

RESEARCH ARTICLE

Putative imbalanced amino acid metabolism in rainbow trout long term fed a plant-based diet as revealed by ¹H-NMR metabolomics

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Abstract

The long-term effect of a plant (P)-based diet was assessed by proton nuclear magnetic resonance (¹H-NMR) metabolomics in rainbow trout fed a marine fish meal (FM)–fish oil (FO) diet (M), a P-based diet and a control commercial-like diet (C) starting with the first feeding. Growth performances were not heavily altered by long-term feeding on the P-based diet. An ¹H-NMR metabolomic analysis of the feed revealed significantly different soluble chemical compound profiles between the diets. A set of soluble chemical compounds was found to be specific either to the P-based diet or to the M diet. Pterin, a biomarker of plant feedstuffs, was identified both in the P-based diet and in the plasma of fish fed the P-based diet. ¹H-NMR metabolomic analysis on fish plasma and liver and muscle tissues at 6 and 48 h post feeding revealed significantly different profiles between the P-based diet and the M diet, while the C diet showed intermediate results. A higher amino acid content was found in the plasma of fish fed the P-based diet compared with the M diet after 48 h, suggesting either a delayed delivery of the amino acids or a lower amino acid utilisation in the P-based diet. This was associated with an accumulation of essential amino acids and the depletion of glutamine in the muscle, together with an accumulation of choline in the liver. Combined with an anticipated absorption of methionine and lysine supplemented in free form, the present results suggest an imbalanced essential amino acid supply for protein metabolism in the muscle and for specific functions of the liver.

Keywords: Fish: Rainbow trout: Nutrition: Plant-based diet: Marine diet: Metabolomics: NMR

The development of fish farming to meet the requirements of the aquatic product market⁽¹⁾ is strongly impacted by the decreasing availability of marine ingredients⁽²⁾. The design of new fish feeds based on sources other than fisheries is, thus,

crucial to ensure that aquaculture production will increase while sustaining breeding performances, product quality, low impacts on the environment and the acceptability of such production⁽³⁾.

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Abbreviations: C, control commercial-like diet; FM, fishmeal; FO, fish oil; M, marine FM–FO diet; ¹H-NMR, proton nuclear magnetic resonance; P, 100 % plant-based diet; PCA, principal component analyses



The incorporation of significant quantities of ingredients of plant origin as a replacement for fishmeal (FM) and fish oil (FO) into the feed of high-trophic-level fish, such as salmonids and marine fish, has progressed rapidly. Albeit the growth performances are still not satisfying enough, salmonids can now be raised experimentally without the need for marine ingredients⁽⁴⁾ and using only plant-based ingredients; this is already being practised for low-trophic-level fish, such as cyprinids, cichlids and silurids⁽³⁾. Investigations into the long-term effects of plant (P)-based diets have already begun,^(5,6) and even if the growth of fish fed the P-based diet was markedly reduced, such investigations appear to be a promising way to assess the potential adaptation of fish nutrition and metabolism to these diets.

Nutritional limits for the total substitution of FM and FO in marine fish require that minimal quantities of ingredients of marine origin be maintained, essentially to cover the specific need of long-chain fatty acids (FAs) of the n-3 series^(7,8). This is not the case in salmonids, although limitations to the adaptation of the fish to plant-based ingredients are also observed at different levels, i.e. ingestion, digestion and metabolism. Replacing marine oil with a mixture of vegetable oils has been shown to have no deleterious effect on salmonids, whether on their growth performance or on product quality, except an alteration of their nutritional quality. The FA composition of lipids in the edible part of fish is generally adjusted to that of the diet, meaning a lower content of long-chain polyunsaturated FA, namely EPA and DHA, as well as a lower n-3/n-6 FA ratio⁽⁹⁾.

The replacement of marine protein sources by plant-based proteins can be easily managed⁽¹⁰⁾, although the supply of essential amino acids for protein metabolism must be fine-tuned^(11,12). Plant-based proteins are supplemented with crystalline amino acids to reach an amino acid composition close to that of FM. We know, however, that the amino acid stability, bioavailability and the uptake of supplemented crystalline amino acid are different from those of intact protein^(13,14). Furthermore, due to the limited potential for carbohydrate utilisation in fish⁽³⁾, proteins are also preferentially used to supply energy substrates for priority metabolic pathways in fish⁽¹⁵⁾ compared with non-carnivorous mammals and birds. Thus, the development of alternative fish feeds has to ensure the adequate supply of amino acids both for energy metabolism and for protein synthesis.

The metabolic pathways of essential nutrients have thoroughly been characterised in main farmed fish species in order to assess their food requirements⁽³⁾. However, metabolic pathways are often studied separately – energy *v.* nutrient metabolism or protein *v.* lipid metabolism – and an integrated analysis of these different metabolic pathways is rarely conducted⁽¹⁶⁾. Furthermore, there is a growing interest in a non-targeted metabolism analysis to decipher the main drivers of fish metabolism and the potential interconnection between pathways under non-limiting farming conditions⁽¹⁷⁾. Significant progress on the impact of P-based diets on nutrition and metabolism has been achieved by using various targeted and non-targeted omic approaches such as transcriptomics and proteomics^(4,7,8,18). Metabolomics is a complementary genomic approach used in human, livestock and bird nutrition^(19–21).

This approach has begun to be used in fish to characterise product quality^(22,23), and it is now widely used to assess the overall metabolic imprint of various environmental stresses such as hypoxia or environmental contaminants^(24,25). Metabolomics has recently begun to be used in fish nutrition to analyse the effects of feeding^(26,27), the impact of a wild *v.* a natural diet⁽²³⁾ and the effects of feed ingredients^(28–32), and more recently, to evaluate the development of new diets^(33–36). The two main metabolomic approaches are proton nuclear magnetic resonance (¹H-NMR) and mass spectrometry, each having their own advantages and technical limitations⁽³⁷⁾. NMR is not sensitive and selective with only a few dozens of metabolites detected and identified requiring a significant sample size. NMR is nevertheless preferentially used in fish nutrition due to minimal sample preparation, easy metabolite identification and good reproducibility⁽³⁷⁾. On the other hand, MS permits the detection and identification of thousands of metabolites, but the interpretation of profiles with numerous biochemical compounds is not easy. Thus, this approach is instead used to target specific compounds and to identify biomarkers of nutrition impairment⁽²⁷⁾. The simultaneous analysis of metabolites involved in various metabolic pathways has begun to identify some of the underlying mechanisms related to changes in metabolism following a dietary manipulation of the fish^(33,34). Thus, this powerful and integrative tool can helpfully be implemented to characterise the impact of P-based diets on metabolism. However, there needs to be a focus more on the fish metabolome that could account for the long-term adaptation of metabolism^(37,38) than on food metabolome.

Finally, limiting factors of plant ingredient quality remain in field conditions related to an imbalance in key nutrients, the stability of feedstuffs, numerous anti-nutritional factors⁽³⁹⁾ and specific contaminants, which could all induce digestion disorders⁽⁴⁰⁾ and impaired metabolism⁽¹⁸⁾. To overcome these difficulties will require significant progress in feed formulation and the further characterisation of feed. Analyses of ingredients and feeds are still based on proximal composition to assess their nutritive value, and, thus, the metabolomic approach, through a more detailed and comprehensive molecular analysis, could offer new perspectives for the development of alternative feeds.

The present study uses ¹H-NMR metabolomics to characterise differences in rainbow trout metabolism fed a P-based diet from the first feeding up to commercial size and that of those fed a marine FM–FO diet (M) and a commercial-like diet (C). Integrative ¹H-NMR metabolomics was implemented to further characterise the diet and to assess a metabolic response at the level of fish plasma, and liver and muscle tissues. The effect on lipid metabolism has already been characterised⁽⁴¹⁾, and this study focuses on the protein and energy metabolism through the characterisation of the metabolome in the polar extract of tissues.

Materials and methods

Source of fish

The fish used in this experiment came from a study supported by the FP7 programme of the European ARRANA project



(Advanced Research Initiatives for Nutrition and Aquaculture) (N_288925). The aim was to study the long-term effects of diets free of FM and FO from the first feeding onwards^(6,41). Rainbow trout was produced at the INRAE experimental fish hatchery of Lées Athas (Pyrénées-Atlantiques, France). At the stage of yolk-sac larvae, the rainbow trout fry was separated randomly into twelve groups and fed three experimental diets in quadruplicate for each diet. The fish were reared from 0.14 g body weight (BW) during 7 months in 50 l tank (310 fish per tank) in the same experimental farm with a natural photoperiod and a constant water temperature of $7 \pm 1^\circ\text{C}$. At an average body weight of 12.5 g, the fish were then transferred to the INRAE experimental fish farm in Donzacq, Landes, France. Fish within each dietary group were split randomly into three groups (nine groups in total) and fed with the three experimental diets. They were reared in 200 l tanks (150 fish per tank maximum stocking densities: 26 kg/m^3), with a natural photoperiod and a constant water temperature of $17 \pm 1^\circ\text{C}$, for another 8 months until final sampling.

Diet and experiment design

The experiment was carried out in strict accordance with the EU legal framework relating to the protection of animals used for scientific purposes (Directive 2010/63/EU) and according to the National Guidelines for Animal Care of the French Ministry of Research (Decree No. 2001-464, 29 May 2001). It was approved by the INRAE ethics committee (INRA 2002-36, 14 April 2002). The INRAE experimental facility is certified for animal services under Permit No. A40 2281 by French veterinary services – the competent authority – while the scientist in charge of the experimentation received training and a personal authorisation (No. B64 10 003).

Fish were fed from the first feeding up to 15 months with one of three diets: two experimental diets *i.e.* a marine (M) and a P-based diet formulated by UMR NuMeA (France) and manufactured at the INRAE experimental facility (Donzacq, Landes, France), and a control C diet from ‘the Cooperative Le Gouessant’ (Lamballe, Côte d’Armor, France). Each treatment was tested in triplicate.

The ingredients and compositions of the diets are presented in Table 1 and Supplementary Table S1 of Supplementary material. The M diet was composed mainly of FM (co-fishery products – all species) and FO (co-fishery products – sardines). The P-based diet was composed exclusively of plant ingredients, *i.e.* plant protein sources supplemented with lysine and methionine to cover the requirement of these essential amino acids, and a blend of vegetable oils (50 % rapeseed oil, 30 % linseed oil and 20 % palm oil) which aimed at providing an overall amount of FA classes similar to those of FO. The C diet contained a mixture of FM, FO and plant ingredients, with 45 % of FM and 50 % of FO replaced by plant ingredients. All diets were isoproteic, isolipidic and isoenergetic (Table 2).

