

An Ediacaran opportunist? Characteristics of a juvenile *Dickinsonia costata* population from Crisp Gorge, South Australia

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Abstract.—Despite 70 years of study, *Dickinsonia* remains one of the Ediacara biota's most enigmatic taxa with both morphological characters and phylogenetic affinities still debated. A large population of relatively small *Dickinsonia costata* present on a semi-contiguous surface from the Crisp Gorge fossil locality in the Flinders Ranges (South Australia) provides an opportunity to investigate this taxon in its juvenile form. This population supports earlier findings that suggest *D. costata*'s early growth was isometric, based on the relationship between measured variables of length and width. The number of body units increases with length, but at a decreasing rate. A correlation between a previously described physical feature, present as a shrinkage rim partially surrounding some specimens and a novel, raised lip in some specimens, suggests that both features may have been the result of a physical contraction in response to the burial process, rather than due to a gradual loss of mass during early diagenesis. A marked protuberance in 15% of the population is also noted in limited specimens within the South Australian Museum collections and appears to be present only in juvenile *D. costata*. Both the abundance and narrow size range of this population support the notion that *Dickinsonia* was a hardy opportunist, capable of rapid establishment and growth on relatively immature textured organic-mat substrates.

Introduction

The Ediacara biota (575–541 Myr) represent the first appearance of complex, macroscopic life on Earth, and body plan diversity spans a broad array from discs and tubes to quilted and fractal forms. Taxonomic affinities and life histories of many of these organisms remain either controversial or poorly understood. The genus *Dickinsonia* has been the subject of near-continual debate since it was first described 70 years ago (Sprigg, 1947). Descriptive and molecular-informed studies have sought to place *Dickinsonia* into various metazoan groups, as well as other kingdoms. Interpretations range from a pelagic cnidarian (Sprigg, 1947; Valentine, 1992) and polychaete worm (Glaessner and Wade, 1966; Wade, 1972) to benthic placozoan (Sperling and Vinther, 2010) and ctenophore (Zhang and Reitner, 2006). Taphonomic and morphological arguments have been used to place *Dickinsonia* in the proposed kingdom Vendobionta as a serially quilted unicellular organism (Seilacher et al., 2003), or as a ground-dwelling lichen (Retallack, 2007). Other studies have focused on *Dickinsonia*'s unique, serially repeating body plan to estimate oxygen requirements, most recently demonstrating that adequate oxygenation may have been achieved via surficial diffusion (see Runnegar, 1982; Gooden, 2014). This body plan has also been utilized to suggest *Dickinsonia* was a bilaterian-grade animal, due to interpreted growth via the terminal addition of units from a posterior growth region (Runnegar, 1982; Gold et al., 2015). Most recently, growth models have been employed to infer a eumetazoan affinity for *Dickinsonia* via studies suggesting both

terminal (Evans et al., 2017) and pre-terminal (Hoekzema et al., 2017) addition of body units. The identification of resting traces associated with several individual specimens indicates that it may have fed on the underlying microbial mat substrate via basal surface absorption (Gehling et al., 2005; Sperling and Vinther, 2010; Ivantsov, 2011).

This study focuses on a population of *Dickinsonia costata* Sprigg, 1947 from Crisp Gorge, South Australia (Fig. 1). The Crisp Gorge population comprises 53.5% of the total abundance on the semi-contiguous surface (Reid et al., 2017), and provides an excellent opportunity to study this taxon in its juvenile form. The remainder of the community comprises seven taxa and the biogenic sedimentary structure 'Mop' (Tarhan et al., 2010). Three additional taxa on the surface are also interpreted as juvenile: *Parvancorina minchami* Glaessner, 1958, *Tribrachidium heraldicum* Glaessner, 1959, and a single specimen of *Rugoconites enigmaticus* Glaessner and Wade, 1966. The Textured Organic Surface (TOS) components of the microbial mat substrate include a range of structures, including round, deep relief bosses and regions of TOS components including 'groove' and 'weave' (Gehling and Droser, 2009). The overall fossil surface, however, is relatively smooth, and is interpreted as immature (Reid et al., 2017).

Geologic setting

In South Australia, the Ediacara Member marks the fossiliferous shallow marine, deltaic succession of the Rawnsley Quartzite.

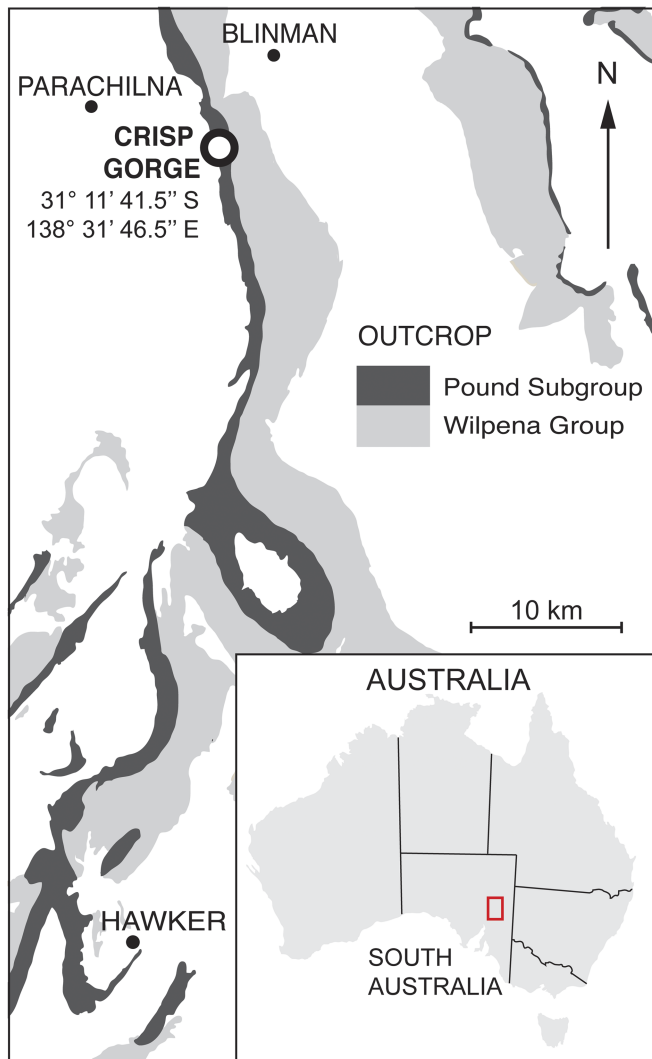


Figure 1. Locality map of the Flinders Ranges and Crisp Gorge fossil site. The Crisp Gorge fossil site is located within the Ediacara Member of the Pound Subgroup.

