

## Bioavailability of vitamin B<sub>12</sub> in cows' milk

J. Jacques Matte<sup>1\*</sup>, Frédéric Guay<sup>2</sup> and Christiane L. Girard<sup>1</sup>

<sup>1</sup>Dairy and Swine Research and Development Centre, Agriculture and Agri-Food Canada, 2000 College Street, PO Box 90, Lennoxville STN, Sherbrooke, QC, Canada J1M 1Z3

<sup>2</sup>Faculté des sciences de l'agriculture et de l'alimentation, Département de sciences animales, Université Laval, Québec, QC, Canada G1V 0A6

(Received 18 November 2010 – Revised 24 March 2011 – Accepted 28 March 2011 – First published online 27 June 2011)

### Abstract

The natural source of vitamin B<sub>12</sub> in human diets comes from animal products. For example, one glass (250 ml) of milk provides approximately 50% of the RDA (2.4 µg/d). It was hypothesised that the provision of vitamin B<sub>12</sub> from milk is more efficiently absorbed than the synthetic form used in vitamin supplements. Pigs (*n* 10) were used as a model for intestinal absorption of vitamin B<sub>12</sub> in humans to compare the net fluxes of vitamin B<sub>12</sub> across the portal-drained viscera (PDV; an indicator of intestinal absorption) after ingestion of meals complemented with conventional and vitamin B<sub>12</sub>-enriched (via injections to cows) milk (raw, pasteurised or microfiltered) or with equivalent amounts of cyanocobalamin, the synthetic form used in supplements or unsupplemented. Net flux of vitamin B<sub>12</sub> across PDV after the ingestion of milk was positive, though not influenced by milk enrichment ( $P > 0.3$ ) or technological processes ( $P = 0.8$ ) and was greater than after ingestion of equivalent amounts of cyanocobalamin (cyanocobalamin *v.* all milk,  $P \leq 0.003$ ). In fact, net fluxes of this vitamin were not different from 0 after either cyanocobalamin or the meal devoid of vitamin B<sub>12</sub> (unsupplemented *v.* cyanocobalamin,  $P = 0.7$ ). The cumulative PDV fluxes during the 24 h following ingestion of meals complemented with milk varied from 5.5 to 6.8 µg. These values correspond to an efficiency of intestinal absorption of vitamin B<sub>12</sub> from milk varying between 8 and 10%. Therefore, vitamin B<sub>12</sub>, which is abundant in cows' milk, is also substantially more available than the most commonly used synthetic form of this vitamin.

**Key words:** Vitamin B<sub>12</sub>: Bioavailability: Cows' milk: Pig model

Among B vitamins, vitamin B<sub>12</sub> occupies a very special niche. This vitamin is produced only by bacteria and archaeobacteria if Co supply is adequate. As opposed to other B vitamins, it is neither synthesised nor used by fungi and plants<sup>(1)</sup>. Therefore, in human diets, the sole natural source of vitamin B<sub>12</sub> comes from animal products. Among animal products, those from ruminants are particularly rich in vitamin B<sub>12</sub>, the vitamin being naturally synthesised by ruminal microflora using Co as an essential precursor and then absorbed and stored in the liver and muscles (meat) of the host or secreted in its milk<sup>(2)</sup>.

In humans, vitamin B<sub>12</sub> deficiency affects cell division and may lead to megaloblastic anaemia and neuropathy<sup>(2)</sup>. In the presence of vitamin B<sub>12</sub> deficiency, increasing folic acid supply cures anaemia but not neurological symptoms, then, by masking haematological symptoms, it could delay the diagnosis of vitamin B<sub>12</sub> deficiency until neurological damages are irreversible. Consequently, over the last decade, since folic acid fortification of flour became mandatory in many Western countries, including Canada and USA, there has been a

renewed interest in the evaluation of vitamin B<sub>12</sub> status in human populations according to folic acid provision<sup>(3,4)</sup>.

Vitamin B<sub>12</sub> status is correlated with vitamin B<sub>12</sub> intake in humans<sup>(5–7)</sup>. Vegetarians had lower vitamin B<sub>12</sub> status than omnivores<sup>(7,8)</sup>. However, dietary sources of the vitamin also seem to matter. For example, vitamin status of vegetarians was positively correlated with their intake of dairy products, especially milk, but not of eggs or seafood<sup>(8)</sup>. Among adults not using vitamin supplements, the relationship between plasma concentration of the vitamin and its intake from dairy products is similar to the relationship observed with the intake from cereals fortified with vitamin B<sub>12</sub>. However, the relationship with intake from meat, poultry or fish is weaker<sup>(5)</sup>. A Norwegian study showed that plasma vitamin B<sub>12</sub> increases with the amounts of vitamin B<sub>12</sub> provided by dairy products or fish but not with those provided by eggs or meat<sup>(6)</sup>. Moreover, for a similar intake, plasma concentrations of vitamin B<sub>12</sub> were higher when the vitamin was supplied by dairy products than by fish, suggesting that the bioavailability of the vitamin from dairy products is higher

**Abbreviations:** AAFC, Agriculture and Agri-Food Canada; PDV, portal-drained viscera.

\* **Corresponding author:** J. J. Matte, email Jacques.Matte@agr.gc.ca

than from the other sources<sup>(6)</sup>. Indeed, these retrospective studies seem to indicate that vitamin B<sub>12</sub> supplied by dairy products is more available than from other natural sources, although in these studies, intake data were obtained by a FFQ.

