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Review Article

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The role of epigenetics in respiratory health in urban populations in low and middle-income countries

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Abstract

As urbanization increases in low- and middle-income countries (LMICs), urban populations will be increasingly exposed to a range of environmental risk factors for non-communicable diseases. Inadequate living conditions in urban settings may influence mechanisms that regulate gene expression, leading to the development of non-communicable respiratory diseases. We conducted a systematic review of the literature to assess the relationship between respiratory health and epigenetic factors to urban environmental exposures observed in LMICs using MEDLINE, PubMed, EMBASE, and Google Scholar searching a combination of the terms: epigenetics, chronic respiratory diseases (CRDs), lung development, chronic obstructive airway disease, and asthma. A total of 2835 articles were obtained, and 48 articles were included in this review. We found that environmental factors during early development are related to epigenetic effects that may be associated with a higher risk of CRDs. Epigenetic dysregulation of gene expression of the histone deacetylase (HDAC) and histone acetyltransferase gene families was likely involved in lung health of slum dwellers. Respiratory-related environmental exposures influence HDAC function and deoxyribonucleic acid methylation and are important risk factors in the development of CRD. Additional epigenetic research is needed to improve our understanding of associations between environmental exposures and non-communicable respiratory diseases.

Introduction

In 2014, 881 million people in low- and middle-income countries (LMICs) lived in urban slums, and this number is expected to grow on average by 9 million people each year [1, 2]. A greater proliferation of low-income settlements, also known as slums, exists with a lack of access to sustainable housing, expansive living space, sanitation, safe drinking water, and security [3, 4]. Disparities are present among slum dwellers but they are not equally distributed worldwide. Specifically, in sub-Saharan Africa, the United Nations Habitat estimated that a majority of urban populations, at about 56%, live in slums [2, 5]. Overall, Africa is shifting to a predominantly urban continent. It is estimated that 40% of Africa's population lives in urban areas, but by 2030 one out of two individuals living in sub-Saharan Africa will live in an urban area [6]. Urbanization, the migration of residents from rural to urban areas often for the economic opportunities offered by urban development, has further contributed to the increase in urban poverty in LMICs [3]. This rapid urbanization has been accompanied by significant shifts in health patterns, increasing the prevalence of non-communicable diseases (NCDs). Lung disease is known as a leading cause of mortality in LMICs, where it is reported to account for 15% of all deaths [7].

As the prevalence of asthma and chronic obstructive airway disease (COPD) rise, the WHO predicts that deaths due to NCDs will increase by 27% by 2030 [7–10]. This is mainly due to demographic transitions and changing lifestyles of populations associated with urbanization. Moreover, populations in urban slums often have inadequate access to health services and seek care after symptoms have advanced and often when life-threatening complications have arisen [11]. The combination of slum conditions in Bangladesh, Kenya, and Egypt has led to severe malnutrition in adolescents, primed by a lower under-five child nutrition status, when compared to individuals living in rural settings [8, 12, 13]. As a result, slum living has been linked to poverty and multiple risk factors for respiratory disease including poor housing

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quality, overcrowding, traffic-related and household air pollution, tobacco smoke, psychological stress, occupational exposures, and exposure to allergens and malnutrition [4, 7, 11, 12].

Environmental exposures associated with living in slums can lead to epigenetic modifications and changes in cell function that begin *in utero* and could last a lifetime, therefore predisposing slum residents to poor respiratory health outcomes. Epigenetics is the study of mitotically-heritable phenotypes not resulting from changes in the deoxyribonucleic acid (DNA) sequence. The term *epi-* is a Greek suffix meaning 'on top or outside', which highlights the nature of the mechanisms outside of the DNA code [14]. The most widely known epigenetic marks include: DNA methylation, histone tail modifications, and noncoding ribonucleic acids (RNAs) [15–17]. These mechanisms control access to DNA to participate in transcription [15, 17]. The same principle applies to histone tail regulatory mechanisms as acetylation by histone acetyltransferases (HATs) leading to active gene transcription, whereas deacetylation by histone deacetylases (HDACs) leads to gene silencing [18, 19]. Non-coding RNAs (ncRNAs) also play a critical role in regulating gene expression by controlling gene expression through binding of 3' untranslated regions of mRNA [20]. The end result is either mRNA degradation or inhibition of protein translation.

Epigenetic markers are important regulators of overall DNA stability, cell differentiation, imprinting, and organismal development. As a result, epigenetic mechanisms, through their key role in cell function and heritable nature, have been implicated in the development of chronic respiratory diseases (CRDs) that include asthma and COPD in people living in LMICs, predisposing these populations to significant negative health outcomes [15]. However, potential molecular mechanisms due to epigenetic modifications from environmental exposure in this population are currently lacking. In this systematic review, we aim to focus on the current knowledge of epigenetic markers related to human environmental exposures that affect lung health and relate this to urban residents, but more specifically slum dwellers, in LMICs as a measure of the impact of urbanization in chronic illness. Also, we aim to identify the knowledge gaps in interactions between epigenetic mechanisms and specific threats to respiratory health in LMIC settings.