The Rawnsley Quartzite is the terminal package of Ediacaran-aged sediments within the Pound Subgroup of the Adelaide Rift Complex (Gehling and Droser, 2012). The Ediacara Member incorporates a number of fossiliferous facies, ranging from shore-face settings, to deeper water delta-front or pro-delta environments (Gehling and Droser, 2013). By far the richest, in situ, fossil assemblages are found within the Oscillation Rippled Sandstone facies (Tarhan et al., 2017). These comprise medium- to coarse-grained feldspathic sands with distinctive rippled bed-tops, representative of a depositional environment between fair-weather and storm wave bases.

Locality information.—The Crisp Gorge fossil locality (31°11'41.5"S, 138°31'46.5"E, Fig. 1) is primarily composed of repeating units of the Oscillation Rippled Sandstone facies (ORSF) and the deeper water Flat Laminated to Linguoid Rippled Sandstone facies, the latter indicative of a delta-front to pro-delta setting (Tarhan et al., 2017). Both facies are present at this locality as interbedded dark red to maroon sands and silts, with the ORSF exhibiting characteristic bed-top combined-flow

ripples and several bed partings revealing a silt layer up to 3 mm thick (Reid et al., 2017). The majority of the sample material examined for this study was removed from the Crisp Gorge fossil locality and now forms a framed rock wall in the 'First Life: Ediacara Biota Gallery' of the South Australian Museum. This material is referred to as the 'Crisp Wall' throughout this study. The Crisp Wall surface is comprised of 16 semi-contiguous slabs of medium- to coarse-grained feldspathic sandstone from the ORSF and gives a surface area of ~6.5 m². A further seven fossiliferous slabs determined to be semi-contiguous with the bedding surface of the existing Crisp Wall slabs have been identified at the Crisp Gorge fossil site and are incorporated into this study.

Materials and methods

The *Dickinsonia costata* specimens identified on the Crisp Wall, combined with the newly identified Crisp Gorge slabs, yielded a total of 150 specimens suitable for analysis. All measurements and observations were made from positive-relief, latex casts taken from the Crisp Wall specimens to aid identification of otherwise negative-relief features and allow for the use of a dissecting microscope. For the purpose of this study, phylogenetically neutral labels are used to describe a number of morphological features of the studied *D. costata*. This is to aid in the identification of features, while avoiding ambiguity or incidental taxonomic affinities that may be inferred from using body terms usually associated with animals. The terms 'A-end' and 'B-end' are used to refer to either end of the specimens as defined by the placement of a midline, with the A-end characterized by the terminal A-end unit (Fig. 2.1). The B-end is located at the opposite end of the body, and is defined by decreasing unit size (Fig. 2.1).

Measurements for each specimen included the maximum length, as the longest distance between A- and B-ends of the specimen, maximum width, taken as the widest measure perpendicular to the midline, and the number of units. Unit counts were not taken for individuals with damaged or indistinct units, defined here as those individuals with any form of mechanical damage to the rock or sand grains that mold them, and/or those that are indistinct from neighboring units. Measurements and unit counts were made with the aid of calipers and a dissecting microscope. Observations also were made of three body features, which have been termed the A-end lip, A-end protuberance, and shrinkage rim (see Results for details).

Statistical analysis of the *D. costata* population was performed using the open access statistical software program PAST (Hammer et al., 2001). A Model II (major axis) regression was used in the analysis of body size (Legendre and Legendre, 1998). A rose diagram of axial orientation was produced using the open access statistical software programs R (RCoreTeam, 2015) and RStudio (RStudioTeam, 2015), and the package 'circular' (Lund and Agostinelli, 2015). Pairwise correlations among three observed features (A-end lip, shrinkage rim, and A-end protuberance) were calculated using the Pearson method for the presence-absence data collected. Significance of relationships was calculated using Chi-squared tests with Yates' continuity correction. To test if the *D. costata*