In terms of the provision of vitamin B<sub>12</sub>, one glass (250 ml) of milk provides more than 1 µg of vitamin B<sub>12</sub><sup>(9)</sup>. According to the Canadian Food Inspection Agency<sup>(10)</sup>, cows' milk could claim to be an 'excellent source' of vitamin B<sub>12</sub>, because one glass of milk (250 ml) provides nearly 50% of the RDA<sup>(11)</sup> for adults and children over 13 years of age (2.4 µg/d). Moreover, vitamin B<sub>12</sub> supplements given to dairy cows via weekly injections increased vitamin B<sub>12</sub> content in milk by 50%<sup>(12)</sup>; in such a case, a glass of milk may provide up to 75% of the RDA.

Cyanocobalamin is the synthetic form of vitamin B<sub>12</sub> present in most supplements, the cyanide group being used to stabilise the molecule. However, cyanocobalamin is not biologically active until the cyanide group is enzymatically removed<sup>(13)</sup>. Bioavailability of the synthetic form of vitamin B<sub>12</sub> is reported to be poor (<4%) in humans and animals<sup>(14)</sup>. In milk, vitamin B<sub>12</sub> is present as adenosylcobalamin, hydroxocobalamin and methylcobalamin<sup>(15,16)</sup>. Hydroxocobalamin is the product of photolysis of light-sensitive cobalamins, whereas adenosylcobalamin and methylcobalamin have a coenzymatic activity in mammal cells and are biologically active<sup>(15)</sup>.

It was therefore hypothesised that the important daily provision of vitamin B<sub>12</sub> brought by unprocessed (raw) or processed milk (pasteurised or microfiltrated) is more efficiently absorbed than the synthetic form, cyanocobalamin, used in vitamin supplements. The present study aimed to compare in pigs, used as an animal model for humans, the net flux of vitamin B<sub>12</sub> across portal-drained viscera (PDV) after ingestion of conventional and vitamin B<sub>12</sub>-enriched (via injections to cows) milk (raw, pasteurised or microfiltrated) to the equivalent amount of cyanocobalamin or to an unsupplemented control diet.

## Experimental methods

### Milk collection and preparation

We collected two types of milk differing in their vitamin B<sub>12</sub> content from the dairy herd of Agriculture and Agri-Food Canada (AAFC) at Sherbrooke. Conventional milk (C; 9801) was obtained from cows not supplemented with vitamin B<sub>12</sub>; it contained 12.0, 3.2 and 3.5% of total solids, crude protein and fat, respectively. Vitamin B<sub>12</sub>-enriched milk (E; 8201) was obtained at the same time from a group of eighteen cows that had received weekly intramuscular injections of 2 ml of a solution of cyanocobalamin (5 mg/ml) during three consecutive weeks as previously employed by Girard & Matte<sup>(17)</sup>. Vitamin B<sub>12</sub>-enriched milk composition was 11.7, 3.2 and 3.3% of total solids, crude protein and fat, respectively. The milk was transported for processing at the AAFC Food Research and Development Centre of St Hyacinthe, QC, Canada. Upon arrival, skimmed milk and cream were separated, and the latter was kept apart. Skimmed milk was separated in three fractions. The first third was not treated (raw skimmed milk; R). The second third was microfiltrated (microfiltrated skimmed milk; F) using the 'Bactocatch'

procedure (cold pasteurisation) as described by Trouvé *et al.*<sup>(18)</sup> in a cross-flow microfiltration unit (Pilot Plant type MFS-7, Alfa-Laval MFSI, Tumba, Sweden) with a ceramic membrane pore size of 1.4 µm, membrane area of 1.4 m<sup>2</sup>, flow rate of 850 l/h per m<sup>2</sup> and temperature of 35°C. The last third of the skimmed milk (pasteurised skimmed milk; P) and the whole cream were pasteurised (73°C for 16 s) using a plate heat exchanger Tetra Plex, C3/-SR type (Alfa-Laval, Tumba, Sweden). All skimmed milk was concentrated separately using a reverse osmosis system with a TFC<sup>®</sup> 3838 RO-N1 spiral membrane of 965 × 96 × 21.1 mm (area of 18.4 m<sup>2</sup>; Koch Membrane Systems, Inc., Wilmington, MA, USA) for final DM content of 25.9, 29.4 and 28.1% for raw, pasteurised and microfiltrated conventional skimmed milk and 27.9, 28.0 and 30.4% for raw, pasteurised and microfiltrated vitamin B<sub>12</sub>-enriched skimmed milk, respectively. Approximately 6 litres of each skimmed milk concentrate were freeze-dried and stored frozen at -20°C. The pasteurised cream and the remaining of the different concentrates of skimmed milk were stored frozen at -20°C in aliquots of 1 and 2 litres, respectively.

