA2H leads to a novel form of neurodegeneration with brain iron accumulation (NBIA). *Ann Neurol.* 2010;68:611–8.
43. Rattay TW, Lindig T, Baets J, et al. FAHN/SPG35: a narrow phenotypic spectrum across disease classifications. *Brain.* 2019;142:1561–72.
44. Mari F, Berti B, Romano A, et al. Clinical and neuroimaging features of autosomal recessive spastic paraplegia 35 (SPG35): case reports, new mutations, and brief literature review. *Neurogenetics.* 2018;19:123–30.
45. Gregory A, Venkateswaran S, Hayflick SJ. Fatty acid hydroxylase-associated neurodegeneration. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. *GeneReviews*®. Seattle (WA): University of Washington; 1993.
46. Alderson NL, Rembiesa BM, Walla MD, Bielawska A, Bielawski J, Hama H. The human FA2H gene encodes a fatty acid 2-hydroxylase. *J Biol Chem.* 2004;279:48562–8.
47. Zöllner I, Meixner M, Hartmann D, et al. Absence of 2-hydroxylated sphingolipids is compatible with normal neural development but causes late-onset axon and myelin sheath degeneration. *J Neurosci.* 2008;28:9741–54.
48. Guo L, Zhou D, Pryse KM, Okunade AL, Su X. Fatty acid 2-hydroxylase mediates diffusional mobility of Raft-associated lipids, GLUT4 level, and lipogenesis in 3T3-L1 adipocytes. *J Biol Chem.* 2010;285:25438–47.
49. Vroegindeweyj LH, Boon AJ, Wilson JP, Langendonk JG. Effects of iron chelation therapy on the clinical course of aceruloplasminemia: an analysis of aggregated case reports. *Orphanet J Rare Dis.* 2020;15:1–8.
50. Ogimoto M, Anzai K, Takenoshita H, et al. Criteria for early identification of aceruloplasminemia. *Intern Med.* 2011;50:1415–8.
51. Vroegindeweyj L, Van Der Beek E, Boon A, et al. Aceruloplasminemia presents as Type 1 diabetes in non-obese adults: a detailed case series. *Diabetic Med.* 2015;32:993–1000.
52. Levi S, Finazzi D. Neurodegeneration with brain iron accumulation: update on pathogenic mechanisms. *Front Pharmacol.* 2014;5:99.
53. McNeill A, Pandolfo M, Kuhn J, Shang H, Miyajima H. The neurological presentation of ceruloplasmin gene mutations. *Eur Neurol.* 2008;60:200–5.
54. Brissot P, Ropert M, Le Lan C, Loréal O. Non-transferrin bound iron: a key role in iron overload and iron toxicity. *Biochim Biophys Acta Gen Subj.* 2012;1820:403–10.

55. Kono S, Yoshida K, Tomosugi N, et al. Biological effects of mutant ceruloplasmin on hepcidin-mediated internalization of ferroportin. *Biochim Biophys Acta*. 2010;1802:968–75.
56. Miyajima H, Takahashi Y, Kono S. Aceruloplasminemia, an inherited disorder of iron metabolism. *Biometals*. 2003;16:205–13.
57. Chinnery PF, Crompton DE, Birchall D, et al. Clinical features and natural history of neuroferritinopathy caused by the FTL1 460InsA mutation. *Brain*. 2007;130:110–9.
58. Vidal R, Ghetti B, Takao M, et al. Intracellular ferritin accumulation in neural and extraneural tissue characterizes a neurodegenerative disease associated with a mutation in the ferritin light polypeptide gene. *J Neuropathol Exp Neurol*. 2004;63:363–80.
59. Kumar N, Rizek P, Jog M. Neuroferritinopathy: pathophysiology, presentation, differential diagnoses and management. *Tremor Other Hyperkin Mov*. 2016;6:355.