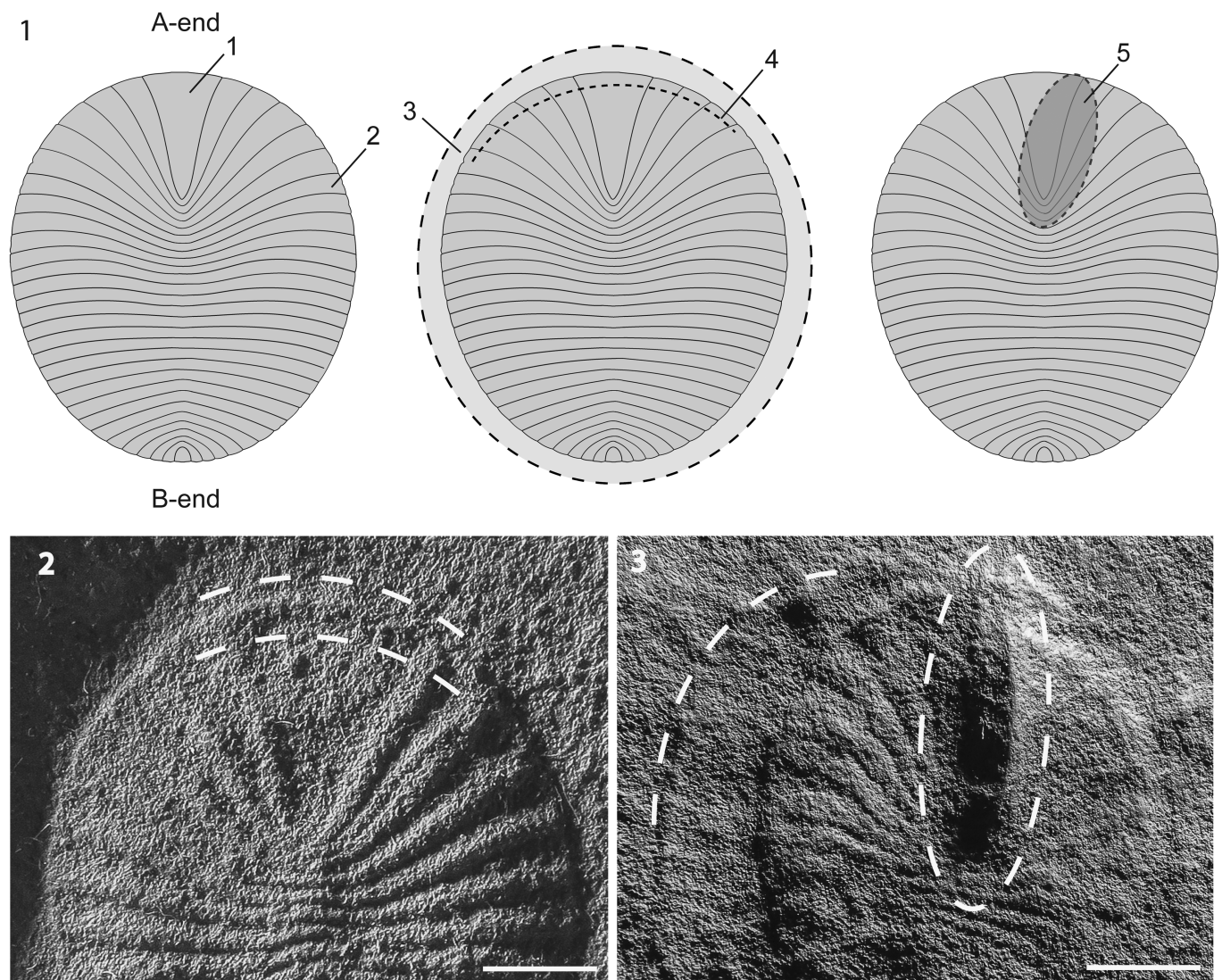


Figure 2. (1) Descriptive labels for *Dickinsonia costata* features: 1 = terminal A-end unit, 2 = body unit, 3 = position of shrinkage rim, 4 = position of A-end lip, 5 = generalized position of A-end protuberance. (2) Detail of A-end lip feature, visible as a narrow, raised rim on the peripheral margin of the terminal A-end unit, SAMP34228. (3) Detail of shrinkage rim feature as an uneven margin exterior to the body fossil, and the A-end protuberance as a high relief, elongate structure involving the terminal A-end unit and several A-end units, SAMP34232. The surface visible is assumed to be the upper surface. Scale bars = 3mm.

size distribution was best described by a single or multiple normal distributions, we fitted univariate Gaussian finite mixture models to specimen length using the function ‘Mclust’ from the R package ‘mclust’ (Fraley et al., 2012), and compared these by optimization (minimization) of the Bayesian Information Criterion (BIC).

Repository and institutional abbreviation.—The majority of specimens examined in this study are housed in the paleontology collections of the South Australia Museum, Adelaide, Australia (Prefix SAMP), and are currently on display in the “First Life: Ediacara Biota Gallery”. The remainder of specimens are located at the Crisp Gorge fossil locality. Specimens figured (Figs. 2, 4) are SAMP32351, SAMP34219, SAMP34224, SAMP34228, SAMP34232, SAMP34235, SAMP34243, SAMP34254 and SAMP34326. Please see Appendix A for the complete list of examined specimens and data set.

Results

Population size and shape.—*Dickinsonia costata* is constructed of repeated, parabolic units intersecting a narrow medial axis. The terminal A-end unit forms a deep V-shape, most noticeable in larger specimens; the curve of subsequent A-end units decreases towards the mid-point of the body. Units intersect the midline and no offset in units across the midline is observed. The midline lies perpendicular to units at the mid-point of the body. Preservation quality ranges from very good, in which all units are clearly visible, to poor, in which there is very little to no differentiation evident between A- and B-ends. Several specimens contained damaged or indistinct units and were excluded from unit-count data. All specimens are preserved in negative hyporelief and occupy a narrow size range, well below typical maximum size for *D. costata* (individuals may reach 25 cm in length). Specimens are 6.3–26.7 mm long, with a mean length of 13.28 mm, and are 5.8–22.7 mm wide, with a mean width of