The vitamin B<sub>12</sub> content of skimmed milk concentrates was determined in duplicate from two hydrolyses of each milk concentrate using a radioassay (Quantaphase B<sub>12</sub>, Bio-Rad Laboratories (Canada) Limited, Mississauga, ON, Canada) as described by Preynat *et al.*<sup>(19)</sup>.

### Animals and surgeries

After weaning at 28 d of age, ten female Yorkshire-Landrace × Duroc piglets were fed *ad libitum* a conventional basal diet

**Table 1.** Ingredients and chemical composition of the basal post-weaning diet

	%
Ingredients	
Maize	65.8
Soyabean meal, 48% crude protein	29.0
Animal fat	1.8
Limestone	1.5
Biophos*	0.8
Salt	0.6
L-Lys	0.17
Met	0.06
CuSO <sub>4</sub>	0.04
Phytase (Natuphos <sup>®</sup> )	0.04
Choline chloride	0.02
Mineral and vitamin premix†	0.2
Nutrient composition (analytical values)	
DM (%)	88.7
Metabolisable energy (MJ/kg)	14.0
Crude protein (%)	18.7
Fat (%)	4.9
Crude fibre (%)	2.6
Ca (%)	0.89
P (%)	0.50
Ash (%)	5.7

\* Biophos (monocalcium phosphate) is an inorganic P source providing 21% available P, 18% Ca and 0.21% F.

† Supplied, per kg of feed: Mn, 52 mg; Zn, 174 mg; Fe, 275 mg; Cu, 130 mg; I, 1 mg; Se, 300 µg; vitamin A, 3008 IU; vitamin D<sub>3</sub>, 900 IU; vitamin E, 41 IU; vitamin K, 1.5 mg; thiamine, 2.0 mg; riboflavin, 3.5 mg; niacin, 20 mg; pantothenic acid, 15 mg; folic acid, 0.5 mg; pyridoxine, 2.0 mg; biotin, 50 µg; choline, 104 mg and vitamin B<sub>12</sub>, 20 µg.

for post-weaned piglets (Table 1). This basal diet was supplemented with cyanocobalamin as recommended by the National Research Council<sup>(20)</sup> for this category of pigs (20 µg/kg). Average body weight at surgery was 48.7 (SEM 1.0) kg. The surgery procedure has been described by Hooda *et al.*<sup>(21)</sup>. Briefly, the animals were equipped with an ultrasonic flow probe (14 mm flow probe, SB-series: Transonic Systems, Inc., Ithaca, NY, USA) around the portal vein and a catheter inside the portal vein at 3.5 and 2.5 cm before its entry into the liver, respectively. Another catheter was inserted through the carotid artery up to the junction between the carotid and subclavian arteries.

The experimental procedures followed the guidelines of the Canadian Council on Animal Care<sup>(22)</sup> and were approved by the Institutional Animal Care Committee of the Dairy and Swine Research and Development Centre of Sherbrooke (Québec, Canada). All animals were cared for according to the recommended code of practice of Agriculture Canada<sup>(23)</sup>.

### Treatments

After surgery, the animals were penned individually (1 m × 1.8 m) and fed a single daily meal of 1.2 kg of the diet described in Table 1. At 10–15 d after surgery, when the animals had recovered full appetite and a normal growth rate, they were gradually (3–5 d) adapted to the metabolic cage (with free access to water) and to the consumption of a mixture of milk concentrate and cereals similar to milk treatments described below. The treatments were tested according to a split-plot design in which the concentration of vitamin B<sub>12</sub> was the first plot. There were five pigs in each plot. Within each plot, each animal received the five experimental treatments according to a cross-over design. The three treatments corresponded to mixtures of concentrated milk (raw, microfiltrated and pasteurised), cream and cereals (67% maize, 16.5% wheat and 16.5% soyabean meal) as described in Table 2. This was done to mimic a human breakfast. In the two remaining treatments, cereals were mixed to provide 1.2 kg of DM and 1.3 litres of water was added to obtain the same volume as the milk treatments; these meals were supplemented with (B<sub>12</sub>-S) or not supplemented with

(B<sub>12</sub>-0) cyanocobalamin (V-2876, Sigma-Aldrich, St Louis, MO, USA), the synthetic form of the vitamin. Cyanocobalamin, approximately 50 µg (84 µl of a solution at 600 µg/ml) or 80 µg (135 µl of a solution at 600 µg/ml) corresponding to the average provision from C or E milk, respectively, was incorporated in a 25 g pellet made of the basal diet mixed with unsweetened apple sauce to induce a rapid consumption just before the meal. It was assumed that the sole source of vitamin B<sub>12</sub> in the mixtures of milk and cereals came from milk because feedstuffs from plants are devoid of that vitamin<sup>(1)</sup>. The analysed values for the cyanocobalamin solution were lower (524.2 (SEM 5.9) µg/ml, *n* 7) than the calculated values. Thus, vitamin B<sub>12</sub> intake for C–S and E–S treatments was 44 and 71 µg, respectively.

