

# MICROSCOPY TODAY

JULY/AUGUST 2002

VOLUME 10 - NUMBER 4





# Imagine Microscopy without Barriers

FEI's 3D Structural Process Management™ solutions for microscopy deliver the most complete data set quickly and efficiently. Whether your interest is in life science, structural biology, materials science or microelectronics, our microscopy solutions help you break through barriers in research, development and manufacturing.

Engineered with your success in mind, our full range of TEM, DualBeam™, ESEM™, and SEM solutions enable you to study samples in three dimensions, including below the surface.

Tecnai™ G<sup>2</sup> TEM: high resolution, three-dimensional TEM analysis down to the atomic level.

Strata™ DB 235M: three dimensional, below the surface data and nanofabrication capabilities.

Quanta™ SEM: All in One SEM delivering high-vac, low-vac, and ESEM flexibility.

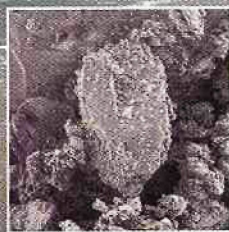
Sirion™ SEM: low kV imaging plus integrated STEM delivering better than 1 nm image resolution.

Contact us and learn how FEI's unique combination of core technologies and more than 70 years of experience combine to remove the obstacles to your microscopy success and enable you to break through nanotech barriers.

3D Structural Process Management™  
Eliminates the Obstacles



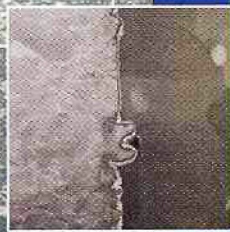
TEM



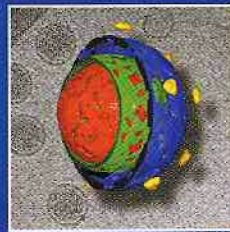
Low-vac SEM / ESEM™



DualBeam™



SEM / STEM



Cryo TEM

[www.feicompany.com](http://www.feicompany.com)  
[sales@feico.com](mailto:sales@feico.com)



## very tiny bar codes

Stephen W. Carmichael,<sup>1</sup> Mayo Clinic  
carmichael.stephen@mayo.edu

The use of barcodes on products, combined with the computer technology to read and track the subsequent data, has revolutionized inventory control in our society. Barcodes are commonly seen on items as large as automobiles and as small as pieces of candy. But if you need to track a large number of items that are microscopic in size, such as biologically relevant molecules (think genomics, proteomics, metabolomics, *etc.*), then you need very small barcodes. Sufficiently small rods that can be marked as barcodes have been developed by Sheila Nicewarner-Peña, Griffith Freeman, Brian Reiss, Lin He, David Peña, Ian Walton, Remy Cromer, Christine Keating, and Michael Natan.<sup>2</sup>

Their starting point was a membrane of alumina with pores that served as a mold for the rods. The backside of the membrane was coated with a silver film that served as the working electrode. Using apparently simple electrochemistry, different metals were sequentially deposited within the pores, creating metallic stripes along the rod. The rod structure was determined by (i) the membrane pore diameter, that determines the diameter of the rod, (ii) the sequence of metal ions introduced into the solution bathing the membrane, that defines the number and pattern of metal stripes, and (iii) the electrical charge passed in each step, that dictates the length of each stripe. Nicewarner-Peña *et al.* have used seven different metals (cobalt, copper, gold, nickel, palladium, platinum, and silver) to create stripes as short as 10 nanometers and as long as several micrometers, and have prepared rods with as many as 13 distinguishable stripes.

After the metals are deposited within the pores, the membrane is dissolved with nitric acid and sodium hydroxide, isolating the rods. The rods were examined by transmission electron microscopy, scanning electron microscopy (SEM), field emission SEM, energy-dispersive x-ray scattering, macroscopic

extinction, and microscopic reflectivity, in addition to chemical analyses typically applied to bulk materials. Most significantly, examination with the light microscope<sup>3</sup> clearly revealed a pattern of stripes when examined at the appropriate wavelength. Various metals have different reflectivities at different wavelength. For example, it was determined that at a wavelength of 430 nanometers, silver was 2.5 times more reflective than gold, whereas other wavelengths worked better for other combinations of metals. This suggests that reading the stripes may require more than one wavelength to optimize the number of unique barcodes.