60. Cozzi A, Rovelli E, Frizzale G, et al. Oxidative stress and cell death in cells expressing L-ferritin variants causing neuroferritinopathy. *Neurobiol Dis*. 2010;37:77–85.
61. Najim al-Din AS, Wriekat A, Mubaidin A, Dasouki M, Hiari M. Pallido-pyramidal degeneration, supranuclear upgaze paresis and dementia: Kufor-Rakeb syndrome. *Acta Neurolog Scand*. 1994;89:347–52.
62. Schneider SA, Paisan-Ruiz C, Quinn NP, et al. ATP13A2 mutations (PARK9) cause neurodegeneration with brain iron accumulation. *Mov Disord*. 2010;25:979–84.
63. Kruer MC, Paudel R, Wagoner W, et al. Analysis of ATP13A2 in large neurodegeneration with brain iron accumulation (NBIA) and dystonia-parkinsonism cohorts. *Neurosci Lett*. 2012;523:35–8.
64. Ramonet D, Podhajska A, Stafa K, et al. PARK9-associated ATP13A2 localizes to intracellular acidic vesicles and regulates cation homeostasis and neuronal integrity. *Hum Mol Genet*. 2012;21:1725–43.
65. Estrada-Cuzcano A, Martin S, Chamova T, et al. Loss-of-function mutations in the ATP13A2/PARK9 gene cause complicated hereditary spastic paraplegia (SPG78). *Brain*. 2017;140:287–305.
66. Park JS, Mehta P, Cooper AA, et al. Pathogenic effects of novel mutations in the P-type ATPase ATP13A2 (PARK9) causing Kufor-Rakeb syndrome, a form of early-onset parkinsonism. *Hum Mutat*. 2011;32:956–64.
67. Ramirez A, Heimbach A, Grundemann J, et al. Hereditary parkinsonism with dementia is caused by mutations in ATP13A2, encoding a lysosomal type 5 P-type ATPase. *Nat Genet*. 2006;38:1184–91.
68. Dehay B, Ramirez A, Martinez-Vicente M, et al. Loss of P-type ATPase ATP13A2/PARK9 function induces general lysosomal deficiency and leads to Parkinson disease neurodegeneration. *Proc Natl Acad Sci USA*. 2012;109:9611–6.
69. Usenovic M, Tresse E, Mazzulli JR, Taylor JP, Krainc D. Deficiency of ATP13A2 leads to lysosomal dysfunction, alpha-synuclein accumulation, and neurotoxicity. *J Neurosci*. 2012;32:4240–6.
70. Grunewald A, Arns B, Seibler P, et al. ATP13A2 mutations impair mitochondrial function in fibroblasts from patients with Kufor-Rakeb syndrome. *Neurobiol Aging*. 2012;33:1843.e1–7.
71. Alazami AM, Al-Saif A, Al-Semari A, et al. Mutations in C2orf37, encoding a nucleolar protein, cause hypogonadism, alopecia, diabetes mellitus, mental retardation, and extrapyramidal syndrome. *Am J Hum Genet*. 2008;83:684–91.
72. Jin J, Arias EE, Chen J, Harper JW, Walter JC. A family of diverse Cul4-Ddb1-interacting proteins includes Cdt2, which is required for S phase destruction of the replication factor Cdt1. *Mol Cell*. 2006;23:709–21.
73. Bohlega S, Abusrair AH, Al-Ajlan FS, et al. Patterns of neurological manifestations in Woodhouse-Sakati Syndrome. *Parkinsonism Relat Disord*. 2019;69:99–103.
74. Jaber E, Rohani M, Shahidi GA, et al. Identification of mutation in GTPBP2 in patients of a family with neurodegeneration accompanied by iron deposition in the brain. *Neurobiol Aging*. 2016;38:216.
75. Woo K-C, Kim T-D, Lee K-H, et al. Modulation of exosome-mediated mRNA turnover by interaction of GTP-binding protein 1 (GTPBP1) with its target mRNAs. *FASEB J*. 2011;25:2757–69.