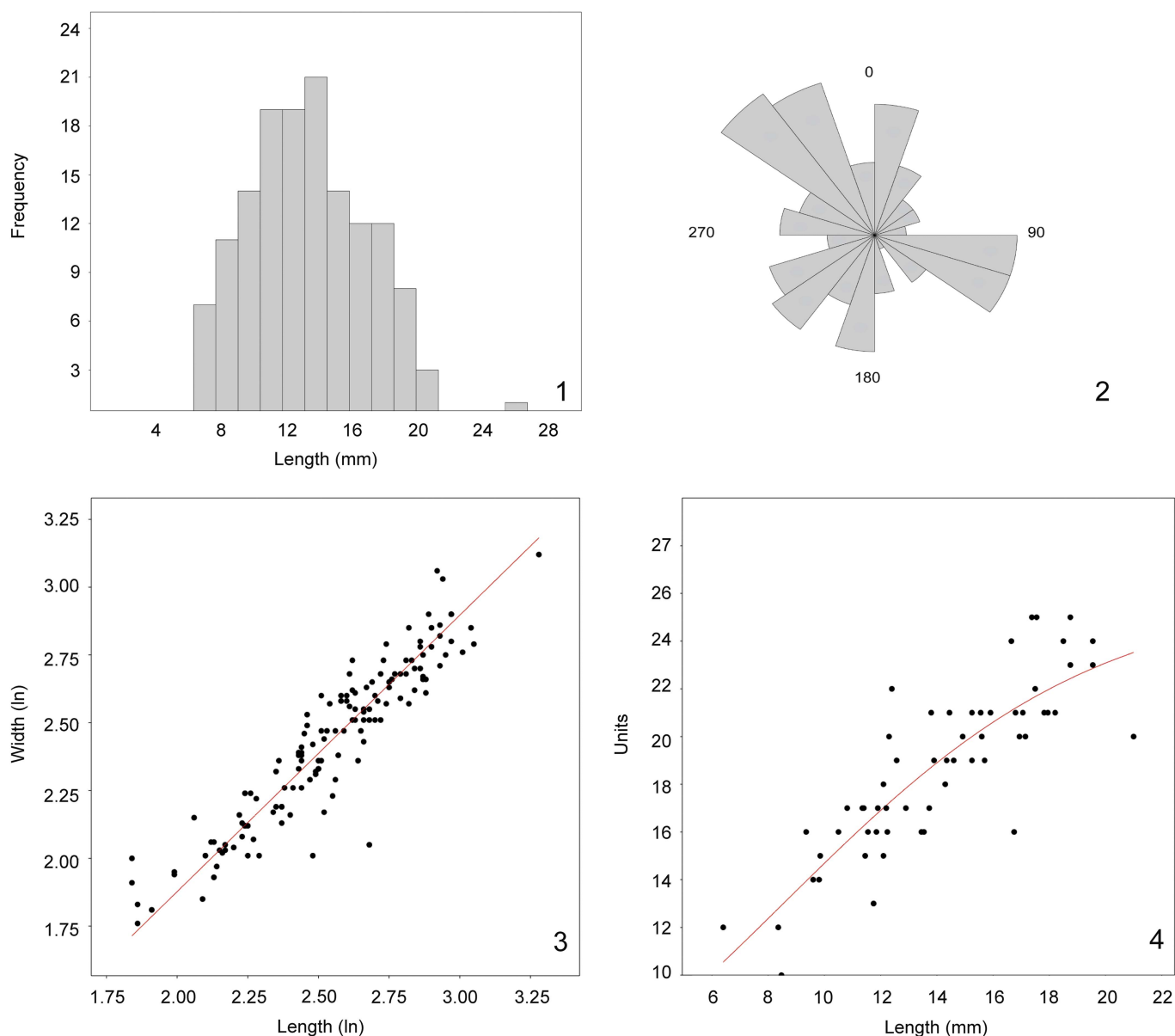


Figure 3. Analyses of *Dickinsonia costata* population. (1) Histogram showing size distribution of population (N = 140); (2) rose diagram showing axial alignment of specimens (N = 118); the ‘0’ is an arbitrary reference and denotes the top of the “Crisp Wall” frame; (3) graph depicting the relationship between (ln) length and (ln) width of specimens (N = 140); (4) logistic graph depicting the relationship between specimen length and number of observable units (N = 58).

11.86 mm. The major axis regression of (ln) length against (ln) width produced a regression slope of 1.019, with a 95% C.I. of (0.953, 1.089), which is not significantly different from one (Fig. 3.3). If a log-log model is best described by a straight line with slope (the allometric coefficient) equal to one, this suggests isometric growth (Hammer and Harper, 2006). This appears to be the case in *D. costata* over the size range examined (Runnegar, 1982; Evans et al., 2017). Comparisons between the linear and several non-linear models using the Akaike Information Criterion (AIC) suggested that a linear model was the best fit for these data (Fig. 3.3; Table 1). Unit number against length was best characterized by models that showed an increase in unit number with size, but at a decreasing rate (particularly for larger specimens). Of those tested, a logistic model best fitted the data based on the AIC (Fig. 3.4; Table 1). This suggests an upper limit on the number of units in *D. costata*, (Evans et al., 2017; Hoekzema et al., 2017).

Table 1. Akaike information criterion (AIC) values for the body-size variables of *Dickinsonia costata* from Crisp Gorge, Flinders Ranges. Length = body length, width = body width, units = number of observable body units. The relationship between body (ln) length and (ln) width is best described by a linear function, suggesting isometric growth. Number of body units present against length is best described by a nonlinear, logistic model.

Function	AIC Values	
	(ln) width ~ (ln) length	units ~ length
Linear	5.73	205.83
Quadratic	7.81	199.11
Power	7.80	201.69
Logistic	7.80	197.89
Gompertz	7.80	198.9

The size frequency histogram, produced using body length measurements (Fig. 3.1), describes the population distribution of *D. costata* from the Crisp Wall slabs and supplementary field

material as normal (Shapiro-Wilk; $W = 0.98$; $p = 0.07$). A single-component Gaussian finite mixture model best fitted the body length data based on optimization of the BIC, suggesting that these data are best described by a single distribution (rather than multiple distributions).

Axial orientation and paleocurrent direction.—Based on primary ripple shape and orientation of the bed top surfaces of the Crisp Wall, a primary paleocurrent direction of $\sim 55^\circ$ from north has been established (Reid et al., 2017). No consistent axial orientation is observed in the Crisp Wall specimens (Fig. 3.2). Hence, there appears to be no relationship between the axial orientation of *D. costata* and either paleocurrent or burial flow direction.

Morphological features.—A marked A-end protuberance occurs in 15% of the Crisp Gorge specimens (Figs. 2, 4). Broadly confined to the terminal A-end unit, this asymmetric bulge appears in deep negative relief relative to the surrounding organism, often developing into a point towards the peripheral margin. Larger protuberances extend for up to 5 mm, in smaller specimens encompassing nearly half the sagittal length. In larger specimens where greater detail is visible, protuberances often observed to be offset from the primary axis, to either the left or the right, towards and sometimes encompassing A-end units. This feature is observed in five other small *Dickinsonia* within the South Australian Museum collections. The deep relief and consistent positioning of the protuberance exclude the possibility that it is the result of the organism overlying a TOS feature within the substrate.

The A-end lip appears as a raised, upper-surface rim in nearly half (45%) of observed specimens (Figs. 2, 4). It is visible as a rim that outlines the body through the outer margin of the units at a thickness of no more than 1 mm. It occurs most commonly through the A-end and mid-body units, and is visible on some specimens distorting the peripheral margins of individual units.

A ‘shrinkage rim’ feature occurs in 38% of Crisp Gorge specimens, appearing as a positive relief rim extending 1–2 mm from the peripheral margin and in contrast to the negative hyporelief of the body mold (Figs. 2, 4). In most specimens, it does not extend the full circumference of the body. In some instances, however, a pronounced continual peripheral rim surrounds the body and can contain a secondary outline, interpreted as representing a second stage of shrinkage (Gehling et al., 2005). In several specimens where shrinkage is most marked, a faint impression of lower-surface units is visible within the shrinkage rim. A test of correlations between these two features shows a weak, but significant, positive relationship between the occurrence of a shrinkage rim and an A-end lip (Pearson’s correlation coefficient = 0.22, p -value < 0.01).

Discussion

The abundance and narrow size distribution of the juvenile population on the Crisp Wall surface suggest that *Dickinsonia costata* may have been an opportunistic species that established on this surface shortly after deposition and stabilization of the underlying substrate (Reid et al., 2017). *Dickinsonia* is noted

throughout all of the fossiliferous facies of the Ediacara Member in a broad range of sedimentary environments and at a range of localities, although more commonly in adult size ranges and usually at lower abundances (Droser and Gehling, 2015; Evans, 2015) than those observed in the Crisp Wall. At the Nilpena fossil locality on the western border of the Flinders Ranges, *Dickinsonia* is the third most abundant body fossil, exceeded only by *Funisia* and *Aspidella*, respectively, both of which represent anchored taxa (Gehling and Droser, 2013).

Opportunistic species display a range of identifying characteristics, many of which are demonstrated by *Dickinsonia*. They are typically generalists, capable of survival within a broad range of environmental and physiological parameters, as demonstrated by the identification of *Dickinsonia* in all fossiliferous facies of the Ediacara Member, and in situ within the majority of these paleoenvironments (Gehling and Droser, 2013). Species utilizing such a life history strategy may display a rapid increase in population numbers, often evidenced by the numerical domination of a given community or the rapid colonization of substrate previously devoid of organisms (Levinton, 1970). *Dickinsonia* comprises >50% of the Crisp Wall community, a trait that is not uncommon in other Ediacara Member communities (Droser and Gehling, 2015).

