

Research Article

Cite this article: Theisen S, Palm HW, Stolz H, Al-Jufaili SH, Kleinertz S (2018). Endoparasitic *Paradiplectanotrema klimpeli* sp. nov. (Monogenea: Ancyrocephalidae) from the Greater Lizardfish *Saurida tumbil* (Teleostei: Synodontidae) in Indonesia. *Parasitology Open* 4, e13, 1–11. <https://doi.org/10.1017/pao.2018.8>

Received: 3 January 2018

Revised: 26 February 2018

Accepted: 26 February 2018

Key words:

Three-dimensional confocal microscopy; co-evolution; common grinner; Dactylogyridae; *Diplectanotrema* group; LSU 28S rDNA sequence genotyping; phylogeny; *Pseudempleurosoma haywardi*; zoogeography

Author for correspondence:

Stefan Theisen, E-mail: Stefan.Theisen@uni-rostock.de

Endoparasitic *Paradiplectanotrema klimpeli* sp. nov. (Monogenea: Ancyrocephalidae) from the Greater Lizardfish *Saurida tumbil* (Teleostei: Synodontidae) in Indonesia

Stefan Theisen¹, Harry W Palm^{1,2}, Hendrik Stolz¹, Sarah H Al-Jufaili^{1,3} and Sonja Kleinertz^{1,4}

¹Aquaculture and Sea-Ranching, University of Rostock, Rostock, Germany; ²Centre for Studies in Animal Diseases, Udayana University, Denpasar, Indonesia; ³Laboratory of Microbiology Analysis, Fishery Quality Control Center, Ministry of Agriculture and Fisheries Wealth, Al Bustan, Sultanate of Oman and ⁴Faculty of Fisheries and Marine Sciences, Bogor Agricultural University, Bogor, Indonesia

Abstract

A new endoparasitic monogenean of *Paradiplectanotrema* Gerasev, Gayevskaya & Kovaleva, 1987, *Paradiplectanotrema klimpeli* sp. nov., is described from the southern Balinese coast, Indonesia. The new species is much larger, wider and characterized by the longest dorsal anchors compared with the congeners. Ventral anchors and ventral bars are the smallest in the genus, with a distinct ratio of 1:1. This is the first species with a gladiator breast-plate-shaped dorsal bar, with a length:width ratio of 1:1. Oesophagi of the Common Grinner *Saurida tumbil* (Bloch, 1795) (Synodontidae) were infected (prevalence = 17%) at an intensity of 12 (1–21). This is the first record of the genus from the eastern Indian Ocean, and lizardfishes represent a new host family. We provide light microscopy (*in situ* in oesophageal folds), three-dimensional confocal illustrations and a morphometric comparison of all congeners, with remarks on the recently described first Indonesian endoparasitic Monogenea *Pseudempleurosoma haywardi* Theisen, Palm, Al-Jufaili & Kleinertz, 2017. First 28S DNA sequences for *Paradiplectanotrema* allocate the new species close to endoparasitic freshwater monogeneans. Its ecology differs from *Pseudempleurosoma* Yamaguti, 1965 by utilizing deep-water fishes instead of coastal, coral reef-associated hosts; however, both are infecting schooling, bottom-dwelling fishes.

Introduction

Monogenea, a class of the Platyhelminthes, are typically host-specific monoxenous ectoparasites of fish, infecting especially their skin and gills. This can lead to mass mortalities when secondary infections (e.g. bacteria, virus and fungi) are triggered, e.g. under aquaculture conditions (Zhang *et al.* 2015; Kotob *et al.* 2016). Cephalopods and, more rarely, amphibians, reptiles and cetaceans can be parasitized, one species infects the hippopotamus.

Only few species became endoparasitic in fishes, utilizing e.g. the urogenital and gastro-intestinal system. Besides showing a simple life cycle, a posterior attachment organ called (opist)haptor, armed with clamps, anchors and/or hooks, is unique and differentiates the hermaphroditic Monogenea from other flatworms (Mehlhorn, 2001). Theisen *et al.* (2017) shed light on the morphology of the often misinterpreted anchor apparatus of *Pseudempleurosoma* Yamaguti, 1965, presenting the first laser confocal images of a '*Diplectanotrema* group' member. Within this group (*Diplectanotrema* Johnston and Tieg, 1922; *Neodiplectanotrema* Gerasev, Gayevskaya & Kovaleva, 1987; *Paradiplectanotrema* Gerasev, Gayevskaya & Kovaleva, 1987; and *Pseudodiplectanotrema* Gerasev, Gayevskaya & Kovaleva, 1987), the species are adapted to attach to the inner epithel cells of the oesophagus, stomach and intestine of a wide range of host species, which is unusual for the commonly ectoparasitic and highly host specific Monogenea. *Pseudempleurosoma haywardi*, Theisen *et al.*, 2017, from the oesophagus of croakers (Sciaenidae) was only recently described as the first endoparasitic Monogenea from Indonesia (Theisen *et al.* 2017).

The genus *Paradiplectanotrema* was established by Gerasev *et al.* in 1987, describing the four species *P. antgoni*, *P. chlorophthalmi*, *P. cytti* and *P. lepidopi*, all from N, NW and SW Africa (Atlantic Ocean). They also transferred *Diplectanotrema trachuri* Kovaleva, 1970 into *Paradiplectanotrema* as the new type species *P. trachuri* (Kovaleva, 1970) Gerasev, Gayevskaya & Kovaleva, 1987. According to the key to the genera by Theisen *et al.* (2017), *Paradiplectanotrema* differs from the above mentioned genera by having a combination of (i) an accessory piece on the male copulatory organ (MCO) and two ventral anchors with (ii) one ventral bar each (one pair of ventral anchors and one pair of ventral bars).

The monogenean species described herein represents a new member of *Paradiplectanotrema*, the first record of this genus from Indonesia, and the second endoparasitic Monogenea species from the eastern Indian Ocean. We document Synodontidae as a new fish host family

© Cambridge University Press 2018. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted re-use, distribution, and reproduction in any medium, provided the original work is properly cited.

CAMBRIDGE
UNIVERSITY PRESS

for *Paradiplectanotrema*. A morphometrical comparison is made among the valid species within this genus, also *via* detailed three-dimensional confocal microscopy, and first DNA sequence data are presented, securing the phylogenetic position of these marine endoparasites. For the first time, *in situ* images of endoparasitic monogeneans attached to tissue of the oesophageal folds inside their hosts are presented.

Materials and methods

Sample collection and processing

The synodontid grinner *Saurida tumbil* (Bloch, 1795) ($n = 24$) were sampled from Kedonganan fish market, South Bali coast, Indonesia (8°45'25.60"S, 115°10'05.94"E) in November 2016 (rainy season). The fishes were transferred on ice to the Marine and Fisheries Faculty Laboratory, Udayana University (UNUD), Kampus Bukit, Jimbaran, Bali, Indonesia.

