

Modification of rheumatic symptoms by diet and drugs

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Physicians and patients have long been intrigued with the possibility that some foods or food-related products may provoke rheumatic diseases, whereas others may alleviate symptoms of arthritis. If this hypothesis were true, then arthritis would respond to appropriate nutritional therapy. However, diet therapy for rheumatic disease has generally been considered a form of 'quack' therapy, for which a large percentage of patients with arthritis spend nearly \$1 billion annually in the USA (Panush, 1987). Surprisingly, despite the fervour of advocates and scepticism of rheumatologists, little objective information exists about nutritional therapy for rheumatic diseases, and virtually all conclusions have been based on inadequate information or improper study design (Ziff, 1983).

Persuasive reasons exist for considering that diet might affect rheumatic disease, and rheumatoid arthritis in particular. Two possible mechanisms suggested which need not be mutually exclusive are: (1) dietary antigens might provoke hypersensitivity responses (food allergies), which would in turn lead to rheumatological symptoms; (2) nutritional modifications might alter immune and inflammatory responses and thus affect manifestations of rheumatic diseases (Panush *et al.* 1986). We will continue by looking at these two concepts in more depth.

Food-induced arthritis

Gout is a rheumatic disease where the aetiological role of diet is established. Over 100 years ago the distinguished physician A. B. Garrod commented that 'fermented liquors' are a powerful predisposing cause of gout. More recently the purine content of the diet has been shown to influence the serum concentration of uric acid, the pathogenic moiety of gout (Seegmiller *et al.* 1961; Griebisch & Zollner, 1974). The influence of diet in gout is so important, that the initial recommended prophylactic treatment is dietary manipulation, before embarking on lifelong drug therapy with allopurinol. What evidence is there of diet causing other rheumatic diseases?

A small number of tentative, yet provocative, observations suggest relationships between foods and rheumatic diseases. These include black walnut (*Juglans nigra*) ingestion with Behcet's syndrome (Marquardt *et al.* 1973), the case of a dermatologist who documented his own palindromic rheumatism to be caused by sodium nitrate hypersensitivity (Epstein, 1969), canavanine in lucerne (*Medicago sativa*) with SLE (Malinow *et al.* 1982), chemical or food challenges (particularly wheat, maize and beef) with symptomatic arthritis (Stroud, 1983), rheumatoid arthritis with dairy products (Parke & Hughes, 1981), food and tartrazine sensitivity with rheumatoid arthritis (Brostoff, 1982; Wraith, 1982) and rheumatoid-like synovitis in rabbits with consumption of cow's milk (Reidenberg *et al.* 1983). How many foods be pathogenic in arthritic disease?

Although immunological mechanisms of tissue injury are important in the pathogenesis of rheumatic disease, the antigen or antigens that trigger these abnormal immune events have not been identified. Microbial agents have received a lot of attention, but food-related antigens should also be considered for the following reasons. Foods may normally evoke immune responses in humans, in the same way as other environmental antigens. Food antigens, food antibodies, and their complexes and sensitized lymphocytes have all been detected in the systemic circulation of normal subjects (Paganalli

et al. 1979; Panush, 1986). Furthermore, some investigators have considered the gut to be more permeable to food antigens in patients with rheumatoid arthritis than in normal individuals (Sundqvist *et al.* 1982; Rooney *et al.* 1983). The role of antigenic absorption from the gut is further implied in the established phenomena of arthropathy following jejunio-ileal bypass (Delamere *et al.* 1983). Foods may cause immunologically-mediated symptoms after ingestion in some persons, symptoms that are usually anaphylactic, cutaneous, respiratory, or gastrointestinal and mediated by mechanisms of immediate hypersensitivity. A new concept of delayed reactions to foods (masked food intolerance) is emerging, in which it is hypothesized that certain clinical symptoms reflect food allergies that may develop after a few hours, days or even longer. It is proposed that this response is mediated by immunological mechanisms other than IgE-mast-cell events. These symptoms might include headaches, behavioural or gastrointestinal disorders, and arthritis (Panush, 1986).