The fish were fed *ad libitum* throughout the experiment continuously during the early stages, and then manually fed three, then two, meals per day. Diet characteristics were maintained during the experiment, except for pellet size, which was adjusted according to the stage of the fish. The experimental

Table 1. Crude diet composition (g/kg)

Ingredients (g/kg)	M	C	P
FM ^a	543	300	—
Maize gluten 48 %	—	102	180
Wheat gluten	—	50	121
Soyabean meal	—	63	43
Soyabean, Estrilvo	—	35	181
White lupin	—	65	50
Dehulled peas	—	69	24
Rapeseed 00 meal	—	63	98
Whole wheat	301	72	28
FO ^b	135	80	—
Rapeseed oil	—	80	73
Linseed oil	—	—	74
Palm oil	—	—	30
Soya lecithin	—	—	20
L-lysine	—	0.5	15
L-methionine	—	—	3
CaHPO ₄ ·2H ₂ O	—	—	26
Mix attractant ^c	—	—	15
Mineral premix ^d	10	10	10
Vitamin premix ^d	10	10	10

^a FM origin co-fishery products – all species.

^b FO origin co-fishery products – sardine.

^c Mix attractant: glucosamine 5 g/kg; taurine 3 g/kg; betaine 3 g/kg; glycine 2 g/kg; alanine 2 g/kg.

^d Mineral premix and vitamin premix compositions are given in Supplementary Table S1 of Supplementary material.

Table 2. Chemical composition of the diet

Ingredients	M	C	P
Dry matter (g/100 g FW)	96.3	96.0	96.3
Crude protein (g/100 g DM)	43.9	46.4	46.1
Crude fat (g/100 g DM)	21.8	24.2	25.0
Energy (kJ/g DM)	24.5	23.8	25.3

FW, fresh weight; DM, dry matter

diets were sampled to analyse their chemical composition during each pellet preparation. Once the pellets reached their final size (6 mm in diameter), the diets were also sampled at 0, 2 and 4 weeks in order to assess the compositional stability of the diet. Samples were stored at -20°C until extraction and analysis by $^1\text{H-NMR}$.

Fish sampling

At the end of the experiment, the fish were starved for 1 day and then fed one meal of the corresponding experimental diet. At 6 and 48 h after the meal, fifteen fish per treatment were randomly sampled (five fish per triplicate group). The fish were first sedated by immersion in a 2 % benzocaine solution, measured and weighed. Blood samples were taken from the caudal vein of each fish by means of a heparinised syringe and centrifuged in a microtube containing 20 μl of heparin solution (1 mg/ml) at 2400 g for 10 min, while the plasma was collected and frozen in liquid nitrogen. Then the fish were killed by sectioning the spinal cord. The fish were then dissected to weight the liver and the visceral mass. The liver was washed in cold saline solution and then dried on wrapped paper. A sample of the dorsal white muscle was taken after removal of the skin and the superficial red muscle. The liver



and the muscle tissues were directly frozen in liquid nitrogen. Plasma and tissues were then stored at -80°C until analysis.

Sample preparation

The diet samples were treated according to a protocol usually applied to plant samples⁽⁴²⁾. Four replicates of each sampling date, at 0, 2 and 4 weeks after the last formulation of the diets, were ground in liquid nitrogen and then freeze-dried. An ethanolic extraction was performed for each diet at each stage of sampling (in quadruplicate) by successive extraction with ethanol–water 80:20, 50:50 and 0:100 (% v/v). Fifty milligrams of the diet were mixed in 2 ml of solvent and then left for 15 min at 80°C before being centrifuged (30 000 g, 10 min, 4°C). The resulting supernatants were collected, combined and resubmitted to centrifugation (30 000 g, 10 min). The supernatant was then freeze-dried (Savant SC210A SpeedVac, Milford, CT, USA) and lyophilised. The extract was dissolved in 500 μl of deuterated phosphate buffer solution (200 mM, apparent pH 7.4), and the pH was adjusted to an apparent pH of 7.40 ± 0.02 when necessary with deuterated KOD deuterated analog of KOH 0.1 M or DCL deuterated analog of HCL 0.1 M solutions, by means of BTpH (Bruker BioSpin GmbH, Rheinstetten, Germany), before being lyophilised once more. For the $^1\text{H-NMR}$ analysis, the hydro-alcoholic extract was dissolved in 500 μl of deuterated water (D_2O) with 5 μl of sodium trimethylsilyl [2,2,3,3- d_4] propionate (TSP; 0.5 mM final concentration for chemical shift calibration), homogenised and centrifuged (14 000 g, 5 min; Centrifuge A14, Jouan, France), and the supernatant was transferred to a 5-mm NMR tube (507-PP-7, Wilmad, Buena Vista Township, NJ, USA) for direct analysis.

Each trout plasma sample was incubated on ice for 2 h to complete thawing. Immediately before analysis, 300 μl of plasma were mixed with 300 μl of D_2O and then transferred to a 5-mm NMR tube for direct $^1\text{H-NMR}$ analysis.

The liver (approximately 2 g, exactly weighed) and muscle frozen tissue samples (approximately 2 g, exactly weighed) were treated using a classical extraction by dichloromethane–methanol (2:1, v/v), containing 0.01 % of butylated hydroxytoluene as antioxidant, according to Folch *et al.*⁽⁴³⁾. The polar phase, containing water–methanol-soluble compounds, and the non-polar phase, containing lipid-soluble compounds, were separated and stored at -80°C before preparation for $^1\text{H-NMR}$ analysis. A calculated volume of polar extracts was sampled according to the initial weight of the liver or muscle samples used for extraction and then freeze-dried (Savant SC210A SpeedVac, Milford, CT, USA). The dried extract was dissolved in 140 μl of deuterated phosphate buffer solution (200 mM, apparent pH 7.4), 460 μl of D_2O and 5 μl of 5 mM TSP solution. After homogenisation and centrifugation (14 000 g, 5 min; Centrifuge A14, Jouan, France), the supernatant was transferred to a 5-mm tube for $^1\text{H-NMR}$ analysis.

Plasma metabolite assay kit analysis

Plasma glucose (Glucose RTU; BioMérieux, Marcy-l'Étoile, France), lactate (Lactate-PAP; BioMérieux, Marcy-l'Étoile,

France) and cholesterol (CHOL100; Sobioda, Montbonnot, France) concentrations were determined by enzymatic analysis using commercial kits adapted to a micro-plate format, according to manufacturer recommendations.

Growth and tissue parameter calculations

The condition factor was calculated using the BW (g)/body length (mm)³ formula. Somatic indices were also calculated: the hepatosomatic index (HSI) was calculated using liver weight (g)/BW (g) %, and the viscerosomatic index (VSI) was calculated using digestive tract weight (g)/BW (g) %.

$^1\text{H-NMR}$ spectra acquisition and processing

$^1\text{H-NMR}$ analyses were performed on a 500-MHz Avance III spectrometer (Bruker, Wissembourg, France) fitted with an ATMA BBI 5-mm Bruker probe at 300 K flushed with nitrogen and a BACS120 autosampler. Acquisition parameters were adapted from Kullgreen *et al.*⁽²⁶⁾ and Gatesoupe *et al.*⁽⁴⁴⁾ using a Carr-Purcell-Meiboom-Gill CPMG – relaxation editing pulse sequence with presaturation (cpmgrp) for plasma – and from Moing *et al.*⁽⁴²⁾ using a quantitative single pulse sequence (zg) for tissue extracts. The acquisition conditions of plasma and extracts (diet, liver and muscle) are summarised in Table 3. To optimise NMR acquisitions, automated tuning and matching, locking, shimming (TopShim) and 90° hard pulse calibration (pulsecal) were performed for each sample. A Fourier transformation was applied to the signal after zero-filling (64k) for extracts and apodization (LB 0.3 Hz) for extract and plasma spectra. The resulting spectra were manually phased with TopSpin (v. 3.0, Bruker).

For plasma, diet and tissue extracts, chemical shift calibration, baseline correction, peak realignment, non-uniform bucketing and signal-to-noise ratio were determined using the NMRProcFlow web tool (www.nmrprocflow.org)⁽⁴⁵⁾. Each spectral region of interest was determined manually, using either the intelligent bucketing module or the variable size

Table 3. $^1\text{H-NMR}$ analytical protocols

NMR parameters	Sample	
	DietLiverMuscle	Plasma
Temperature	27°C	27°C
Sequence ^a	Standard zg	CPMG cpmgrp1 ^b
Pulse angle	90	90
Number of dummy scans	0	8
Number of scans	64	64
Number of time-domain points	32 K	64 K
Relaxation delay	15 s	2 s
Acquisition time	2.73 s	5.46 s
Additional parameters		Spin-echo delay 500 μs Number of loops: 150
Experiment duration	19 min	9 min 10 s
Acquisition	Quantitative	Semi-quantitative

^a Bruker denomination.

^b Presaturation of water signal.



bucketing module of the NMRProcFlow for each dataset. For plasma spectra, this was performed within the 8.7–0.1 ppm region, and 212 spectral regions were determined for the initial analysis of time after-feeding effect, and then 398 spectral regions were determined for the analysis of the diet effect at each time after feeding, excluding the residual signals of water, lipids and lipoprotein resonances. The spectral regions were named according to the following example: B5_1811 for a spectral region centred on 5.1811 ppm. Total sum normalization was applied to minimise minor differences due to the total plasma metabolite concentration and/or variation in acquisition conditions among samples. The determination of spectral regions for tissue extract ¹H-NMR spectra was also performed as described above in diet (*n* 74), in liver tissue (*n* 252) and in muscle tissue (*n* 268) using the NMRProcFlow with the exclusion of the residual water area. The data were then standardised and normalised to the total intensity of the spectra.

Compounds were identified by comparing the spectra of individual compounds – analysed under the same conditions of preparation (solvent and/or pH) – to those available in the home database, the NMR HMDB (www.hmdb.ca), BMRB (<http://www.bmrwisc.edu/metabolomics/>) and Chemomx (Chemomx Metabolite Reference Libraries, www.chemomx.com) databases, and based on data from the literature^(24,26) and their annotation status according to MSI Metabolomic Standards Initiative⁽⁴⁶⁾. For a selection of metabolites, identification was confirmed by spiking commercially available reference compounds on representative samples and by the acquisition of 2D ¹H-NMR spectra on a selection of samples. Patterns of resonance that could not be identified were named according to the following example: unkDD6.15, meaning the unknown doublet of a doublet pattern of resonances centred on 6.15 ppm.

For the diet ¹H-NMR spectra, resonance groups were selected and integrated using the NMRProcFlow⁽⁴⁵⁾, then the absolute concentration of each compound was calculated using the electronic reference (TopSpin ERETIC2 module), and the calibration of signals with glucose (2.5–100 mM) and glutamate (2.5–30 mM) standard curves. The concentration in mol/l was then converted to g/l and transformed into a content by dividing it by the amount of lyophilised powder used for extraction. The content of each organic or amino acid was expressed as g of the acid form per weight unit. The content of NMR unknown compounds was calculated, hypothesising that the observed resonance corresponded to one proton and using an arbitrary molecular weight of 100 Da.