On experimental days, the animals were placed in metabolic cages and fed one of the experimental meals. Blood samples (4 ml) were collected simultaneously from the two catheters every 45 min for the first 3 h post-feeding, and every hour for the following 21 h. Portal blood flow was recorded continuously during 24 h using the WinDaq<sup>®</sup> software (Dataq Instruments, Inc., Akron, OH, USA). Between experimental days, the animals were moved back to their pen for 3–4 d and fed the basal diet described in Table 1.

### Blood collection and analysis

Immediately after sampling, the blood was transferred from syringes into EDTA-treated tubes (Vacutainer<sup>®</sup>, Becton Dickinson, Franklin Lakes, NJ, USA). Packed cell volume was measured in duplicate on fresh blood by microcentrifugation. An aliquot of blood was immediately frozen for Hb determination according to the method of Drabkin<sup>(24)</sup>. Plasma was collected after centrifugation at 1800 g for 10 min at 4°C and frozen at –20°C for further analysis. Plasma concentrations of vitamin B<sub>12</sub> were measured in duplicate by radioassay in two different assays (SimulTRAC-S Radioassay kit, Vitamin B<sub>12</sub> (<sup>57</sup>Co)/Folate (<sup>125</sup>I), MP Biomedicals, Diagnostics Division, Orangeburg, NY, USA). Validation tests for these measurements showed satisfactory parallelism (CV = 5.0%), and recovery tests (92.8%, CV = 4.0%) between 50 and 800 pg/ml. The intra- and inter-assay CV were 4.1 (*n* 13) and 5.6% (*n* 23), respectively.

**Table 2.** Composition of the milk-based experimental meals and their provision of endogenous vitamin B<sub>12</sub>

	C			E		
	C–R	C–P	C–F	E–R	E–P	E–F
Cream (g DM)*	150	151	151	148	161	174
Cereal mix (g)†	362	309	328	334	319	270
Skimmed milk concentrate (g DM)‡	688	740	721	718	720	756
Vitamin B <sub>12</sub> (ng/g DM skimmed milk concentrate)	69.4	76.8	68.6	99.1	120.3	113.9
B <sub>12</sub> intake (µg)	47.7	56.9	49.5	71.1	86.7	86.1

C, conventional milk; E, enriched milk; R, raw milk; P, pasteurised milk; M, microfiltrated milk.

\* Amount of cream added to obtain an equivalent of 2% fat milk.

† The mixture of skimmed milk and cream was complemented with cereals (67% maize, 16.5% wheat and 16.5% soyabean meal) to provide a meal of 1.2 kg DM. The nutrient composition (analytical values) of the cereal mix was as follows: DM, 88.2; metabolisable energy, 14.1 MJ/kg; crude protein, 15.7%; fat, 3.5%; crude fibre, 4.1%; Ca, 0.15%; phosphorus, 0.54%; ash, 3.6%.

‡ Whole DM intake from skimmed milk concentrate in experimental meals (1.5 litres of liquid + 300 g of freeze-dried).

**Table 3.** Average portoarterial difference and portal-drained viscera (PDV) flux of vitamin B<sub>12</sub> during the 24 h post-meal according to the treatments (Least square means with their standard errors)

	Unsupplemented	Cyanocobalamin	Raw milk	Pasteurised milk	Microfiltrated milk	SEM	P values
<i>n</i>	9	9	9	9	9		
Portoarterial difference (pg/ml)*	-1.9	-2.7	5.3	3.8	5.8	1.9	0.004
PDV flux of vitamin B <sub>12</sub> (ng/min)†	-1.8‡	-2.9‡	3.8	3.9	4.7	1.9	0.02

\* Values for unsupplemented did not differ from those of cyanocobalamin ( $P=0.76$ ), cyanocobalamin differed from those of all milks (raw, pasteurised and microfiltrated;  $P=0.001$ ), raw milk did not differ from those of treated (pasteurised + microfiltrated) milks ( $P=0.83$ ) and pasteurised milk did not differ from those of microfiltrated milk ( $P=0.45$ ).

† Values for unsupplemented did not differ from those of cyanocobalamin ( $P=0.68$ ), cyanocobalamin differed from those of all milks (raw, pasteurised and microfiltrated) ( $P=0.003$ ), raw milk did not differ from those of treated (pasteurised + microfiltrated) milks ( $P=0.83$ ) and pasteurised milk did not differ from those of microfiltrated milk ( $P=0.76$ ).

