

Monitoring of seasonality of norovirus and other enteric viruses in Cameroon by real-time PCR: an exploratory study

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SUMMARY

We studied the seasonal fluctuation of norovirus and other enteric viruses in Cameroon. Two hundred participants aged between 1 and 69 years were prospectively followed up. Each participant provided monthly faecal samples over a 12-month period. A total of 2484 samples were tested using multiplex real-time PCR assay for the detection of norovirus, rotavirus and enterovirus. The effect of weather variables and risk factors were analysed by Pearson correlation and bivariate analysis. Overall, enterovirus was the most commonly detected virus (21.6% of specimens), followed by norovirus (3.9%) and rotavirus (0.4%). Norovirus and enterovirus were detected throughout the year with a peak of norovirus detection at the beginning of the rainy season and a significant alternation of circulation of norovirus genogroups from one month to the next. Age <5 years and consumption of tap water were risk factors for norovirus infection. Better understanding of factors influencing transmission and seasonality may provide insights into the relationship between physical environment and risk of infection for these viruses.

Key words: Enterovirus, epidemiology, Norwalk agent and related viruses, rotavirus, virus infection.

INTRODUCTION

Noroviruses, rotaviruses, and enteroviruses are among the most diverse and clinically important enteric viruses [1–3]. Norovirus is the most frequent cause of non-bacterial gastroenteritis in both children and adults (in industrialized countries) [4, 5]. Usually, the infection is self-limiting and symptoms subside within 2–3 days [6]. However, severe forms of the

disease may occur in young children, the elderly and immunocompromised individuals [7, 8]. Rotaviruses are considered to be the major cause of severe infantile diarrhoea worldwide, and are thought to be responsible for 60% of all diarrhoeal episodes in developing countries and 40% in developed countries [9–11]. Both rotavirus and norovirus show marked wintertime seasonality in temperate countries [12, 13].

Enteroviruses are second only to rhinoviruses as the most common viral infectious agents in humans [14]. Most enterovirus infections are typically mild or subclinical. The presentation of symptomatic infection is very variable, including hand, foot and mouth

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disease, myocarditis, aseptic meningitis, encephalitis and acute flaccid paralysis [15–17]. A characteristic feature in the epidemiology of these viruses is their seasonality, with peaks in late summer or early autumn in temperate regions [18].

Thus, these three enteric viruses all have seasonal epidemiology. The seasonality is less pronounced in Sub-Saharan Africa, and the climate variables influencing their transmission or other factors explaining this seasonal pattern are not well understood, in part due to the fact that laboratory capacity for their detection is often lacking.

These viruses are highly transmissible, primarily via the faecal–oral route, contaminated food or water or person-to-person transmission via airborne droplets [19, 20]. Waterborne transmission may occur in the context of contamination of drinking or recreational water by waste water [21]. The waterborne route constitutes a significant mode of transmission of these viruses, especially of norovirus genogroup GI while norovirus GII is mostly foodborne and also a common cause of outbreaks in the hospital setting [22]. As climate change in the form of increased precipitation sets in, it is estimated that the incidence of waterborne infectious diseases will increase [23]. Therefore, knowledge of the seasonality and influence on weather condition on the transmission of these viruses in the community could provide the information necessary to alert healthcare providers on when a peak period of infection in the community is likely to occur. This will also be important for prioritization of public health actions and programme planning interventions.

Over the years, molecular diagnostic methods such as real-time polymerase chain reaction (PCR) have improved viral detection and have often replaced traditional diagnostics such as antigen detection by immunofluorescence or enzyme immunoassays [24]. The recent advances of multiplex real-time PCR based on primers and probes designed to bind and amplify specific but conserved genomic regions of target genes has grown in importance due to their higher sensitivity. These tests can also detect more than one pathogen in a single reaction, thereby reducing cost and time [25]. The aim of this prospective study was to determine the seasonal fluctuations of the prevalence of norovirus, rotavirus and enterovirus in children and adults in Cameroon, as assayed by a highly sensitive molecular diagnostic assay. We also analysed the potential impact of weather variables such as temperature, relative humidity and rainfall,

as well as demographic and other risk factors for infection. Better understanding of factors enhancing transmission and seasonality may provide insights into the relationship between physical environment and risk of infection of these viruses.

METHODS

Study region

The study was conducted in Limbe, located at the foot of Mount Cameroon, close to the Atlantic Ocean, with a population of about 84 000 (Fig. 1). Detailed demographic data on this study population was not available. The Limbe equatorial climate experiences the dry season from December to May, and the rainy season with heavy rainfall between June and November, with the peak period in August. Temperatures range from 23 °C to 33 °C and relative humidity often reaches as high as 86%. Sources of drinking water are mainly tap water and borehole wells.