Data reduction and statistical analyses of spectra

The size of fish sampling (*n* 15) was selected to permit a potential 20 % dropout for fish outliers (maturing male) or for abnormal spectral distribution (liver and muscle). The resulting sample size (*n* 11 per treatment) is sufficient to detect a minimum 20 % difference between diets with a 95 % confidence for NMR variables. Statistical analyses were performed using BioStatFlow (v. 2.8, www.biostatflow.org). Multivariate principal component analyses (PCAs) were performed on

normalised data to visualise the discrimination between treatments and diets and to identify the compounds or spectral regions related to the discrimination. Supervised analysis such as ANOVA were performed on raw data to assess the significance of the differences observed between diets and, when relevant, MANOVA to assess the combined effect of other factors (a duration of storage for diet and time after feeding for plasma), as well as their interactions. A statistical threshold level of 5 % was retained. Data are presented as mean values with their standard deviation.

Results

Fish characteristics

The BW and the body length of fish fed a P-based diet were significantly lower than those of fish fed a M diet and a C diet (Table 4), but the condition factor was not significantly different between the diets (data not shown). The relative weight of the liver (HSI) was significantly higher in fish fed the M diet than in those fed the P-based diet and the C diet. The gut relative weight (VSI) was significantly higher in fish fed the C diet than in those fed the two other diets.

Contrasted ¹H-NMR quantitative profiling of diets

¹H-NMR spectra annotations of water–ethanol diet extracts made it possible to identify thirty-six soluble compounds (Fig. 1; Table 5) comprising amino acids and derivatives, sugars, organic acids, nucleosides, alkaloids and vitamin precursors. Thirty-one patterns of resonances remain unknown (Supplementary Table S2 of Supplementary material).

The ¹H-NMR profiles differed based on the diets. Some signals related to alanine, lysine, methionine, betaine, glycine, sucrose and stachyose were more specific to the P-based diet. In the 6–9 ppm range, signals corresponding to anserine, adenosine, xanthine and hypoxanthine were more specific to the M diet and the C diet. The signals corresponding to pterin and biopterin were very specific to the P-based diet.

The PCA, performed on the main identified and unknown compound contents determined by ¹H-NMR, demonstrated a

Table 4. Characteristics of rainbow trout fed three different diets from the first feeding: marine M, plant-based P and commercial C diets (mean (SD) *n* 30)

	Marine (M)	Commercial (C)	Plant-based (P)	ANOVA on diet effect
Total BW (g)	350 (37) ^a	359 (36) ^a	263 (52) ^b	**
Total body length BL (mm)	287 (11) ^a	283 (12) ^a	259 (17) ^b	**
HSILiver weight/100 g BW	1.22 (0.16) ^a	0.94 (0.11) ^b	0.89 (0.18) ^b	***
VSI Gut weight/100 g BW	8.23 (1.37) ^b	9.00 (1.63) ^a	8.14 (2.16) ^b	*

This is the whole set of experimental fish sampled at 6 and 48 h after the meal. ANOVA on diet effect: ****P* < 0.001; ***P* < 0.01; **P* < 0.05. Data with different letters are significantly different (Student's *t* test).

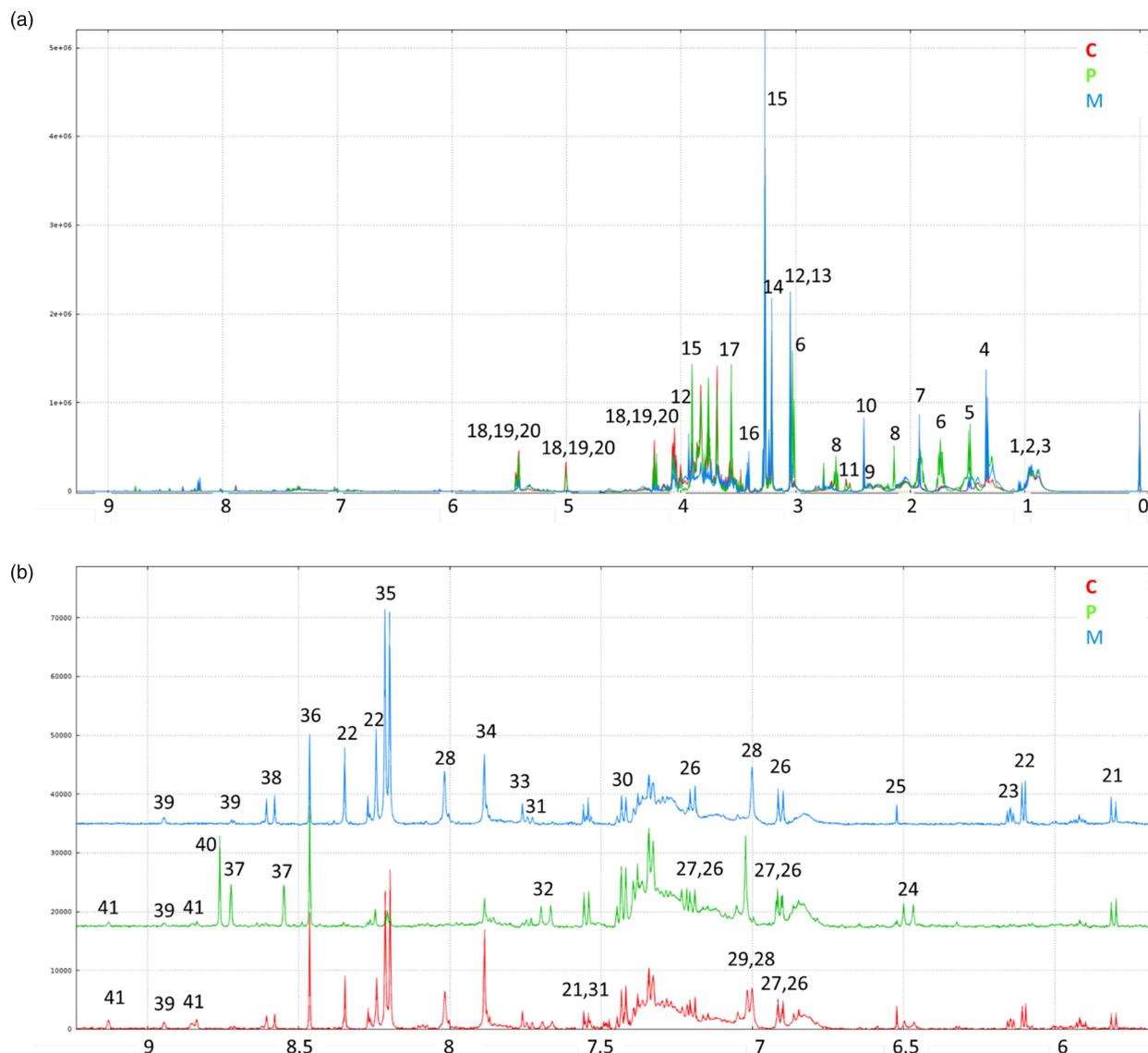


Fig. 1. (a) Representative 500 MHz $1D$ 1H -NMR spectra of ethanolic extracts of C, P-based and M diets and (b) representative expanded spectra of the (δ 9.2–5.7 ppm) range. Legend: 1 LEU; 2 ILEU; 3 VAL; 4: lactate; 5 ALA; 6 LYS; 7 acetate; 8 MET; 9 GLU; 10 succinate; 11 citrate; 12 creatine; 13 unkS3.04; 14 choline; 15 betaine; 16: taurine; 17 GLY; 18: sucrose; 19: stachyose; 20: raffinose; 21: uracil; 22 adenosine; 23 unkDD6.15; 24 unkD6.48; 25 fumarate; 26: TYR; 27 tyramine; 28 anserine; 29 HIS; 30 PHE; 31 TRP; 32 unk D7.68; 33 unkS7.76; 34 xanthine; 35 hypoxanthine; 36 formate; 37 pterin; 38 unkD8.59; 39 niacinamide; 40 bioplerin; 41 trigonelline (see Table 5).

clear discrimination between the three diets on the first two PCs (89 % of the total variability; Fig. 2). The main PC1 differences (69 % of total variability) were related to the M and P-based diets, C showing intermediate results, and secondary differences were observed on PC2 (20 % of the total variability) between the P and M diets, on the one hand, and the C diet, on the other hand. Differences due to diet storage for 2 weeks or 1 month were relatively low and non-significant, although such differences related to the PC2 axis seemed to be higher in the C and the P-based diets than in the M diet.

The main identified compounds (Table 6) as well as unknown compounds (Supplementary Table S2 of Supplementary material) were quantified with putative 100 g/mol for unknown compounds. Sugars –stachyose and sucrose – were present at a very high content (>9.8 g/kg of dry matter (DM) and >4.6 g/kg DM, respectively, in the P

and C diets compared with the M diet approximately 1 g/kg DM), as well as lysine (>10 g/kg DM in the P-based diet). Other compounds were also present at a high content (>1 g/kg DM): choline, taurine and lactate in all three diets; anserine, creatine, glutamate, alanine and hypoxanthine in the M diet; and alanine, glycine, lysine, methionine and betaine in the P-based diet. Some compounds, such as raffinose, were detected only in the M diet at a relatively high content of >1 g/kg DM. On the contrary, two other compounds – pterin and bioplerin – were detected only in the P-based diet at a low content.