Morphometrical data for each fish were taken [total and standard lengths (TL and SL) to the nearest 0.1 cm, total and gutted weight (TW and GW) to the nearest 0.1 g]. The body cavity was opened, the stomach was cut at the level of the pharynx right behind the gill rakers and transferred to Petri dishes containing NaCl solution (0.9%), and studied for parasites under a Zeiss Stemi DV4 binocular microscope. Endoparasitic monogenean parasites were isolated from the pharyngeal/oesophageal folds, washed in saline solution and pre-identified under a Novel XSZ-107BN microscope. Most worms were fixed using AFA (Alcohol: Formalin: Acetic acid) or 4% neutral buffered formalin and then transferred and stored in 70% EtOH for further morphological analyses (light microscopy). Some were either compressed and directly transferred to 95% ethanol for confocal laser scanning microscopy or to 99.8% EtOH for DNA analysis. Type specimens are deposited in the Berlin Natural History Museum, Germany ['Museum für Naturkunde', catalogue 'Entozoa', collection 'Vermet', numbers E.7439 (holotype) and E.7440a–E.7440 h (paratypes)] and the Zoological Museum Bogor, Indonesia (MZBTr240–MZBTr245, paratypes). The ecological parameters for fish infections with *Paradiplectanotrema klimpeli* sp. nov. (prevalence, intensity and abundance) were calculated following Bush *et al.* (1997).

Light microscopy, drawing and morphological investigation

Following standard parasitological methods, specimens were cleared and mounted with glycerin after fixation. Some compressed samples from 70% ethanol were prepared as whole mounts, stained with acetic carmine and studied for sclerotized organs. Measurements are given as the range followed by mean in parentheses (in micrometers), according to the genus description by Gerasev *et al.* (1987) and Kovaleva (1970), respectively. A camera lucida drawing tube was used to illustrate the new species while photographs were taken with a digital camera Olympus UC30 attached to an Olympus BX53 DIC light microscope or a Zeiss SZX10 binocular magnifier, supported with Cellsens Dimension software (Olympus Soft Imaging Solutions GmbH) for measurements.

Confocal microscopy

Confocal laser scanning microscopy was performed following the procedure described in Galli *et al.* (2006) and Marchiori *et al.* (2015) with specimens fixed in 95% ethanol, using a Leica TCS SP2 confocal microscope (laser with HeNe at 543 nm wavelength, minimum setting: 200 z section) equipped with an inverted Leica DMIRE2 microscope and a PL APO 363 oil immersion objective

(×63 magnification, numerical aperture 5 1.4) at the Live Cell Imaging Center, Department of Biology, University of Rostock.

DNA analysis

For molecular analysis, total genomic DNA was extracted from individual adult worms fixed in 99.8% ethanol using QIAamp mini DNA kit (Qiagen) according to the manufacturer's instructions with some modification. In order to increase the DNA yield, the obtained DNA was completely evaporated using Concentrator plus/Vacufuge® (Eppendorf), dehydrated DNA was then re-suspended with 50 µL of elution buffer and stored in a –20°C freezer. Amplification of D1–D3 fragments of the large subunit region (LSU) was achieved using the following primers: C1 (forward; 5'-ACCCGCTGAATTTAAGCAT-3') and D2 (reverse; 5'-TGGTCCGTGTTTCAAGAC-3') (Mendlová *et al.* 2010). The reaction was performed using illustra™ puReTaq Ready-To-Go PCR beads (0.2 ml tubes, 96 reactions), containing 5 pmol of each primer, 5 µL of DNA suspension and nuclease-free water to a total volume of 30 µL. Obtained PCR products (1 µL) were viewed on a 0.8% agarose gel stained with ethidium bromide. Contiguous sequences were aligned and assembled using BioEdit v.7.0.9 (Hall, 1999). The generated sequences [GenBank accession numbers (GBAN) MG763101] were aligned with their closest matches in GenBank [15 ingroup and *Actinocleidus recurvatus* (GBAN AJ969951) as an outgroup taxa]. Phylogenetic analysis were performed in MEGA version 7 (Kumar *et al.* 2016) based on the best scoring model (Bayesian Information Criterion), maximum likelihood was used for the best fitting tree based on the Tamura–Nei model, rates among sites were gamma distributed with invariant sites (G + I), the number of discrete gamma categories was 5, and using complete deletion of gaps as gaps missing data treatment. The robustness of the inferred phylogeny was assessed using a bootstrap procedure with 1000 replications (Wu *et al.* 2005).

Nomenclatural acts

The description of *P. klimpeli* sp. nov. (Life Science species Identifier number (ZooBank LSID): urn:lsid:zoobank.org:act:18989757-3EE3-462E-A389-491A132A6672) complies with the requirements of the International Commission on Zoological Nomenclature (ICZN). The electronic edition of this article conforms to the requirements of the amended International Code of Zoological Nomenclature, and hence the new names contained herein are available under that Code from the electronic edition of this article. This published work and the nomenclatural acts it contains have been registered in ZooBank, the online registration system for the ICZN. The ZooBank LSIDs (Life Science Identifiers) can be resolved and the associated information viewed through any standard web browser by appending the LSID to the prefix '<http://zoobank.org/>'. The LSID for this publication is: urn:lsid:zoobank.org:pub:F44C225F-795C-462F-BD33-D1E4BBED9067. The electronic edition of this work was published in a journal with an ISSN, and has been archived and is available from the following digital repositories: PubMed Central and ResearchGate. DNA sequences are available in GenBank under the GBAN MG763101.

Ethics

No experiments were performed on live vertebrates. Freshly caught dead fish was used and therefore no ethics statement is required. Samples were taken in cooperation with Udayana University, Denpasar, Bali under a valid Memorandum of Understanding (MoU). The fishes were obtained from an official fish market (see the 'Material and Methods' section), national laws regulate captures and fishermen/salesmen are

licensed, sampled fish species are common in Indonesia, not protected, and the number of sampled 24 fish were all available individuals.

Results

Taxonomy and description

Family Ancyrocephalidae Bychowsky & Nagibina, 1968

Paradiplectanotrema Gerasev, Gayevskaya & Kovaleva, 1987

History of type species: *Diplectanotrema trachuri* Kovaleva, 1970 became *P. trachuri* (Kovaleva, 1970) Gerasev, Gayevskaya & Kovaleva, 1987

Paradiplectanotrema klimpeli sp. nov.

urn:lsid:zoobank.org:act:18989757-3EE3-462E-A389-491A132A6672

Type-host: Common/Greater Lizardfish/Grinner, *Saurida tumbil* (Synodontidae)

Type-locality: Kedonganan fish market, South Bali coast, Indonesia (8°45'25.60"S, 115°10'05.94"E)

Habitat: oesophageal/pharyngeal folds

Type-material: holotype E.7439; paratypes E.7440a–E.7440h (paratypes) and additional paratypes MZBTr240–MZBTr245

Deposition of specimens: Berlin Natural History Museum, Germany ['Museum für Naturkunde', catalog 'Entozoa', collection 'Vermetes', numbers E.7439 (holotype) and E.7440a–E.7440h (paratypes)], the Zoological Museum Bogor, Indonesia (MZBTr240–MZBTr245, additional paratypes)

Infection: four of 24 fishes harboured 47 specimens, prevalence 17%, mean intensity 12, intensity 1–21, mean abundance 2