Participants and longitudinal sampling

We conducted a prospective longitudinal exploratory study over a period of 1 year (from September 2011 to August 2012). The age distribution of participants was children aged 1–17 years (mean age 6.4 years) and adults aged 18–69 years (mean age 32 years) (Table 1). Inclusion criteria included healthy participants without any signs or symptoms of acute gastroenteritis and not receiving chemotherapy at the time of recruitment. A sample size calculation based on the population size and 95% confidence interval (CI) was used to determine the sample size. A two-stage sampling strategy was used. First, there was random selection of the households and then random sampling of child and adult participants in the household.

A total of 154 children and 146 adults were randomly selected, of which 115 and 111 children and adults, respectively, met the inclusion criteria and were recruited. In the course of the study 13 children and 11 adults dropped out and two children died. Each participant received monetary remuneration of US\$48 to discourage dropping out of the study. All participants visited the centre and were given containers to provide faecal samples. Samples were separated into 2-ml aliquots and stored at –20 °C prior to analysis.

One year surveillance data of both children and adults was analysed (each adult participant was

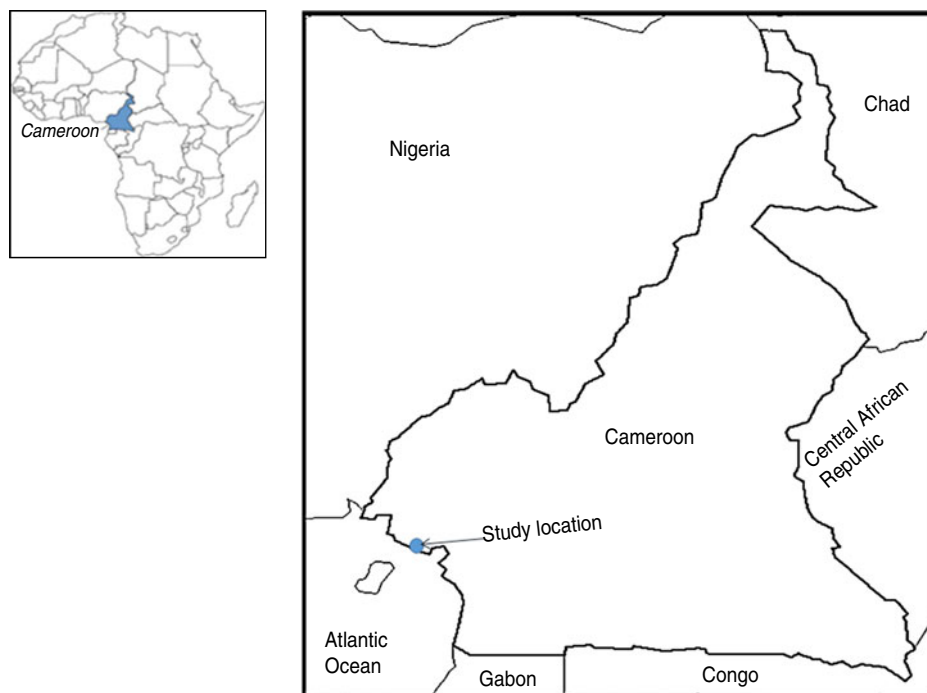


Fig. 1 [colour online]. Map of Cameroon with neighbouring countries showing study location. Top left panel is a map of Africa showing the location of Cameroon.

Table 1. Characteristics of study participants in Limbe, Cameroon, September 2011–August 2012

Variable	Age group (years)	(N=200) n (%)	Mean age (years)
Children	1–5	47 (23.5)	6.4
	6–17	53 (26.5)	
Adults	18–69	100 (50.0)	32
Gender			
Male		97 (48.5)	
Female		103 (51.5)	
Source of water*			
Tap water only		50 (25.0)	
Borehole wells		150 (75.0)	

* Ten participants used both tap water and borehole wells and were classified under use of borehole wells.

matched with a child from the same household). Surveillance included monthly collection of a faecal specimen. Each participant responded to questions in a questionnaire regarding age, gender, household size, and source of domestic water. It was observed that 50 participants used tap water only while 140 participants used borehole wells only. An additional 10 participants used both tap water and borehole wells.

The surveillance protocol was reviewed and approved by the South West Regional Delegation of

Public Health in Cameroon, and participants (parents or guardians of children) provided written or oral informed consent.

Meteorological data collection

Weather information was recorded according to guidelines of the World Meteorological Organization [26]. Briefly, average daily temperature and relative humidity were recorded. Monthly mean values were calculated by adding the daily averages and dividing by the number of days in each calendar month. Total daily rainfall on the other hand was recorded and added up to give the monthly rainfall.

Stool preparation and nucleic acid extraction

Stool suspensions were prepared by adding 1 g stool sample to 5 ml phosphate-buffered saline containing 1 g glass beads (Corning Inc., USA). The mixture was shaken for 2 min, and centrifuged at 1500 g for 25 min at 4 °C. Stool suspensions (supernatant) were aspirated and stored in aliquots. Nucleic acid (NA) from 130 µl faecal sample was extracted into an elution volume of 100 µl by the MagNA Pure LC robot (Roche Molecular Systems, Germany), using the Total Nucleic Acid external lysis protocol.