76. Bourne HR, Sanders DA, McCormick F. The GTPase superfamily: a conserved switch for diverse cell functions. *Nature*. 1990;348:125.
77. Kaziro Y, Itoh H, Kozasa T, Nakafuku M, Satoh T. Structure and function of signal-transducing GTP-binding proteins. *Annu Rev Biochem*. 1991;60:349–400.
78. Ferdinandusse S, Kostopoulos P, Denis S, et al. Mutations in the gene encoding peroxisomal sterol carrier protein X (SCPx) cause leukoencephalopathy with dystonia and motor neuropathy. *Am J Hum Genet*. 2006;78:1046–52.
79. Horvath R, Lewis-Smith D, Douroudis K, et al. SCP2 mutations and neurodegeneration with brain iron accumulation. *Neurology*. 2015;85:1909–11.
80. Seedorf U, Raabe M, Ellinghaus P, et al. Defective peroxisomal catabolism of branched fatty acyl coenzyme A in mice lacking the sterol carrier protein-2/sterol carrier protein-x gene function. *Genes Dev*. 1998;12:1189–201.
81. Wanders R, Vreken P, Ferdinandusse S, et al. Peroxisomal fatty acid α - and β -oxidation in humans: enzymology, peroxisomal metabolite transporters and peroxisomal diseases. *Biochem Soc Trans*. 2001;29:250–67.
82. Drecourt A, Babbord J, Dussiot M, et al. Impaired transferrin receptor palmitoylation and recycling in neurodegeneration with brain iron accumulation. *Am J Hum Genet*. 2018;102:266–77.
83. Levi S, Tiranti V. Neurodegeneration with brain iron accumulation disorders: valuable models aimed at understanding the pathogenesis of iron deposition. *Pharmaceuticals*. 2019;12:27.
84. Tüysüz B, Bilguvar K, Koçer N, et al. Autosomal recessive spastic tetraplegia caused by AP4M1 and AP4B1 gene mutation: expansion of the facial and neuroimaging features. *Am J Med Genet Part A*. 2014;164:1677–85.
85. Roubertie A, Hieu N, Roux C-J, et al. AP4 deficiency: a novel form of neurodegeneration with brain iron accumulation? *Neurol Genet*. 2018;4:e217.
86. Najmabadi H, Hu H, Garshasbi M, et al. Deep sequencing reveals 50 novel genes for recessive cognitive disorders. *Nature*. 2011;478:57–63.
87. Correa-Vela M, Lupo V, Montpeyó M, et al. Impaired proteasome activity and neurodegeneration with brain iron accumulation in FBXO7 defect. *Ann Clin Transl Neurol*. 2020;7:1436–42.
88. Shojae S, Sina F, Banihosseini SS, et al. Genome-wide linkage analysis of a Parkinsonian-pyramidal syndrome pedigree by 500 K SNP arrays. *Am J Hum Genet*. 2008;82:1375–84.
89. Cheon Y, Kim HW, Igarashi M, et al. Disturbed brain phospholipid and docosahexaenoic acid metabolism in calcium-independent phospholipase A(2)-VIA (iPLA(2)beta)-knockout mice. *Biochim Biophys Acta*. 2012;1821:1278–86.
90. Iankova V, Karin I, Klopstock T, Schneider SA. Emerging disease-modifying therapies in neurodegeneration with brain iron accumulation (NBIA) disorders. *Front Neurol*. 2021;12:629414.
91. Haeri G, Haji Akhond F, Alavi A, Abdi S, Rohani M. Endocrine abnormalities in a case of neurodegeneration with brain iron accumulation. *Mov Disord Clin Pract*. 2020;7:706–7.
92. Hashemi H, Rokni YH, Adibi A. Atypical pantothenate-kinase associated neurodegeneration (PKAN) in two Iranian patients. *Iran J Radiol*. 2008;5:e78978.