‡ These values are not different from 0 ( $P>0.13$ ).

DM, crude protein, fat, crude fibre, Ca, P and ash in the conventional basal diet and in the cereal mix were analysed following AOAC<sup>(25)</sup> methods (930.15, 990.03, 920.39, 962.09, 985.01, 985.01 and 942.05, respectively).

### Calculations and statistical analysis

In the C group, one pig had to be removed from the study. Signals from the flow probes were lost for the last period for one pig (treatment C-R) and for the three last periods for another pig (treatments E-P, E-F and E-S). For these two pigs, the calculations of PDV fluxes for these periods were done using the average plasma flow at each sampling time recorded during the previous periods. The estimated values for these periods were not included in statistical analysis of blood and plasma flows.

Statistical analyses of blood and plasma flows, arterial concentrations, venoarterial difference and PDV flux of vitamin B<sub>12</sub> were conducted on postprandial values that were averaged across the 24 h post meal. The net flux of vitamin B<sub>12</sub> across PDV was calculated as described by Girard *et al.*<sup>(26)</sup> for each sampling time. As time intervals between the blood samples were unequal, the average net flux of vitamin B<sub>12</sub> across PDV during the 24 h post meal was calculated as the summation of PDV net flux at each time point multiplied by the interval of time between the two consecutive samples divided by 1440 min. A positive net flux indicates a release of a nutrient from PDV, whereas a negative flux indicates an uptake.

All variables were analysed using the MIXED procedure of SAS (SAS Institute Inc., Cary, NC, USA)<sup>(27)</sup> according to a split-plot design with vitamin B<sub>12</sub> concentration and period as the main plots, treatment and the vitamin B<sub>12</sub> concentration × treatment interaction as the subplots and the vitamin B<sub>12</sub> concentration × period and treatment × period within vitamin B<sub>12</sub> concentration interactions as the random effects. When the treatment effects reached a level of significance of 95%, the following *a priori* comparisons were used: (1) unsupplemented *v.* cyanocobalamin; (2) cyanocobalamin *v.* all milk (raw, pasteurised and microfiltrated); (3) raw *v.* treated (pasteurised + microfiltrated) milk; (4) pasteurised milk *v.* microfiltrated milk. The results are reported as least square means and standard errors of the means. The significance level was defined at  $P\leq 0.05$ , and trends towards significance were considered at  $0.05 < P < 0.10$ .

### Results

The pigs in the E group had lower ( $P=0.04$ ) blood and plasma flows than those in the C group (1.08 (SEM 0.05) *v.* 1.31 (SEM 0.06) and 0.75 (SEM 0.04) *v.* 0.91 (SEM 0.04) litres/min, respectively). There was no treatment or vitamin B<sub>12</sub> concentration × treatment interaction effect ( $P\geq 0.7$ ) on these two variables.

There was no effect ( $P\geq 0.3$ ) of vitamin B<sub>12</sub> concentration, treatment or their interaction on arterial plasma concentration of vitamin B<sub>12</sub> (186 (SEM 9) pg/ml).

Average portoarterial difference and PDV flux of vitamin B<sub>12</sub> during the 24 h post meal were not affected by vitamin B<sub>12</sub> concentration ( $P\geq 0.5$ ) but differed among treatments ( $P\leq 0.02$ ; Table 3). Average portoarterial difference in vitamin B<sub>12</sub> during the 24 h post meal was higher ( $P=0.001$ ) when the pigs were fed milk (raw + pasteurised + microfiltrated) than those fed cyanocobalamin. However, there was no difference ( $P\geq 0.5$ ) between raw and treated (pasteurised + microfiltrated) milk or between pasteurised and microfiltrated milk. The portoarterial difference of vitamin B<sub>12</sub> was not different ( $P=0.8$ ) when the pigs were fed the unsupplemented diet or cyanocobalamin supplements.

Similarly, PDV fluxes of vitamin B<sub>12</sub> during the 24 h post meal were not different ( $P=0.68$ ) when the pigs were fed the unsupplemented diet or cyanocobalamin supplements, and these fluxes were not different from 0 ( $P\geq 0.13$ ; Table 3). PDV flux of the vitamin was higher ( $P=0.003$ ) when the pigs were fed milk than those fed cyanocobalamin. PDV fluxes of vitamin B<sub>12</sub> were different from 0 ( $P\leq 0.05$ ) when the pigs were fed milk, but it did not differ ( $P=0.8$ ) between raw and treated milk or between milk treatments (Table 3). The cumulative PDV fluxes for 24 h varied from 5.5 to 6.8 μg. These values correspond to an efficiency of intestinal absorption of vitamin B<sub>12</sub> from milk varying between 8 and 10%.