Table 2. Primers and probes used in the panel of real-time PCR tests

Mix	Target	Primer/ probe	Sequence (5'–3')*	Source
A	Norovirus GI	FP	TGGCAGGCCATGTTCCGCT	[27]
		RP1	CGCTTGATGTAGCGTCCTTAGAC	
		RP2	TTTGKTGGGGCGTCCTTAGAC	
		Probe	VIC-ATTGCGATCTCCTGTCCA-MGB	
B	Norovirus GII Rotavirus	FP	TGGAYTTTTAYGTGCCAG	[27]
		RP	CCACGCCATCTTCATTCAC	[28]
		Probe	VIC-AGCCAGATTGCGATCGCCC-TAMRA	
		FP1	AACCATCTACACATGACCCTCTATGA	
		FP2	AACCATCTTCACGTAACCCTCTATGA	
		RP	GGTCACATAACGCCCTATAGC	
		Probe	FAM-CAATAGTTAAAAGCTAACACTGTCAAAA-MGB	
C	Enterovirus	FP	IGGTGYGAAGAGICTATTGAGCTA	[28]
		RP	GGACACCCAAAGTAGTCGGTTC	
		Probe	VIC-CGGCCCTGAATGCGGCTAATC-TAMRA	

* FAM, 6-carboxyfluorescein; TAMRA, 6-carboxytetramethylrhodamine; MGB, minor groove-binding quencher; VIC, fluorescent label (Applied Biosystems, USA).

Real-time PCR

All assays were run on an ABI 7300 real-time PCR platform (Applied Biosystems, USA). The target viruses were detected in 25 μ l reaction volumes containing 5 μ l NA, 13 μ l 2 \times reaction mix with ROX (Invitrogen Ltd, UK), 0.5 μ M each primer and probe, 20 U RNase OUT and 0.5 μ l Superscript III platinum one-step, following the manufacturer's instructions [27]. After a reverse transcription step at 48 °C for 25 min and initial denaturation at 95 °C for 10 min, 45 cycles of two-step (95 °C for 15 s, 60 °C for 60 s) PCR was performed in three parallel reactions, targeting four different viruses as described in Table 2. A cycle threshold (C_t) value <40 indicated that the specimen was positive. This method enables the semi-quantitative detection of target viruses, with the C_t value being inversely proportional to the amount of virus in each sample [25]. The limit of detection was about 50 copies per reaction as determined by serial dilution of the plasmid-containing inserts of the target region. The quality of the assays was ensured by performing parallel runs of positive and negative controls samples.

Statistical analysis

Monthly prevalence was calculated as the number of monthly cases of infection divided by the sample size multiplied by 100. Pearson regression analysis was performed to evaluate the effect of meteorological parameters and incidence of infection, and a bivariate

analysis was performed to determine the relationship of demographic and risk factors for infection. Two-sided Fisher's test was used to compare proportions with alpha set at 0.05. Odd ratios (ORs) and 95% CIs were calculated using the SPSS software package v. 17.0 for Mac (SPSS Inc., USA).

RESULTS

Demographic characteristics of participants

During the 12-month study period from September 2011 to August 2012, 200 participants were prospectively followed-up; 48.5% were males and 50% adults aged between 18 and 69 years. Children aged 1–17 years constituted 50% of the study population. The mean age of adults and children was 32 years and 6.4 years, respectively (Table 1). A total of 2484 faecal samples were collected; 1298 (52.2%) during the dry season and 1186 (47.8%) during the rainy season.

Virus detection and risk factors

Figure 2(a, b) shows the temporal pattern of virus detection. Enterovirus was the most frequently identified virus and was detected with a monthly prevalence ranging from 8% to 47% in children and 1–31% in adults. The prevalence of norovirus ranged from 1% to 16% in children and 1–12% in adults. In contrast, rotavirus detection was low with prevalence between 0% and 3% (Fig. 2a, b). In a bivariate analysis,