The Crisp Wall population displays minimal variance in size and is normally distributed, in contrast with other observed *D. costata* populations. Broadly speaking, *Dickinsonia* records a right-skewed population and a spread of body sizes spanning juvenile to adult (Zakrevskaya, 2014; Evans, 2015). There are two potential reasons for this discrepancy: (1) this population was the result of a single, isolated reproductive event; and (2) this population was buried in the very early stages of continuous or recurring, episodic recruitment, but was too immature to record a bimodal, clustered, or skewed distribution. Regardless, the distribution of this juvenile population does not negate the likelihood of longer-term continuous recruitment within the taxon, and supports the notion that the Crisp Wall surface records an immature, pioneer-stage community (Reid et al., 2017).

Based on the length to width ratio, *Dickinsonia costata* displays a juvenile isometric growth pattern (Fig. 3.3). This is consistent with previous findings, which have focused on larger individuals of the taxon and suggest that overall growth was isometric, with length and width increasing at a steady ratio throughout the majority of the organism’s lifecycle (Runnegar, 1982; Evans et al., 2017). This is despite the observed decrease in unit addition, indicating that overall shape is maintained independent of unit number (Evans et al., 2017; Hoekzema et al., 2017). Although all specimens are broadly ovoid in shape, variation in shape does occur. This may be accounted for by flexibility of the organism, and may be related to flexure resulting from movement of the organism in relation to the underlying microbial substrate. It may also be due to natural variation within the population. Likewise, a minor amount of size variation may be accounted for by the presence of the shrinkage rim in organisms that display this feature.

The lack of axial orientation noted in *Dickinsonia* both in this study and previously is in contrast to observations made for other Ediacaran forms (see Evans et al., 2015). Several Ediacaran taxa, including *Thectardis avalonensis*

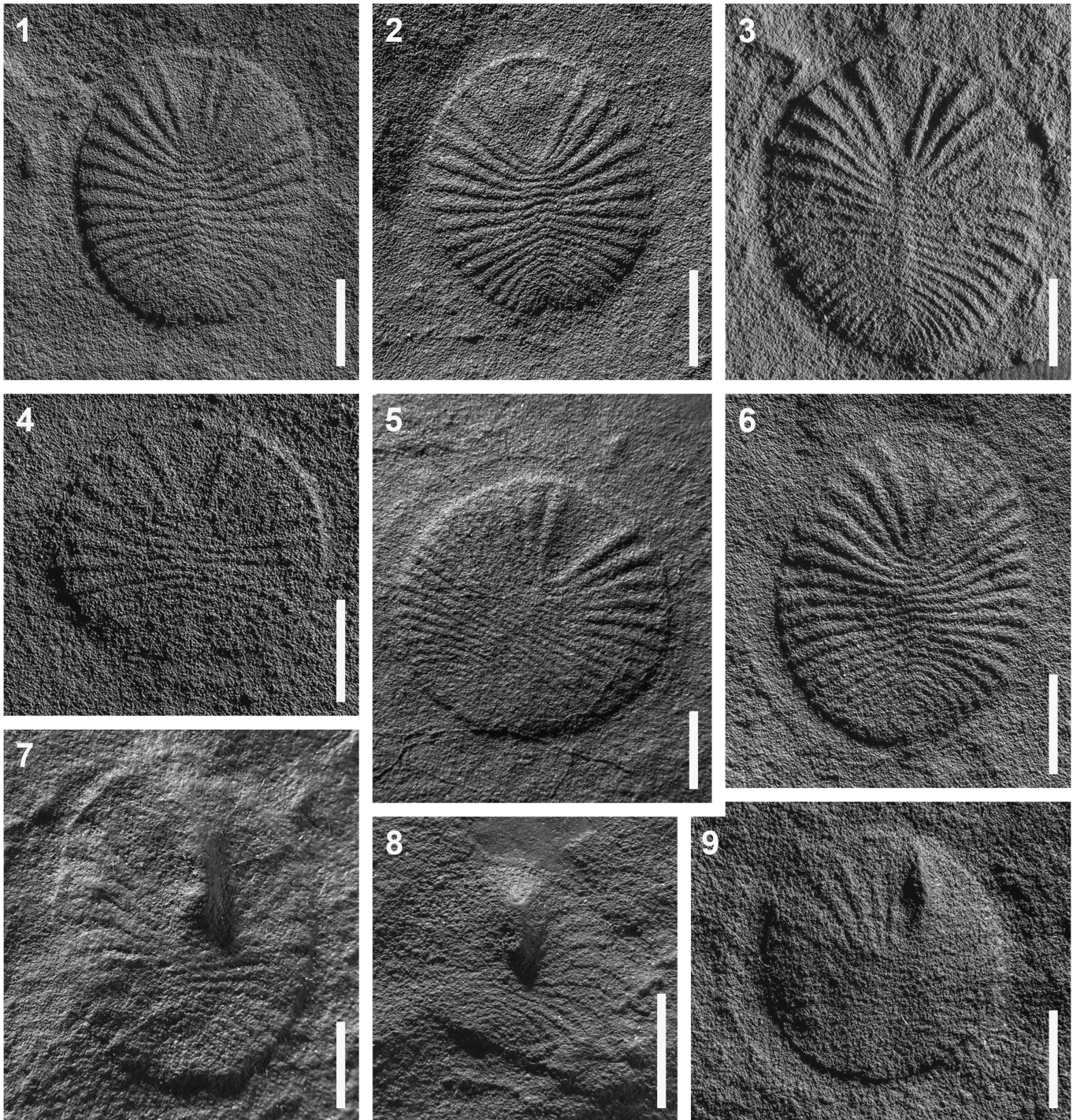


Figure 4. Photographs of *Dickinsonia costata* Sprigg 1947 from the Crisp Gorge fossil locality, Flinders Ranges (latex rubber casts). (1) Well-defined, non-continuous shrinkage rim and A-end lip, SAMP34235; (2) well-developed A-end lip present as narrow, high relief band in A-end units, SAMP34228; (3) continuous shrinkage rim visible around entirety of body, SAMP34243; (4) A-end lip visible on right side of body distorting perimeter of A-end units, SAMP32351; (5, 6) displaying continuous, uneven shrinkage rims SAMP34224, SAMP34219; (7, 8) A-end protuberance present as a bold, high relief structure encompassing terminal A-end units and involving subsequent A-end units, SAMP34232, SAMP34254; (9) A-end protuberance and shrinkage rim, SAMP34326. Scale bars = 5mm.