### Discussion

In the present study, pigs were used as a model to estimate the bioavailability of vitamin B<sub>12</sub> in humans. In a recent review, Guilloteau *et al.*<sup>(28)</sup> highlighted the numerous similarities (anatomy, physiology, digestion, absorption and metabolism) between the gastrointestinal tract of humans and pigs. More specifically for vitamin B<sub>12</sub>, the bioavailability of dietary

supplements of cyanocobalamin has recently been studied<sup>(29)</sup> in this species using two different methods. A first one, used as a reference method, measured whole-body deposition of vitamin B<sub>12</sub> after several days of dietary supplementation, whereas the second method measured the net flux of vitamin B<sub>12</sub> across PDV following a single meal supplemented with cyanocobalamin. The second method requires sophisticated surgical preparation, but it has the great advantage of measuring the bioavailability of different concentrations or sources of vitamin B<sub>12</sub> within the same animal. Although estimates of bioavailability varied according to the two methods, both showed an inverse relationship between dietary concentrations of vitamin B<sub>12</sub> and bioavailability of this vitamin as reported previously for human subjects<sup>(30)</sup>. Those results are in accordance with the observations that the mechanisms of absorption of vitamin B<sub>12</sub> in pigs (an omnivorous species) are similar to those described in human subjects<sup>(2,31)</sup> supporting that the pig is an appropriate model to estimate vitamin B<sub>12</sub> availability for human subjects.

In the present study, the mean PDV flux of vitamin B<sub>12</sub> during the 24 h following a meal not supplemented with vitamin B<sub>12</sub> was not different from 0, as observed previously in pigs by Matte *et al.*<sup>(29)</sup>. However, in this last study, the cumulative net PDV fluxes of vitamin B<sub>12</sub> during the 24 h following boluses of 25 and 250 µg of dietary cyanocobalamin were positive at 2.4 and 5.1 µg, respectively, and differed between dietary concentrations. In contrast, in the present study, the actual mean net PDV fluxes of vitamin B<sub>12</sub> after ingestion of meals supplemented with 44 and 71 µg of cyanocobalamin were not different from 0. The major difference between Matte *et al.*<sup>(29)</sup> and the present study was the composition of the experimental meals. In Matte *et al.*<sup>(29)</sup>, a semi-purified diet, containing 16% of vitamin-free casein (derived from cows' milk), 67% maize starch, 11% sucrose and 6% mineral and vitamin supplements, was used, whereas, in the present study, the experimental meal was based on plant-derived feedstuffs (maize, wheat and soyabean meal). It can be hypothesised that a milk component, which survived the technological process for production of vitamin-free casein, improves intestinal absorption of vitamin B<sub>12</sub>. In fact, casein itself could be involved because fractions of this protein were identified as major components of the protein-binding capacity of vitamin B<sub>12</sub> in cows' milk<sup>(32)</sup>. Globally, these explanations are in accordance with the numerically greater, although not statistically significant, efficiency of absorption of dietary <sup>58</sup>Co-labelled cyanocobalamin when given in milk (65%) rather than in water or bread (55%)<sup>(33)</sup>. Nevertheless, it cannot be ruled out that the current lack of vitamin B<sub>12</sub> absorption after ingestion of meals supplemented with cyanocobalamin (C-S and E-S) was related to the fibre content of feedstuffs, which could interfere with the intestinal bioavailability of dietary cyanocobalamin as observed by Cullen & Oace<sup>(34)</sup>.

In milk treatments, the cumulative net PDV flux of vitamin B<sub>12</sub> for 24 h varied between 5.5 and 6.8 µg and was not significantly affected within the range of dietary intakes of conventional (48–57 µg) and enriched milk (71–87 µg). It cannot be ruled out that the present experimental approach based on the

quantification of the net flux of vitamin B<sub>12</sub> through the PDV was not sensitive enough to detect the difference within this range of vitamin B<sub>12</sub> intakes ingested in one single supplemented meal. Such PDV flux values for vitamin B<sub>12</sub> were previously observed following ingestion of 250 µg of cyanocobalamin incorporated in a semi-purified diet containing vitamin-free casein<sup>(29)</sup>. Enhanced PDV flux of vitamin B<sub>12</sub> from milk could also be related to the forms of cobalamins present in milk. In milk, vitamin B<sub>12</sub> is mostly present as adenosylcobalamin and hydroxocobalamin whereas methylcobalamin and cyanocobalamin are present in lesser amounts<sup>(15,16)</sup>. Rapid pasteurisation had few effects on vitamin B<sub>12</sub> content in milk<sup>(35)</sup>. To the best of our knowledge, there is no information on the fate of the different cobalamins in milk after microfiltration. Information on the relative intestinal availability of the different cobalamins is scarce. The only data available compared whole-body retention of crystalline radioactive forms of different cobalamins in human subjects<sup>(36,37)</sup>. At doses between 100 and 1000 µg, Weisberg & Glass<sup>(36)</sup> observed no difference between cyanocobalamin and hydroxocobalamin. At 25 µg, the whole-body retention of vitamin B<sub>12</sub> was higher after the ingestion of crystalline forms of adenosylcobalamin, hydroxocobalamin and methylcobalamin than cyanocobalamin<sup>(37)</sup>. Therefore, along with the above-mentioned possible interaction with casein, cobalamin forms naturally present in milk could also contribute to explain the increased net PDV flux of the vitamin following ingestion of milk compared with cyanocobalamin.