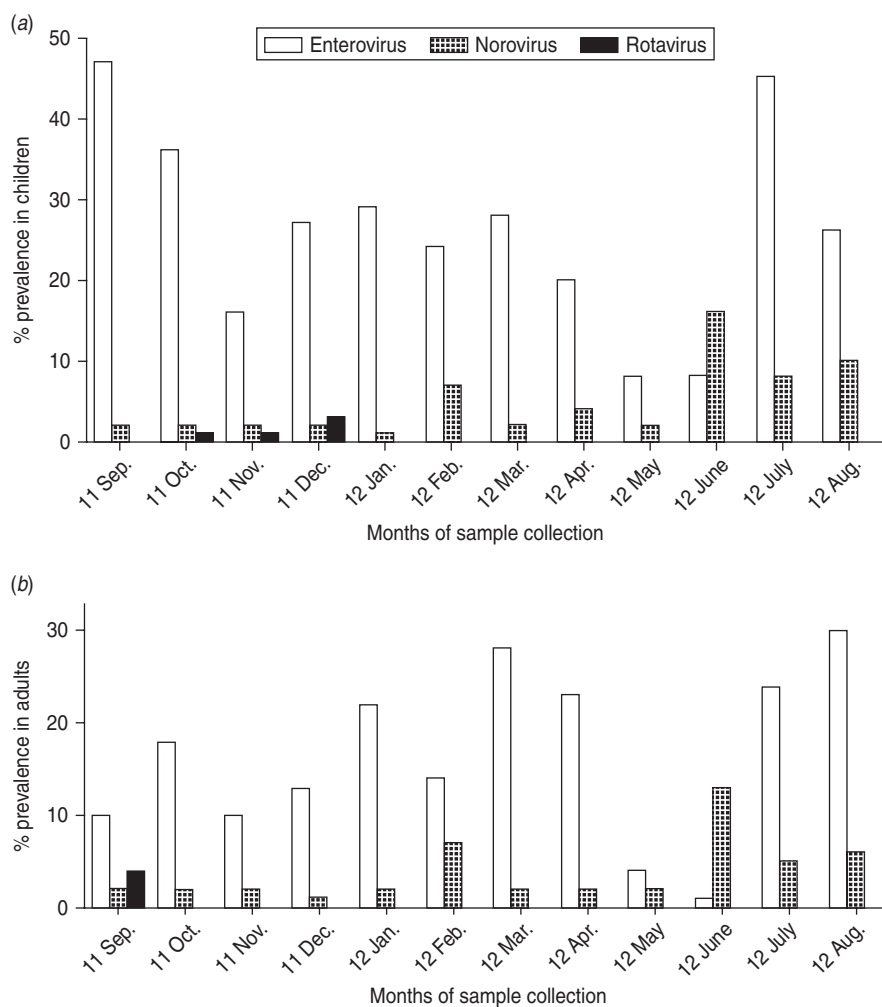


Fig. 2. Proportion (%) of samples positive for norovirus, rotavirus and enterovirus by real-time PCR in (a) children and (b) adults in Limbe, Cameroon, sampled monthly from September 2011 to August 2012.

we observed that children aged <5 years were associated with an increased risk of norovirus infection. Risk of infection was also higher due to consumption of tap water by both children and adults (Table 3). Older children aged 6–17 years were also at higher risk of enterovirus infection. No significant differences in norovirus, rotavirus and enterovirus infection between males and females was observed.

Seasonality

We observed norovirus and enterovirus circulation throughout the study period in both children and adults. However, a markedly low enterovirus activity was observed in May and June. Norovirus peak prevalences were observed from June to August (Fig. 3). This peak of norovirus circulation coincided with an increase in total rainfall (Fig. 4). Few rotavirus

infections were detected during September–December with no further detection during January–August. Rotavirus was therefore excluded from the statistical analyses. In all, detection of norovirus infection was statistically higher in the rainy season than in the dry season for both children and adults ($P=0.02$) (Table 4). A weak positive correlation of temperature and relative humidity with respect to norovirus, rotavirus and enterovirus infections was observed ($r \leq 0.4$, $P > 0.05$).

Demographic and seasonal pattern of norovirus genogroups GI and GII circulation

Out of the 100 cases of norovirus detected during the study period, 45 were of GI and 55 were of GII. Moreover, out of the 55 cases of GII, 30 were detected in children and 25 in adults. There was no significant

Table 3. Risk factors for norovirus, rotavirus and enterovirus infection in Limbe Cameroon, September 2011–August 2012

Variable	Total	Norovirus (<i>n</i> = 100)		Enterovirus (<i>n</i> = 160)		Rotavirus (<i>n</i> = 9) <i>n</i> (%)
		<i>n</i>	OR (95% CI)	<i>n</i>	OR (95% CI)	
Age (years)*						
1–5	47	31	2.3 (1.2–4.6)†	34	0.5 (0.2–1.2)	3 (6.3)
6–17	53	28	1.1 (0.6–2.1)	48	3.0 (1.1–8.1)†	2 (3.7)
18–69	100	41	0.4 (0.2–0.8)	78	0.7 (0.3–1.5)	4 (4.0)
Gender						
Male	97	50	1.1 (0.6–1.9)	72	0.5 (0.2–1.0)	5 (5.1)
Female	103	50	Reference	88	Reference	4 (3.8)
Water source						
Tap water	50	33	2.4 (1.2–4.7)†	41	1.1 (0.5–2.7)	1 (2.0)
Borehole wells	150	67	Reference	119	Reference	8 (5.3)

OR, Odds ratio; CI, confidence interval.

* To calculate the odds of infection for an age group, the rest of the other age groups combined were used as the reference. Both demographic and risk factors for infection were determined by bivariate analyses.