Clapham et al., 2004 and fronds, including *Charnia* Ford, 1958 have been demonstrated to have been manipulated by the uni-directional current flow at the time of their burial, effectively recording current direction of the burial flow with their final resting position (Wood et al., 2003; Clapham et al., 2004; Wilby et al., 2011). Recent evidence of rheotaxis in *Parvancorina*

suggests that an active response to current flow was employed, with individuals demonstrated to align with the current within two different paleoenvironments, most likely to minimize drag and ensure that current flow was directed over feeding structures (Paterson et al., 2017; Darroch et al., 2017). The absence of current alignment in *Dickinsonia* may be due in part to its

inferred life habit in near-constant contact with the underlying microbial substrate on which it likely fed (Sperling and Vinther, 2010). This close relationship with the dense, often ropey, substrate may have made *Dickinsonia* more or less impervious to current action, with the exception of higher-energy events lifting a small portion of the body away from the substrate (Evans et al., 2015).

The correlation between the shrinkage rim and A-end lip suggests that these features may have represented a physical response by the organism to the burial process. The shrinkage rim is also observed in *Dickinsonia* specimens from a number of localities across a range of sizes and has been interpreted as representing the shrinkage of the organism, due to a loss of mass associated with desiccation during early diagenesis (Gehling et al., 2005). Its correlation with the A-end lip suggests that this feature may also be due to the burial process, and that these features combined could be the result of a muscle-like contraction in response to the burial event, indicating displacement rather than loss of mass, and resulting in a thickening of the peripheral margin of the organism. The occurrence of this lip most commonly through the terminal A-end unit may be due to this unit representing the thinnest section through the fossil, and therefore making it most apparent in this location.

In contrast to these features, the striking A-end protuberance noted in this population is observed in only a limited number of small or juvenile *Dickinsonia* elsewhere in the South Australian Museum collections. Although broadly limited to the terminal A-end unit, placement of the protuberance within this region appears to be random. The deep-relief nature of the feature suggests that it was largely impervious to the burial process and was therefore likely constructed of the same material as the rest of the organism. Why it appears in such a high concentration in the Crisp Gorge material, but remains relatively rare in other *Dickinsonia* populations such as those at the Nilpena fossil locality, remains unclear. Given the juvenile size range of this population, however, it appears most likely that this feature is an artifact of early growth and may reflect a biological structure found exclusively in juveniles. Were it to be observed in a single individual, it may be dismissed as a preservational abnormality. It is unlikely, however, that this structure is taphonomic because there is nothing to suggest that the mode of preservation varied based on the size of the organism preserved.

Conclusions

The *Dickinsonia costata* population from Crisp Gorge provides an excellent opportunity to examine this taxon in juvenile form, and investigate the physical characteristics of such a population. *Dickinsonia* was likely a hardy opportunist with a single, juvenile size mode, which grew isometrically by maintaining its overall size ratio. The presence of the morphological features observed, including the shrinkage rim and A-end lip, requires further investigation in adult specimens, with the study of larger individuals perhaps providing higher-resolution material for examination. The A-end protuberance described in this population is most likely an artifact of early growth, however, further examination of juvenile *Dickinsonia* from other Ediacara surfaces will be helpful to confirm this.

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References

- Clapham, M.E., Narbonne, G.M., Gehling, J.G., Greentree, C., and Anderson, M.M., 2004, *Tectardis avalonensis*: a new Ediacaran fossil from the Mistaken Point biota, Newfoundland: *Journal of Paleontology*, v. 78, p. 1030–1036.
- Darroch, S.A., Rahman, I.A., Gibson, B., Racicot, R.A., and Laflamme, M., 2017, Inference of facultative mobility in the enigmatic Ediacaran organism *Parvancorina*: *Biology Letters*, v. 13, e20170033.
- Droser, M.L., and Gehling, J.G., 2015, The advent of animals: the view from the Ediacaran: *Proceedings of the National Academy of Sciences*, v. 112, p. 4865–4870.
- Evans, S.D., 2015, *Ecology and Biology of Dickinsonia*, an Iconic Member of the Ediacara Biota From Nilpena, South Australia [MSc Thesis]: Riverside, CA, University of California, Riverside, 43 p.
- Evans, S.D., Droser, M.L., and Gehling, J.G., 2015, *Dickinsonia* liftoff: evidence of current derived morphologies: *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 434, p. 28–33.
- Evans, S.D., Droser, M.L., and Gehling, J.G., 2017, Highly regulated growth and development of the Ediacara macrofossil *Dickinsonia costata*: *PLoS One*, v. 12, e0176874.
- Fraley, C., Raftery, A.E., Murphy, T.B., and Scrucca, L., 2012, package “mclust”: Version 4 for R: Normal Mixture Modeling for Model-Based Clustering, Classification, and Density Estimation, v. 5.3, <https://cran.r-project.org/web/packages/mclust/index.html>
- Ford, T.D., 1958, Pre-Cambrian fossils from Charnwood Forest: *Proceedings of the Yorkshire Geological and Polytechnic Society, Geological Society of London*, v. 31, p. 211–217.
- Gehling, J.G., and Droser, M.L., 2009, Textured organic surfaces associated with the Ediacara biota in South Australia: *Earth-Science Reviews*, v. 96, p. 196–206.
- Gehling, J.G., and Droser, M.L., 2012, Ediacaran stratigraphy and the biota of the Adelaide Geosyncline, South Australia: *Episodes - Newsmagazine of the International Union of Geological Sciences*, v. 35, p. 236.
- Gehling, J.G., and Droser, M.L., 2013, How well do fossil assemblages of the Ediacara Biota tell time?: *Geology*, v. 41, p. 447–450.
- Gehling, J.G., Droser, M.L., Jensen, S., and Runnegar, B.N., 2005, Ediacara organisms: Relating form to function, in Briggs, D.E.G., ed., *Evolving Form and Function: Fossils and Development*: New Haven, CT, Yale University, p. 43–66.
- Glaessner, M.F., 1958, New fossils from the base of the Cambrian in South Australia: *Transactions of the Royal Society of South Australia*, v. 81, p. 185–188.
- Glaessner, M.F., 1959, Precambrian Coelenterata from Australia, Africa and England: *Nature*, v. 183, p. 1472–1473.
- Glaessner, M.F., and Wade, M., 1966, The late Precambrian fossils from Ediacara, South Australia: *Palaentology*, v. 9, p. 599–628.
- Gold, D.A., Runnegar, B., Gehling, J.G., and Jacobs, D.K., 2015, Ancestral state reconstruction of ontogeny supports a bilaterian affinity for *Dickinsonia*: *Evolution and Development*, v. 17, p. 315–324.
- Gooden, B., 2014, Segmentation and oxygen diffusion in the ediacaran ‘*Dickinsonia*’: an applied analysis: *Journal and Proceedings of the Royal Society of New South Wales*, v. 147, p. 107.
- Hammer, Ø., and Harper, D.A., 2006, *Paleontological Data Analysis*: Oxford, Wiley-Blackwell, 356 p.