The differences between the M and the P-based diets were highly significant ($P < 0.001$) for most of the compound content, except for tryptophan (Table 6). The content of creatine, anserine, adenosine, xanthine, hypoxanthine, niacinamide and various unknown compounds (unkS2.74, unkS3.03, unkS3.23,

**Table 5.** Cumulative list of annotated compounds in the ¹H-NMR spectra of hydro-alcoholic extracts of the three diet pellets

Compound	Resonances used for quantification (chemical shift in ppm) and multiplicity	Other chemical shifts (ppm) and multiplicity	MSI status
Amino acids and derivatives			
Tryptophan	7.73 (d)	7.54 (d); 7.28 (t); 7.2 (t)	1
Phenylalanine	7.43 (m)	7.39 (m); 7.35 (m); 3.14 (dd)	1
Tyrosine	7.2 (d); 6.91 (1/2d)	6.91 (d)	1
Histidine	7.02 (d)	6.99 (d); 8.02 (d) with anserine	2
Lysine	1.74 (m)	7.92 (m); 3.76 (t); 3.03 (t); 1.49 (m)	1
Glycine	3.56 (s)		1
Glutamic acid	2.35 (4/6 m)	2.35 (m); 2.14 (m); 2.07 (m)	1
Methionine	2.64 (1/3 t)	2.64 (t); 2.20 (m); 2.14 (m); 2.14 (s)	1
Alanine	1.49 (d)	3.79 (q)	1
Valine	1.05 (d)	3.62 (d); 2.28 (m); 0.99 (d)	1
Isoleucine	1.01 (1/2d)	1.01 (d); 0.94 (t)	1
Leucine	0.97 (3/4t)	1.72 (m); 0.97 (t)	1
Anserine	4.48 (dd)	8.02 (s); 7.00(s); 3.74 (s); 3.2 (m);	1
Taurine	3.43 (t)	3.27 (t)	1
Creatine	3.93 (s); 3.04 (s)		1
Betaine (trimethylglycine)	3.90 (s)	3.27 (s)	1
Carbohydrates			
Glucose	5.24 (1/2d); 4.65 (d 1/2)	5.24 (d); 4.65 (d)	1
Stachyose	5.00 (m)	5.44 (d); 4.24 (d); 4.15 (dd); 3.68 (s)	1
Sucrose	5.42 (d)	4.22 (d); 4.06 (t); 3.82 (m); 3.77 (t); 3.68 (s); 3.57 (dd); 3.48 (t)	1
Raffinose	5.00 (m)	5.44 (d); 4.24 (d); 4.15 (dd); 3.68 (d)	1
Organic acids			
Formate	8.47 (s)		1
Fumarate	6.52 (s)		1
Malate	4.3 (1/2 dd)	4.3 (dd); 2.67 (dd); 2.37 (dd)	1
Lactate	4.11 (q)	1.33 (d)	1
Citrate		2.61 (dd)	2
Succinate	2.41 (s)		1
Acetate	1.92 (s)		1
Nucleoside/nucleobases			
Hypoxanthine	8.20 (s)	8.21 (s)	1
Xanthine	7.89 (s)		1
Uracil	5.8 (d)	7.54 (d)	2
Adenosine	6.10 (d)	8.35 (s); 8.24 (s)	2
Vitamin, vitamin precursor, alkaloid and other compounds			
Choline	3.21 (s)	4.07 (m); 3.53 (m)	1
Trigonelline	9.135 (s); 8.84 (m)3.21 (s)	9.135 (s); 8.84 (m); 8.09 (t); 4.44 (s)4.07 (m); 3.53 (m)	1
Pterin	8.72 (d); 8.54 (d)	9.135 (s); 8.84 (m); 8.54 (d); 8.09 (t); 4.44 (s)	2
Biopterin	8.76 (s)8.72 (d)	8.54 (d)	2
Niacinamide	8.94 (d)	8.71 (dd); 7.60 (dd)	1
Tyramine	7.22 (d); 6.91 (d); 8.94 (d)	8.71 (dd); 7.60 (dd); 2.93 (t)	2

s, singlet; d, doublet; dd, doublet of doublets; t, triplet; q, quadruplet; m, multiplet (not solved). MSI status from Sumner *et al.* (40): 1 refers to an identified compound; 2 refers to a putatively annotated compound.

unkS7.76, unkS7.87 and unkS8.59; Supplementary Table S2 of Supplementary material) was significantly higher in both the M diet and the C diet than in the P-based diet, whereas the content of histidine and various unknown compounds (unkS2.86, unkD5.17, unkD5.20, unkD6.48; unkD7.15 and unkD7.68) was significantly higher in both the P-based and the C diets than in the M diet. The C diet generally showed intermediate results between the M and the P-based diets, except for the content of stachyose, sucrose, choline, xanthine and several unknown compounds (unkS5.89, unkS5.91, unkS5.93, unkS5.92, unkD7.15 and unkS7.86), which was lower in the C diet than in the other two diets.

Lysine and methionine were added to the P-based diet to compensate for the imbalanced amino acid composition of plant-based proteins. The ratio between the contents of

compounds measured by ¹H-NMR in the diet and the content in the diet was calculated. The ratios were, respectively, of 0.73 and 0.68 for lysine and methionine. Other compounds were added to the P-based diet as a mix attractant. Only four of these compounds – taurine, betaine, glycine and alanine – were detected and quantified with a recovery ratio of 0.96, 0.72, 0.75 and 1.32, respectively.

Significant differences in a classical set of circulating metabolites in plasma

Glucose, lactate and cholesterol concentrations in plasma were not significantly different between 6 and 48 h after feeding. Glucose concentrations were significantly higher in the M diet than in the P-based diet and the C diet (Fig. 3) at 48 h

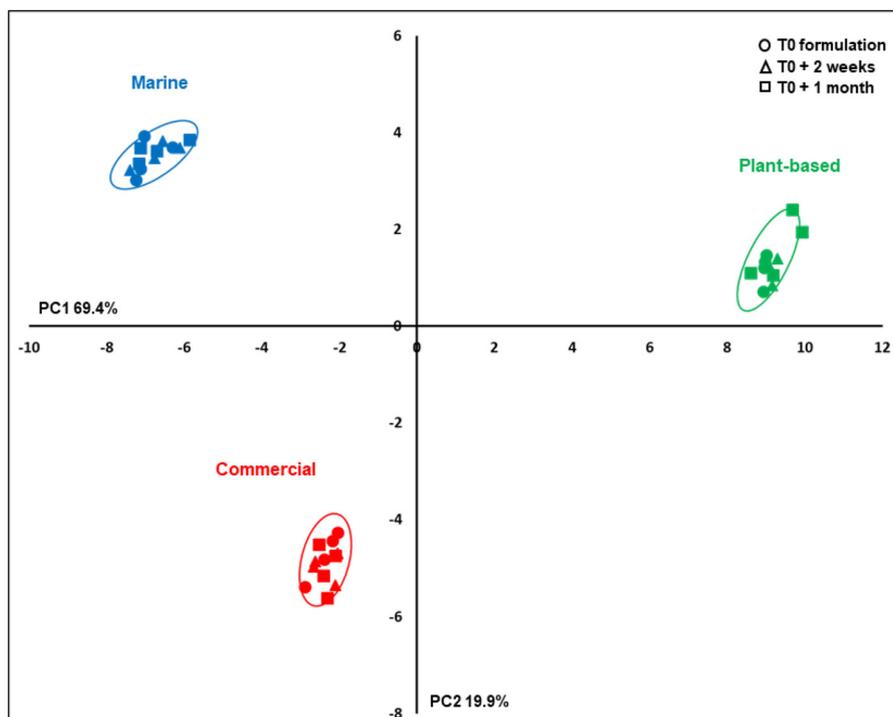


Fig. 2. PCA on compound contents determined by $^1\text{H-NMR}$ spectra of polar extracts of experimental diets. Scores of the three diets, M, C and P-based, sampled at 0 (circle), 2 weeks (triangle) and 1 month (square) after the last formulation.

after feeding, whereas the concentration of cholesterol in the M diet was significantly higher than that in the P-based diet at both 6 and 48 h after feeding. Lactate concentration was not significantly different between diets.

$^1\text{H-NMR}$ profiling of plasma relevant to both post-prandial changes and long-term diet effects

The $^1\text{H-NMR}$ plasma spectra using a 'water presaturation' sequence showed generally broad resonances of the circulating macromolecules (lipids and lipoproteins); thus, a 'cpmg' sequence was used to attenuate the signal of these macromolecules and to highlight small metabolite molecules. Twenty-seven circulating compounds were detected, including main amino acids, sugars, organic acids, vitamins, nucleotide and nucleosides, while residual signals of lipids and lipoproteins were still observed (Supplementary Fig. S1 and Table S3 of Supplementary material).

Each spectrum was segmented into 212 variable spectral regions on which putative specific compounds were assigned according to the annotation. The PCA demonstrated a significant discrimination of plasma profiles between 6 and 48 h after the last meal (in Fig. 4(a), PC1 represents 32 % of the total variability). The compounds related to post-prandial differences are the essential amino acids, such as methionine, phenylalanine and tyrosine, which were higher at 6 h post feeding, and glucose and non-essential amino acids, such as alanine, which were higher at 48 h post feeding (Fig. 4(b)). The status of branched-chain amino acids on this first PCA axis was, however, contrasted, with leucine showing higher values at 6 h post feeding and valine showing higher values at 48 h post feeding. The PCA did not show a significant

discrimination between the fish fed different diets on the first two axes (cumulated 44 % of the total variability), although the third axis (PC3 9 % of the total variability; Supplementary Fig. S2A of Supplementary material) suggested a discrimination between the fish fed the P-based diet and those fed the M diet, while the fish fed the C diet showed intermediate results. These differences were due to higher scores of creatine, dimethylamine and serine in the M diet and higher scores of glucose, lysine, leucine, inositol and pterin in the P-based diet (Supplementary Fig. S2B of Supplementary material).

Due to highly significant interactions between diet and time after feeding for most of the spectral regions (54 % of the whole set of spectral regions), a separate analysis was performed at 6 h post feeding and 48 h post feeding. At 6 h after feeding, the PCA (Fig. 5(a)) showed significant profile discriminations between the fish fed different experimental diets, particularly on the second principal axis (12.6 %). Volcano plot analyses (Fig. 5(c)) show a few compounds with a higher content in the fish fed the M diet, such as dimethylamine and creatine, while the content of numerous compounds was significantly higher in the fish fed the P-based diet, such as glucose, lysine, methionine, inositol, pterin and various unknown compounds (Tables 7; Supplementary Table S4 of Supplementary material). At 48 h after feeding, the PCA (Fig. 5(b)) did not show a significant discrimination between the fish fed different diets, although the fish fed the M diet and those fed the P-based diet seemed to be on opposite sides of the first principal axis (25.6 %). The volcano plot analysis (Fig. 5(d)) shows numerous compounds with content that was significantly higher in fish fed the P-based diet, such as inositol, alanine, lysine, valine, isoleucine, leucine, pterin and

