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First report of apparent praziquantel resistance in Dipylidium caninum in

Europe

Andreas W. Oehm¹, Anton Reiter², Angela Binz³, Manuela Schnyder¹

¹ Institute of Parasitology, Vetsuisse Faculty of Zurich, University of Zurich, Zurich,

Switzerland

² Dammweg 94, 5000 Aarau, Switzerland

³ Tierklinik Aarau West, Oberentfelden, Switzerland

Corresponding author: Manuela Schnyder, manuela.schnyder@uzh.ch

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Abstract

Dipylidium caninum is a common tapeworm of dogs. Two cases of praziquantel resistance have been described in *D. caninum* in the United States. No further reports have been published to the authors' knowledge. Here, the case of a dog imported to Switzerland from Spain with a history of chronic excretion of tapeworm proglottids and unresponsiveness to praziquantel treatments is reported. Clinical signs were mild (restlessness, tenesmus, anal pruritus, squashy faeces) and flea infestation could be ruled out. Infection with *D. caninum* was confirmed through morphological and genetic parasite identification. Different subsequently applied anthelmintic compounds and protocols, including epsiprantel, did not confer the desired effects. Proglottid shedding only stopped after oral mebendazole administration of 86.2 mg/kg body weight for five consecutive days. Clinical signs resolved and the dog remained coproscopically negative during a follow-up period of almost ten months after the last treatment. This case represents the first reported apparent praziquantel and epsiprantel resistance in *D. caninum* in Europe. Treatment was extremely challenging especially due to the limited availability of efficacious alternative compounds.

Key words: Dipylidium caninum; anthelmintic resistance; praziquantel/epsiprantel; dog; cestode

Introduction

Dipylidium caninum (Cyclophyllidea, Dipylidiidae) is a common intestinal tapeworm of dogs with a worldwide distribution and with zoonotic potential (Gates and Nolan, 2009; Deplazes *et al.*, 2021). Especially infants are prone to become infected after ingestion of infected fleas or lice (Elmonir *et al.*, 2021) which act as intermediate hosts. In dogs, infections remain subclinical, or manifest with mild and unspecific clinical signs in most cases. Clinical signs may include anal pruritus causing an animal to rub its bottom along the ground, diarrhoea, weight loss, general restlessness, or tenesmus (Wani *et al.*, 2015; Saini *et al.*, 2016). The primary drug of choice to combat infections with *D. caninum* in dogs is praziquantel (at a single dose of at least 5 mg/kg body weight (BW) *per os* (p.o.) which usually shows high levels of efficacy (Schroeder *et al.*, 2009; Saini *et al.*, 2016; ESCCAP, 2021).

Up to date, anthelmintic resistance in canine and feline parasites has been of minor relevance and rather confined to regions and limited in scope, whereas it is a considerable problem in livestock and horses (Raza *et al.*, 2018; von Samson-Himmelstjerna *et al.*, 2021). Recently, an increasing frequency of reports on multiple anthelmintic resistance of *Ancylostoma caninum* in dog kennels in North America has been addressed as a major concern (Marsh and Lakritz, 2023). Moreover, two cases of possible praziquantel resistance have been reported in *D. caninum* infected dogs in the United States recently (Jesudoss Chelladurai *et al.*, 2018; Loftus *et al.*, 2022), emphasising the importance of this matter in companion animals as well. In Europe, no cases of anthelmintic resistance in companion animals have been reported yet. This case represents the first description of clinical resistance to praziquantel in *D. caninum* in Europe.

