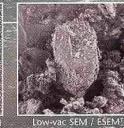
MICROSCOPY TODAY

NOVEMBER / DECEMBER 2002

VOLUME IO - NUMBER 6

Imagine Microscopy without Barriers

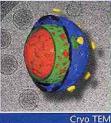
3D Structural Process Management™ Eliminates the Obstacle











www.feicompany.com sales@feico.com

FEI's 3D Structural Process Management[™] solutions for microscopy deliver the most complete data set quickly and efficiently. Whether your interest is in life science, structural biology, materials science or microelectronics, our microscopy solutions help you break through barriers in research, development and manufacturing.

Engineered with your success in mind, our full range of TEM, DualBeam™, ESEM™, and SEM solutions enable you to study samples in three dimensions, including below the surface.

Tecnai[™] G² TEM: high resolution, three-dimensional TEM analysis down to the atomic level.

Strata[™] DB 235M: three dimensional, below the surface data and nanofabrication capabilities.

Quanta[™] SEM: All in One SEM delivering high-vac, low-vac, and ESEM flexibility.

Sirion[™] SEM: low kV imaging plus integrated STEM delivering better than 1 nm image resolution.

Contact us and learn how FEI's unique combination of core technologies and more than 70 years of experience combine to remove the obstacles to your microscopy success and enable you to break through nanotech barriers.

When Dinosaurs Became Extinct, What Happened to the Insects?

Stephen W. Carmichael¹ carmichael.stephen@mayo.edu

It is widely accepted that approximately 65 million years ago, an extraterrestrial object slammed into the Yucatán Peninsula of Mexico, creating a worldwide climatic shift that wiped out the dinosaurs, and many other terrestrial and marine species. But what happened to the insects? They undoubtedly represented a larger biomass than the dinosaurs, but it's fair to say they haven't captured the public's imagination in the same way. The reason the fate of the insects has not been adequately explored is due to the paucity of the available fossil record of the insect bodies. There are a few records of insects embedded in amber or fossilized, but practically none are available from the time of mass extinction referred to as the Cretaceous-Paleocene boundary. Recently, Conrad Labandeira, Kirk Johnson, and Peter Wilf found an ingenious way to examine indirect evidence and show what happened to the insects during this time.2

Whereas the insect fossil record is sparse, there is an abundance of plant fossils. For those of us who are weekend gardeners, we are all too familiar with the damage that insects can do to plants. We also recognize that some of this damage is general (a nibbled leaf can be caused by several different insects) to specific (a leaf miner leaves a distinctive trail within a leaf of a particular species of plant). Labandeira et al. had the clever idea to examine plant fossils for insect damage.

At a site in North Dakota known for plant fossils that spanned the Cretaceous-Paleocene boundary, the team used 10X or 20X hand lenses to examine specimens in situ. In all, 13,441 fossil plant specimens (9,292 from the Cretaceous period, 4,149 from the Paleocene) were examined back in the laboratory with a standard stereomicroscope at magnifications ranging from 14X to 80X, with particularly interesting insect damage patterns examined at higher magnifications. In contrast, only five insect body fossils have been found within the same collections. Labandeira et al. took advantage of the statistical robustness of the large sample size to document the trophic associations between plants and their insect herbivores.

Each specimen was examined for damage from insects, and the damage was assigned to one or more of 51 insect damage types, ranging from general to very specific. Each damage type was classified into one of four functional feeding groups (external foliage feeding, galling, mining, and piercing and sucking) or nonfeeding damage due to deposition of eggs. After extensive use of sophisticated statistical methods, Labandeira et al. showed significant variations of insect-plant associations during the latest Cretaceous Period, with a dramatic decrease at the boundary and into the earliest Paleocene Epoch. Their observations are consistent with a genuine extinction of many herbivorous insect species at the same time the dinosaurs disappeared. They proposed two nonexclusive scenarios. First is an outright extermination of insects caused by adverse environmental conditions. Alternatively, a secondary extinction of insects occurred after the demise of plant hosts.

Whereas this study is limited to a single site, it nevertheless presents compelling evidence for the extinction of a large number of insect species after a historically important event. Labandeira et al. are to be congratulated for taking a shrewd and indirect approach to providing a definitive answer to a perplexing question.

Footnotes

- ¹ The author gratefully acknowledges Dr. Conrad Labandeira for supplying information on the use of microscopy in this study and reviewing the manuscript
- ² Labandeira, C.C., K.R. Johnson, P. Wilf, Impact of the terminal Cretaceous event on plant-insect associations, Proc. Nat. Acad. Sci. 99(4):2061-2066, 2002.

INDEX OF ARTICLES

When Dinosaurs Became Extinct, What Happened To The Stephen W. Carmichael, Mayo Clinic

A Comparison Of Grain Size Measurements In Al-Cu Thin Films: Imaging Vs. Diffraction Techniques......5 L.M. Gignac,* C.E. Murray,* K.P. Rodbell,* M. Gribelyuk* *IBM T.J. Watson Research Center. +IBM Microelectronics Division

- Using a Sony Cyber-Shot Digital Camera for Photomicrography.....10 Gregor Overney, Agilent Technologies Inc.
- Use Adobe Acrobat to Keep Original Resolutions and to Make TIFF Files From Any Program16 Jerry Sedgewick, University of Minnesota
- Confocal Microscopy System Performance: Laser Power Measurements20 Robert M. Zucker, PhD, U.S. Environmental Protection Agency

Imaging of Shallow Surface Topography by the Low-Loss ElectronMethod in the Scanning Electron Microscope24 Oliver C. Wells, IBM Research Division

New and Interesting at Microscopy & Micranalysis-2002.....28

Bryan R. Hewlett,

McMaster University Medical Centre

James Ekstrom, Phillips Exeter Aca	32
Designing A Microscopy/analytical Step By Step Procedure	

Judy A. Mulphy, San Joaquin Dena Conege	
Preparing Ultra-Smooth SEM Stud Surfaces Dr. Carole Hickman, University of California, Berkeley	42
Protection from Sulfur Hexafluoride Leaks Mick Thomas, Cornell University	42
Industry News	44
Index of Advertisers	46

ABOUT THE COVER

By Gregor Overney Agilent Technologies Inc., California, USA.

See his article on photomicrography with a consumer digital camera on page 10. Photomicrograph of a healthy human kidney section is shown. The tangential section goes through renal corpuscles of the cortical zone. A modified Masson's trichrome stain is used. The digital image is obtained with a SONY DSC-S70 digital camera connected with a Nikon Eclipse E200-F microscope with a 40x CFI on Plan Achromat objective.

MICROSCOPY TODAY November/December 2002 3