93. Dezfouli MA, Jaber E, Alavi A, et al. Pantothenate kinase 2 mutation with eye-of-the-tiger sign on magnetic resonance imaging in three siblings. *Iran J Neurol*. 2012;11:155.
94. Aryani O, Houshmand M, Fatehi F. A novel PANK2 gene mutation in a Persian boy: the first report from Iran. *Clin Neurol Neurosurg*. 2013;115:1170.
95. Dezfouli MA, Alavi A, Rohani M, et al. PANK2 and C19orf12 mutations are common causes of neurodegeneration with brain iron accumulation. *Mov Disord*. 2013;28:228–31.
96. Malakouti-Nejad M, Shahidi G-A, Rohani M, et al. Identification of p. Gln858* in ATP13A2 in two EOPD patients and presentation of their clinical features. *Neurosci Lett*. 2014;577:106–11.
97. Ghafouri-Fard S, Yassaee VR, Rezayi A, Hashemi-Gorji F, Alipour N, Miryounesi M. A novel nonsense mutation in PANK2 gene in two patients with pantothenate kinase-associated neurodegeneration. *Int J Mol Cell Med*. 2016;5:255.

98. Dastsooz H, Nemati H, Fard MAF, Fardaei M, Faghihi MA. Novel mutations in PANK2 and PLA2G6 genes in patients with neurodegenerative disorders: two case reports. *BMC Med Genet.* 2017;18:87.
99. Fasano A, Shahidi G, Lang AE, Rohani M. Basal ganglia calcification in a case of PKAN. *Parkinsonism Relat Disord.* 2017;36:98–9.
100. Rohani M, Shahidi G, Alavi A, et al. Tremor-dominant pantothenate kinase-associated neurodegeneration. *Mov Disord Clin Pract.* 2017;4:772–4.
101. Rohani M, Fasano A, Lang AE, et al. Pantothenate kinase-associated neurodegeneration mimicking Tourette syndrome: a case report and review of the literature. *Neurol Sci.* 2018;39:1797–800.
102. Alavi A, Mokhtari M, Hajati R, et al. Late-onset mitochondrial membrane protein-associated neurodegeneration with extensive brain iron deposition. *Mov Disord Clin Pract.* 2019;7:120.
103. Habibi AH, Razmeh S, Aryani O, et al. A novel homozygous variation in the PANK2 gene in two Persian siblings with atypical pantothenate kinase associated neurodegeneration. *Neurol Int.* 2019;11:7959.
104. Rohani M, Fasano A, Akhoundi FH, et al. Beta-propeller protein associated neurodegeneration (BPAN); the first report of three patients from Iran with de novo novel mutations. *Parkinsonism Relat Disord.* 2019;61:231–3.
105. Rohani M, Shahidi G, Vali F, et al. Oculogyric crises in PLA2G6 associated neurodegeneration. *Parkinsonism Relat Disord.* 2018;52:111–2.
106. Saeedi Y, Kazemi F, Habibi SAH, et al. Tongue protrusion dystonia in pantothenate kinase-associated neurodegeneration. *Pediatr Neurol.* 2020;103:76–8.
107. Sina F, Shojaee S, Elahi E, Paisan-Ruiz C. R632W mutation in PLA2G6 segregates with dystonia-parkinsonism in a consanguineous Iranian family. *Eur J Neurol.* 2009;16:101–4.
108. Rohani M, Lang AE, Sina F, et al. Action myoclonus and seizure in Kufor-Rakeb syndrome. *Mov Disord Clin Pract.* 2018;5:195–9.
109. Eslami M, Razmeh S, Abdollahi M, Ghurchian F, Maher M. A 7-year-old girl with progressive gait disturbance. *MOJ Biol Med.* 2017;1:130–1.
110. Bertoli-Avella AM, Garcia-Aznar JM, Brandau O, et al. Biallelic inactivating variants in the GTPBP2 gene cause a neurodevelopmental disorder with severe intellectual disability. *Eur J Hum Genet.* 2018;26:592–8.