Materials and Methods

Case presentation

A male, mixed breed Can de Chira dog imported from Spain of approximately 10 months of age and weighing 16.7 kg was presented with a history of chronic excretion of tapeworm proglottids after arrival in Switzerland. The dog was probably born in December 2021 around Monzón or Huesca (municipality of Aragon, Spain) and had been picked up from the street at the approximate age of four months together with other dogs of the same age. Subsequently, it had been kept at an animal shelter in Monzón. Clinical signs upon presentation were mild with general restlessness, tenesmus, slight anal pruritus, and occasionally squashy faeces. No evidence of flea infestation was present, and the dog had received treatment against ectoparasites (fluralaner, unknown dose) and helminths (praziquantel, febantel, pyrantel, unknown dose) before entering Switzerland. In Switzerland, the dog had been presented to the primary veterinarian and treated orally with 5.9 mg/kg body weight (BW) praziquantel, 5.9 mg/kg BW pyrantel, 18.0 mg/kg BW febantel (DrontalPlus®, Vétoquinol) twice within a period of two weeks as well as with 14.7 mg/kg BW fluralaner (Bravecto®, MSD Animal Health). Flea prophylaxis with fluralaner was pursued every 12 weeks as recommended by the manufacturer. Subsequently, proglottid excretion ceased, but re-started in an unchanged manner three weeks after the last praziquantel/pyrantel/febantel treatment. Hence, the dog received a single dose of 0.7 mg/kg BW milbemycinoxime and 7.4 mg/kg BW praziquantel (Milbemax®, Elanco Animal Health). Proglottid shedding continued and fenbendazole (Panacur®, MSD Animal Health) was administered orally for five consecutive days at a dose of 44 mg/kg BW. For four days, proglottid shedding ceased and 0.7 mg/kg BW milberrycinoxime and 7.4 mg/kg BW praziquantel (Milberrax®, Elanco Animal Health) were orally given twice within two weeks. Excretion of proglottids continued and 0.7 mg/kg BW milbemycinoxime and 7.4 mg/kg BW praziquantel (Milbemax®, Elanco Animal Health) was given orally once a week for a period of four weeks, yet excretion of cestode segments persisted.

Investigations

The presence of eggs/proglottids was examined via the adhesive tape method (Deplazes *et al.*, 2021) and proglottids were also directly collected from the surface of the faecal samples. Furthermore, a combined sedimentation-flotation using saturated sodium-chloride solution with a specific weight of 1.2 g/cm³ was performed on faecal samples (Deplazes *et al.*, 2021). Proglottids were assessed microscopically and egg packets were pressed out from the proglottids in squash preparations. A multiplex PCR targeting the mitochondrially encoded 12S ribosomal RNA of non-*Echinococcus* cestodes (267 base pairs (bp) (Trachsel *et al.*, 2007) was carried out to molecularly confirm the microscopic diagnosis.

Treatments and follow-up

The patient received 31.1 mg/kg BW pyrantel and 12.0 mg/kg BW epsiprantel (Dosalid[®], Zoetis) in the first place. Next, 50.3 mg/kg BW mebendazole (Lendue Maxi[®], Teknofarma S.r.l.) were administered on five consecutive days. Subsequently, 6.0 mg/kg BW praziquantel, 6.0 mg/kg pyrantel, and 24.0 mg/kg oxantel (Dolpac 10[®], Vétoquinol) was initiated for a total of six days. One month after the last treatment with Dolpac 10[®], a second round of mebendazole (Lendue Maxi[®], Teknofarma S.r.l.) was given for five days, this time at an increased dose of 86.2 mg/kg. Contemporaneously, the animal owner recorded the development of proglottid excretion.

All the compounds mentioned in this section are not commercially available in Switzerland and were purchased in Portugal, Italy, or an international pharmacy after considering the regulations of off-label use in veterinary medicine in Switzerland and following the Swiss compendium of veterinary medicinal products (Tierarzneimittelkompendium CliniPharm, <u>www.vetpharm.uzh.ch</u>). The extension from three to five days of mebendazole treatment was derived from Miro *et al.* (2007). A detailed temporal pattern of anthelmintic treatments and proglottid excretion is compiled in Table 1.