Methods

Using MEDLINE, PubMed, EMBASE, and Google Scholar and following the Preferred Reporting Items for Systematic Reviews and Meta-Analyses guidelines [21], we conducted a systematic search of literature published between 1 January 1995 and 9 September 2017 using the following search terms: epigenetics, Africa, Asia, Latin America, Caribbean, LMICs, CRDs, NCDs, urban slum, lung development, and asthma using a combination of 'AND' and 'OR' search settings among various combinations of terms. The primary outcomes of the review are epigenetic markers and mechanisms related to the following human environmental exposures that influence lung health: cigarette smoke, second hand smoke, ambient or household air pollution, traffic-related pollution, maternal nutrition, exposure to allergens, and nutritional status. The generated abstracts were limited to publications in English and to research papers, editorials, reviews, original articles, and reports. The first reviewer (NR) screened titles and abstracts for relevance, selecting papers for abstract analysis. The inclusion and exclusion criteria were considered during this

process. Only research articles relating to settings in LMICs or studies that could be applied to populations in LMICs were included. Articles published before 1995 were excluded from our search to limit our review to recent literature. The first reviewer (NR) selected 82 papers. Reference lists of selected papers were manually searched for further related studies. Then, the second reviewer (AK) performed the final selection of 48 articles. Any disagreement between the two authors resulted in a discussion and joint review of the article with reconciliation. Quality criteria were assessed using the Newcastle-Ottawa Scale (NOS) [22]. The aim of the search was to assess the relationship between respiratory health and epigenetic factors in environments similar to that of urban areas of LMICs. The methodological summary of this literature search is outlined in Fig. 1.

Results

A total of 8543 articles were obtained from the searches, with 48 articles ultimately included in this review. Abstracts were manually sorted, and unrelated and duplicated papers were removed. Papers were excluded based on quality, relevance to systematic literature review aims, and reported methodology and results (Fig. 1). Studies that presented results not conducted in LMICs or that could not be applied to LMIC settings were excluded. Papers that did not report methodology or results were excluded. Other reasons that papers were excluded were duplication or abstracts and publications that were not written in English. The quality of papers was assessed utilizing the NOS, Table 1. The articles included in this review span a diverse geographical distribution and are included in Supplementary Table S1 with specific relationships between epigenetic factors and lung health further outlined in Table 2. Our systematic literature review yielded articles relating to epigenetic modifications due to environmental exposures including cigarette smoke, air pollution, maternal nutrition, exposure to allergens, and malnutrition, among others. Few studies related specifically to how these predisposing environmental factors were associated with common CRDs including COPD and asthma.

DNA methylation and non-coding RNAs

Our search revealed several slum-related exposures *in utero*, in childhood, and adulthood, which have been associated with altered DNA methylation. We found that cigarette smoke and second-hand smoke (SHS) are both associated with altered epigenome methylation patterns. *In utero* exposure to maternal tobacco smoke has also been shown to result in both global and site-specific DNA methylation levels during fetal development [33–36]. Exposure to SHS has shown increased methylation of CpG sites in T effector cell IFN- γ promoters relative to unexposed individuals [26]. Traffic-related air pollution exposure increased locus-specific methylation in the *TET1* promoter region, specifically at CpG (cg23602092) site, which was significantly associated with childhood asthma [37]. Studies *in utero* have shown that maternal famine was associated with higher methylation of *IL10*, *LEP*, *ABCA1*, *GNASAS*, and *MEG3* genes, compared to the same-sex unexposed siblings, potentially increasing risk of asthma later in life [38–40]. In the current literature, there is a gap regarding ncRNA's role in regulating gene expression emphasizing the need for greater research in this area.