**Table 6.** Compound contents ($\mu\text{g/g DM}$) in diet polar extracts

Compounds	Diet (contents in $\mu\text{g/g DM}$)			ANOVA on diet effect	
	M	C	P-based		
Sugars	Glucose	168 (34) ^c	255 (40) ^b	415 (60) ^a	***
	Stachyose	0 ^c	13 451 (374) ^a	9836 (189) ^b	***
	Sucrose	1105 (27) ^b	4724 (175) ^a	4683 (118) ^a	***
	Raffinose	1103 (142)	0	0	<i>det. only in M</i>
Amino acids and metabolites	Phenylalanine	187 (10) ^c	241 (10) ^b	390 (9) ^a	***
	Tyrosine	76 (1) ^a	57 (0) ^c	69 (14) ^b	***
	Glycine	358 (14) ^b	172 (11) ^c	1506 (87) ^a	***
	Lysine	566 (48) ^c	845 (62) ^b	10 921 (157) ^a	***
	Methionine	0	0	2029 (55) ^a	***
	Glutamate	1014 (41) ^a	719 (42) ^b	140 (20) ^c	***
	Alanine	1053 (35) ^b	728 (21) ^c	1984 (62) ^a	***
	Valine	341 (9) ^a	269 (6) ^b	193 (4) ^c	***
	Isoleucine	177 (8) ^a	141 (5) ^b	134 (4) ^c	***
	Leucine	244 (14) ^b	219 (14) ^c	261 (18) ^a	***
	Tryptophan	75 (11)	70 (12)	78 (7)	NS
	Histidine	0	315 (14) ^b	564 (18) ^a	***
	Taurine	3291 (42) ^a	1862 (34) ^c	2885 (70) ^b	***
	Anserine	1067 (58) ^a	739 (93) ^b	147 ((44) ^c	***
	Betaine	574 (25) ^b	294 (20) ^c	2170 (58) ^a	***
	Creatine	1793 (39) ^a	1134 (25) ^b	0	***
	Organic acids	Tyramine	19 (5) ^c	34 (3) ^b	59 (6) ^a
Formate		66 (7) ^c	88 (7) ^b	148 (15) ^a	***
Acetate		827 (69) ^a	501 (180) ^b	317 (40) ^c	***
Lactate		1656 (33)	1644 (178)	1516 (141)	*
Fumarate		40 (2) ^b	47 (3) ^a	19 (3) ^c	***
Malate		156 (11) ^c	377 (28) ^b	523 (12) ^a	***
Vitamins, precursors and metabolites	Succinate	517 (17) ^a	360 (10) ^b	153 (3) ^c	***
	Pterin	0	0	262 (14) ^a	***
	Bioperin	0	0	501 (54) ^a	***
	Choline	1054 (39) ^b	1179 (51) ^a	1012 (37) ^b	***
	Trigonelline	0	65 (5) ^a	32 (6) ^b	***
Nucleosides nucleobases	Niacinamide	47 (3) ^a	44 (4) ^a	28 (4) ^b	***
	Xanthine	428 (40) ^b	535 (51) ^a	164 (10) ^c	***
	Hypoxanthine	983 (23) ^a	667 (12) ^b	53 (6) ^c	***
	Uracile	114 (6) ^b	84 (5) ^c	164 (10) ^a	***
	Adenosine	596 (24) ^a	371 (19) ^b	0	***

The number 0 is not detectable (mean (sd); n 12).

ANOVA significance of diet effect: *** P < 0.001; ** P < 0.01; * P < 0.05.

various unknown compounds, while only a few compounds had a significantly higher content in fish fed the M diet, such as glutamine and glucose (Tables 7; Supplementary Table S4 of Supplementary material). The significant interactions between post-feeding time and diet were related mainly to glucose, inositol, glutamine, alanine, methionine, valine, lysine, phenylalanine, tyrosine and various unknown compounds.

¹H-NMR liver tissue analysis

The ¹H-NMR spectra of aqueous polar extracts in liver made it possible to identify eighteen water–methanol-soluble compounds, including main amino acids (phenylalanine, tryptophan, histidine, lysine, isoleucine, leucine, valine, glutamic acid, glutamine, alanine and taurine), carbohydrates (glucose and mannose), organic acids (lactate and acetate), peptides and amino acid derivatives (anserine and phosphocreatine), vitamin precursors and derivatives (niacinamide, choline and betaine) and nucleoside (inosine) (Supplementary Fig. S3 and Table S5 of Supplementary material). Some compounds were difficult to identify because their NMR signals

overlapped each other, and this is the case for the arginine signal that overlapped the choline signal.

The whole spectrum was segmented in 252 variable spectral regions on which putative specific compounds were assigned according to the annotation. The PCA showed a clear discrimination between the fish fed different experimental diets (in Fig. 6(a), PC1 represents 26 % of the variability). The volcano plot analysis (Fig. 6(b)) shows that numerous spectral regions contributed to the differences observed between the fish fed the M diet and those fed the P-based diet.

The relative quantification of the more representative spectral regions (Table 8; Supplementary Table S6 of Supplementary material) revealed at least twenty-four signals significantly higher in the M diet and forty-three signals significantly higher in the P-based diet. The compounds more specifically related to the M diet were glutamine, mannose and uridine derivatives, while some signals were related to a mixture of compounds and numerous signals were not associated with any compounds. Some of the latter were observed in a relatively high content in liver tissue (unkS3.91, unkD3.79 unkS3.43 and unkD1.32). The compounds specifically related to the

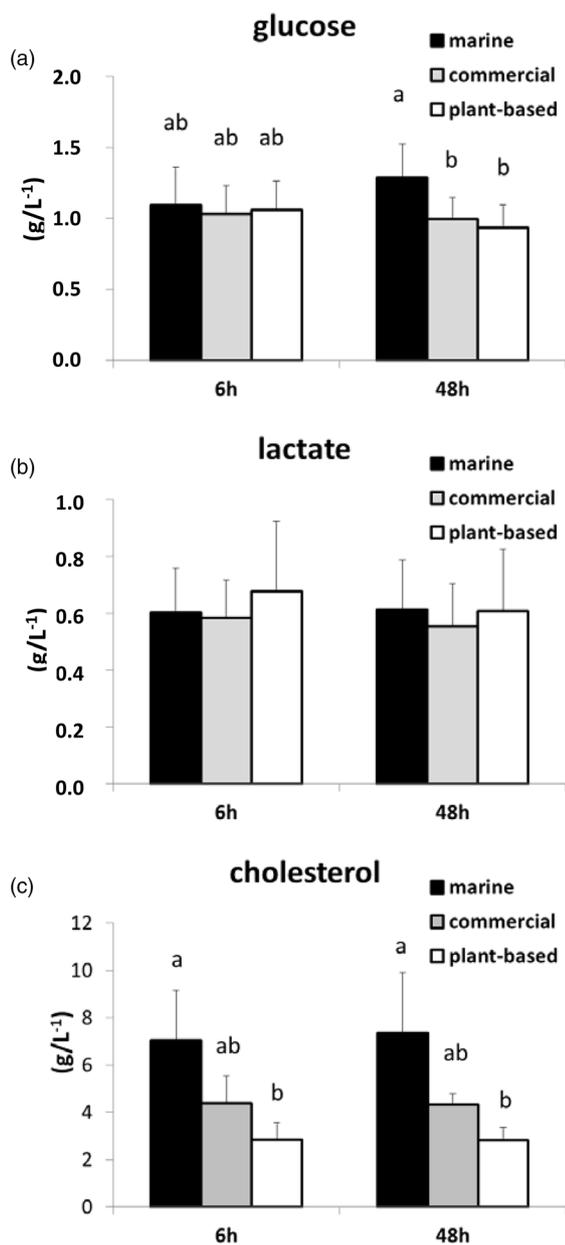


Fig. 3. Concentration (g/L) determined by assay kits of selected metabolites (a) glucose, (b) lactate and (c) cholesterol in plasma at 6 and 48 h after feeding in rainbow trout fed either a M diet, a P-based diet or a C diet. Mean and standard deviation: $n = 15$. Data affected by different letters were significantly different (two factors: MANOVA analysis and Student's t test; $P < 0.05$ for pairwise comparison), knowing that the time effect and the interaction with diet effect were not significant.

P-based diet were glucose, threonine, isoleucine, tryptophan, glutamate, uracil, inosine, putrescine, formate and ethanol, and there were numerous signals for which no associated compounds were identified. Some of these unknown compounds were shown to be in a relatively high content in liver tissue such as unkS3.42 and unkS3.27.

¹H-NMR muscle tissue analysis

The ¹H-NMR spectra of polar aqueous extracts of muscle tissue made it possible to detect sixteen water-methanol-soluble compounds identical to those detected in the liver tissue

(mannose and ethanol were not detected in muscle tissue) (Supplementary Fig. S4 and Table S4 of Supplementary material).

The whole spectra were segmented in 268 variable spectral regions on which putative specific compounds were assigned according to the annotation. The PCA showed a clear discrimination between diets (in Fig. 7(a), PC1 represents 32 % of the variability). Numerous spectral regions contributed to differences between the fish fed the M diet and those fed the P-based diet, as indicated on the volcano plot figure (Fig. 7(b)).

The quantification of the more representative spectral regions (Table 9; Supplementary Table S6 of Supplementary material) revealed at least forty-two signals significantly higher in fish fed the M diet and seventy-eight signals significantly higher in fish fed the P-based diet. Signals more specifically related to fish fed the M diet were assigned to the following compounds: niacinamide, choline and uridine derivatives. One strong signal was associated with a mixture of compounds (choline, anserine, taurine, betaine and histidine) and was difficult to solve, although it could be associated mainly with taurine. Numerous signals had no associated compounds, and some of these were detected only in the M diet (unkD9.14, unkD8.86 and unkD5.45) at low levels. Signals specifically related to fish fed the P-based diet were attributed to the following compounds: histidine, threonine, lysine, leucine, valine, isoleucine, glutamate, taurine, creatine, lactate and inosine, and there were also numerous signals for which no associated compounds were identified. A few of these unidentified compounds were in a relatively high content in muscle tissue (unkM3.79).

Interconnections between metabolites in plasma, and liver and muscle tissues

The global PCA on ¹H-NMR profiles of plasma, and liver and muscle tissues at 48 h post feeding showed a clear discrimination of the fish fed different diets, particularly between those fed the M diet and those fed the P-based diet (Fig. 8). The differences in signals related to amino acid accumulations in the plasma of fish fed the P-based diet at 48 h post feeding were correlated with signals associated with amino acid accumulations in muscle tissue. The differences in signals related to plasma glucose in the fish fed the M diet were correlated with signals associated with the presence of glucose in the liver and of niacinamide in muscle tissue.

Discussion

This study was performed on rainbow trout whose growth performance, after having been fed a full P-based diet for 15 months, was relatively good, although it was still significantly lower than that of the control fish fed the FM/FO diet (M) and the reference C diet. The growth reduction due to the P-based diet was observed since the first feeding stage but was still effective in the on-growing stage of the present study⁽⁴¹⁾.