Results

Based on gross morphological features of the proglottids (size of ~ 12mm x 3mm, mature genital organs, double genital pores slightly behind the middle of the lateral margins of each proglottid) and the eggs (capsulated and clustered in packets of $120 - 200 \mu m$, with hexacanth embryo), D. caninum was diagnosed (Deplazes et al., 2021) (Fig. 1). PCR confirmed an infection with D. caninum: a DNA segment of 259 bp was successfully sequenced and nucleotide basic local alignment search tool (BLAST) against the nucleotide collection of GenBank showed an identity of 98.07% (254/259 bp) with D. caninum from a canine host MH182479.1) (Jesudoss Chelladurai et al., (accession number 2018). After epsiprantel/pyrantel (Dosalid®, Zoetis) treatment, proglottid shedding continued in an unchanged manner (Tab. 1). Mebendazole at a dose of 50.3 mg/kg BW on five consecutive days reduced the quantity of the excreted tapeworm segments. These were not superficially located on the faeces anymore but mixed within the faecal matter. Moreover, the proglottids appeared macerated and without motility. Re-appearance on the surface, however, occurred final administration of mebendazole. weeks after the The combination two praziquantel/pyrantel/oxantel (Dolpac 10[®], Vétoquinol) did not stop the excretion of tapeworm segments. Eventually, proglottid shedding stopped from the second day of treatment with mebendazole 86.2 mg/kg BW (Tab. 1) and clinical signs (general restlessness, tenesmus, slight anal pruritus, occasionally squashy faeces) resolved. Moreover, the shedding of proglottids stopped and the dog remained coproscopically negative for a follow-up period of ten months.

Discussion

This case represents the first written report of apparent praziquantel resistance in *D. caninum* in Europe. During the investigations for potential alternative anthelmintic treatments, further oral reports from veterinary parasitologists of Italy and Spain were mentioned to the authors (M. Schnyder, personal communication). To confirm that the infection was indeed caused by *D. caninum*, a combination of traditional parasitological and genetic methods was implemented.

Anthelmintic resistance is defined as the ability of helminth parasites to survive the administration of a certain previously effective drug (Prichard et al., 1980; Sangster et al., 2018). According to the VICH (Veterinary International Conference on Harmonization) guidelines (VICH, 2000, 2001), efficacy is described as a reduction of \geq 90% of *D. caninum* scolices in controlled terminal studies. In the current case resistance may be suspected by the fact that multiple doses of praziquantel at the standard label treatment dose of 5 mg/kg BW (Schmid et al., 2010) did not stop the continuous excretion of proglottids. Investigating the above-mentioned threshold for a drug to be considered efficacious in a controlled study would have required experimental infections: several factors (i.e. deriving from the individual dog, or being related to the environment, etc.) may in fact influence the presence and quantity of proglottids in faeces. Therefore, the absence of proglottids in this case did not allow to finally conclude on the efficacy of an anthelmintic agent, still, this represented the goal for the animal owner. Consequently, in the context of the current case, it was not possible to establish the resistance of the isolate by experimental infection of laboratory animals to further fathom the nature of this resistance, comparable to Jesudoss Chelladurai et al. (2018), who did also not establish the isolates in experimentally infected laboratory animals for further investigations either. The efficacy of anthelmintic treatment in the here reported case was instead

continuously assessed by documenting the presence and quantity of proglottids excreted in faeces and by final determination of egg absence in faeces and by adhesive tape method. Accordingly, the isolate was not eliminated by the administration of praziquantel at the label dose of 5 mg/kg BW (Lloyd and Gemmell, 1992; Altreuther et al.; 2009, Schroeder et al., 2009) nor by fenbendazole at a dosage of 44 mg/kg BW (Burke and Roberson, 1978) nor by epsiprantel (combined with pyrantel) at a dose of 12.0 mg/kg BW, i.e. more than twice as high as the recommended dose of 5.5 mg/kg BW (Corwin et al., 1989). Interestingly, oral administration of praziquantel/pyrantel/febantel twice within a period of two weeks induced a 3-week suspension of proglottid excretion, suggesting some efficacy. The administration of praziquantel/pyrantel/oxantel had previously been successful in D. caninum infections (Grandemange et al., 2007, Jesudoss Chelladurai et al., 2018) and it was hypothesised that oxantel would exert a synergistic effect with the other compounds due to differences in drug action (Martin et al., 2004, Jesudoss Chelladurai et al., 2018). Yet, no reduction in the excretion of proglottids was noticed. In contrast, with mebendazole at the dosage of 50.3 mg/kg BW for five days, a clear reduction of the number of proglottids on faecal samples was observed, but single proglottids were present also after the last day of treatment. Previous studies have indicated an efficacy of oral mebendazole against cestodes (Vanparijs and Thienpont, 1973; Genchi et al., 1990). Side effects of mebendazole such as vomiting and diarrhoea can occur already when the apeutic doses are administered. Moreover, the compound has been associated with hepatotoxicity visible as icterus, depression or anorexia and side effects commonly occur one day up until two weeks after administration (Polzin et al., 1981; Swanson and Breider, 1982). In the current case, the dog was closely monitored, and no side effects of any kind were observed even when mebendazole was given at the increased dose. A thorough surveillance of dogs being treated with this compound seems reasonable, especially if administered off-label. As mebendazole had at least induced an apparent reduction of the number of excreted