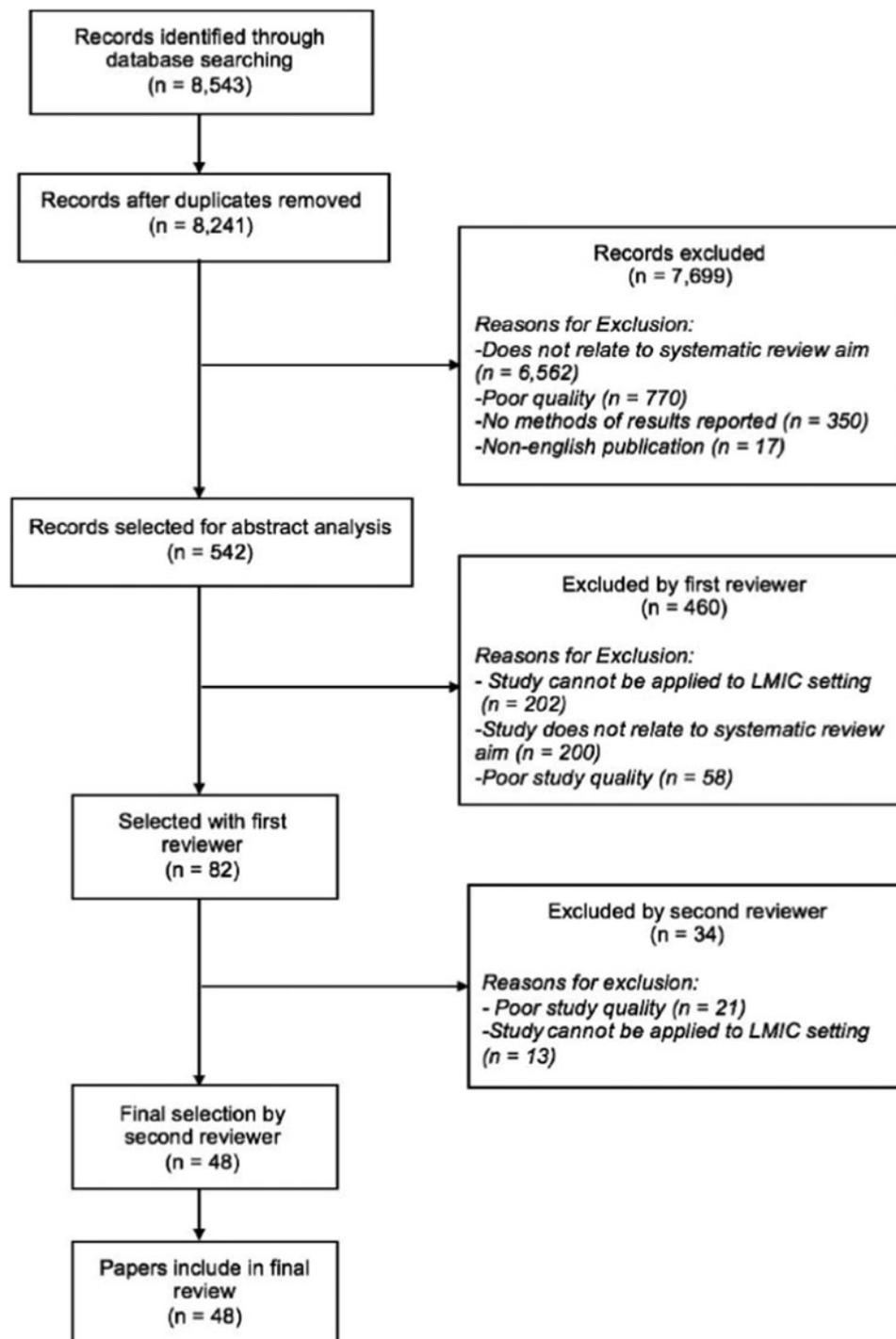


Fig. 1. Literature search process to identify papers to include in this review.

Histone deacetylases and histone acetyltransferases

Two major enzymes, HDAC and HATs, potentially play a role in epigenetic factors related to the lung health of slum dwellers. The HDAC family facilitates local histone deacetylation and transcriptional repression by binding to DNA substrates. In Class I of the HDAC family, specifically HDAC 1, 2, 3, and 8, have been shown at reduced levels in individuals with COPD or asthma or individuals that smoke tobacco. Marwick *et al.*, observed decreased HDAC2 activity due to cigarette smoke exposure. Ito *et al.*, expanded on these results by demonstrating that cigarette smoke

exposure decreased overall HDAC activity in bronchial biopsies and alveolar macrophages of smokers compared to non-smokers [29, 30]. Moreover, the HAT families acetylate the lysine residues at the N-terminus of histone proteins by removing positive charges. As a result, the affinity between histones and DNA is reduced, facilitating access to the promoter region of target genes by RNA polymerase and transcription factors. Among the Gcn5-related N-acetyltransferase (GNAT) family, HAT activity was associated with higher pro-inflammatory gene expression among individuals with asthma [27]. A summary of these findings included studies is contained in Table 2 and in Supplementary Table S1.