Etymology: the specific name is to honor Dr Sven Klimpel for his work on marine fish parasites

Description (all μm) (Figs 1–3): based on 17 specimens from the type host *Saurida tumbil*

Body slender, unspined, 2050–2880 (2390) long; maximum width 600–980 (800) (posterior to testis). Pair of small head glands anterior to oesophagus lateral to eye spots; second, larger pair lateral at level of intestinal bifurcation. Two pairs of eye-spots. Oral aperture ventral; pharynx globular, 106–164 \times 112–167 (140 \times 142.5) (length \times width) (Fig. 1). Intestinal caeca overlaid posterior to testis, coexisting with vitelline follicles, with lateral diverticula lacking haematin pigment. Haptor, 92–131 \times 100–147 (116.5 \times 125.5) (length \times width), not distinctly set off from body, but laterally slightly lobed; with two pairs of dissimilar hamuli (~anchors) and 14 marginal hooks (~hooklets) (Figs 1–3). Dorsal hamuli, from the tip of the ventral root to the height of curve of the blade (with the inner dorsal root being shorter than the outer ventral root in this genus), 55–62 (58) in length, connected by a single gladiator breast-plate shaped dorsal bar with a 1:1 length:width ratio (Figs 1–3), 15–19 (17) from end to end (=width), with a distance of 14–19 (16.5) from anterior to posterior border (=length) (Figs 1 and 3). Ventral hamuli 9–11 (10) in length, each one with one wide (>5) (connected) bar of 9–13 (11) length (Figs 1 and 3). Marginal hooks 11–15 (13) long (Figs 1 and 3). MCO sclerotized, tubular, 67–96 (79.5) long, with seminal vesicle and prostatic reservoir at base; accessory piece sclerotized, irregular in shape, 25–32 (27.5) long (Figs 1 and 3). Testis post-ovarian, shaped like a rounded cumulus cloud, widest anterior, median or posterior, 120–237 \times 110–225 (183 \times 180) (length \times width) (Fig. 1). Ovary turned back on itself, giving an appearance of being kidney shaped, wider than long, 122–159 \times 176–239 (143.5 \times 207) (length \times width) (Fig. 1). Uterus hose shaped, ends as muscular aperture of 33–63 \times 40–59 (51.5 \times 49) (length \times width), in a smooth genital atrium, almost round in shape (Fig. 1). Vagina simple, opens at the level of muscular genital atrium. Vitelline follicles arranged longitudinally in lateral fields along body (Figs 1 and 2). Vitelline ducts unite in

a slender line directly anterior to ovary. Eggs oviform, 50–104 \times 46–66 (84 \times 52) (length \times width) (Fig. 1).

Remarks

Based on Santos *et al.* (2001) and Theisen *et al.* (2017), the new species belongs into *Paradiplectanotrema* due to the presence of an accessory piece on the MCO and one bar associated with each ventral anchor as well as one bar connecting the two dorsal anchors (one pair of dorsal anchors, one pair of ventral anchors, one pair of ventral bars, one dorsal bar).

The specimens represent a new species, as seen in Table 1. While the sizes of the marginal hooklets, MCO and eggs are similar in all species of this genus (Table 1), other morphological measurements are distinct. *Paradiplectanotrema klimpeli* sp. nov. differs from all congeners by having a longer body with a mean of 2390 (2050–2880). So far, only few specimens of *P. trachuri* have been reported being longer than 2 mm; however, the mean length of *P. trachuri* [1680 (1030–2220)] is smaller (see Table 1). The body width of *P. klimpeli* sp. nov. [800 (600–980)] is wider than the congeners' (mean of 450–500) (Table 1). The widest part of the body lies half way between testis and opisthaptor (while in *Pseudempleurosoma* it lies at the level of the testis). With a length of 55–62 (58), the dorsal anchors are the longest in the genus, even though the size of the dorsal anchor of *P. antigoni* is almost as large (up to 60 with a mean of 57.5). Thus, the dorsal anchor length alone is characteristic to distinguish *P. klimpeli* sp. nov. from its congeners except *P. antigoni*.

A unique character of the new species is the shape of the dorsal bar being gladiator breast-plate shaped, with a 1:1 length:width ratio, while all so far described dorsal bars of the congeners have different shapes and ratios (Gerasev *et al.* 1987). Thus, the two structures of the dorsal anchor complex, namely, the dorsal hooks and the dorsal bar of *P. klimpeli* sp. nov., are unique. The ventral anchor is the smallest in this new species. With sizes of 9–11 (10), it differs from its congeners with means between 12.5 and 15. The bar of the ventral hamuli has smaller sizes (mean 11) compared with the congeners (means between 12.5 and 16) (Table 1).

The amount of eggs in *P. klimpeli* sp. nov. is noteworthy, with up to four eggs per specimen, similar to other species within the genus (up to three eggs), but in comparison with *P. haywardi*, that had nil to a maximum of one egg *in utero*. Other differences from the congeners is the host species, with the Synodontidae representing a new host family for the genus, and the geographic range in Indonesia, South-East Asia. The sampled grinders had sizes of 14.5–34.2 cm SL, with TW of 32.6–447.2 g. The infected four-type host fish specimens were comparably large, with 26.6/239.5, 29.9/285.1, 29.6/304.4 and 34.2/447.2 [SL (cm)/TW (g)].

DNA

The partial LSU rDNA sequences were 838 bp long (with primers). A NCBI GenBank blast showed closest similarity to the only so far available sequence of a member of the *Diplectanotrema* group, namely *P. haywardi* (GBAN MF115714–MF115717), followed by freshwater endoparasitic monogeneans, namely *Enterogyrus* 'sp. 1 AS-2010', *Enterogyrus* 'sp. 2 AS-2010' [both *ex Sarotherodon galilaeus* (L.)], and *Enterogyrus coronatus* Pariselle, Lambert & Euzet, 1991 *ex Coptodon zillii* (Gervais, 1848) (GBAN: HQ010032, HQ010031, HQ010030). The obtained sequence of *P. klimpeli* sp. nov. formed a well-supported sister clade with *P. haywardi* (bootstrap value of 100%) (Fig. 4). The new sequence is available under the GBAN MG763101.



Fig. 1. Drawings of *Paradiplectanotrema klimpeli* sp. nov. Holotype from the type fish host *Saurida tumbil* (A), with detailed view of reproductive system with genital atrium/disc (mga), male copulatory organ (MCO) with accessory piece, and prostatic reservoir (pr) and gland at base; and egg in uterus (B) and opisthaptor with anchors (one dorsal and one ventral pair), bars (one between dorsal anchors and one attached to each ventral anchor = three) and seven pairs of hooklets (C); scale bars A: 100 μ m; B & C: 10 μ m.