The number of unique barcode patterns of  $n$  number of metals with  $s$  number of stripes is on the order of  $n^s$ . Although the limit of resolution in a typical light microscope is around 143 nanometers, Nicewarner-Peña *et al.* suggested stripes on the order of half a micron. With a rod a few microns in length, this gives tens of thousands of unique combinations, far more than any other tagging method yet developed.

Fortunately, there are many chemical methods for attaching biologic molecules to the barcoded rods. Nicewarner-Peña *et al.* demonstrated this by attaching DNA molecules to rods that were then used in hybridization assays, and immunoglobulins used in a sandwich immunoassay. As the field of bioinformatics explodes and interest grows in conducting comprehensive analyses on classes of molecules, there will be a demand for very high-level multiplexing in small samples. This in turn will depend on a suitably small readout system. The barcodes developed by Nicewarner-Peña *et al.* may have fulfilled this need of the future.

### References

- 1 The author gratefully acknowledges Dr. Christine D. Keating for reviewing this article.
- 2 Nicewarner-Peña, S.R., R.G. Freeman, B.D. Reiss, L. He, D.J. Peña, I.D. Walton, R. Cromer, C.D. Keating, and M.J. Natan, Submicrometer metallic barcodes, *Science* **294**:137-141, 2001.
- 3 They used a Nikon TE-300 inverted microscope equipped with a bright-field reflectance filter, using 100X oil immersion lens, and a bandpass filter to vary the wavelength of the excitation beam.

## INDEX OF ARTICLES

<b>very tiny bar codes</b> .....	<b>3</b>
Stephen W. Carmichael, Mayo Clinic	
<b>Prog. Towards More Realistic In-Situ Microscopy Observations</b> .....	<b>5</b>
A. Howie, Cavendish Laboratory, University of Cambridge	
<b>Spectral Image Analysis: Getting The Most From All That Data</b> .....	<b>8</b>
P.G. Kotula, M.R. Keenan, and R. Loehman, Sandia National Laboratories	
<b>New Fluorescent Labeling Technologies for Ultrasensitive Cytochemical and Histochemical Imaging</b> .....	<b>12</b>
Iain Johnson, Molecular Probes, Inc	
<b>Artifacts &amp; Non-Local Effects In SPM Potential Measurements</b> .....	<b>16</b>
Sergei V. Kalinin and Dawn A. Bonnell, University of Pennsylvania	
<b>Vibration, Resonance and the Effect on Microscopes</b> .....	<b>22</b>
Douglas A. Anderson, Schnabel Engineering Associates	
<b>The Basics of Immunoglobulins and Immunostaining</b> .....	<b>26</b>
W. Gray (Jay) Jerome, Vanderbilt University Medical Center	
<b>Swapping Atoms For Bits: Managing The Digital Evolution In The Microscopy Laboratory</b> .....	<b>28</b>
Judy A. Murphy, San Joaquin Delta College	
<b>A Note on Iodine and Vacuum Pumps</b> .....	<b>34</b>
Scott D. Walck, PPG Industries, Inc.	
<b>A Simple Way to Eliminate Frost Build-up on Cryo-SEM Samples</b> .	<b>34</b>
Gib Ahlstrand, University of Minnesota	

<b>Mousescope™</b> .....	<b>35</b>
Lee van Hook, Plietdown Research Institute, Munchhausen University	
<b>A Method for Safely Restraining Mouse Pups for Microscopy</b> .....	<b>36</b>
Michael J. Herron, University of Minnesota	
<b>A Simple Image Archive That's Cheap, Too!</b> .....	<b>36</b>
Tim Morken, Centers for Disease Control and Prevention	

## ABOUT THE COVER

**Iain Johnson, Molecular Probes, Inc. See his article on labelling technologies on page 12**

A cryosection of mouse intestine showing basement membranes labeled with an anti-fibronectin antibody and Alexa Fluor 488 goat anti-chicken secondary antibody (green fluorescence). Goblet cells and crypt cells are labeled with Alexa Fluor 594 wheat germ agglutinin (red fluorescence). The microvillar brush border and smooth muscle layer are visualized with Alexa Fluor 680 phalloidin