Table 1. Example of NOS for assessment of quality of included studies-cross-sectional studies (1 indicates individual criterion within the subsection was fulfilled)

| Quality assessment criteria | White <i>et al.</i> [23] | Waterland <i>et al.</i> [24] | Ezzie <i>et al.</i> [25] | Kohli <i>et al.</i> [26] |
|---|--------------------------|------------------------------|--------------------------|--------------------------|
| Selection (maximum 4) | | | | |
| Case definition adequate | 1 | 1 | 1 | 1 |
| Representativeness of the cases | 1 | 1 | 1 | 1 |
| Selection of controls | 1 | 1 | 1 | 1 |
| Definition of controls | 1 | 1 | 0 | 1 |
| Comparability (maximum 2) | | | | |
| Comparability of cohorts on the basis of the design of analysis (controls used) | 1 | 1 | 1 | 1 |
| Confounding factors are controlled. | 1 | 1 | 1 | 1 |
| Exposure (maximum 3) | | | | |
| Ascertainment of exposure | 1 | 1 | 1 | 1 |
| Same method cases and controls? | 1 | 1 | 1 | 1 |
| Non-response rate | 0 | 1 | 0 | 0 |
| Overall quality score (maximum 9) | 8 | 9 | 7 | 8 |

Discussion

In this review, maternal nutrition, tobacco smoking, ambient and household air pollution, and traffic-related air pollution during early development were risk factors associated with epigenetic changes in LMICs. In particular, the HDAC, HAT, and GNAT families, specifically, were associated with lung health outcomes.

The Barker hypothesis proposes that environmental factors acting during the phase of early development interact with the genome to change the capacity of the organism to cope with its environment in later life [38, 41]. This hypothesis has expanded to find other immunological, mental health, and reproductive diseases that can result from malnutrition or over-nutrition [38]. Current research points to evidence that epigenetic marks alter gene expression in the lung, which may be associated with common CRDs [42]. Because the epigenome is dynamic and changes accordingly to the environment and ageing process, associations between cigarette smoke and SHS and epigenetic marks have been demonstrated in recent studies in high-income settings [33, 43]. For example, intergenerational DNA methylation associated with *in utero* cigarette smoke exposure is associated with a high risk of asthma development in offspring [33, 44]. *In utero* exposure to maternal tobacco smoking has been shown to be significantly associated with increased newborn epigenome-wide alterations and methylation at 26 CpGs mapped at 10 genes and reduced mean methylation for AluYb8 repetitive elements [33–35]. SHS exposure has also been associated with a higher percentage of methylation of CpG sites in the T effector cell IFN- γ promoter with a concomitant reduction in IFN- γ expression from T effector cells when compared to individuals not exposed to SHS [26]. Exposure to cigarette smoke also affects miRNA expression. Ezzie *et al.*, found that *miR-223* and *miR-1274a* were expressed almost three-fold in COPD lung samples compared to non-COPD lung samples [25]. Moreover, 28 miRNAs were differentially expressed in airway epithelial cells, with most of these miRNAs down-regulated when exposed to tobacco smoke compared to unexposed cells [45]. Because childhood years have been found to be a critical time for rapid lung

development, reducing exposure to cigarette smoke and SHS may reduce the risk of developing chronic respiratory conditions later in life [46].

Air pollution is a ubiquitous environmental exposure, especially in urban settings. Previous research in high-income settings has found that DNA methylation of long interspersed nucleotide element (LINE)-1 was lower with higher levels of ambient particulate matter [47]. Diesel exhaust particle exposure has been found to impact DNA methylation at 2827 CpG sites compared to filtered air; with CpG sites, such as *GSTP1*, becoming significantly less methylated [48]. Asthma was significantly associated with loss of methylation at a single CpG site in the *TET1* promoter (cg23602092) and increased global 5hmC in bronchial epithelial cells and human participants [37]. Diesel exhaust particle exposure was associated with alterations in Alu and LINE-1 elements and the CpG site within *miR-21*, and increased *FOXP3* was significantly associated with increased diesel exhaust particle exposure, which was associated with increased risk of asthma development in children [48, 49]. Allergen exposure, diesel exhaust particle exposure, and co-exposure showed alterations in 7 CpG sites in bronchial epithelial cells after two days. Spacing out allergen and diesel exhaust particle exposure by four weeks resulted in alterations in over 500 CpG sites including four Hox family of genes such as *HOXA3*, *HOXA4*, *HOXB1*, and *HOXB3* involved in fetal lung development, and global DNA methylation suggesting sequential exposure has a pronounced impact on DNA methylation [50]. Moreover, methylation of the *ACSL3* 5'-CGI was significantly associated with maternal airborne polycyclic aromatic hydrocarbon exposure above 2.41 ng/m³ in umbilical cord white blood cells potentially related to the development of traffic-related air pollution exposure asthma [51].