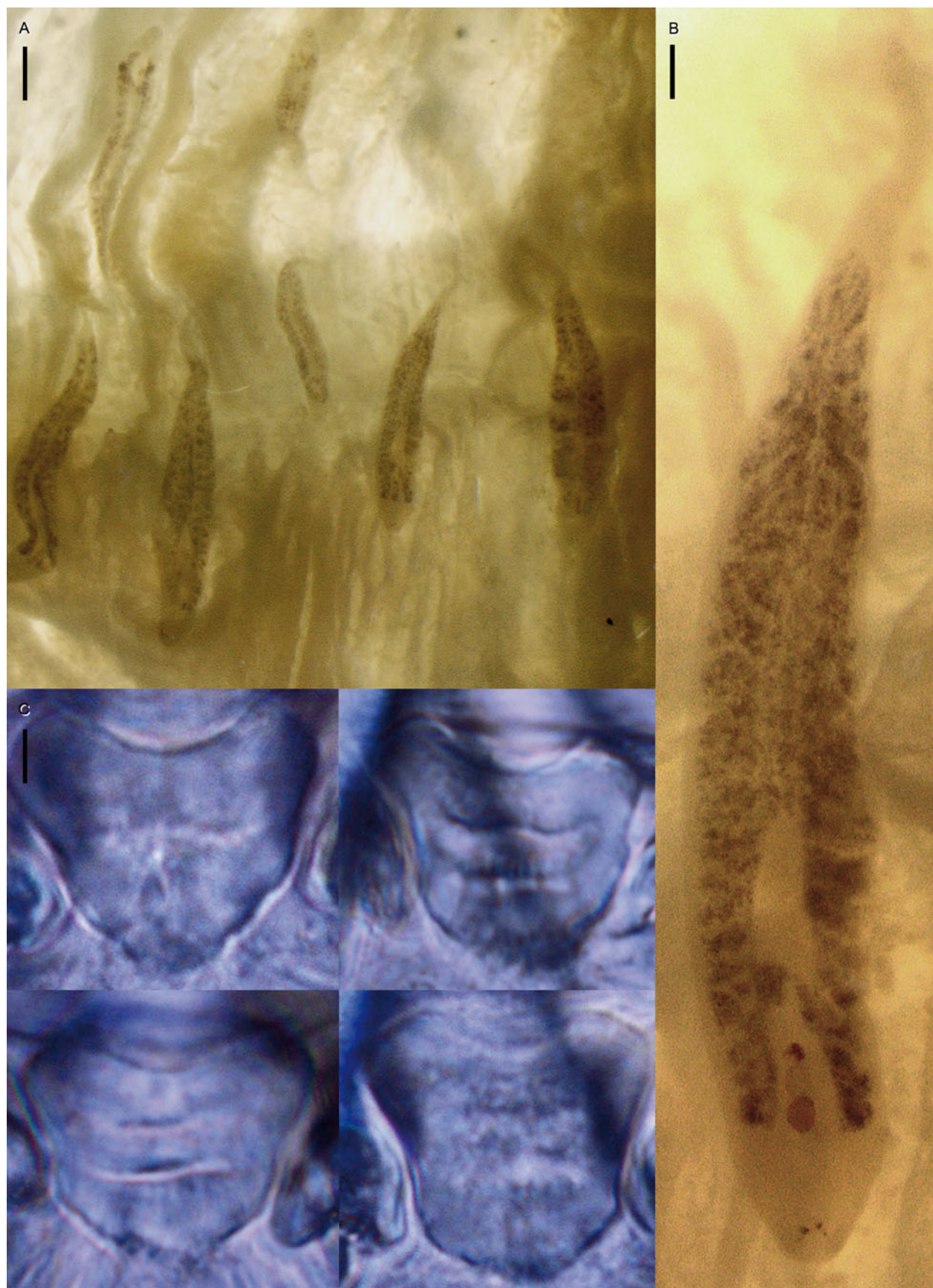


Fig. 2. Light microscopy illustrations of *Paradiplectanotrema klimpeli* sp. nov. Worms *in situ*, attached to its hosts oesophageal epithel folds (A, binocular), detailed individual *in situ* (B), gladiator breast-plate shaped dorsal bar of four different individuals (C, DIC).

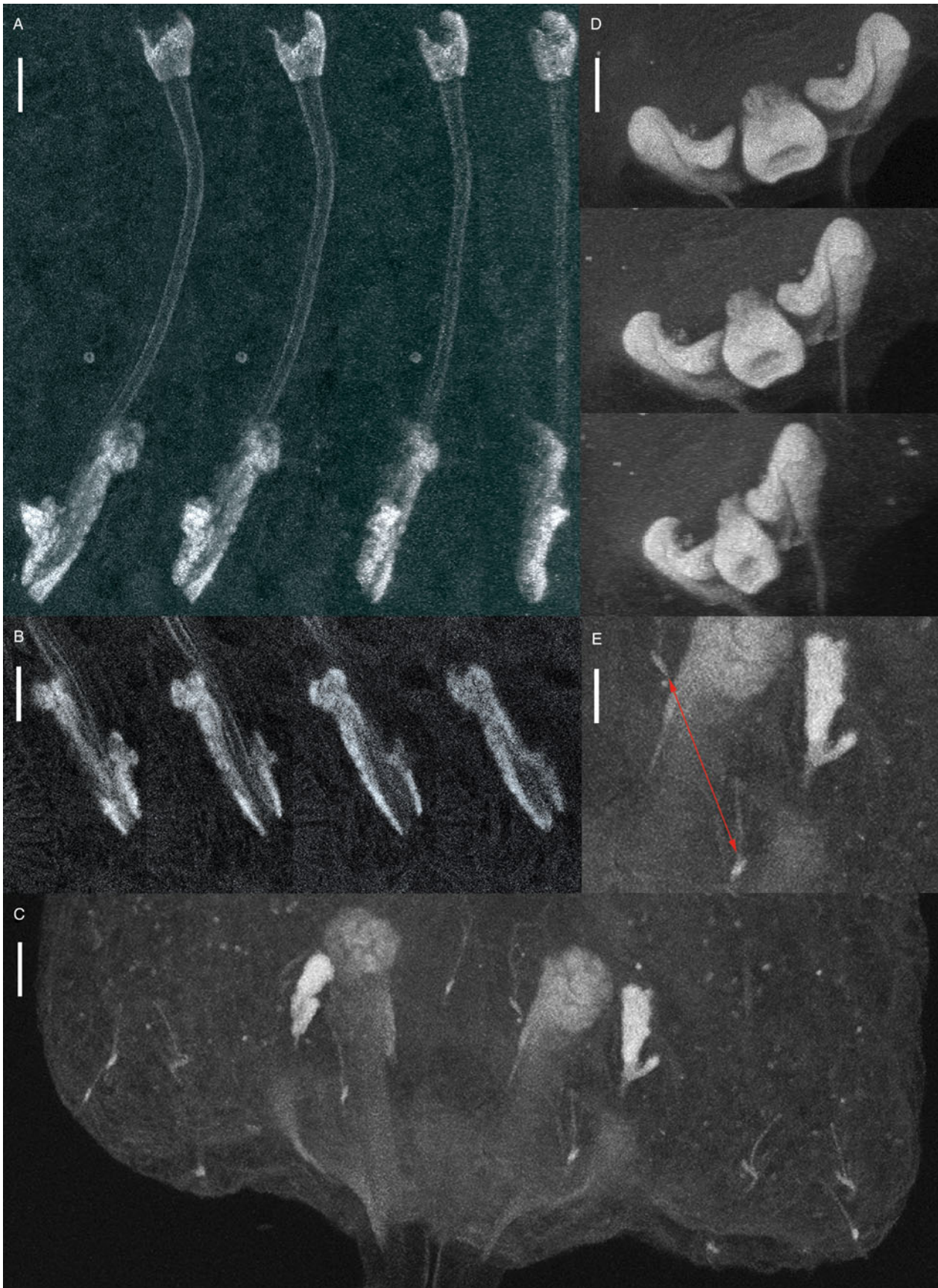


Fig. 3. Confocal laser scanning illustrations of *Paradiplectanotrema klimpeli* sp. nov. Male copulatory organ (MCO) in different angles (A) and levels (B), haptor (C) with dorsal anchors' saucer-type excrescences and dorsal bar stained, from different angles (D), and detailed view of two central hooks (red arrow), base of a dorsal anchor and a ventral anchor with its associated, irregular ventral bar (E); scale bars: A–D: 10 μ m; E: 15 μ m.