The plant ingredients were supplemented with specific compounds, such as lysine and methionine, to re-equilibrate

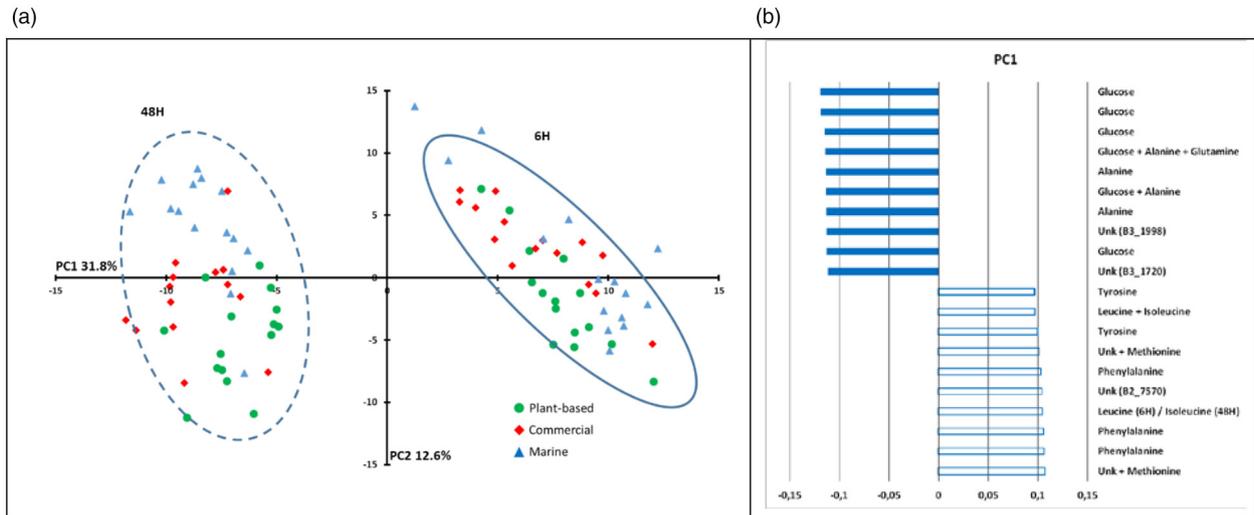


Fig. 4. Multidimensional PCA of ^1H -NMR spectra ('CPMG' sequence) of plasma segmented into 212 variable size spectral regions: (a) representation on the first two PC axes of the scores of plasma of individuals fish fed a M diet (triangle), a P-based diet (circle) and a C diet (diamond) either 6 h post feeding (straight line) or 48 h post feeding (dotted line) n 15 per treatment and (b) representation of the main significant spectral regions involved in the PC1-axis differences related to time after feeding.

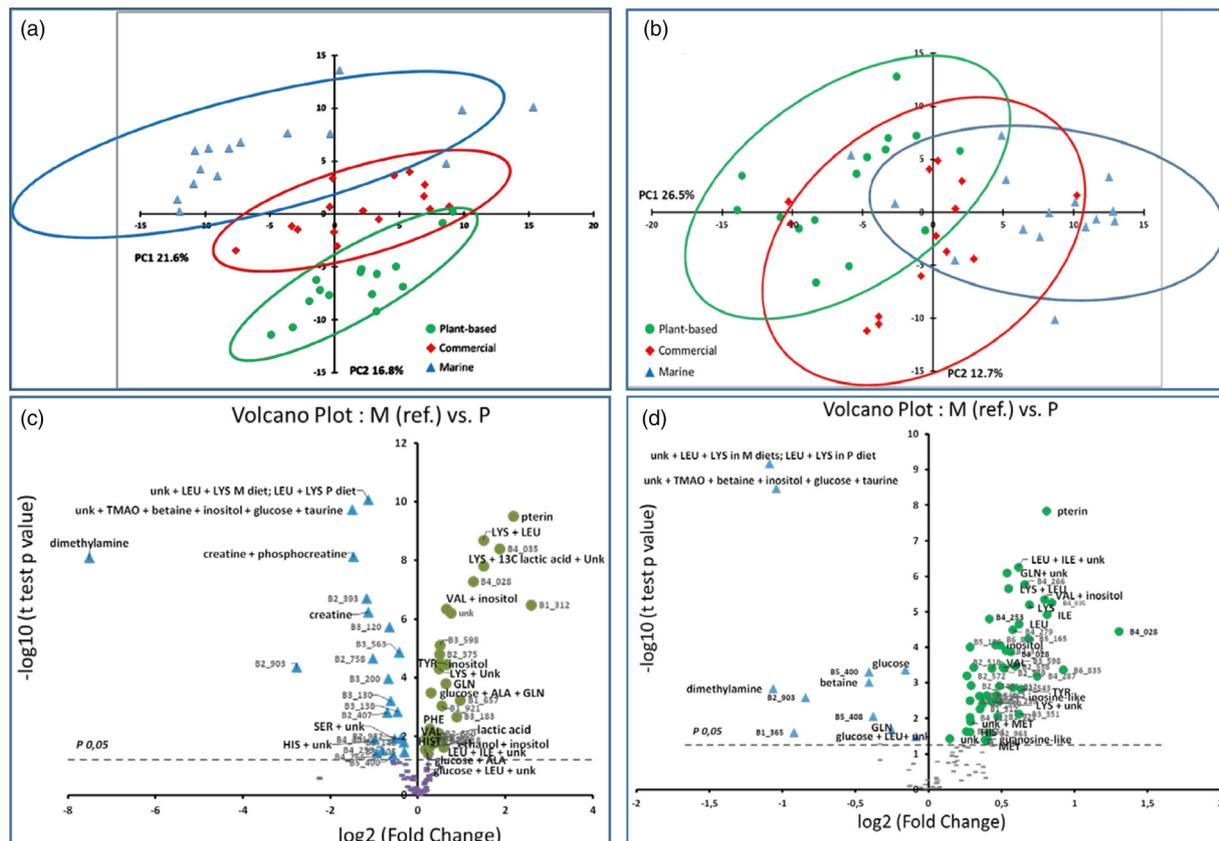


Fig. 5. Multidimensional PCA of ^1H -NMR spectra (CPMG sequence) of plasma segmented in 388 spectral regions. (a) Score plot on the first two principal components of individual rainbow trout at 6 h after feeding on a M diet (triangle), a P-based diet (circle) and a C diet (diamond) n 15 per treatment. (b) Score plot on the first two principal components of individual rainbow trout at 48 h after feeding on a M diet (triangle), a P-based diet (circle) and a C diet (diamond) n 15 per treatment. (c) Volcano plot representations of the differences at 6 h between M and P-based diets with the M diet used as a reference. Circle, significantly higher signal ($P < 0.05$); triangle, significantly lower signal ($P < 0.05$); dash, non-significant differences. Each bucket was assigned to a given metabolite compound or a group of metabolite compounds, except for unidentified signals: unks2.18 (singlet at 2.18), unkt3.99 (triplet at 3.99 ppm), unkd1.008 (doublet at 1.008 ppm) and unkM (multiplet of resonances).



Table 7. Significant effects of diet (M v. P-based) on semi-quantitative assessment of representative annotated spectral regions or bucket of the ¹H-NMR spectra of plasma at 6 and 48 h after feeding

Bucket	Metabolite resonance annotation	Significance diet
6 h M > P-Based diet		
B1_753	unk + leucine + lysine ^a	***
B2_732	Dimethylamine	***
B3_044	Creatine + phosphocreatine	***
B3_270	unk + TMAO + betaine + inositol + Glucose	***
B3_936	Creatine	***
B4_068	Inositol	***
6 h P-based diet > M		
B1_447	Lysine + unk	***
B1_459	Lysine + ¹³ C lactic acid+ unk	***
B1_681; B1_694; B1_702; B1_694	Lysine + leucine	***
B2_436; B2_444; B2_421; B2_460; B2_468; B2_479	Glutamine	***
B3_607	Valine + inositol	***
B3_628; B4_060; B4_068	Inositol	***
B3_763	Glucose + alanine + glutamine	***
B6_906; B6_888, B7_207	Tyrosine	***
B8_549; B8_721	Pterin	***
48 h M > P-Based diet		
B1_753	unk + leucine + lysine ^a	***
B2_436; B2_444	Glutamine	***
B3_270	unk + TMAO + betaine + inositol + glucose	***
B3_483; B3_488; B5_216; B5_248	Glucose	***
48 h P-based diet > M		
B0_966	Leucine + isoleucine + unk (M diet)	***
B0_966; B0_976	Leucine	***
B0_987; B0_999; B1_037; B1_052	Valine	***
B1_009; B1_021	Isoleucine	***
B1_447	Lysine + unk	***
B1_681; B1_694; B1_702; B1_714	Lysine + leucine	***
B1_914	Lysine	***
B3_628; B4_058; B4_066; B4_076	Inositol	***
B3_607	Valine + inositol	***
B3_676	Ethanol	***
B3_796; B3_808	Alanine	***
B5_186	unk sugar mannose like	***
B8_549; B8_721	Pterin	***

unk, resonance(s) from unknown metabolite(s) (an un-annotated bucket with significant effects of diet is listed in Supplementary Table S4 of Supplementary material). ANOVA on diet effect (*n* 15): ****P* < 0.001; ***P* < 0.01; **P* < 0.05. a unk + leucine + lysine diet C and M; leucine + lysine diet P.

essential amino acid profile. The differences in the growth performance of the P-based diet is often related to reduce feed intake^(4,7). To mitigate such a negative effect, a set of selected compounds comprising glucosamine, taurine and betaine, *a priori* not found in plant-based ingredients, complemented with glycine and alanine was also intentionally added to mimic the expected composition in the M diet⁽⁶⁾. Betaine and glucosamine are known to feed attractants and growth-promoting compounds in fish^(47,48). Furthermore, some of

these compounds have specific biological functions such as preserving tissue osmotic pressure for taurine⁽⁴⁹⁾ and contributing to C1 metabolism for betaine⁽⁵⁰⁾. Taurine is now considered as an essential nutrient, especially in species that could not synthesise taurine⁽⁵¹⁾. Thus, the adequate supplementation with specific compounds most likely contributes to the relatively good performances of fish fed with the P-based diet⁽⁴¹⁾ offering optimum condition to assess the long-term effects of the P-based diet.

The objective of this study was to characterise feeds and fish metabolic response by NMR metabolomic approaches in order to explain the differences observed in the growth performance. Metabolomic approaches in plasma and tissues helped to identify which tissues and which metabolic pathways were mainly altered in fish fed the P-based diet and how this could be used for making further improvements to the alternative diets.

Soluble compounds in experimental feeds as putative diet biomarkers

Although ¹H-NMR analyses are often used to characterise food and feed ingredients⁽⁵²⁾ and to assess fish product quality and origin⁽⁵³⁾, they have only just started to be implemented on fish feed^(38,54,55). The alcoholic extracts of the diets contained a rich and complex mixture of soluble compounds representative of free soluble compounds originally present in marine products and plant products. Some compounds constitute markers of feedstuffs and ingredients of the diet. Creatine, anserine, niacinamide and adenosine detected only in M and C diets are known to be present in FM^(56,57). These compounds were also detected in the plasma of fish fed M and C diets as well as dimethylamine. Secondary amines, dimethylamine and trimethylamine, which are specific to marine resources fish and seafood^(49,54,58), have been specifically detected in the plasma of fish fed an FM-based diet⁽⁵⁹⁾. Some other compounds, such as sugar compounds, trigonelline and biopterin, are specific to plant-based ingredients. Indeed, the P-based and the C showed large quantities of sugar sucrose and stachyose. Stachyose is suspected to be involved in the gut enteritis related to soyabean meal ingestion, but in the present conditions, it did not induce any differences in the growth of fish fed with the C diet compared with the M diet. Further investigations are required to conclude on the effect of this oligosaccharide in gut alteration⁽⁶⁰⁾.