† Statistically significant data ($P < 0.05$), rotavirus data were sparse, and were thus excluded from analyses.

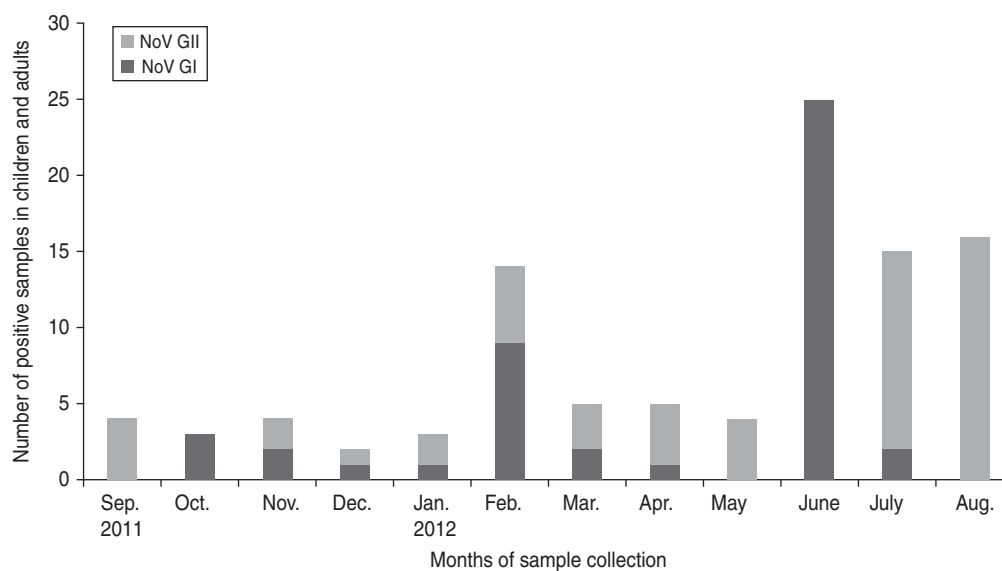


Fig. 3. Seasonal pattern of the prevalence of norovirus (NoV) genogroups GI and GII in children and adults in Limbe, Cameroon sampled from September 2011 to August 2012.

difference in infection by norovirus genogroups GI and GII in children and adults ($P = 0.7$). Although the two genogroups of norovirus circulated throughout the year, each month was characterized by the predominance of a particular genogroup. In June a predominance of GI was observed while a shift to GII predominance was seen during the subsequent months from July to September and then replaced again by genogroup GI in October (Fig. 3).

DISCUSSION

In this prospective study, the seasonality of norovirus, rotavirus, and enterovirus in Cameroon was investigated. The high prevalence of enterovirus in this study is consistent with that of our previous report [28]; however, our enterovirus seasonality is in slight contrast with a report by Njouom *et al.* [29] who found that enterovirus in Cameroon was mostly