Panush *et al.* (1986) have investigated a patient in depth, whose rheumatoid arthritis flared when dairy products were eaten. There was marked and consistent relief of these exacerbations (both subjective and objective) during fasting, which was sustained with elemental nutrition supplements. Four different blinded challenges with milk reproducibly exacerbated symptoms, whereas placebo and other food challenges had no effect. Symptoms peaked 24–28 h after each challenge and resolved after 1–3 d. Immunological studies suggested both delayed and immediate cutaneous reactivity to milk, no elevation of IgE anti-milk, marked increases of IgG and IgG₄ anti-milk levels, marginally increased IgG–milk circulating immune complexes, and *in vitro* cellular sensitivity to milk. This study related symptomatic exacerbation of inflammatory arthritis with immunologic hypersensitivity to milk (Panush *et al.* 1986).

Animal experimental work has shown that nine out of twenty-five Old English rabbits drinking cow's milk for 12 weeks, developed rheumatoid-like synovial lesions. This was associated with a raised percentage of T lymphocytes in the synovial fluid and high titres of serum and synovial fluid C1q binding activity, due to specific antibody C1q produced in response to C1q in cow's milk (Welsh *et al.* 1985a,b).

Fasting and its effects on rheumatoid arthritis

Impressive evidence for an effect of diet restriction on autoimmune disease was obtained by Fernandez *et al.* (1976, 1978) who showed that (NZB × NZW) F1 'lupus' mice lived up to twice as long when total food intake was decreased. This was accompanied by a marked reduction in anti-DNA levels. It has also been demonstrated that the autoimmune disease of NZB mice is slowed by deprivation of zinc in the diet (Beach *et al.* 1981). Claims are sometimes made that fasting relieves arthritic symptoms in rheumatoid arthritis. It is known that malnutrition is immunosuppressive (Chandra, 1981), and studies have shown that fasting by healthy subjects is accompanied by altered neutrophil bacterial killing and depressed lymphocyte response to mitogens (Palmlblad, 1976), decreased serum levels of acute-phase reactants and complement (Palmlblad *et al.* 1977a) and increased serum cortisol concentrations (Palmlblad *et al.* 1977b). In accordance with these findings, an association between improvement in inflammatory activity of rheumatoid joints, fall in the ESR and enhancement of neutrophil bactericidal capacity, following a 7 d fast has been reported (Uden *et al.* 1983). Skoldstrom *et al.* (1979) also noted symptomatic improvement of patients with rheumatoid arthritis during fasting.

Rheumatoid arthritis and polyunsaturated fatty acids

Adverse publicity surrounding the drugs used to treat rheumatoid arthritis and growing consumer enthusiasm for alternative medicine have increased interest in dietary

manipulation as a form of therapy. Patients have long believed that fish oils in general, and cod liver oil in particular, may relieve arthritis. More recently it has been suggested that evening primrose (*Oenothera biennis*) oil may also be beneficial. What is the scientific evidence to support these claims?

Experimental evidence. Critical in the considerations involved in the possible effect of fatty acids on arthritis has been the amount of the 2-double-bonded polyunsaturated fatty acid (PUFA), linoleic acid (LA), in the diet. The level of intake of this fatty acid has a number of important ramifications above and beyond the fact that it is an essential fatty acid (EFA) whose absence from the diet can lead to deficiency states. First it is the precursor of arachidonic acid (AA) which is, in turn, the precursor of the prostaglandins (PG) via the cyclo-oxygenase pathway and leukotrienes via the 5-lipoxygenase (*EC* 1.13.11.12) pathway. Thus, decreased intake of LA leads to decreased formation of PG and leukotrienes, and increased intake of LA leads to increased amounts of these inflammatory agents. The E series PG (PGE) and leukotrienes have a pro-inflammatory effect (Kuehl & Egan, 1980). The PGE also have important suppressive effects on both the cellular and humoral immune responses (Goodwin & Webb, 1980) and on the response of the fibroblast to stimulation by immunologic mediators (Korn *et al.* 1980). So the PG may be expected to increase or decrease inflammation, immunity or collagen synthesis, depending on their site of action and their concentration at that site. As a corollary, diets which contain either large amounts of LA, leading to increased synthesis of PG, or small amounts of LA, leading to reduced PG synthesis, may be expected to affect inflammation, immunity and fibrosis.