Diet and nutrition may also exert epigenetic changes. Indeed, the relationship between epigenetic maternal and either early childhood nutrition or dietary supplementation have been studied in populations in the Dutch Hunger Winter Famine in Europe, in Nepal, and in The Gambia [24, 40, 52, 53]. In individuals exposed to *in utero* famine conditions *INSIGF* methylation was significantly reduced and increased in *IL10*, *LEP*, *ABCA1*, *GNASAS*,

Table 2. Results of the relationship between epigenetic factors and lung health in included studies in this systematic literature review

| Epigenetic factors | Activity in asthma, COPD, or smoking | Reference | Sample size | Type of sample | Country |
|-----------------------------------|---|-----------|-------------|-------------------------------|----------------|
| Class I HDAC Family | | | | | |
| HDAC 1 | Decreased in asthma, not yet studied in smoking and COPD patients | [27] | 40 | Bronchial biopsy samples | UK |
| | | [28] | 28 | Alveolar macrophages | UK |
| | | [29] | 24 | Rat lung tissue | UK |
| HDAC 2 | Decreased in asthma, smoking and COPD | [27] | 40 | Bronchial biopsy samples | UK |
| | | [28] | 28 | Alveolar macrophages | UK |
| | | [29] | 24 | Rat lung tissue | UK |
| | | [30] | 29 | Human bronchial biopsies | UK |
| HDAC 3 and 8 | Decreased in COPD, no difference in asthma, limited published data in smoking | [28] | 28 | Alveolar macrophages | UK |
| | | [31] | 159 | Human lung tissue | USA and Canada |
| Class II HDAC family | | | | | |
| HDAC 5 | Decreased in COPD, no difference in asthma, limited published data in smoking | [28] | 28 | Alveolar macrophages | UK |
| | | [31] | 159 | Human lung tissue | USA and Canada |
| Class III HDAC family | | | | | |
| SIRT1 | Limited published data | | | | |
| Class IV HDAC family | | | | | |
| HDAC 11 | Limited published data | | | | |
| GNATs-HAT family | | | | | |
| HAT1, GCN5, GCN5L, Elp3, PCAF | Increased in asthma patients, limited published data on smoking and COPD | [27] | 40 | Bronchial biopsy samples | UK |
| | | [32] | – | Cell culture-mature monocytes | USA |
| General transcription factor-HATs | | | | | |
| TAF250, TFIIC | Increased in asthma, limited published data on smoking and COPD | [27] | 40 | Bronchial biopsy samples | UK |
| | | [28] | 28 | Alveolar macrophages | UK |

HDAC, histone deacetylase; SIRT1, sirtuin; HAT, histone acetyl transferase; GCN5, general control of amino acid synthesis protein 5; GCN5L, general control of amino acid synthesis protein 5 ligand; Elp3, elongation complex protein; PCAF, protein300/CREB-binding protein-associated factor; TAF250, TBP-associated factor 250 kDa; TFIIC, transcription factors IIC.

and *MEG3* genes compared to unexposed same-sex siblings [40]. Seasonal changes in maternal methyl-donor nutrient intake during conception also have been found to influence 13 plasma biomarkers and systemic epigenetic changes in human metastable epialleles in Gambian populations with metastable epialleles having increased DNA methylation levels for individuals conceived in the rainy season when nutritional intake is low [24, 52]. Moreover, the lack of maternal dietary vitamin A may alter *in utero* lung development potentially increasing susceptibility of postnatal lung diseases. In Nepal, children aged 9–13 years born to mothers who were assigned to receive 7000- μ g retinol-equivalents of vitamin A weekly before, during, and after pregnancy were shown to have a larger forced expiratory volume at 1 s and forced vital capacity than children of the same age who were born to mothers assigned to receive a placebo [53].

Emerging evidence of the relationship between environmental exposures and epigenetic markers coupled with the function of epigenetic regulation in T-cell differentiation demonstrates epigenetic alterations that may contribute to a higher prevalence of asthma in LMICs [54]. Current research is focusing on the effect of maternal nutrition on the development of atopy and asthma in children [55, 56]. For example, epigenetic changes, specifically increasing site-specific methylation levels of the genome, during pregnancy shifts towards a Type II helper phenotype, increasing the risk of asthma [38, 39]. Previous epidemiological studies have shown that increasing folate levels, which is a precursor for the methyl group supply that is used to generate DNA methylation marks, are associated with an increased risk of developing asthma [38]. Additionally, acetylation of histones may influence the onset of asthma because increased levels of acetylation of H4 has been found in asthmatic individuals and is associated

with increased inflammatory gene expression in lung tissue [57]. The level of acetylation of histones has been shown to be associated with enhanced inflammatory gene expression by HATs and reduced inflammatory gene expression by HDACs [58]. The altered HAT/HDAC ratio from inflammation in peripheral blood cells has been shown in adults and children that correlates with alterations in bronchial hyperresponsiveness as in asthma and in patients with COPD [59, 60].

The degree of increase in the acetylation of histones associated with the promoter region of inflammatory genes in peripheral lung tissue has been found to be associated with the severity of COPD [61]. Alterations in histone acetylation patterns and other epigenetic changes could mean promising therapies for anti-inflammatory conditions such as corticosteroid resistant cases of asthma [57]. Glucocorticoids change acetylation patterns of histones via mechanisms that regulate inflammatory and anti-inflammatory genes [57]. In addition to maternal nutrition, cigarette smoke exposure, for example, reduces the expression of HDAC2, a glucocorticoid receptor corepressor, at the protein and mRNA levels meaning maternal secondhand smoke exposure may influence the HDAC2 promoter, which could lead to asthma progression *in utero* [62].