Table 1. Morphological measurements of the now six species of *Paradiplectanotrema*

Measurements (all μm)	<i>P. antigoni</i> Gerasev, Gayevskaya & Kovaleva, 1987 <i>n</i> = 2	<i>P. chlorophthalmi</i> Gerasev, Gayevskaya & Kovaleva, 1987 <i>n</i> = 7	<i>P. cytti</i> Gerasev, Gayevskaya & Kovaleva, 1987 <i>n</i> = 1	<i>P. lepidopi</i> Gerasev, Gayevskaya & Kovaleva, 1987 <i>n</i> = 23	<i>P. trachuri</i> (Kovaleva, 1970) Gerasev, Gayevskaya & Kovaleva, 1987 <i>n</i> = 16 in re-description	<i>P. klimpeli</i> sp. nov., present study <i>n</i> = 17
Body (length \times width)	1000 ^a \times 360–550 (1000 \times 450)	700–1660 \times 280–970 (1330 \times 500)	1620 \times 280 ^a	780–2190 \times 280–620 (1400 \times 470)	1030–2220 \times 300–650 (1680 \times 450)	2050–2880 \times 600–980 (2390 \times 800)
Opisthaptor (length \times width)						92–131 \times 100–147 (116.5 \times 125.5)
Pharynx (length \times width)						106–164 \times 112–167 (140 \times 142.5)
Ovary (length \times width)						122–159 \times 176–239 (143.5 \times 207)
Testis (length \times width)						120–237 \times 110–225 (183 \times 180)
Dorsal anchor	55–60 (57.5)	40–50 (45.5)	55 ^a	48–58 (51.5)	45–55 (50)	55–62 (58)
Dorsal bar (length \times width)	20 \times 12.5 ^a	12.5–20 \times 10–17.5 (19 \times 11.5)	15 \times 12.5 ^a	15–20 \times 9–18 (17.5 \times 11.5) ^b	12.5–20 \times 7.5–15 (17 \times 11)	14–19 \times 15–19 (16.5 \times 17)
Ventral anchor	15 ^a	14.5 ^a	15 ^a	12–15 (14) ^b	11.5–15 (12.5)	9–11 (10)
Ventral bar	16 ^a	12.5 ^a	13.5 ^a	10–16 (13.5) ^b	10–16 (12.5)	9–13 (11)
Marginal hooks (14 pcs)	14.5 (/15) ^{a,b}	12.5–15 (14)	12 ^a	10–16 (13) ^b	10.5–15.5 (13.5)	11–15 (13)
Male copulatory organ (MCO)	79–97.5 (88.5) \times 1	85.5–110 (99) \times 1.5	80 \times 1 ^a	86–92 (88.5) \times 1	70–80 (75) \times 2	67–96 (79.5)
Accessory piece of MCO						25–32 (27.5)
Muscular genital atrium (length \times width)						33–63 \times 40–59 (51.5 \times 49)
Egg (length \times width)		77–90 \times 42.5–80 (86.5 \times 63)	72.5 \times 57.5 ^a	67.5–75 \times 42.5–62 (67 \times 62) (/70 \times 48) ^b	65–77.5 \times 45–67 (67.5 \times 50)	50–104 \times 46–66 (84 \times 52)
Hosts	<i>Antigonia capros</i> (Caproidae)	<i>Chlorophthalmus atlanticus</i> , <i>C. agassizi</i> (Chlorophthalmidae)	<i>Cyttus</i> cf. <i>traversi</i> ^c (Zeidae)	<i>Lepidopus caudatus</i> (Trichiuridae)	<i>Trachurus trachurus</i> , <i>T. mediterraneus</i> , <i>T. picturatus</i> (Carangidae)	<i>Saurida tumbil</i> (Synodontidae)
Geographical area	Atlantic: Off Guinea-Bissau (NW Africa)	Atlantic: Off Guinea-Bissau & Angola (NW & SW Africa)	Atlantic: Off N Africa	Atlantic: Off Western Sahara (NW Africa)	Atlantic: Off Western Sahara; Mediterranean: Gibraltar channel (NW Africa)	Pacific: Off South Bali, Indonesia

Main differences are in bold.

^aOnly mean/one value is given in Gerasev *et al.* (1987) (thus, no statistical tests could be performed); note that details of former measurements are only available for the species *P. lepidopi* and *P. trachuri* (both show similar values and were sampled in the same region), although all species were (re-)described in one publication (see text) with *n* > 1 for each species except for *P. cityi* (see also discussion).

^bGerasev *et al.* (1987) provided different values for this character within one species (partially shown in *italics*), resulting from mounting (squeezing) techniques and/or omitting juvenile specimens from the measurements.

^cGerasev *et al.* (1987) identified *Cyttus* sp. as host, however, out of the three valid *Cyttus* spp., only *C. traversi* is known from the African West coast (Atlantic).