In conclusion, the present results, reporting that vitamin B<sub>12</sub> naturally and abundantly present in cows' milk is also more available at the intestinal level than its synthetic form, are in accordance with retrospective studies in human subjects showing that vitamin B<sub>12</sub> status is highly correlated with dairy product intake<sup>(5,6,8)</sup>. Therefore, cows' milk could become a unique prophylactic tool to prevent vitamin B<sub>12</sub> deficiencies in human nutrition.

### Acknowledgements

The authors are grateful to M. Guillette, G. Danyod and G. Bélanger for their invaluable technical assistance and to C. Perreault, S. Bourgault, C. Roy and D. Morrissette for their animal care assistance. None of the authors had a financial or personal conflict of interest in relation to the present study. The study was carried out with support from Dairy Farmers of Canada and the Matching Investment Initiative program of Agriculture and Agri-Food Canada. The study sponsors had no role in the design and conduct of the study; the collection, management, analysis and interpretation of the data; the preparation, approval and submission of the manuscript. J. J. M., F. G. and C. L. G. contributed to the design of the study, interpretation of the results, writing of the manuscript, revision of its content and approval of the final version submitted for publication. J. J. M. was the principal investigator and supervised data collection; C. L. G. supervised laboratory analysis and performed the statistical analysis.

## References

- Martens J-H, Barg H, Warren MJ, *et al.* (2002) Microbial production of vitamin B<sub>12</sub>. *Appl Microbiol Biotechnol* **58**, 275–285.
- Combs GF Jr (1998) *The Vitamins: Fundamental Aspects in Nutrition and Health*, 2nd ed. San Diego, CA: Academic Press.
- Selhub J, Morris MS & Jacques PF (2007) In vitamin B<sub>12</sub> deficiency, higher serum folate is associated with increased total homocysteine and methylmalonic acid concentrations. *Proc Natl Acad Sci U S A* **104**, 19995–20000.
- Selhub J, Morris MS, Jacques PF, *et al.* (2009) Folate-vitamin B-12 interaction in relation to cognitive impairment, anemia, and biochemical indicators of vitamin B-12 deficiency. *Am J Clin Nutr* **89**, 702S–706S.
- Tucker KL, Rich S, Rosenberg IH, *et al.* (2000) Plasma vitamin B-12 concentrations relate to intake source in Framingham offspring study. *Am J Clin Nutr* **71**, 514–522.
- Vogiatzoglou A, Smith AD, Nurk E, *et al.* (2009) Dietary sources of vitamin B-12 and their association with plasma vitamin B-12 concentrations in the general population: the Hordaland Homocysteine Study. *Am J Clin Nutr* **89**, 1078–1087.
- Bor MV, von Castel-Roberts KM, Kauwell GPA, *et al.* (2010) Daily intake of 4 to 7 µg dietary vitamin B-12 is associated with steady concentrations of vitamin B-12-related biomarkers in a healthy young population. *Am J Clin Nutr* **91**, 571–577.
- Miller DR, Specker BL, Ho ML, *et al.* (1991) Vitamin B<sub>12</sub> status in a macrobiotic community. *Am J Clin Nutr* **53**, 524–529.
- United States Department of Agriculture (USDA) (2004) National Nutrient Database for Standard, Reference, Release 16. Composition of Foods, Raw, Processed, Prepared. U.S. Department of Agriculture Agricultural Research Service, Beltsville, Maryland 20705, USA. [www.nal.usda.gov/fnic/foodcomp/Data/SR16/sr16\\_doc.pdf](http://www.nal.usda.gov/fnic/foodcomp/Data/SR16/sr16_doc.pdf)
- Canadian Food Inspection Agency (CFIA) (2010) Guide to Food Labelling and Advertising. Chapter 7. 7.25 Vitamin and Mineral Nutrient Claims. <http://www.inspection.gc.ca/english/fssa/labete/guide/ch7be.shtml#tab7-15>
- Health Canada (2006) Food and Nutrition, Dietary Reference Intakes, Tables, ISBN: 0-662-41134-X, Cat. No.: H44-87/2005E-HTML. <http://www.hc-sc.gc.ca/fn-an/nutrition/reference/table/index-eng.php>
- Preynat A, Lapierre H, Thivierge MC, *et al.* (2009) Influence of methionine supply on the responses of lactational performance of dairy cows to supplementary folic acid and vitamin B<sub>12</sub>. *J Dairy Sci* **92**, 1685–1695.