While there is a significant amount of research focused on the genetics and epigenetic factors surrounding populations in high-income countries, there is little priority on the interaction between environmental factors and genetics in LMICs, where the burden of disease is greatest. The lack of research data, as evident in a large number of papers excluded during the systematic review process, hinders health systems from allocating resources and targeting policy efforts towards short-term and long-term CRD prevention and management services in LMICs (Fig. 1). Further epigenetic research could lead to identifying epigenetic targets stemming from living in urban slums. This research could identify potential therapies that target epigenetic modifications in urban slum dwellers along with associated poor health outcomes that result from epigenetic modifications.

Currently, there is little focus on these silent killers known as NCDs such as CRDs in LMICs. As is evident in this review, environmental factors afflicting residents of slums and urban areas of developing settings can have a significant impact on lung health and development that can lead to adverse health outcomes. Environmental exposures *in utero* and during adolescent years can have long lasting and possibly irreversible effects later in life. Researching and targeting the upstream factors of CRD onset and the biological mechanisms is not only economical but will improve the quality of life for these vulnerable, at-risk populations.

Supplementary material. The supplementary material for this article can be found at <https://doi.org/10.1017/gheg.2019.7>

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Conflict of interest. We declare no competing interests.

References

1. **UN-Habitat** (2016) *Planning Sustainable Cities: Global Report on Human Settlements 2009*. London: Routledge.
2. **UN-Habitat** (2016) *World Cities Report*. Nairobi, Kenya: UN-Habitat.
3. **Nolan LB** (2015) Slum definitions in urban India: implications for the measurement of health inequalities. *Population and Development Review* **41**, 59–84.
4. **Gilbert A** (1998) *An Urbanizing World: Global Report on Human Settlements, 1996*. Oxford: United Nations Centre for Human Settlements (Habitat), Oxford University Press, Pergamon.
5. **Artuso M** (2013) *UN Habitat, State of the World's Cities 2012/13—Prosperity of Cities*. London: Earthscan, 184 pp., ISBN 13: 978-0-415-83888-7. Tab. Graph. Images. 2014, Taylor & Francis.
6. **United Nations** (2014) Department of Economic and Social Affairs, Population Division. *World Urbanization Prospects: The 2014 Revision*.
7. **Brashier Bill, Londhe J, Madas S, Vincent V and Salvi S** (2012) Prevalence of self-reported respiratory symptoms, asthma and chronic bronchitis in slum area of a rapidly developing Indian city. *Open Journal of Respiratory Diseases* **2**, 73.
8. **Mberu BU, Haregu TN, Kyobutungi C et al.** (2016) Health and health-related indicators in slum, rural, and urban communities: a comparative analysis. *Global Health Action* **9**, 33163.
9. **Adeloye D, Basquill C, Papan A et al.** (2015) An estimate of the prevalence of COPD in Africa: a systematic analysis. *Journal of Chronic Obstructive Pulmonary Disease* **12**, 71–81.
10. **Mathers CD and Loncar D** (2006) Projections of global mortality and burden of disease from 2002 to 2030. *PLoS Medicine* **3**, e442.
11. **Riley LW, Ko AI, A Unger et al.** (2007) Slum health: diseases of neglected populations. *BMC International Health and Human Rights* **7**, 2.
12. **Checkley W, Pollard SL, Siddharthan T et al.** (2016) Managing threats to respiratory health in urban slums. *The Lancet Respiratory Medicine* **4**, 852–854.
13. **Bousquet J, Mantzouranis E, Cruz AA et al.** (2010) Uniform definition of asthma severity, control, and exacerbations: document presented for the World Health Organization consultation on severe asthma. *Journal of Allergy and Clinical Immunology* **126**, 926–938.
14. **Allis CD, Jenuwein T and Reinberg D** (eds) (2009) *Epigenetics*. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press.
15. **Yang IV and Schwartz DA** (2011) Epigenetic control of gene expression in the lung. *American Journal of Respiratory and Critical Care Medicine* **183**, 1295–1301.
16. **Egger G, Liang G, Aparicio A et al.** (2004) Epigenetics in human disease and prospects for epigenetic therapy. *Nature* **429**, 457.
17. **Bannister AJ and Kouzarides T** (2011) Regulation of chromatin by histone modifications. *Cell Research* **21**, 381.
18. **Agalioti T, Chen G and Thanos D** (2002) Deciphering the transcriptional histone acetylation code for a human gene. *Cell* **111**, 381–392.
19. **De Ruijter AJ, van Gennip AH, Caron HN et al.** (2003) Histone deacetylases (HDACs): characterization of the classical HDAC family. *Biochemical Journal* **370**, 737–749.
20. **Wang KC and Chang HY** (2011) Molecular mechanisms of long non-coding RNAs. *Molecular cell* **43**, 904–914.
21. **Shamseer L, Moher D, Clarke M et al.** (2015) Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015: elaboration and explanation. *BMJ* **349**, 7647.
22. **Wells GA, Shea B, O'Connell D et al.** (2016) The Newcastle-Ottawa Scale (NOS) for Assessing the Quality of Nonrandomised Studies in Meta-Analyses. Accessed from: http://www.ohri.ca/programs/clinical_epidemiology/oxford.asp.
23. **White GP, Hollams EM, Yerkovich ST et al.** (2006) CpG methylation patterns in the IFN gamma promoter in naive T cells: variations during Th1 and Th2 differentiation and between atopics and non-atopics. *Pediatric Allergy and Immunology* **17**, 557–564.
24. **Waterland RA, Kellermayer R, Laritsky E et al.** (2010) Season of conception in rural Gambia affects DNA methylation at putative human metastable epialleles. *PLoS Genetics* **6**, e1001252.
25. **Ezzie ME, Crawford M, Cho JH et al.** (2012) Gene expression networks in COPD: microRNA and mRNA regulation. *Thorax* **67**, 122–131.
26. **Kohli A, Garcia MA, Miller RL et al.** (2012) Secondhand smoke in combination with ambient air pollution exposure is associated with increased CpG methylation and decreased expression of IFN- γ in T effector cells and Foxp3 in T regulatory cells in children. *Clinical Epigenetics* **4**, 17.
27. **Ito K, Caramori G, Lim S et al.** (2002) Expression and activity of histone deacetylases in human asthmatic airways. *American Journal of Respiratory and Critical Care Medicine* **166**, 392–396.