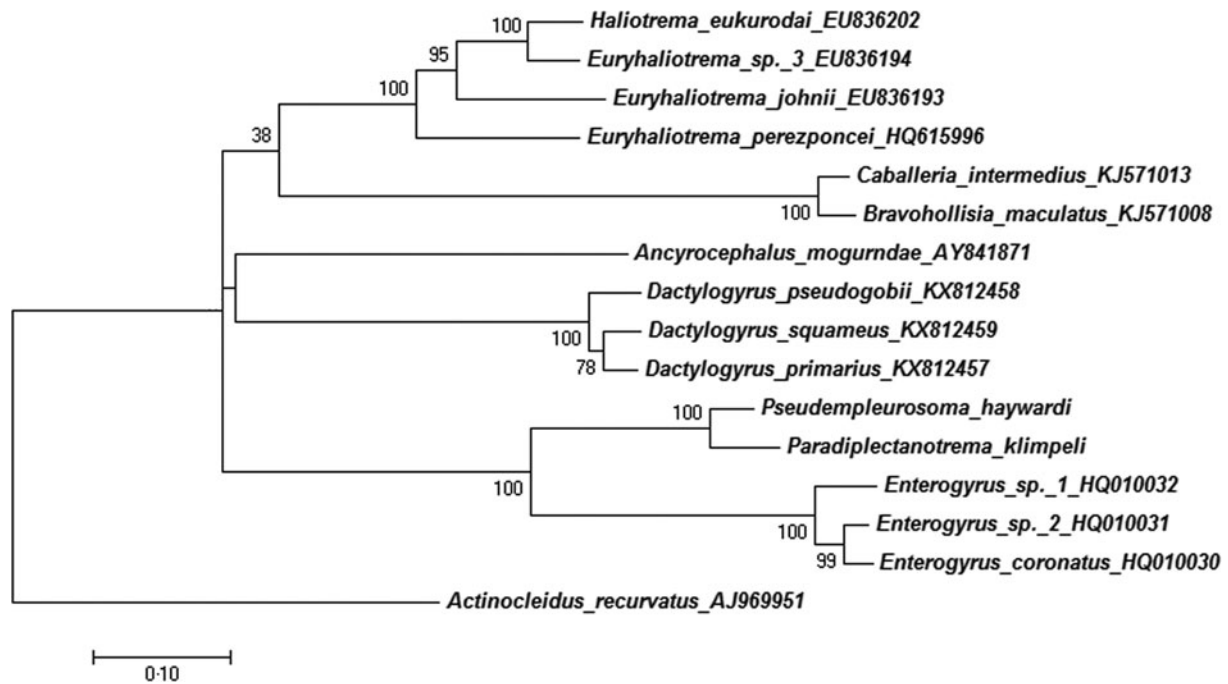


Fig. 4. Maximum-likelihood tree inferred from the analysis of LSU rDNA. The generated sequences were aligned with their closest matches in GenBank (15 ingroup and *Actinocleidus recurvatus* as outgroup) and formed a sister clade with *Pseudempleurosoma haywardi* (bootstrap value 100%), and with a branch of freshwater endoparasitic Monogenea from Cichlidae. Phylogenetic analysis based on the Tamura–Nei model with complete deletion used as gaps missing data treatment. The robustness was assessed using a bootstrap procedure with 1000 replications (Wu et al. 2005; Kumar et al. 2016).

Discussion

Morphological characterization

Gerasev et al. (1987) described several species of the *Diplectanotrema* group and named them after their respective teleost host, as seen in the description of the five *Paradiplectanotrema* spp. The morphological measurements of these species, all documented from the same region (African Atlantic coast), appear similar (Table 1). However, the descriptions were based on few specimens ($n=2$ *P. antigoni*, $n=7$ *P. chlorophthalmi* and $n=1$ *P. cytti*), making detailed comparisons rather difficult. In addition, Gerasev et al. (1987) provided a broader range of intraspecific measurements for the same character, possibly resulting from mounting (squeezing) techniques and/or omitting juveniles. Future DNA analyses of the West African species of *Paradiplectanotrema* are needed to confirm their validity and so far recorded host specificity (see Table 1). On the other hand, *P. klimpeli* sp. nov. can be clearly distinguished by the body and dorsal anchor length, the shape of the dorsal bar being gladiator breast-plate shaped (in upside-down view, see Fig. 2), and further parameters as summarized in Table 1. A number of misinterpretations of the morphology of the anchor apparatus of the *Diplectanotrema* group has been published over the years (compare Theisen et al. 2017). Confocal images can clearly illustrate these important structures for diagnoses, and we herewith provide the first confocal images for a *Paradiplectanotrema* species (Fig. 2). Confocal microscopy enabled us to position structures, providing angle of views for illustrating and comparison as well as accurate measurements, for example the saucer-type excrescences, the exact length of the curved MCO, the irregular shape of its accessory piece, the connection of the hooks and bars or the dorsal bar, showing its length, width and shape accurately (Fig. 3). This methodology is found to be suitable to detect minor, often difficult to describe differences in the haptor structures of these small monogeneans (also see Theisen et al. 2017).

The (sub-)generic (re-)descriptions of *Diplectanotrema* by Johnston and Tiegs (1922) and Price (1937) are based on the type species *Diplectanotrema balistes* (MacCallum, 1915) Johnston and

Tiegs, 1922. According to Gerasev (pers. comm. 2017), all (re-) descriptions seem to have misinterpreted the anchor apparatus (discussion on *Pseudempleurosoma* in Theisen et al. 2017), with *D. priacanthi* Lebedev, 1968 still awaiting re-examination in the future. The first adequate diagnosis for *Pseudempleurosoma* as well as *Paradiplectanotrema* were provided by Gerasev et al. (1987). However, confocal illustrations of specimens belonging to all so far described genera within the *Diplectanotrema* group, describing the sclerotized structures of the anchor apparatus, are needed to distinguish the so far five valid genera (including *Diplectanotrema*, see Theisen et al. (2017), especially their Table 1) and clarify the synonymies within this group of monogeneans.

Zoogeography and host range

The five so far described *Paradiplectanotrema* species have been reported exclusively from the Atlantic Ocean, off the coasts of West Africa [Guinea-Bissau, Western Sahara, Strait of Gibraltar (NW) and Angola (SW, also W Mediterranean)], along with their to date reported hosts (a caproid fish for *P. antigoni*, two chlorophthalmid species for *P. chlorophthalmi*, a zeid fish for *P. cytti*, a trichiurid fish for *P. lepidopi* and three carangid hosts for *P. trachuri*) (Table 1). This is the second description of an endoparasitic marine monogenean in South East Asia, the Indian Ocean respectively, suggesting a wider distribution also for other endoparasitic monogeneans of the *Diplectanotrema* group.

Here we establish a new host and family record for *Paradiplectanotrema*. It was already pointed out by Santos et al. (2001) and Theisen et al. (2017) that, although monogeneans are known for being host specific, site selecting and well adapted, the closely related *Pseudempleurosoma* (see below the ‘Phylogeny and Evolution’ section) has a variety of different host families and internal sites (from the beginning to the end of the gastrointestinal system), and was also randomly collected from the gills. Theisen et al. (2017) summarized 14 different host species from seven families reported for the seven *Pseudempleurosoma* spp. worldwide (circum-equatorial). It is possible that

Paradiplectanotrema might also have a lower host specificity (see Table 1 and Table 1 in Theisen *et al.* 2017) and a worldwide equatorial occurrence as well (Atlantic and Pacific, see also below the 'Phylogeny and Evolution' section). The oesophagus and pharynx are widely neglected host sites in the research of endoparasitic monogeneans, and the worms are hidden in the oesophageal folds (Fig. 2), covered by mucus. Consequently, further samples are needed to answer the real host and site specificity within this genus.

Concerning the host ecology, the so far known host species of *Paradiplectanotrema* spp. are, just like in *Pseudempleurosoma* spp., associated with muddy bottoms, infecting bottom-dwelling and sometimes schooling fish. However, while all hosts of *Pseudempleurosoma* spp. are associated with coastal waters and reefs, the host species of *Paradiplectanotrema* are deep-water fishes (see Table 2 here and Table 3 in Theisen *et al.* (2017)).