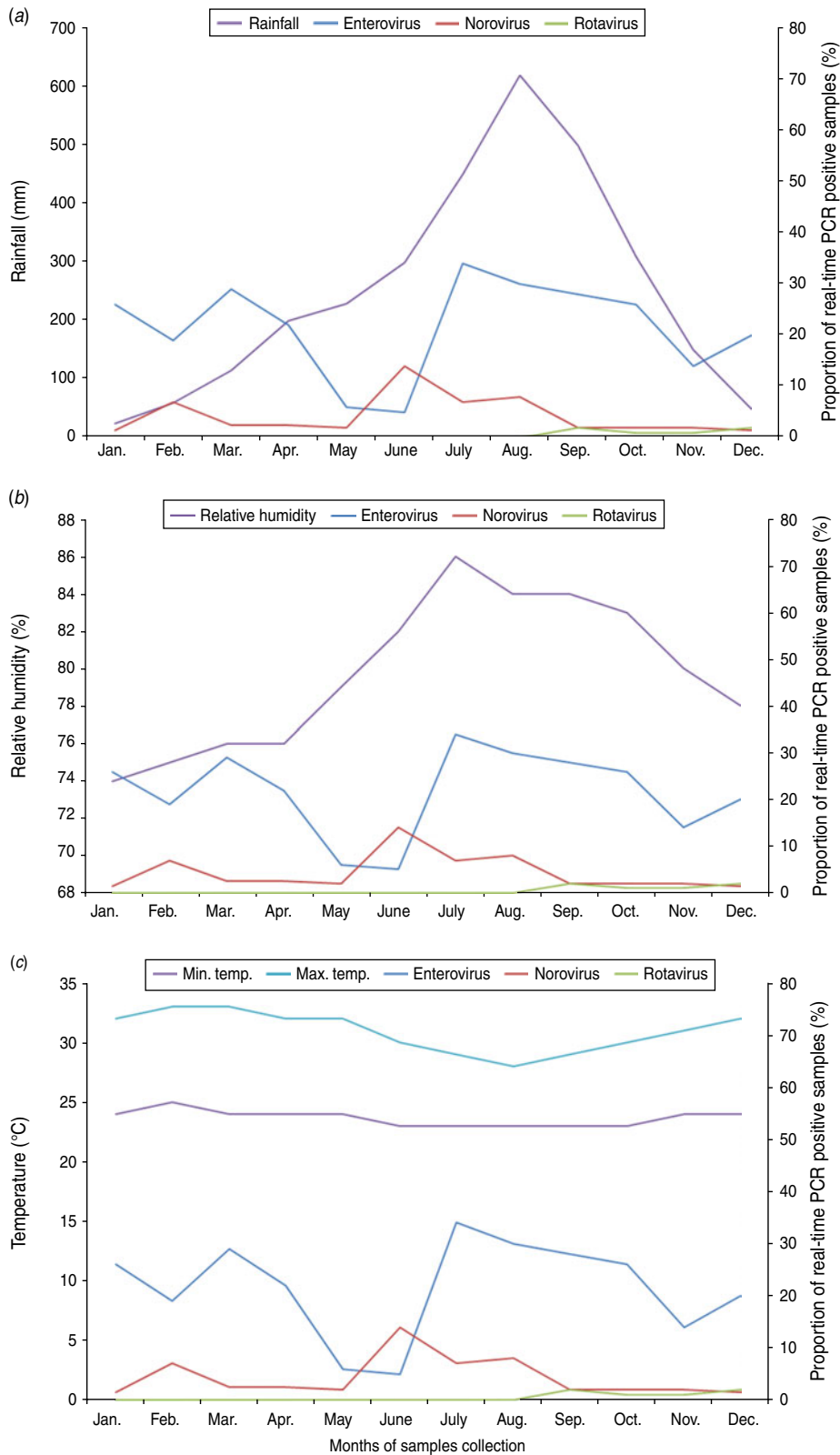


Fig. 4. Variation of weather factors and real-time PCR positivity (%) of norovirus, rotavirus and enterovirus in Limbe, Cameroon from September 2011 to August 2012. (a) Rainfall, (b) relative humidity, (c) temperature. Norovirus and enterovirus prevalence correlated positively with rainfall ($r \geq 0.6$, $P < 0.05$). Moreover, the correlation with relative humidity and temperature was positive although weak ($r \leq 0.4$, $P > 0.05$).

Table 4. *Norovirus, rotavirus and enterovirus detection during the dry and rainy season in Limbe, Cameroon, September 2011–August 2012*

Participants*	Season†	No. of samples	Viral agents‡				
			Norovirus		Enterovirus		Rotavirus
			<i>n</i> (%)	OR (95% CI)	<i>n</i> (%)	OR (95% CI)	
Children	Rainy	607	39 (6.4)	2.4 (1.3–4.2)	182 (30.0)	1.4 (1.0–1.7)	2 (0.3)
	Dry	650	18 (2.7)	Reference	154 (23.6)	Reference	3 (0.4)
	Total	1257	57 (4.5)		336 (26.7)		5 (0.4)
	<i>P</i> value		0.002		0.012		
Adults	Rainy	579	28 (4.8)	2.1 (1.1–4.0)	91 (15.7)	0.9 (0.6–1.2)	4 (0.7)
	Dry	648	15 (2.3)	Reference	111 (17.1)	Reference	0 (0.0)
	Total	1227	43 (3.5)		202 (16.4)		4 (0.3)
	<i>P</i> value		0.019		0.53		

OR, Odds ratio; CI, confidence interval.

* Children (1–17 years), adults (18–69 years).

† Dry season (December–May), rainy season (June–November).

‡ $P < 0.05$ was considered to be statistically significant (two-sided Fisher's test). Rotavirus data were sparse, and were thus excluded from analyses.

detected from January to June. We found an all-year-round circulation of enterovirus with relatively low activity in May and June. This difference in enterovirus seasonality between the two studies could be due to difference in specimen type. Njouom *et al.* investigated respiratory enterovirus infections (by throat and nasopharyngeal swab sampling), and these infections may well have a different epidemiology and seasonality than enteric enterovirus infections [30].

Similarly norovirus circulation was observed throughout the year with a major peak during the beginning of the rainy season from June to August. This is the first comprehensive prospective study of norovirus seasonality in tropical Africa. Our observation that circulation of norovirus occurs throughout the year is in accord with other surveys [31, 32]. However, unlike the cold weather peak demonstrated in most studies performed in temperate regions of the Northern Hemisphere [13, 33], our data demonstrate a sharp increase in norovirus prevalence, dominated by GI, at the beginning of the rainy season. Since this virus is highly stable in the environment, abundant rainfall is likely to wash sewage and household waste into the environment, thereby favouring norovirus transmission [34].