28. Cosio BG, Mann B, Ito K *et al.* (2004) Histone acetylase and deacetylase activity in alveolar macrophages and blood monocytes in asthma. *American Journal of Respiratory and Critical Care Medicine* **170**, 141–147.
29. Marwick JA, Kirkham PA, Stevenson CS *et al.* (2004) Cigarette smoke alters chromatin remodeling and induces proinflammatory genes in rat lungs. *American Journal of Respiratory Cell and Molecular Biology* **31**, 633–642.
30. Ito K, Lim S, Caramori G *et al.* (2001) Cigarette smoking reduces histone deacetylase 2 expression, enhances cytokine expression, and inhibits glucocorticoid actions in alveolar macrophages. *The FASEB Journal* **15**, 1110–1112.
31. Hogg JC, Chu F, Utokaparch S *et al.* (2004) The nature of small-airway obstruction in chronic obstructive pulmonary disease. *New England Journal of Medicine* **350**, 2645–2653.
32. Yang SR, Chida AS, Bauter MR *et al.* (2006) Cigarette smoke induces proinflammatory cytokine release by activation of NF-kappaB and post-translational modifications of histone deacetylase in macrophages. *American Journal of Physiology-Lung Cellular and Molecular Physiology* **291**, L46–L57.
33. Breton CV, Byun HM, Wenten M *et al.* (2009) Prenatal tobacco smoke exposure affects global and gene-specific DNA methylation. *American Journal of Respiratory and Critical Care Medicine* **180**, 462–467.
34. Joubert BR, Häberg SE, Bell DA *et al.* (2014) Maternal smoking and DNA methylation in newborns: in utero effect or epigenetic inheritance? *Cancer Epidemiology, Biomarkers & Prevention* **23**, 1007–1017.
35. Markunas CA, Xu Z, Harlid S *et al.* (2014) Identification of DNA methylation changes in newborns related to maternal smoking during pregnancy. *Environmental Health Perspectives* **122**, 1147–1153.
36. Suter M, Abramovici A, Showalter L *et al.* (2010) In utero tobacco exposure epigenetically modifies placental cyp11a expression. *Metabolism: Clinical and Experimental* **59**, 1481–1490.
37. Sominen HK, Zhang X, Biagini Myers JM *et al.* (2016) TET1 methylation is associated with childhood asthma and traffic-related air pollution. *The Journal of Allergy and Clinical Immunology* **137**, 797–805.e5.
38. Heindel JJ and Vandenberg LN (2015) Developmental origins of health and disease: integrating environmental influences. *Endocrinology* **156**, 3416–3421.
39. Hollingsworth JW, Maruoka S, Boon K *et al.* (2008) *In utero* supplementation with methyl donors enhances allergic airway disease in mice. *The Journal of Clinical Investigation* **118**, 3462.
40. Tobi EW, Lumey LH, Talens RP *et al.* (2009) DNA methylation differences after exposure to prenatal famine are common and timing- and sex-specific. *Human Molecular Genetics* **18**, 4046–4053.
41. Gluckman PD and Hanson MA (2004) Developmental origins of disease paradigm: a mechanistic and evolutionary perspective. *Pediatric Research* **56**, 311–317.
42. Jirtle RL and Skinner MK (2007) Environmental epigenomics and disease susceptibility. *Nature Reviews Genetics* **8**, 253.
43. Wilson AG (2008) Epigenetic regulation of gene expression in the inflammatory response and relevance to common diseases. *Journal of Periodontology* **79**, 1514–1519.
44. Li YF, Langholz B, Salam MT *et al.* (2005) Maternal and grandmaternal smoking patterns are associated with early childhood asthma. *Chest* **127**, 1232–1241.
45. Schembri F, Sridhar S, Perdomo C *et al.* (2009) MicroRNAs as modulators of smoking-induced gene expression changes in human airway epithelium. *Proceedings of the National Academy of Sciences* **106**, 2319–2324.