Infection pattern

We provide infection rates with SLs of the host fishes, also for the sciaenid *P. haywardi* hosts (Theisen *et al.* 2017, including unpublished data). Within the sample of grinders [14.5–34.2 SL (in cm), 32.6–447.2 TW (in g)], the infected four type host fish specimens were comparable in size. With four of 24 fishes harbouring 47 specimens, the prevalence was 17%, with a mean intensity of 12 and mean abundance of 2. According to own observations, the infections of Indonesian croakers with *P. haywardi* showed higher prevalences of 63% (22 of 35 fishes infected with 65 worms for *Nibea soldado*) and 46% (16 of 35 fishes infected with 32 worms for *Otolithes ruber*), but lower mean intensities of 3 (*N. soldado*) and 2 (*O. ruber*), intensities of 1–7 (*N. soldado*) and 1–6 (*O. ruber*), mean abundances of 2 (*N. soldado*) and 1 (*O. ruber*). Analysed soldier croakers had an SL of 20.0 (17.5–22.5) and a TW of 155.0 (99.5–211.4), the infected ones were of 17.5–21.8 SL and 99.5–203.3 TW, and the largest individual was negative. Investigated tiger-toothed croakers had an SL of 15.2 (13.7–17.1) and TW of 54.8 (41.6–78.1), the infected ones were of 13.7–16.7 SL and 41.6–71.0 TW, with the largest investigated individual being also negative (Theisen *et al.* 2017, including unpublished data). Only two *J. amblycephalus* (SL 22.6/24.6, TW 258.2/202.2) were screened for *P. haywardi* (see Theisen *et al.* 2017), and both were found to be infected (with three and five worms, respectively). Therefore, we can summarize that Indonesian fishes infected with endoparasitic monogeneans of the *Diplectanotrema* group are bottom-dwelling, are generally schooling, and of a middle size range, showing a TL/TG of 13.7–24.6/41.6–258.3 for croakers and 26.6–34.2/239.5–447.2 for grinders. It is interesting to note that the eggs in this monogenean group are few (maximum four eggs per specimen) and might infect the host of origin. However, bottom-dwelling schooling fish often ingest and disgorge bottom material such as mud and sand while searching for food, which might be a possible way for transmitting the eggs and parasite infection to another host.

Theisen *et al.* (2017) sampled *P. haywardi* in May and July 2011 (dry season) off the South coast of Central Java, and again during sampling of *P. klimpeli* sp. nov. in November 2016 (rainy season) off South Bali. The re-covering of the species within only two examined potential host organisms, almost 1000 km remote off the type locality, and 5 years later, during the rainy season, demonstrate the real abundance of *P. haywardi* in coastal Indonesian sciaenids. We conclude that, firstly, Indonesia is a marine reef fish biodiversity hotspot, secondly, *Pseudempleurosoma* spp. show low host specificity, and thirdly, fish parasitology research is still limited in Indonesian waters and the infection site, the oesophagus and pharynx, has been

neglected in Monogenea research surveys. Furthermore, the tiny transparent worms are regularly covered by mucus in the pharyngeal folds of their host and are difficult to detect (compare Fig. 2A: magnified, transilluminated and without mucus). Taking deep-sea fishes into account, which also show a high biodiversity in Indonesia, we suggest that many more host records and probably species of these endoparasitizing genera might be present in the region.

Phylogeny and evolution

Theisen *et al.* (2017) provided the first DNA sequences for a marine endoparasitic monogenean, namely *P. haywardi*, a member of the *Diplectanotrema* group, and ensured its position within the Dactylogyridae. Thus far, it was assumed that evolution towards an endoparasitic lifestyle must have happened multiple times within the Monogenea, for freshwater and marine species separately. This might still be possible, because lacking DNA sequence data of *Montchadskyella*, marine endoparasitic Monogenea of armorheads (Histiopterinae) in Southern Australia, are not affiliated to the *Diplectanotrema* group, and might have been evolved separately. However, Theisen *et al.* (2017) demonstrated the close relationship of *Pseudempleurosoma* and *Enterogyryus*, the latter is a freshwater endoparasitic monogenean from African Cichlidae fishes.

With the present study, we provide the first DNA sequences for the genus *Paradiplectanotrema*. The 28S region is found to be suitable for differentiation of diplectanotrem species and genera, however, the detected differences between the two diplectanotrem species of 2% in query cover and 8% in identity (according to NCBI GenBank Nucleotide Blast) can be primarily attributed to the two analysed species belonging to different genera, with similar generic differences occurring for example between respective species in the genus *Dactylogyryus* (or *Enterogyryus*, see Fig. 4). The relationship of two different genera within the *Diplectanotrema* group, was only based on morphology and ecology, but could be also now confirmed genetically (Fig. 4). The necessity of analysing further diplectanotrem species and genera will (i) help to understand the distribution patterns and evolution of a monogenean endoparasitic lifestyle through the world's oceans, and (ii) help to confirm validity and synonymy within the diplectanotrem taxa of less described species.

Conclusion

A new species of endoparasitic monogeneans in the marine lizardfish *S. tumbil* from Indonesian waters was described as *P. klimpeli* sp. nov., based on unique morphological characteristics, zoogeography, host distribution and DNA markers. Indonesian fish infections with endoparasitic monogeneans of the *Diplectanotrema* group can be abundant, their hosts known so far show medium sizes (13.7–34.2 cm SL, 41.6–447.2 g TW). All hosts of *Paradiplectanotrema* and *Pseudempleurosoma* are related to muddy bottoms, show schooling behaviour, and some are of commercial value. While the genus *Pseudempleurosoma* is associated with coastal reefs, *Paradiplectanotrema* occurs strictly in deep waters.

The phylogeny of marine endoparasitic *Paradiplectanotrema* was postulated for the first time, securing the position of these endoparasites and supporting an African freshwater origin. The *Diplectanotrema* group is ideally suited to analyse the spreading of a respective lifestyle and the worldwide colonization out of African waters into the world's oceans.

Table 2. *Paradiplectanotrema* spp. fish host species ecology and economic value

Fish host species	Host ecology ^a	Host economic value ^a	<i>Paradiplectanotrema</i> species ^b						
			1	2	3	4	5	6	
<i>Antigonia capros</i> (Caproidae)	Demersal, depth range 50– 900 m , usually 100–300 m, adults close to bottom , juveniles in mid-water	Minor commercial	x						
<i>Chlorophthalmus atlanticus</i> (Chlorophthalmidae)	Bathy demersal , depth range 240– 270 m (probably deeper) , inhabits deep-water of the continental shelf	N.e.		x					
<i>Chlorophthalmus agassizi</i> (Chlorophthalmidae)	Bathy demersal , depth range 50– 1000 m , 5–13 °C, on continental shelf and upper slope over mud and clay bottom , schooling habit indicated, juveniles pelagic near surface, adults demersal	Minor commercial		x					
<i>Cyttus traversi</i> ^c (Zeidae)	Bathy demersal , depth range 200– 1000 m , 17 °C, offshore waters, juveniles pelagic in surface waters	Commercial fisheries				x			
<i>Lepidopus caudatus</i> (Trichiuridae)	Benthopelagic , oceanodromous, depth range 40– 620 m , 24 °C, continental shelf, along edge and upper slope, over sandy and muddy bottoms , in midwater at night, schooling	Highly commercial fisheries and gamefish					x		
<i>Saurida tumbil</i> (Synodontidae)	reef-associated, amphidromous, depth range 20– 700 m , 27 °C, on muddy bottoms and trawling grounds	Commercial fisheries						x	
<i>Trachurus trachurus</i> (Carangidae)	Pelagic-neritic, oceanodromous, depth range 0– 1050 m , usually 100–200 m, 17 °C, adults schooling coastal over sandy substrate	Highly commercial fisheries and gamefish							x
<i>Trachurus mediterraneus</i> (Carangidae)	pelagic-oceanic, oceanodromous, depth range 0– 500 m , usually 5–250 m, 19 °C, adults near bottom , at times also in surface waters, migratory in large schools	Commercial fisheries and gamefish							x
<i>Trachurus picturatus</i> (Carangidae)	Benthopelagic , oceanodromous, depth range (of adults) 300– 370 m , 20 °C, neritic zones of island shelves, banks and sea mounts, schooling	Commercial fisheries							x