- Herbert V (1988) Vitamin B-12: plant sources, requirements, and assay. *Am J Clin Nutr* **48**, 852–858.
- Zittoun J (1996) Cobalamins: an update of fundamental and clinical data. *Hématologie* **2**, 119–129.
- Farquharson J & Adams JF (1976) The forms of vitamin B<sub>12</sub> in foods. *Br J Nutr* **36**, 127–136.
- Fie M, Zee JA & Amiot J (1994) Séparation et quantification des isomères de la vitamine B<sub>12</sub> dans le lait et certains produits laitiers par chromatographie liquide haute performance et par radio-essai. *Sciences des aliments* **14**, 763–775.
- Girard CL & Matte JJ (2005) Effects of intramuscular injections of vitamin B<sub>12</sub> on lactation performance of dairy cows fed dietary supplements of folic acid and rumen-protected methionine. *J Dairy Sci* **88**, 671–676.
- Trouvé E, Maubois JL, Piot M, *et al.* (1991) Rétention de différentes espèces microbiennes lors de l'épuration du lait par microfiltration en flux tangentiel. *Lait* **71**, 1–13.
- Preynat A, Lapierre H, Thivierge MC, *et al.* (2009) Effects of supplements of folic acid, vitamin B<sub>12</sub> and rumen-protected methionine on whole body metabolism of methionine and glucose in lactating dairy cows. *J Dairy Sci* **92**, 677–689.
- NRC (1998) *Nutrient Requirements of Swine*, 10th ed. Washington, DC: National Academy Press.
- Hooda S, Matte JJ, Wilkinson CW, *et al.* (2009) Technical note: an improved surgical model for the long-term studies of kinetics and quantification of nutrient absorption in swine. *J Anim Sci* **87**, 2013–2019.
- Canadian Council on Animal Care (1993) *Guide to the Care and Use of Experimental Animals*, vol. 1. Ottawa, ON, Canada: Canadian Council on Animal Care.
- Agriculture Canada (1993) *Recommended Code of Practice for Care and Handling of Pigs*. Publication no. 1771E. Ottawa, ON, Canada: Agriculture Canada.
- Manet L (1969) Techniques usuelles de biologie clinique. In *Hématologie*, p. 39. Paris, France: Éditions médicales Flammarion.
- AOAC (Association of Official Analytical Chemists) (1990) *Official Methods of Analysis*, 16th ed. Arlington, VA: AOAC Int.
- Girard CL, Lapierre H, Desrochers A, *et al.* (2001) Net flux of folates and vitamin B<sub>12</sub> through the gastrointestinal tract and the liver of lactating dairy cows. *Br J Nutr* **86**, 707–715.
- SAS Institute (2004) *SAS/STAT User's Guide*, vol. 1–7. Cary, NC: SAS Inst. Inc.
- Guilloteau P, Zabielski R, Hammon HM, *et al.* (2010) Nutritional programming of gastrointestinal tract development. Is pig a good model for man? *Nutr Res Rev* **23**, 4–22.
- Matte JJ, Guay F, Le Floch N, *et al.* (2010) Bioavailability of dietary cyanocobalamin (vitamin B<sub>12</sub>) in growing pigs. *J Anim Sci* **88**, 3936–3944.
- Herbert V (1987) Recommended dietary intakes (RDI) of vitamin B-12 in humans. *Am J Clin Nutr* **45**, 671–678.
- Schneider Z & Stroiński A (1987) *Comprehensive B<sub>12</sub>. Chemistry, Biochemistry, Nutrition, Ecology, Medicine*. Berlin, Germany: Walter de Gruyter.
- Gizis E, Kim YP, Brunner JR, *et al.* (1965) Vitamin B<sub>12</sub> content and binding capacity of cow's milk proteins. *J Nutr* **87**, 349–352.
- Russell RM, Baik H & Kehayias JJ (2001) Older men and women efficiently absorb vitamin B-12 from milk and fortified bread. *J Nutr* **131**, 291–293.
- Cullen RW & Oace SM (1978) Methylmalonic acid and vitamin B<sub>12</sub> excretion of rats consuming diets varying in cellulose and pectin. *J Nutr* **108**, 640–647.
- Karlin R, Hours C, Vallier C, *et al.* (1969) Sur la teneur en folates des laits de grand mélange. Effets de divers traitements thermiques sur les taux de folates, B<sub>12</sub> et B<sub>6</sub> de ces laits. *Internationale Zeitschrift für Vitaminforschung* **39**, 359–371.
- Weissberg H & Glass GBJ (1966) Hydroxocobalamin. VI. Comparison of intestinal absorption in man of large doses of hydroxocobalamin and cyanocobalamin. *Proc Soc Exp Biol Med* **122**, 25–28.
- Adams JF, Ross SK, Mervyn L, *et al.* (1971) Absorption of cyanocobalamin, coenzyme B 12, methylcobalamin, and hydroxocobalamin at different dose levels. *Scand J Gastroenterol* **6**, 249–252.