46. Gauderman WJ, Urman R, Avol E *et al.* (2015) Association of improved air quality with lung development in children. *The New England Journal of Medicine* **372**, 905–913.
47. Baccarelli A, Wright RO, Bollati V *et al.* (2009) Rapid DNA methylation changes after exposure to traffic particles. *American Journal of Respiratory and Critical Care Medicine* **179**, 572–578.
48. Jiang R, Jones MJ, Sava F *et al.* (2014) Short-term diesel exhaust inhalation in a controlled human crossover study is associated with changes in DNA methylation of circulating mononuclear cells in asthmatics. *Particle and Fibre Toxicology* **11**, 71.
49. Brunst KJ, Leung YK, Ryan PH *et al.* (2013) FOXP3 hypermethylation is associated with diesel exhaust exposure and risk for childhood asthma. *The Journal of Allergy and Clinical Immunology* **131**, 592–594.e3.
50. Clifford RL, Jones MJ, Malsaac JL *et al.* (2017) Inhalation of diesel exhaust and allergen alters human bronchial epithelium DNA methylation. *Journal of Allergy and Clinical Immunology* **139**, 112–121.
51. Perera F, Tang WY, Herbstman J *et al.* (2009) Relation of DNA methylation of 5'-CpG island of ACSL3 to transplacental exposure to airborne polycyclic aromatic hydrocarbons and childhood asthma. *PLoS ONE* **4**, e4488.
52. Dominguez-Salas P, Moore SE, Baker MS *et al.* (2014) Maternal nutrition at conception modulates DNA methylation of human metastable epialleles. *Nature Communications* **5**, 3746.
53. Checkley W, West Jr KP, Wise RA *et al.* (2010) Effects of maternal vitamin A supplementation on lung function in preadolescent offspring: a follow-up study of a randomized, double-blinded, placebo-controlled trial cohort in rural Nepal. *New England Journal of Medicine* **362**, 1784–1794.
54. Eder W, Ege MJ, von Mutius E *et al.* (2006) The asthma epidemic. *New England Journal of Medicine* **355**, 2226–2235.
55. Zeiger RS and Heller S (1995) The development and prediction of atopy in high-risk children: follow-up at age seven years in a prospective randomized study of combined maternal and infant food allergen avoidance. *Journal of Allergy and Clinical Immunology* **95**, 1179–1190.
56. Shaheen S, Northstone K, Newson RB *et al.* (2009) Dietary patterns in pregnancy and respiratory and atopic outcomes in childhood. *Thorax* **64**, 411–417.
57. Barnes PJ, Adcock IM, Ito K *et al.* (2005) Histone acetylation and deacetylation: importance in inflammatory lung diseases. *European Respiratory Journal* **25**, 552–563.
58. Rahman IJ, Marwick J, Kirkham P *et al.* (2004) Redox modulation of chromatin remodeling: impact on histone acetylation and deacetylation, NF-kB and pro-inflammatory gene expression. *Biochemical Pharmacology* **68**, 1255–1267.
59. Hew M, Bhavsar P, Torrego A *et al.* (2006) Relative corticosteroid insensitivity of peripheral blood mononuclear cells in severe asthma. *American Journal of Respiratory and Critical Care Medicine* **174**, 134–141.
60. Su RC, Becker AB, Kozyrskyj AL *et al.* (2009) Altered epigenetic regulation and increasing severity of bronchial hyperresponsiveness in atopic asthmatic children. *The Journal of Allergy and Clinical Immunology* **124**, 1116–1118.
61. Ito K, Ito M, Elliott WM *et al.* (2005) Decreased histone deacetylase activity in chronic obstructive pulmonary disease. *New England Journal of Medicine* **352**, 1967–1976.
62. Islam KN and Mendelson CR (2008) Glucocorticoid/glucocorticoid receptor inhibition of surfactant protein-A (SP-A) gene expression in lung type II cells is mediated by repressive changes in histone modification at the SP-A promoter. *Molecular Endocrinology* **22**, 585–596.