- Hammer, Ø., Harper, D.A.T., and Ryan, P.D., 2001, PAST: Paleontological statistics software package for education and data analysis: *Palaeontologia Electronica*, v. 4, no. 1, p. 1–9 http://palaeo-electronica.org/2001_1/past/issue1_01.htm
- Hoekzema, R.S., Brasier, M.D., Dunn, F.S., and Liu, A.G., 2017, Quantitative study of developmental biology confirms *Dickinsonia* as a metazoan: *Proceedings of the Royal Society B*, v. 284, p. 20171348.
- Ivantsov, A.Y., 2011, Feeding traces of proarticulata—the Vendian metazoa: *Paleontological Journal*, v. 45, p. 237–248.
- Legendre, P., and Legendre, L., 1998, *Numerical Ecology*: Amsterdam, Elsevier, *Developments in Environmental Modelling*, v. 20, 853 p.
- Levinton, J.S., 1970, The paleoecological significance of opportunistic species: *Lethaia*, v. 3, p. 69–78.
- Lund, U., and Agostinelli, C., 2015, package “circular”: Circular Statistics, v. 0.4–7, <https://cran.r-project.org/web/packages/circular/circular.pdf>
- Paterson, J.R., Gehling, J.G., Droser, M.L., and Bicknell, R.D., 2017, Rheotaxis in the Ediacaran epibenthic organism *Parvancorina* from South Australia: *Scientific Reports*, v. 7, 45539.
- RCoreTeam 2015, R: A language and environment for statistical computing: Vienna, Austria, R Foundation for Statistical Computing, <https://www.r-project.org>
- Reid, L.M., García-Bellido, D.C., Payne, J.L., Runnegar, B., and Gehling, J.G., 2017, Possible evidence of primary succession in a juvenile-dominated Ediacara fossil surface from the Flinders Ranges, South Australia: *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 476, p. 68–76.
- Retallack, G.J., 2007, Growth, decay and burial compaction of *Dickinsonia*, an iconic Ediacaran fossil: *Alcheringa*, v. 31, p. 215–240.
- RStudioTeam 2015, RStudio: Integrated Development for R: Boston, MA, RStudio, Inc., <https://www.rstudio.com>
- Runnegar, B., 1982, Oxygen requirements, biology and phylogenetic significance of the late Precambrian worm *Dickinsonia*, and the evolution of the burrowing habit: *Alcheringa*, v. 6, p. 223–239.
- Seilacher, A., Grazhdankin, D., and Legouta, A., 2003, Ediacaran biota: the dawn of animal life in the shadow of giant protists: *Paleontological Research*, v. 7, p. 43–54.
- Sperling, E.A., and Vinther, J., 2010, A placozoan affinity for *Dickinsonia* and the evolution of late Proterozoic metazoan feeding modes: *Evolution and Development*, v. 12, p. 201–209.
- Sprigg, R.C., 1947, Early Cambrian (?) jellyfishes from the Flinders Ranges, South Australia: *Transactions of the Royal Society of South Australia*, v. 71, p. 212–224.
- Tarhan, L.G., Droser, M.L., and Gehling, J.G., 2010, Taphonomic controls on Ediacaran diversity: uncovering the holdfast origin of morphologically variable enigmatic structures: *Palaios*, v. 25, p. 823–830.
- Tarhan, L.G., Droser, M.L., Gehling, J.G., and Dzaugis, M.P., 2017, Microbial mat sandwiches and other anactualistic sedimentary features of the Ediacara Member (Rawnsley Quartzite, South Australia): implications for interpretation of the Ediacaran sedimentary record: *Palaios*, v. 32, p. 181–194.
- Valentine, J.W., 1992, *Dickinsonia* as a polypoid organism: *Paleobiology*, v. 18, p. 378–382.
- Wade, M., 1972, *Dickinsonia*: polychaete worms from the late Precambrian Ediacara fauna, South Australia: *Memoirs of the Queensland Museum*, v. 16, p. 171–190.
- Wilby, P.R., Carney, J.N., and Howe, M.P.A., 2011, A rich Ediacaran assemblage from eastern Avalonia: evidence of early widespread diversity in the deep ocean: *Geology*, v. 39, p. 655–658.
- Wood, D.A., Dalrymple, R.W., Narbonne, G.M., Gehling, J.G., and Clapham, M.E., 2003, Paleoenvironmental analysis of the late Neoproterozoic Mistaken Point and Trepassy formations, southeastern Newfoundland: *Canadian Journal of Earth Sciences*, v. 40, p. 1375–1391.
- Zakrevskaya, M., 2014, Paleocological reconstruction of the Ediacaran benthic macroscopic communities of the White Sea (Russia): *Palaeogeography, Palaeoclimatology, Palaeoecology*, v. 410, p. 27–38.
- Zhang, X., and Reitner, J., 2006, A fresh look at *Dickinsonia*: removing it from Vendobionta: *Acta Geologica Sinica (English Edition)*, v. 80, p. 636–642.

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Appendix

Physical characteristics and orientation of *Dickinsonia costata* on Crisp Gorge “wall” surface. ¹Axial Orientation is measured relative to a north proxy (top horizontal bar of the Crisp Wall frame). Axial orientation was not

measured for specimens in which orientation was indistinct. ²Width and/or length measurements are not included for specimens that were damaged or the outline of the fossil was unclear.

SAMP Number	Axial orientation ¹	Width (mm) ²	Length (mm)	Units	Shrinkage rim	Anterior lip	Protuberance
P34200	168	11.65	11.55	16	0	1	0
P34202	73	10.55	12.15		1	0	1
P34203	198	13.15	15		0	0	0
P34205	216	10.54	13.99		0	1	0
P34206	332	14.65	15.25	19	0	0	0
P34207	357	14.2	14.8		0	0	0
P34209	241	11.5	12.4	22	1	1	0
P34212	103		15.85		1	1	0
P34213	320				0	0	0
P34217	212	9.55	11.45	15	0	1	0
P34218		10.8	13.1		0	0	1
P34219	103	13.75	17.15	20	1	1	0
P34220	142	13.15	13.45	16	1	1	0
P34221	110	12.3	14.35	19	0	0	0
P34222	58	7.5	11.9	14	1	1	0
P34223	360	9.4	9.6	14	0	0	0
P34224	335	17.3	18.1		1	1	0
P34226	173	10.3	11.4	17	0	1	0
P34228		12.65	14.3	18	0	1	0
P34229	268	10.2	12.1	15	0	1	1
P34230	118	8.4	9.3		0	0	0
P34232	323	13.9	15.7	19	1	0	1
P34233		12.75	14.3		0	0	1
P34234	142	15	18.75		0	0	0
P34235	109	14.2	15.6	20	1	1	0
P34236	189	16.8	18.75	25	0	0	0
P34237	83	8.95	10.7		0	0	0
P34238	353	17.25	16.75	16	1	0	0
P34239		18.25	19.55	23	1	1	0
P34240	110	13.45	13.25		0	1	0