There is a growing literature demonstrating the importance of the LA-AA-PG axis in inflammatory disease. Suppression of adjuvant arthritis in rats by injection of PGE₂ (Aspinall & Cammarata, 1969) and PGE₁ (Zurier & Quagliata, 1971) has been reported. Subsequent work demonstrated a beneficial effect of PGE₁ on the survival of NZB/NZW mice (Zurier *et al.* 1977b) and on the course of renal disease in these mice (Zurier *et al.* 1977a). Kunkel *et al.* (1982) found that evening primrose oil, which is very rich in LA, markedly inhibited the development of adjuvant-induced arthritis in rats. Stackpoole & Mertin (1981) showed that experimental allergic encephalomyelitis in guinea-pigs was almost completely inhibited by oral supplements of EFA. These reports have established the suppressive effects of PG or increased intake of PUFA on immunologically-mediated disease.

Conversely a number of studies have described benefits of EFA-deficient diets in animal models of immunologically-mediated inflammatory disease. Adjuvant arthritis was ameliorated in animals deficient in EFA (Denko, 1976). An EFA-deficient diet diminished humoral response to T cell-dependent and T cell-independent antigens in mice. Full restoration of these responses occurred on switching to a control diet (DeWille *et al.* 1978). Interestingly, Hurd *et al.* (1981) found that NZB/NZW mice fed on a diet in which all fat was in the form of saturated fat (coconut oil) had marked prolongation of life, and delay in the development of renal abnormalities. This improvement appeared to be associated with a deficiency of dietary PUFA. Conversely Prickett *et al.* (1981) reported that when NZB/NZW mice were fed on a diet rich in eicosapentaenoic acid (EPA) (the predominant fatty acid in the fish oil-rich diet of Greenland Eskimos), they experienced a similar marked prolongation of life and delay in onset of renal abnormalities. More recently Lee *et al.* (1985) showed that diets enriched with fish oil may have anti-inflammatory effects by inhibiting the 5-lipoxygenase pathway in neutrophils and monocytes, and by inhibiting the leukotriene B₄-mediated function of neutrophils. McColl *et al.* (1986) reported that dietary supplementation with EPA reduced the severity of adjuvant arthritis in rats. There is, therefore, evidence that substitution of

PUFA in the diet by saturated fatty acid or 'unphysiological' fish oil-derived PUFA, leads to a marked reduction in the severity of autoimmune disease in these mice.

Fish oils are rich in long-chain PUFA, particularly the ω -3 series: EPA and docosahexaenoic acid. These are unlike LA which is an ω -6 PUFA and is the predominant PUFA in the Western diet. One theory behind fish oil therapy is that fatty acids, such as α -linolenic acid from fish oils, are converted into PG of the third series, whereas the saturated fats commonly consumed in the Western diet are converted via AA into PG of the second series. Because PG of the second series are actively involved in the rheumatoid process there may well be some benefit in diverting PG synthesis to the less inflammatory third series.

Thus, theoretically there appear to be three beneficial ways in which to modify fatty acid dietary intake in rheumatic diseases: (1) a PUFA-deficient diet aimed at reducing PG formation and decreasing lymphocyte and macrophage cell membrane reactivity. This diet could lead to the problem of EFA deficiency, so supplements may be required; (2) a diet rich in PUFA designed to increase the synthesis of suppressor PG; (3) a diet rich in fish oils which would produce relatively inactive PG of the '3' series in place of the pro-inflammatory PG of the '2' series.