proglottids, it was decided to increase the dose to 86.2 mg/kg BW. This eventually turned out to be effective after the failure of multiple agents as well as mebendazole at a lower dose. However, it cannot be excluded that shedding of proglottids stopped due to reaching of the natural lifetime of the parasite (Jesudoss Chelladurai *et al.*, 2018). Yet, the macroscopic appearance of the proglottids and their motility evidently changed after initiation of mebendazole administration supporting the resistance of this *D. caninum* isolate to previous anthelmintic treatments and regardless of potential natural termination of infection.

One aspect to consider was the potential of flea infestation as a source of continuous reinfection of the patient. Consequently, the inability to eliminate the parasite could have been mistakenly interpreted as anthelmintic resistance with only temporary elimination of the infection. Yet, reinfection due to continued flea infestation of the dog appeared extremely unlikely as the dog was repeatedly treated against ectoparasites and in accordance with the information provided by the manufacturer the treatments were still exerting their fleainsecticidal activity during the investigations and their follow-up. Moreover, owners implemented intensive environmental surveillance for potential presence of fleas as well as environmental decontamination, although indications of flea infestation were absent. This is further corroborated by the fact that the dog remained negative for a follow-up period of more than three months after the last anthelmintic treatment as well as for an extended follow-up period of almost 10 months.

Owner compliance is an obviously relevant aspect in the context of suspected drug resistance when the administration of anthelmintics is delegated to the pet owner. This may include anthelmintics that are not administered in the right dose or frequency (Jesudoss Chelladurai *et al.*, 2018) or if the animal, unobserved, expels orally administered drugs. In the present case, owner compliance, involvement, and engagement were exceptionally high and due to the background of the owner as a medical practitioner, correct observation of the case,

meticulous documentation as well as appropriate administration of medications was ensured.

Infections with *D. caninum* in humans are rare, associated with mild clinical signs such as discomfort or gastrointestinal disturbances (Taylor and Zitzmann, 2011; Portokalidou *et al.*, 2018), and mainly limited to cases where flea infestations and oral ingestion of fleas are present, e.g. in infants (Chappell *et al.*, 1990; Molina *et al.*, 2003; Jesudoss Chelladurai *et al.*, 2018). Given the interconnectedness of human and animal health, the emergence of anthelmintic resistance in *D. caninum* could pose a minimal risk to human health as well.

Conclusions

Future investigations are necessary to understand the extent of potential anthelmintic resistance present in *D. caninum* infecting dogs and cats, and the mechanisms involved to confer this resistance. Importantly, notifying the occurrence of similar cases with pharmacovigilance authorities will contribute to better data collection. Major challenges are represented by the limited availability of alternative effective compounds and their restricted availability depending on the country.

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Methodology, MS, AR, AOE; Resources, MS, AR; writing-original draft, AOE, MS; writing – review and editing, AR, AB, AOE, MS. All authors have read and agreed to the submitted version of the article.

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Ethical standards. Informed and written consent for the anthelmintic treatments and the coproscopic analyses was obtained from the animal owner.



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Figure 1. Eggs clustered in packets with hexacanth embryo typical for *Dipylidium caninum*.

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Table 1. Observed proglottid shedding and administered medications after antiparasitic treatments with pyrantel, febantel, milbemycinoxime, fenbendazole and fluralaner, and unsuccessful treatments with praziquantel administered to a dog infected with *Dipylidium caninum*. Proglottid shedding stopped for at least 10 months after the last mebendazole administration.