In contrast to our expectation, an increased risk of norovirus infection was associated with consumption of tap water. It should be noted that the municipality

supplies tap water to the homes. The stream, which supplies the municipality water treatment plant, is likely to be contaminated upstream by human sewage containing viral particles. Moreover, considering that norovirus is fairly resistant to chlorine disinfection which is used for tap water [35, 36], it is plausible that water from borehole wells may be relatively safer for consumption with regard to norovirus infection in Limbe, Cameroon. It should be noted that human norovirus infections are caused by genogroups GI, GII and to a lesser extent by GIV [37]. Genogroup GII is the most common cause of norovirus outbreaks worldwide and these infections peak during the winter season in temperate regions [13], and are often designated as 'winter vomiting disease'. On the other hand, GI viruses have been linked to outbreaks associated with contaminated water source [38]. In our study, the significant increase in norovirus GI at the beginning of the rainy season showed a different seasonality to that described in temperate regions, and supported the notion of water as the main vehicle of transmission of these viruses. The results of the current study failed to show a statistical difference in norovirus genogroup infection between children and adults, and between males and females. Average temperature and relative humidity also correlated positively with an increase in the prevalence of norovirus and enterovirus infection, although the correlation was weak.

Overall, rotavirus detection was low and restricted to the end of the rainy season and the beginning of the dry season (September–December). This finding is compatible with a report by Oluwatoyin Japhet *et al.* in Nigeria who found a peak of rotavirus infection from October to January [39]. Considering the variability of incidence of enteric viruses within the same population during a period of 1 year, results of epidemiological studies at a single time point should be interpreted with caution as different periods of the year may show different patterns of infection.

Several limitations may be underscored in the current study. Perhaps the most compelling is that the study period of 1 year does not allow testing the consistency of seasonal trends of these viruses, which may change from one year to another. Second, the study does not distinguish between symptomatic and asymptomatic infections with these viruses because our focus was on seasonality and meteorological influence on transmission. Third, due to the small sample size and the very few cases of rotavirus detected, rigorous statistical analyses were lacking.

Despite these limitations, this study is the first of its kind in Sub-Saharan Africa and extends knowledge on the seasonality and risk factors of norovirus, rotavirus, and enterovirus infection in South West Cameroon and suggests that rainfall significantly increases the transmission of noroviruses and enteroviruses in Limbe, Cameroon. Taken together, we hypothesize that as climate change occurs with an increase in precipitation, high incidence of norovirus and enterovirus infection may occur and a corresponding increase in the burden of disease due to these infections is likely.

These data may provide valuable information necessary to alert healthcare providers on when a peak period of infection in the community is likely to occur. Further studies with a larger sample size and covering a greater area in Cameroon are needed to test the consistency of this seasonal trend and to evaluate the potential contribution of other risk factors for enteric virus infections.

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DECLARATION OF INTEREST

None.

REFERENCES

1. **Wilhelmi I, Roman E, Sanchez-Fauquier A.** Viruses causing gastroenteritis. *Clinical Microbiology and Infection* 2003; **9**: 247–262.
2. **Tapparel C, et al.** Picornavirus and enterovirus diversity with associated human diseases. *Infection, Genetics and Evolution* 2013; **14**: 282–293.
3. **Hansman GS, et al.** Genetic and antigenic diversity among noroviruses. *Journal of General Virology* 2006; **87**: 909–919.
4. **Lopman BA, et al.** Viral gastroenteritis outbreaks in Europe, 1995–2000. *Emerging Infectious Diseases* 2003; **9**: 90–96.
5. **Kaplan JE, et al.** Epidemiology of Norwalk gastroenteritis and the role of Norwalk virus in outbreaks of acute nonbacterial gastroenteritis. *Annals of Internal Medicine* 1982; **96**: 756–761.
6. **Koopmans M, et al.** Foodborne viruses. *FEMS Microbiology Reviews* 2002; **26**: 187–205.
7. **Mattner F, et al.** Risk groups for clinical complications of norovirus infections: an outbreak investigation. *Clinical Microbiology and Infection* 2006; **12**: 69–74.
8. **Murata T, et al.** Prolonged norovirus shedding in infants ≤ 6 months of age with gastroenteritis. *Pediatric Infectious Diseases Journal* 2007; **26**: 46–49.
9. **Thapar N, Sanderson IR.** Diarrhoea in children: an interface between developing and developed countries. *Lancet* 2004; **363**: 641–653.
10. **Simpson R, et al.** Infantile viral gastroenteritis: on the way to closing the diagnostic gap. *Journal of Medical Virology* 2003; **70**: 258–262.
11. **Parashar UD, et al.** Rotavirus and severe childhood diarrhea. *Emerging Infectious Diseases* 2006; **12**: 304–306.
12. **Koopmans M, Van Asperen I.** Epidemiology of rotavirus infections in The Netherlands. *Acta Paediatrica* (Suppl.) 1999; **88**: 31–37.
13. **Mounts AW, et al.** Cold weather seasonality of gastroenteritis associated with Norwalk-like viruses. *Journal of Infectious Diseases* 2000; **181** (Suppl. 2): S284–287.
14. **Pasquinelli L.** Enterovirus infections. *Pediatric Reviews* 2006; **27**: e14–15.
15. **Palacios G, Oberste MS.** Enteroviruses as agents of emerging infectious diseases. *Journal of Neurovirology* 2005; **11**: 424–433.
16. **Grimwood K, et al.** Acute flaccid paralysis from echovirus type 33 infection. *Journal of Clinical Microbiology* 2003; **41**: 2230–2232.
17. **Yamashita K, et al.** Epidemics of aseptic meningitis due to echovirus 30 in Japan. A report of the National Epidemiological Surveillance of Infectious Agents in Japan. *Japanese Journal of Medical Science and Biology* 1994; **47**: 221–239.
18. **Fisman D.** Seasonality of viral infections: mechanisms and unknowns. *Clinical Microbiology and Infection* 2012; **18**: 946–954.
19. **Parashar UD, Monroe SS.** ‘Norwalk-like viruses’ as a cause of foodborne disease outbreaks. *Reviews in Medical Virology* 2001; **11**: 243–252.