Clinical studies. Kremer *et al.* (1985) reported the first double-blind study to show that a diet high in PUFA and EPA in rheumatoid arthritis, led to improvement in morning stiffness, number of tender joints, and grip strength compared with controls who were eating an American diet high in saturated fats plus placebo EPA. These favourable results were not supported by laboratory markers of disease activity. More recently Kremer *et al.* (1987) studied the effects of supplementing an unchanged diet with fish oil supplements (15 'Max-EPA' (Duncan Flockhart) tablets/d) in rheumatoid arthritis. The leukotriene B₄ levels fell substantially and statistically significant improvement was seen in fatigue time and number of tender joints during the treatment period, but there was again no change in laboratory markers of disease activity. Sperling (1986) reported a synergistic effect of fish oils and non-steroidal anti-inflammatory drugs (NSAID) in rheumatoid arthritis in both clinical and biological indices.

It is believed that treatment with γ -linolenic acid from evening primrose oil, may produce a similar fall in PG of the second series as is found with fish oils, and also produce an increase in PG of the first series (e.g. PGE₁), which have anti-inflammatory activity. Hansen *et al.* (1983) found no benefit in twenty patients with active rheumatoid arthritis undergoing treatment with evening primrose oil for 12 weeks. Belch *et al.* (1986) investigated the effect both of evening primrose oil alone, and in combination with fish oil in rheumatoid arthritis. In contrast they found that both groups produced significant clinical improvement which allowed some patients to reduce or even stop their NSAID. They found no evidence of disease modification.

Many of these studies have been criticized for studying patients with early, often mild disease and response being clouded by hospitalization. Nash *et al.* (1988) studied twenty-six patients with well-established rheumatoid arthritis, treated as outpatients with E028 (an elemental diet consisting of essential amino acids, trace elements and vitamins, free of protein and considered hypo-allergic). A control group supplemented their diet with E028, whilst diet-treated patients used E028 alone for 4 weeks. After 4 weeks individual foods were re-introduced and disease activity was monitored. The results showed that this elemental diet improved functional score and a thermographic joint index, but not ESR or C-reactive protein.

The therapeutic implications of these studies may be somewhat disappointing to many rheumatologists because the quantitative changes were modest and reflected mainly subjective benefit. However, these observations are among the few that provide us with

useful information about diet therapy, and point us in new directions for studying the pathogenesis of rheumatic diseases and ultimately developing new therapies.

Another area of nutritional interest in rheumatic diseases which is open to therapeutic manipulation is iron. This is a major interest of our group.

Fe and rheumatic disease

The potential role of Fe in rheumatoid disease is well recorded (Blake *et al.* 1981). Any sustained inflammatory reaction causes changes in Fe metabolism, with a drop in serum Fe and a redistribution of Fe to the activated reticulo-endothelial system, including, in rheumatoid disease, the inflamed synovium. It has been suggested that this synovial Fe deposition contributes to the joint inflammation (Muirden & Senator, 1968), and Blake *et al.* (1985a) found that high levels of synovial Fe anticipated a poor prognosis in early rheumatoid patients. Furthermore, treatment of the anaemia of chronic disease associated with rheumatoid arthritis, with intravenous Fe dextran (Blake *et al.* 1985b) or with oral Fe (Blake & Bacon, 1982), has been shown to exacerbate joint symptoms.

The crucial role of Fe in inflammation is its ability to act as a redox agent, and hence take part in electron transfer pathways. Fe is thus able to catalyse the production of highly toxic reactive oxygen species (ROS) via the Fenton reaction. During inflammation it is thought that ROS produced by stimulated phagocytes contribute to tissue damage by inducing lipid peroxidation (Halliwell & Gutteridge, 1984). Lipid peroxidation products have been found in synovial fluid both in rheumatoid patients (Lunec *et al.* 1981) and adjuvant arthritis (Yoshikawa *et al.* 1983), suggesting such a process may contribute to joint inflammation. High levels of PGE₂ have been reported in inflamed tissue and perfusates of adjuvant arthritic ankle joints, and the levels measured reflected the severity (Barbier *et al.* 1984). Both periosteal proliferation and bone resorption may be induced by PGE₂ (Galasko & Bennett, 1976), and *in vitro* studies have shown that Fe nitilotriacetate will augment the release of PGE₂ from synovial cells in culture (Okasaki *et al.* 1981). Fe chelation may, therefore, reduce PGE₂ production, inhibiting both the periosteal reaction of adjuvant disease and the erosive consequences of synovial inflammation. It is interesting that this is an effect that can also be produced by dietary manipulation of fatty acids as mentioned earlier.