Two compounds, identified as pterin and biopterin, specific to plants⁽⁶¹⁾ were detected only in the P-based diet and in the plasma of fish fed the P-based diet. The metabolic effects of pterin and biopterin are not known, contrary to those of folic acid, an analogue⁽⁶²⁾. Pterin found in plants is supposed to be distinct from biopterin and neopterin, which are metabolites of guanosine found in animals and particularly in fish, where they contribute to skin pigmentation⁽⁵⁷⁾. Pterin detected in plasma could, thus, be considered as a specific biomarker of the P experimental diet. Other unknown compounds detected only in the plasma of fish fed P-based diet are also putative biomarkers of plant feedstuffs, as they get to be revealed by means of metabolomic approaches^(38,50).

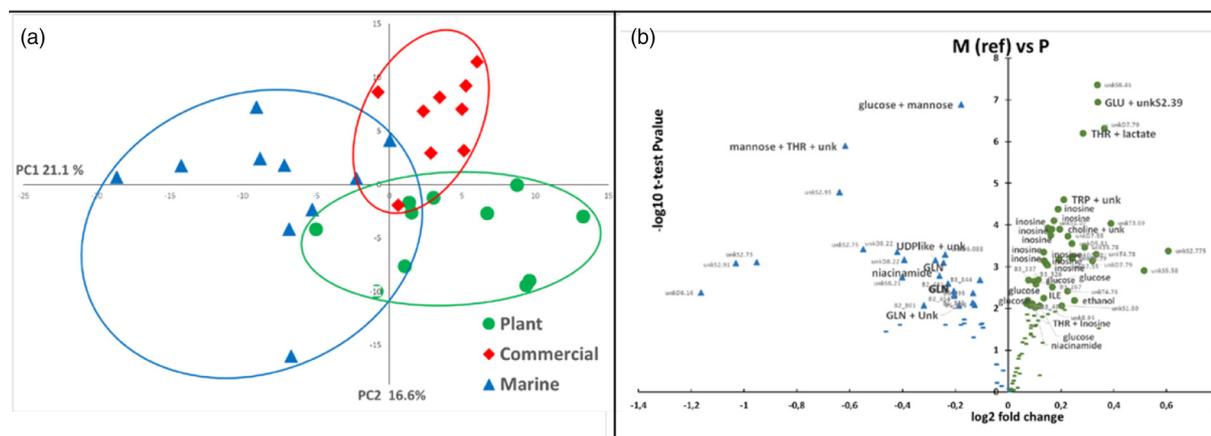


Fig. 6. Multidimensional PCA of $^1\text{H-NMR}$ spectra of liver aqueous polar extracts segmented in 341 spectral regions. (a) Representation of the scores on the two first principal components of rainbow trout 48 h after feeding on a M diet (triangle n 10), a P-based diet (circle n 11) and a C diet (diamond n 9). (b) Volcano plot representation of the differences of the signals of spectral regions between M and P-based diets with the M diet used as a reference. Circle, significantly higher signal ($P < 0.05$); triangle, significantly lower signal ($P < 0.05$); dash, non-significant differences.

The combined metabolomic analysis of the diet and the plasma, thus, appears to be promising tools to identify a set of plant-based soluble compounds presumably accounting for known and suspected alterations or stimulation in the ingestion and digestion of the P-based diet.

Table 8. Significant effects of diet (M v. P-based) on semi-quantitative assessment of representative spectral regions or bucket of the $^1\text{H-NMR}$ spectra of a hydro-alcoholic extract of liver in rainbow trout

Bucket	Metabolite resonance annotation	Significance diet
M > P-based diet		
B2_445; B2_478; B2_487	Glutamine	***
B3_587	unk + mannose + threonine	***
B3_774	Glucose + mannose	***
B4_312	unk + uridine	***
B8_267	Niacinamide	**
P-based diet > M		
B0_928	Isoleucine	*
B1_216	Ethanol	**
B1_350	Threonine + lactic acid	***
B1_802	Putrescine	*
B3_390	Glucose	*
B2_391	unkS2.39 + glutamate	***
B8_46	Formic acid	***
B3_255; B3_390; B3_557; B3_566; B3_920	Glucose	***
B4_073	unk + choline	***
B4_285; B4_294; B4_303; B4_438; B4_448; B4_453; B4_462; B6_105; B6_124; B8_361	Inosine	***
B4_776; B4_787; B4_798	Inosine	**
B5_804; B5_824; B7_548; B7_563	Uracile	***
B7_310	unk + tryptophan	***
B8_940	Niacinamide	**

ANOVA on the diet effect (for M diet, n 10; for P-based diet, n 11): *** $P < 0.001$; ** $P < 0.01$; * $P < 0.05$.

Delay in amino acid absorption and imbalanced supply of essential amino acids to tissues in fish fed the P-based diet

The substitution of marine feedstuffs by plant-based feedstuffs, particularly plant-based protein feedstuffs, is generally associated with limitations in fish digestion and metabolism⁽³⁹⁾. These limits are thought to be attributed to the physical and chemical characteristics of plant-based feedstuffs with regard to specific anti-nutritional factors⁽³⁹⁾. Our data strongly support this hypothesis.

On the one hand, the metabolite plasma profile at 6 h post feeding is mainly influenced by the diet. It depends essentially on diet composition reflecting both food metabolome combined with the efficiency of the digestion and absorption processes of the last meal. This was illustrated by the status of lysine and methionine, which were supplemented in their free form in the P-based diet. These two purified amino acids are absorbed before the bolus of amino acids from intact protein digestion^(13,14), and this was attested in the present work by their higher plasma content at 6 h post feeding and a significant interaction between post-feeding time and diet.

On the other hand, the plasma profile at 48 h post feeding, equivalent in fish to a short fasting period, is mainly influenced by fish metabolism and, thus, reflects rather the fish metabolome. Moreover, there could be still an influence of the diet, as at the end of the trial, only a single meal was distributed, and this could slow down the transit. The higher concentration and accumulation of compounds – mainly amino acids – in the plasma of fish fed the P-based diet observed at 48 h post feeding compared with those fed the M diet could, thus, be the result of a delay in the delivery of these essential nutrients in the fish fed the P-based diet compared with those fed the M diet. These results are in accordance with previous studies performed using classical analytical approaches in rainbow trout^(63–65). The late accumulation of amino acids is suspected to be induced by the physical matrix of plant-based feedstuffs, which reduced amino acid availability and delayed their delivery. Indeed, this could induce a reduced metabolism, either due to an imbalance in amino acid supply to tissues or due

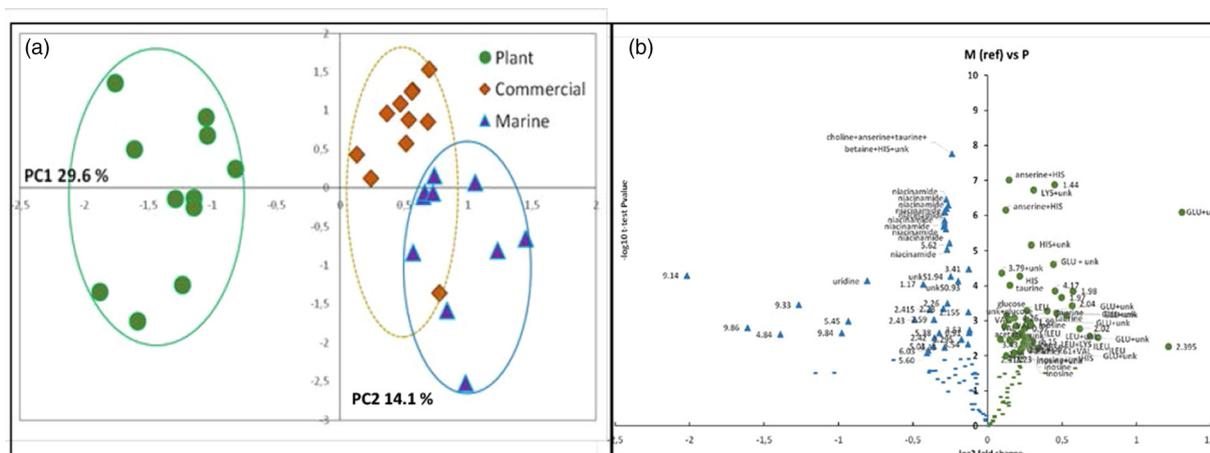


Fig. 7. Multidimensional PCA of $^1\text{H-NMR}$ spectra of muscle tissue aqueous polar extracts segmented in 276 spectral regions. (a) Representation of the scores on the first two principal components of rainbow trout 48 h after feeding on a M diet (triangle n 10), a P-based diet (circle n 11) and a C diet (diamond n 9). (b) Volcano plot representation of the differences of the signals of spectral regions between M and P-based diets with the M diet used as a reference. Circle, significantly higher signal ($P < 0.05$); triangle, significantly lower signal ($P < 0.05$); dash, non-significant differences.

Table 9. Significant effects of diet (M v. P-based) on semi-quantitative assessment of representative spectral regions or bucket of the $^1\text{H-NMR}$ spectra of a hydro-alcoholic extract of muscle in rainbow trout

Bucket	Metabolite resonance annotation	Significance diet
M > P-based diet		
B3_2250	Choline + anserine + taurine + betaine + histidine + unk	***
B3_5253; B3_5356; B4_0635; B4_0690 ^a	Choline	***
B7_5894; B7_6039; B7_6120; B7_6242; B8_7136; B8_7199; B8_7250; B8_7323; B8_9420; B8_9526	Niacinamide	***
P-based diet > M		
B0_9268; B0_9445; B0_9592; B1_0026; B1_0123	Isoleucine	** & ***
B0_9531; B0_9786	Leucine	***
B0_9856; B0_9933; B1_0245; B1_0407; B2_3088; B3_6127	Valine	* & ** & ***
B1_3117; B1_3538; B4_0979; B4_1119; B4_1269; B4_1459	Lactic acid	*
B1_4512; B1_4655	^{13}C lactic acid	
B1_6944; B1_7107	Leucine + unk	***
B1_8731; B1_8889	Lysine + unk	***
B1_9244	Acetate	***
B1_9660; B1_9913	Glutamate + unk	***
B2_0087; B2_0237; B2_0385; B2_0488	Glutamate	***
B2_0563; B2_3557; B2_3707; B4_1674	Glutamate + unk	** & ***
B2_0711; B2_0845; B2_0986; B2_1097; B2_3258; B2_3390; B2.4073 ^b	Glutamate	***
B3_0570	Creatine + P-creatine	*
B3_4242; B3_4403; B3_4539	Taurine	**
B3_5630	Glucose	**
B3_6127	unk + valine	*
B3_9967; B4_0049; B4_0121	Histidine	***
B4_2769; B4_2846; B4_4367; B4_4488; B4_4591	Inosine	** & ***
B6_9027; B6_9150	Tyrosine	* & **
B7_1465; B8_1235	Histidine + anserine	***

ANOVA on the diet effect (M: n 10; P: n 11): *** $P < 0.001$; ** $P < 0.01$; * $P < 0.05$.