20. **Arvelo W, et al.** Norovirus outbreak of probable waterborne transmission with high attack rate in a Guatemalan resort. *Journal of Clinical Virology* 2012; **55**: 8–11.
21. **Nenonen NP, et al.** Marked genomic diversity of norovirus genogroup I strains in a waterborne outbreak. *Applied and Environmental Microbiology* 2012; **78**: 1846–1852.
22. **Lysen M, et al.** Genetic diversity among food-borne and waterborne norovirus strains causing outbreaks in Sweden. *Journal of Clinical Microbiology* 2009; **47**: 2411–2418.
23. **Shuman EK.** Global climate change and infectious diseases. *New England Journal of Medicine* 2010; **362**: 1061–1063.
24. **Wolffs PF, et al.** Replacing traditional diagnostics of fecal viral pathogens by a comprehensive panel of real-time PCRs. *Journal of Clinical Microbiology* 2011; **49**: 1926–1931.
25. **Brittain-Long R, et al.** Multiplex real-time PCR for detection of respiratory tract infections. *Journal of Clinical Virology* 2008; **41**: 53–56.
26. **World Meteorological Organization.** *Guide to Meteorological Instruments and Methods of Observation*, 7th edn. Geneva: American Meteorological Society, 2008.
27. **Nenonen NP, et al.** Molecular analysis of an oyster-related norovirus outbreak. *Journal of Clinical Virology* 2009; **45**: 105–108.
28. **Ayukekbong J, et al.** Enteric viruses in healthy children in Cameroon: viral load and genotyping of norovirus strains. *Journal of Medical Virology* 2011; **83**: 2135–2142.
29. **Njouom R, et al.** Viral etiology of influenza-like illnesses in Cameroon, January–December 2009. *Journal of Infectious Diseases* 2012; **206** (Suppl. 1): S29–35.
30. **Solomon T, et al.** Virology, epidemiology, pathogenesis, and control of enterovirus 71. *Lancet Infectious Diseases* 2010; **10**: 778–790.
31. **Dedman D, et al.** Surveillance of small round structured virus (SRSV) infection in England and Wales, 1990–5. *Epidemiology and Infection* 1998; **121**: 139–149.
32. **Lopman BA, et al.** A summertime peak of ‘winter vomiting disease’: surveillance of noroviruses in England and Wales, 1995 to 2002. *BMC Public Health* 2003; **3**: 13.
33. **McSwiggan DA, Cubitt D, Moore W.** Calicivirus associated with winter vomiting disease. *Lancet* 1978; **1**: 1215.
34. **Cannon JL, et al.** Surrogates for the study of norovirus stability and inactivation in the environment: a comparison of murine norovirus and feline calicivirus. *Journal of Food Protection* 2006; **69**: 2761–2765.
35. **Duizer E, et al.** Inactivation of caliciviruses. *Applied and Environmental Microbiology* 2004; **70**: 4538–4543.
36. **Estes MK, Prasad BV, Atmar RL.** Noroviruses everywhere: has something changed? *Current Opinion in Infectious Diseases* 2006; **19**: 467–474.
37. **Zheng DP, et al.** Norovirus classification and proposed strain nomenclature. *Virology* 2006; **346**: 312–323.
38. **Riera-Montes M, et al.** Waterborne norovirus outbreak in a municipal drinking-water supply in Sweden. *Epidemiology and Infection* 2011; **139**: 1928–1935.
39. **Oluwatoyin Japhet M, et al.** Molecular epidemiology of rotavirus and norovirus in Ile-Ife, Nigeria: high prevalence of G12P[8] rotavirus strains and detection of a rare norovirus genotype. *Journal of Medical Virology* 2012; **84**: 1489–1496.