Manipulation of the diet by the removal of Fe thus has a sound theoretical basis for reducing the inflammation of rheumatoid arthritis. Andrews *et al.* (1987a) studied the effect of Fe chelation by using desferrioxamine (DFX) in adjuvant disease in rats. They found that DFX induced mild Fe deficiency, and reduced the incidence and severity of joint inflammation, but did not alter the local primary inflammatory response of adjuvant or the systemic sequelae. The joints showed a reduction in soft tissue swelling and bone erosion. These findings suggested an apparent selective influence of Fe on joint-mediated inflammation. They extended their studies to look at the effect of mild nutritional Fe deficiency on adjuvant disease in rats and also on models not involving the joint (carrageenan pleurisy, urate pleurisy, pyrophosphate foot pad; Andrews *et al.* 1987b). Mild nutritional Fe deficiency was found to significantly reduce the severity of adjuvant arthritis assessed by histology, radiology and subjective scoring. Again systemic features were unaffected, and there was no suppression of the other models of acute inflammation, supporting the selectivity of the phenomena. In Glynn-Dumonde synovitis of guinea-pigs, DFX stimulated the acute phase of inflammation, but repeated administration depressed the chronic phase (Blake *et al.* 1983).

These effects of DFX on chronic inflammation were sufficiently encouraging for preliminary trials on human rheumatoid patients. Of seven patients given DFX (3 g/d, 5 d/week for 1–3 weeks), four acquired retinal abnormalities that were reversed on drug

withdrawal (Blake *et al.* 1985c). Also, two of the patients received the phenothiazine prochlorperazine during DFX therapy and subsequently lost consciousness for 48–72 h, possibly because this combination of drugs removes essential Fe from the nervous system. Thus there is a need for new Fe-chelating drugs that can be administered orally, and are safer than DFX.