Note that all host species are – in contrast to hosts of the members of coastal, reef-associated *Pseudempleurosoma* – deep-water fishes, sometimes schooling, and – similar to *Pseudempleurosoma* – associated with muddy bottoms; some fish host species have commercial value, N.e.: not evaluated, bold font marks similarities.

^aData from Froese and Pauli (2017).

^b1 = *P. antigoni*, 2 = *P. chlorphthalmi*, 3 = *P. cytti*, 4 = *P. lepidopi*, 5 = *P. klimpeli* **sp. nov.**, 6 = *P. trachuri*.

^cGerasev et al. (1987) identified *Cyttus* sp. as host; however, out of the three valid *Cyttus* spp., only *C. traversi* is known from the African West coast (Atlantic) (Froese and Pauli, 2017).

Supplementary material

The supplementary material for this article can be found at <https://doi.org/10.1017/pao.2018.8>

Acknowledgements. We are thankful to Endang Wulandari Suryaningtyas, Pande Gde Sasmita Julyantoro and I Ketut Wija Negara from Udayana Universitas, for their help with issuing authorities, sampling, transport, laboratory work and providing work space. This is publication no. 11 under the Memorandum of Understanding between the Faculty of Veterinary Medicine, Udayana University, Bali, and the Faculty of Agricultural and Environmental Sciences, Aquaculture and Sea-Ranching, University of Rostock, Germany, in order to promote fish parasite and biodiversity research in Indonesia. Finally, we thank the colleagues from the University of Rostock for providing the chance for confocal microscopy (PD Dr Sergei A. Kuznetsov from the Live Cell Imaging Center (LMZ), Department of Biology), and our colleague from our institute for providing and translating Russian literature, namely Dr Ekaterina Pikalov.

Financial Support. Open access article processing charge was paid by the open access publication fund of Rostock University, funded by the Deutsche Forschungsgemeinschaft (German Research Foundation).

Conflict of Interest. None.

References

- Bush O, Lafferty AD, Lotz JM and Shostak AW** (1997) Parasitology meets ecology on his own terms: Margolis *et al.* revisited. *Journal of Parasitology* **83**, 575–583.
- Froese R and Pauly D** (2017) Editors. FishBase World Wide Web electronic publication (cited 10 December 2017). Available at <http://www.fishbase.org>.
- Galli P, Strona G, Villa AM, Benzoni F, Fabrizio S, Dogila SM and Kritsky DC** (2006) Three-Dimensional imaging of monogenean sclerites by laser scanning confocal fluorescence microscopy. *Journal of Parasitology* **92**, 395–399.
- Gerasev PI, Gaevskaya AV and Kovaleva AA** (1987) New genera of monogeneans of the group of diplectanotremes (Ancyrocephalinae). *Parazitologicheskii Sbornik* **34**, 192–210.
- Hall TA** (1999) Bioedit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acid Symposium Series* **41**, 95–98.
- Johnston TH, Tiegs OW** (1922) New gyrodactylid trematodes from Australian fishes, together with a reclassification of the superfamily Gyrodactyloidea. *Proceedings of the Linnean Society of New South Wales* **47**, 83–131.
- Kotob MH, Menanteau-Ledouble S, Kumar G, Abdelzaher M and El-Matbouli M** (2016) The impact of co-infections on fish: a review. *Veterinary Research* **47**(98). doi: 10.1186/s13567-016-0383-4. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5050641/pdf/13567_2016_Article_383.pdf
- Kovaleva A** (1970) Gelmintofauna stavrid roda Trachurus (Carangidae, Perciformes) bazejna atlanticeskogo okeana. *Biologia Morja* **20**, 37–66 (in Russian).
- Kumar S, Stecher G and Tamura K** (2016) MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution* **33**, 1870–1874.
- Marchiori NC, Pariselle A, Pereira J, Agnèse J, Durand J and Vanhove MPM** (2015) A comparative study of *Ligophorus uruguayense* and *L. saladensis* (Monogenea: Ancyrocephalidae) from *Mugil liza* (Teleostei: Mugilidae) in southern Brazil. *Folia Parasitologica* **62**, 024.
- Mehlhorn H** (2001) *Encyclopedic Reference of Parasitology: Diseases, Treatment, Therapy*, 2nd edn. Berlin/Heidelberg: Springer, GER. ISBN: 3-540-66239-1.
- Mendlová M., Pariselle A, Vyskočilová M and Šimková A** (2010) Molecular phylogeny of monogeneans parasitizing African freshwater Cichlidae inferred from LSU rDNA sequences. *Rarasitology Research* **107**, 1405–1413.
- Price EW** (1937) North American monogenetic trematodes. I. The superfamily Gyrodactyloidea. *Journal of the Washington Academy of Science* **27**, 146–164.
- Santos C, Mourão ED and Cárdenas MQ** (2001) *Pseudempleurosoma gibsoni* n. sp., a new ancyrocephalid monogenean from *Paralonchurus brasiliensis* (Sciaenidae) from off the southeastern coast of Brazil. *Memórias do Instituto Oswaldo Cruz* **96**, 215–219.
- Theisen S, Palm HW, Al-Jufaili SH and Kleinertz S** (2017) *Pseudempleurosoma haywardi* sp. nov. (Monogenea: Ancyrocephalidae (*sensu lato*) Bychowsky & Nagibina, 1968): an endoparasite of Croakers (Teleostei: Sciaenidae) from Indonesia. *PLoS ONE* **12**(9), e0184376.
- Wu XY, An XL, Xing QZ and Ming QX** (2005) Description of *Pseudorhabdosynochus seabassi* sp. n. (Monogenea: Diplectanidae) from *Lates calcarifer* and revision of the phylogenetic position of *Diplectanum grouperi* (Monogenea: Diplectanidae) based on rDNA sequence data. *Folia Parasitologica* **52**, 231–240.
- Zhang C, Li DL, Chi C, Ling F and Wang GX** (2015) *Dactylogyrus intermedius* parasitism enhances *Flavobacterium columnare* invasion and alters immune-related gene expression in *Carassius auratus*. *Diseases of Aquatic Organisms* **116**, 11–21.