Date	Number of proglottids Assessment time		Anthelmintic treatments (compounds, commercial
			name, company, country of origin)
	Morning	Afternoon/	
		Evening	×
16/02/2023	Not observed	1	*.·O`
17/02/2023	9	0	
18/02/2023	10	0	C
19/02/2023	1	3	S
20/02/2023	13	0	
21/02/2023	5	0	
22/02/2023	16	1	
23/02/2023	0	>20	
24/02/2023	8	0	
25/02/2023	8	0	
26/02/2023	>16	9	
27/02/2023	12	6	Epsiprantel, 12.0 mg/kg BW /
			Pyrantel 31.1 mg/kg BW p.o. (Dosalid [®] , Zoetis,
28/02/2023		0	Portugal)
01/03/2023	4	0	
02/03/2023	9	2	
03/03/2023	5	0	
04/03/2023	15	0	
05/03/2023	21	Not observed	
06/03/2023	13	0	Mebendazole 50.3 mg/kg BW p.o.
07/03/2023	>7 ^b	0	Mebendazole 50.3 mg/kg BW p.o. (Lendue
			Maxi [®] , Teknofarma S.r.l., Italy)
08/03/2023	0	0	Mebendazole 50.3 mg/kg BW p.o.
09/03/2023	0	0	Mebendazole 50.3 mg/kg BW p.o.
10/03/2023	0	0	Mebendazole 50.3 mg/kg BW p.o.
23/03.2023	1	0	
24/03.2023	0	0	
25/03.2023	1	0	

26/03.2023	1	0	
27/03.2023	3	0	
28/03.2023	2	0	
29/03.2023	2	0	
30/03.2023	2	0	
01/04/2023	0	0	
02/04/2023	0	0	
03/04/2023	0	0	
04/04/2023	8	0	Praziquantel 6.0 mg/kg BW
			Pyrantel 6.0 mg/kg BW
			Oxantel 24.0 mg/kg p.o. (Dolpac 10 [®] , Vétoquinol, international pharmacy)
05/04/2023	5	5	Praziquantel 6.0 mg/kg BW
			Pyrantel 6.0 mg/kg BW
06/04/2023	2	2	Oxantel 24.0 mg/kg p.o.
00/04/2023	2	Z	Praziquantel 6.0 mg/kg BW Pyrantel 6.0 mg/kg BW
			Oxantel 24.0 mg/kg p.o.
07/04/2023	2	0	Praziquantel 6.0 mg/kg BW
			Pyrantel 6.0 mg/kg BW Oxantel 24.0 mg/kg p.o.
08/04/2023	4	1	Praziquantel 6.0 mg/kg BW
			Pyrantel 6.0 mg/kg BW
09/04/2023	8	0	Oxantel 24.0 mg/kg p.o. Praziquantel 6.0 mg/kg BW
0710412025	0	0	Pyrantel 6.0 mg/kg BW
		U	Oxantel 24.0 mg/kg p.o.
10/04/2023	0	0	
11/04/2023	20	0	
12/04/2023	0	0	
13/04/2023	0	0	
14/04/2023	6	0	
15/04/2023	7	0	
16/04/2023	9	0	
17/04/2023	4	4	
18/04/2023	9	0	
19/04/2023	28	0	
22/04/2023	9	0	
23/04/2023	5	0	
24/04/2023	10	0	
25/04/2023	15	0	
26/04/2023	1	0	
27/04/2023	10	0	
28/04/2023	2	0	

29/04/2023	10	0	
30/04/2023	11	0	
01/05/2023	10	0	
02/05/2023	1	0	
03/05/2023	8	0	
04/05/2023	8	0	
05/05/2023	7	0	
06/05/2023	5	0	
07/05/2023	5	0	
08/05/2023	8	0	
09/05/2023	6	0	
10/05/2023	3	0	×
11/05/2023	5	0	Mebendazole 86.2 mg/kg BW p.o. (Lendue
			Maxi [®] , Teknofarma S.r.l., Italy)
12/05/2023	0	0	Mebendazole 86.2 mg/kg BW p.o.
13/05/2023	0	0	Mebendazole 86.2 mg/kg BW p.o.
14/05/2023	0	0	Mebendazole 86.2 mg/kg BW p.o.
15/05/2023	0	0	Mebendazole 86.2 mg/kg BW p.o.
16/05/2023	0	0	Mebendazole 28.7 mg/kg BW p.o.

^aBW body weight

^b at his point, proglottids were not superficially located on the faeces but rather mixed within the faecal matter and they seem macerated without motility

Accepteo