REFERENCES

- Andrews, F. J., Morris, C. J., Kondratowicz, G. & Blake, D. R. (1987a). *Annals of Rheumatic Diseases* **46**, 327–333.
- Andrews, F. J., Morris, C. J., Lewis, E. J. & Blake, D. R. (1987b). *Annals of Rheumatic Diseases* **46**, 859–865.
- Aspinall, R. L. & Cammarata, P. S. (1969). *Nature* **224**, 1320–1321.
- Barbier, A., Navarro, J., Brehere, J. C. & Roncucci, R. (1984). *Agents and Actions* **15**, 103–110.
- Beach, R. S., Gershman, M. E. & Hurley, L. S. (1981). *Journal of Immunology* **126**, 1999–2006.
- Belch, J. J. F., Ansell, D., Madhok, R. & Sturrock, R. D. (1986). *British Journal of Rheumatology* **25**, Suppl., 75 Abstr.
- Blake, D. R. & Bacon, P. A. (1982). *Lancet* **i**, 623.
- Blake, D. R., Gallagher, P. J., Potter, A. R., Bell, M. J. & Bacon, P. A. (1985a). *Arthritis and Rheumatism* **27**, 495–501.
- Blake, D. R., Hall, N. D., Bacon, P. A., Dieppe, P. A., Halliwell, B. & Gutteridge, J. M. C. (1981). *Lancet* **ii**, 1141–1144.
- Blake, D. R., Hall, N. D., Bacon, P. A., Dieppe, P. A., Halliwell, B. & Gutteridge, J. M. C. (1983). *Annals of Rheumatic Diseases* **42**, 89–93.
- Blake, D. R., Lunec, J., Ahern, M., Ring, E. F. J., Bradfield, J. & Gutteridge, J. M. C. (1985b). *Annals of Rheumatic Diseases* **44**, 183–188.
- Blake, D. R., Winyard, P., Lunec, J. & Williams, A. (1985c). *Quarterly Journal of Medicine* **56**, 345–355.
- Brostoff, J. (1982). *Fourth International Food Allergy Symposium of the American College of Allergy*, Vancouver.
- Chandra, R. K. (1981). *British Medical Bulletin* **37**, 89–94.
- Delamere, J. P., Baddeley, R. M. & Walton, K. W. (1983). *Annals of Rheumatic Diseases* **42**, 553–557.
- Denko, C. W. (1976). *Agents and Actions* **6**, 636–641.
- DeWille, J. W., Fraker, P. J. & Romsos, D. R. (1978). *Journal of Nutrition* **109**, 1018–1027.
- Epstein, S. (1969). *Annals of Allergy* **27**, 343–349.
- Fernandez, G., Friend, P., Yunis, E. J. & Good, R. A. (1978). *Proceedings of the National Academy of Sciences, USA* **75**, 1500–1504.
- Fernandez, G., Yunis, E. J. & Good, R. A. (1976). *Proceedings of the National Academy of Sciences, USA* **73**, 1279–1283.
- Galasko, C. S. B. & Bennett, A. (1976). *Nature* **263**, 508–510.
- Goodwin, J. S. & Webb, D. R. (1980). *Clinical Immunology and Immunopathology* **15**, 106–122.
- Griebisch, A. & Zollner, N. (1974). *Advances in Experimental Medicine and Biology* **41**, 443–449.
- Halliwell, B. & Gutteridge, J. M. C. (1984). *Biochemical Journal* **219**, 1–14.
- Hansen, T. M., Lerche, A., Kassis, V., Lorenzen, I. & Sondergaard, J. (1983). *Scandinavian Journal of Rheumatology* **12**, 85–88.
- Hurd, E. R., Johnston, J. M., Okita, J. R., MacDonald, P. C., Ziff, M. & Gilliam, J. N. (1981). *Journal of Clinical Investigation* **67**, 467–485.
- Korn, J. H., Haluska, P. V. & LeRoy, E. C. (1980). *Journal of Clinical Investigation* **65**, 543–554.
- Kremer, J. M., Bigauoette, J. & Michalek, A. V. (1985). *Lancet* **i**, 184–187.
- Kremer, J. M., Jubiz, W. & Rynes, R. I. (1987). *Annals of Internal Medicine* **106**, 497–503.
- Kuehl, F. A. & Egan, R. W. (1980). *Science* **210**, 978–984.
- Kunkel, S. L., Ogawa, H., Ward, P. A. & Zurier, R. B. (1982). *Progress in Lipid Research* **20**, 885–888.
- Lee, T. H., Hoover, R. L. & Williams, J. D. (1985). *New England Journal of Medicine* **312**, 1217–1224.
- Lunec, J., Halloran, S. P., White, A. G. & Dormandy, T. L. (1981). *Journal of Rheumatology* **8**, 233–245.
- McCull, S. R., Whitehouse, M. W., Cleland, L. G. & Hurst, N. P. (1986). *British Journal of Rheumatology* **25**, 106.
- Malinow, M. R., Bardana, E. J., Pirofsky, B., Craig, S. & McLoughlin, P. (1982). *Science* **216**, 415–417.
- Marquardt, J. L., Snyderman, R. & Oppenheim, J. J. (1973). *Cellular Immunology* **9**, 263–272.
- Muirden, K. D. & Senator, G. B. (1968). *Annals of Rheumatic Diseases* **27**, 38–47.