(Continued)

SAMP Number	Axial orientation ¹	Width (mm) ²	Length (mm)	Units	Shrinkage rim	Anterior lip	Protuberance
P34241	25	7.55	8.7		0	0	0
P34242	204	12.25	14.65		1	0	0
P34244	249	14.35	17.6		1	1	0
P34245	204	16.4	17.4		1	0	0
P34257	263	8.45	10.75		0	1	0
P34258	213	16.25	21.1		1	1	0
P34259		12.75	14.6	19	1	1	0
P34260	113	12.75	13.9		0	0	0
P34261	78	8.9	10.5	16	1	1	0
P34262	112	9.6	11.1		0	1	0
P34263	218	9.35	9.4		0	1	1
P34264	342	11.25	11.9	17	1	1	0
P34266	77	7.5	9.45		0	1	0
P34267	19	8.35	9.35	16	0	0	0
P34268	188	18.2	17.95	21	1	0	0
P34269	106	14.65	16.65		0	0	1
P34270		9.9	11.85	16	0	1	0
P34277	347	13.75	13.8	21	1	1	0
P34278	60	11.8	14.2		0	0	0
P34279		13.55	13.9		0	1	0
P34280	127	11.1	11.45		1	0	1
P34281	326	10.55	12.3		0	0	0
P34282	348	10.6	11.45		0	0	0
P34283	329	10.55	10.55		1	1	0
P34285	142	13.5	12.3	20	0	1	0
P34287	187	8.9	10.75		0	1	0
P34288	51	7.65	8.8		0	1	0
P34289	263	12.35	14.9		1	1	0
P34291	347	11.8	12.3		1	0	1
P34294	114	10.87	11.45		0	0	0
P34295	126	10.85	11.45		0	0	0
P34297	126	15.7	17.55	25	1	0	0
P34299	252	14.29	17.8		0	0	0
P34300	328	15.35	13.75		1	0	0
P34301	72	11.4	14.3		0	0	0
P34302	321	11.8	12.9	17	0	1	0
P34303	86	12.3	15.25	21	0	1	0
P34304	67	13.05	16.8	21	0	0	1
P34305	276	17.45	18.75	23	1	0	0
P34312	270	18.18	19.55		0	1	1
P34313		6.25	6.45		0	0	0
P34314		6.35	8.1		1	0	0
P34316					1	1	0
P34317	357	14.3	15.85		0	0	0
P34318	263	8.7	11		1	0	0
P34320		12.6	11.75	13	0	1	0
P34321	224	16.4	19.55	24	0	0	0
P34322	45	10.1	12.1	18	0	1	0
P34323	137	7.65	8.6		0	0	0
P34324		5.8	6.4	12	0	0	0
P34325	260	22.65	26.7		1	1	0
P34326	138	13.25	13.2		1	0	1
P34327		8.75	12.45		0	1	0
P34328		7.5	9.85	15	0	1	0
P34329	229	13.35	16.3		1	0	1
P34330		7.8	8.75		0	1	0
P34331		15.8	20.3		1	0	0
P34333	222	7.9	9.7		1	1	0
P34337	42	8.75	10.35		0	0	0
P34339	73	12.35	13.9	19	0	1	0
P34340		9.5			0	0	0
P34341	89	10.3	12.2	17	1	1	0
P34342	243	14.9	17.5	22	1	1	0
P34345	126	15.65	19.05		1	0	1
P34346	179	8.7	9.25		1	1	0
P34347	124	7.8	14.6		1	1	0
P34348	152	7.45	8.15		0	0	0
P34349		9.3	12.85		0	0	0
P34204		7.03	7.35		0	0	0
P34208	243	9.88	12.88		0	0	0
P34210	330	12.89	13.54		0	1	0
P34215		7.69	9.03		0	0	0
P34216	126	8.6	7.84		0	0	0
P34217	212	10.32	12.24	16	1	1	0
P34225	158	15.37	15.32		1	0	0
P34226	173				0	0	0
P34227	262	17.27	21	20	1	1	0
P34231	291				0	0	0
P34243	238	14.94	17.06	21	1	1	0
P34246	97	8.34	9.4		0	0	0

(Continued)

SAMP Number	Axial orientation ¹	Width (mm) ²	Length (mm)	Units	Shrinkage rim	Anterior lip	Protuberance
P34247		6.14	6.78		1	0	0
P34248		13.09	15.49		1	1	1
P34249	111	14.58	15.91	21	0	0	0
P34250		6.98	7.31		0	0	0
P34251	156	16.3	15.56	21	0	1	1
P34252	143	14.64	13.54	16	0	1	0
P34253	153	10.94	11.35	17	0	1	0
P34256		6.88	8.39		0	0	0
P34265	171				0	0	0
P34273	3	15.36	16.94	20	1	1	0
P34274	223	12.33	13.73	17	0	0	0
P34275	331	8.33	9.49		0	0	0
P34284		6.75	6.32		0	0	0
P34286	155	13.5	13.47		0	0	1
P34290					0	0	0
P34293	325	7.88	8.41		0	1	0
P34294	114				0	0	0
P34296	96	16.06	17.38	25	1	0	1
P34306	13	14.45	17.58		0	0	0
P34307	78	7.99	9.32		0	0	0
P34308	306	12.08	11.65		1	1	0
P34309	66	13.54	17.82	21	1	0	1
P34310	328	7.88	8.36	12	0	1	0
P34311	132	20.7	18.92		1	0	0
P34332	270				0	0	0
P34334	125	11.82	12.57	19	1	1	0
P34335	213	13.92	14.45	21	0	1	0
P34336	222	14.52	16.3		0	0	0
P34343	267	16.16	18.21	21	1	1	0
P34344	31	11.83	13.27		0	1	0
P34356	58	9.55	10.81	17	0	0	0
P34357	287	7.19	8.47	10	0	0	0
P34254.1	168	7.36	6.3		0	1	0
P34254.2	354	10.14	10.51		1	0	1
Additional Crisp Gorge specimens							
B6		21.39	18.5	24	0	0	0
B7		13.13	12.69		0	0	0
C8		13.5	14.92	20	1	1	0
C9		9.17	9.81	14	0	0	0
C10		10.78	11.35		0	0	1
D11		15.38	16.65	24	1	1	1