- Nash, P., Workman, E., Smith, M., Hazleman, B. L. & Hunter, J. O. (1988). *British Journal of Rheumatology* **27**, Suppl. 1, 55 Abstr.
- Okasaki, I., Brinckerhoff, C. E., Sinclair, J. F., Sinclair, P. R., Bronkowsky, H. L. & Harris, E. D. (1981). *Journal of Laboratory and Clinical Medicine* **97**, 396–402.
- Paganalli, R., Levinsky, R. J., Brostoff, J. & Wraith, D. G. (1979). *Lancet* **i**, 1270–1272.
- Palmblad, J. (1976). *Scandinavian Journal of Haematology* **17**, 217–226.
- Palmblad, J., Cantell, K., Holm, G., Norberg, R., Strander, H. & Sunblad, L. (1977a). *Clinical and Experimental Immunology* **30**, 50–55.
- Palmblad, J., Levi, L. & Burger, A. (1977b). *Acta Medica Scandinavica* **201**, 15–22.
- Panush, R. S. (1986). *Annals of Allergy* **56**, 500–503.
- Panush, R. S. (1987). *Annals of Internal Medicine* **106**, 619–621.
- Panush, R. S., Stroud, R. M. & Webster, E. M. (1986). *Arthritis and Rheumatism* **29**, 220–226.
- Parke, A. L. & Hughes, G. R. V. (1981). *British Medical Journal* **282**, 2027–2029.
- Prickett, J. D., Robinson, D. R. & Steinberg, A. D. (1981). *Journal of Clinical Investigation* **68**, 556–559.
- Reidenberg, M. M., Durant, P. J., Harris, R. A., Boccardo, G. P., Lahita, R. & Stenzel, R. H. (1983). *American Journal of Medicine* **75**, 365–370.
- Rooney, P. J., Jenkins, R. T., Goodacre, R. L. & Sivakumaran, T. (1983). *Clinical Research* **31**, 160A.
- Seegmiller, J. E., Grayzel, A. I., Laster, L. & Liddle, L. (1961). *Journal of Clinical Investigation* **40**, 1304–1314.
- Skoldstrom, L., Larsson, L. & Linstrom, F. D. (1979). *Scandinavian Journal of Rheumatology* **8**, 249–255.
- Sperling, R. (1986). *Advances in Rheumatology* **1**, 8.
- Stackpoole, A. & Mertin, J. (1981). *Progress in Lipid Research* **20**, 649–654.
- Stroud, R. M. (1983). In *Current Topics in Rheumatology*, pp. 145–157 [B. H. Hahn, F. C. Arnett, T. M. Zizic and M. C. Hochberg, editors]. Kalamazoo, MI: Upjohn Press.
- Sundqvist, T., Linstrom, F., Magnusson, K. E., Skoldstrom, L., Stjernstom, I. & Tagesson, C. (1982). *Scandinavian Journal of Rheumatology* **11**, 33–38.
- Uden, A. M., Trang, L., Venizelos, N. & Palmblad, J. (1983). *Annals of Rheumatic Diseases* **42**, 45–51.
- Welsh, C. J. R., Hanglow, A. C., Conn, P., Barker, T. H. W. & Coombs, R. R. A. (1985a). *International Archives of Allergy and Applied Immunology* **78**, 145–151.
- Welsh, C. J. R., Hanglow, A. C., Conn, P., Barker, T. H. W. & Coombs, R. R. A. (1985b). *International Archives of Allergy and Applied Immunology* **78**, 152–157.
- Wraith, D. G. (1982). *Fourth International Food Allergy Symposium of the American College of Allergy*, Vancouver.
- Yoshikawa, T., Tanaka, H. & Kondo, M. (1983). *Journal of Applied Biochemistry* **5**, 382–387.
- Ziff, M. (1983). *Arthritis and Rheumatism* **26**, 457–461.
- Zurier, R. B., Damjanov, I., Sayadoff, D. M. & Rothfield, N. F. (1977a). *Arthritis and Rheumatism* **20**, 1449–1456.
- Zurier, R. B., Sayadoff, D. M., Torrey, S. B. & Rothfield, N. F. (1977b). *Arthritis and Rheumatism* **20**, 723–728.
- Zurier, R. B. & Quagliata, F. (1971). *Nature* **234**, 304–305.