^a B4_0690 = choline + ^{13}C creatine.

^b B2.4073 = unkS2.39 + glutamate.

to the non-synchronicity of amino acid supply with other energy substrate supplies. This hypothesis has begun to be formalised in human nutrition through the concept of 'fast' and 'slow' proteins that account for differences in the digestive and metabolic efficiency of meat v. plant-based protein, especially in elderly people⁽⁶⁶⁾. This concept needs to be taken into consideration in fish nutrition for the development of

alternative diets such as the P-based diets but also of new ingredients (micro-algae, insects, etc.). Alternatively, the higher concentration of amino acids in plasma could also result in an accumulation of amino acids in tissues, as was observed in muscle tissue, suggesting an alteration in amino acid utilisation.

Thus, the early delivery of supplemented essential amino acid in fish fed the P-based diet, associated with the delay

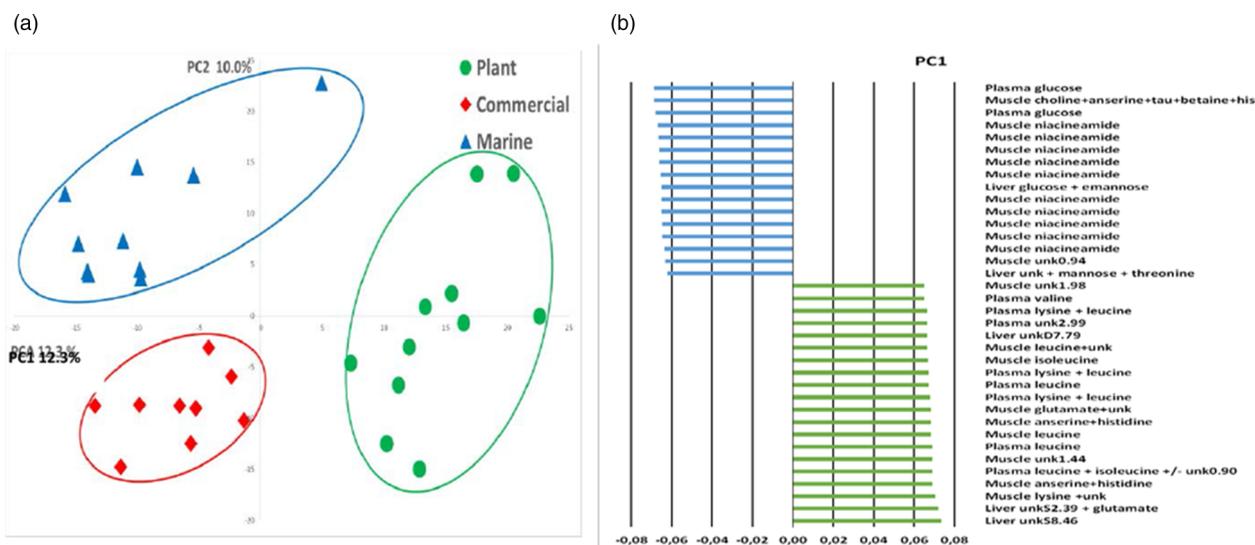


Fig. 8. Multidimensional PCA analysis of $^1\text{H-NMR}$ spectra of plasma, liver and muscle polar extracts. (a) Representation of the scores on the first two principal components of rainbow trout 48 h after feeding on a M diet (triangle n 10), a P-based diet (circle n 11) and a C diet (diamond n 9). (b) Representation of the main significant spectral regions involved in the PC1-axis differences.

observed in the delivery of amino acids from the digestion of plant-based protein, would induce an imbalanced supply of essential amino acids to the tissues.

Minimised but targeted contribution of liver to whole-body metabolism

Liver is orchestrating the delivery of metabolites to other tissues, but, in the present study, its contribution seems to be minimal if one considers the faint diet separation between diets in this tissue. It means that liver metabolism was slightly affected by P-based diet compared with plasma and muscle even taking into account that at 48 h post feeding, the contribution of liver to metabolite exchanges could be largely reduced.

The increased status of choline in liver and the corresponding decrease in the muscle tissue of fish fed the P-based diet should be considered. Choline is a precursor of phospholipid metabolism, and we found in a previous study that the metabolisms of lipid and cholesterol are stimulated in the liver of fish fed the P-based diet⁽⁴¹⁾. On the other hand, choline and related compounds are well known to contribute to one-carbon metabolism in interaction with methionine metabolism. It has been demonstrated in fish that the sub-deficiency of methionine requires an increased supply of choline to maintain methionine status⁽⁶⁷⁾. Choline and its metabolites are specifically affected in other fish species fed P-based diet diets^(54,68). Betaine (trimethylglycine), dimethylglycine and serine are intermediate metabolites of the choline metabolic pathway, where they act as $-\text{CH}_3$ and $-\text{CH}_2\text{OH}$ donors, particularly for methionine synthesis⁽⁶⁹⁾. Furthermore, early absorption of free-form methionine probably stimulates its use as a methyl donor in the liver. This will contribute to the preservation of choline but could, however, induce, a further deficiency of methionine in other tissues such as muscle.

Long term adaptation of muscle tissue to unbalanced supplies of amino acids

The adaptation of fish to long term feeding with only plant-based ingredients has been demonstrated in salmonids^(5,6,18) and several marine fish⁽⁷⁾, but growth performances are generally lower than with marine ingredients. Muscle metabolism would, thus, be specifically affected by a P-based diet and in the present study, muscle demonstrated a better discrimination of the P-based diet with the M and C diets than in plasma and liver. The main differences observed in muscle tissue at 48 h after feeding were an accumulation of essential amino acids in the fish fed the P-based diet. This suggests an amino acid imbalance for protein synthesis due to the limited supply of certain essential amino acids to muscle tissue. Changes observed in muscle tissue were related to other differences observed, i.e. a delay in amino acid release in fish plasma fed the P-based diet and an imbalanced supply of essential amino acids observed in plasma and liver tissue due to the early absorption of methionine and lysine in fish fed the P-based diet. These results clearly show that both an overall balanced supply of essential amino acids and the synchronic delivery of all essential amino acids to muscle tissue are crucial for protein turnover and muscle growth.

Further alterations in energy and vitamin status

The P-based diet also induced alterations in energy metabolism. It is noteworthy that glucose levels are not significantly different at 6 and 48 h after feeding, and this could be due to the fact that all diets comprise a significant amount of starch ingredients. Due to the low utilisation of dietary glucose, it is generally observed that plasma glucose can be maintained at high levels during post-prandial time, even 12–24 h after the last feeding in rainbow trout fed either M or P-based diet^(18,40). All diets contained high levels of starch, and, thus, the differences observed in plasma glucose at 48 h between M and P-based diets could not be attributed simply to the



plant-based ingredients. Furthermore, the same reduction in plasma glucose was observed in cobia fed experimental diets with an increased substitution of FM (25 and 75 %) by plant-based protein⁽³²⁾. Specific changes in the liver metabolism of fish fed a P-based diet have also been observed using transcriptomic and proteomic approaches. These changes concern mainly the intermediary metabolism, through lipid and carbohydrate metabolism interactions for neoglucogenesis, and lipid and protein metabolism interactions for the control of protein catabolism^(4,7). In our analysis of liver tissue, 48 h after feeding, the main differences concerned glucose, inosine and even ethanol, which were found to be higher in the liver of fish fed the P-based diet. This is consistent with an alteration of the liver's energy metabolism towards neoglucogenesis from amino acids, as demonstrated in previous transcriptomic studies^(4,7). Furthermore, the lower content of glutamine in the liver of fish fed the P-based diet could also account for a higher contribution of amino acids to the energy metabolism. The accumulation of amino acids observed in fish plasma fed the P-based diet could lead to their use to produce energy substrate in the liver, particularly through glutamate as an entry point into the TCA cycle. Finally, the creatine and the niacinamide status of muscle were altered in fish fed the P-based diet. This has already been observed for creatine in fish fed a P-based diet^(30,36) but had not yet been demonstrated for niacinamide, a vitamin B metabolite involved in the NAD synthesis. The global analysis showed that the main differences observed in plasma, liver and muscle energy status, and those observed in liver choline cycle and muscle protein metabolism were connected. Thus, these alterations in energy metabolism could account for the adaptation of fish metabolism to an imbalanced and delayed supply of nutrients.

Conclusions

This study shows the great interest of implementing metabolomic approaches on biological fluids and tissues in order to assess the long-term effects of P-based diets on rainbow trout metabolism. Metabolomics helps to draw a coherent picture on the concomitant alterations of amino acid metabolism by feeding P-based diets: (i) an early supply of free-form lysine and methionine, (ii) an accumulation of amino acids in fish fed a P-based diet observed at the plasma level, (iii) an alteration of choline-related metabolites for $-CH_3$ supply to the liver and (iv) an imbalance in amino acids for protein synthesis in muscle tissue. This constitutes relevant targets for future research on protein digestion, amino acid metabolism and protein metabolism that should be considered to improve P-based diets for rainbow trout and farmed fish in general.

Supplementary material

The supplementary material for this article can be found at <https://doi.org/10.1017/jns.2021.3>.

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G.C. and F.M. have designed and managed the fish nutrition part; B.M.H. and B.F. have contributed to biofluid and tissue sampling; B.M.H. has contributed to tissue extraction; B.M.H., C.D. and M.M. have managed the metabolomic part (NMR acquisition, spectral analysis, quantification and annotation) under the supervision of A.M.; D.J. has contributed to the statistics for the metabolomic part, including the development of software such as BioStatFlow and NMRProcFlow; and B.F. has supervised this work, contributed to tissue sampling, spectral analysis and statistical analysis and written the manuscript. All authors read and approved the final manuscript.

The authors declare that they have no conflicts of interest.

References

1. Food and Agriculture Organisation of the United Nations (2014) *The State of World Fisheries and Aquaculture: Opportunities and Challenges*, p. 223, Rome: FAO Press.
2. Tacon AGJ & Metian M (2008) Global overview on the use of fish meal and fish oil in industrially compounded aquafeeds. *Trends Future Prospects Aquaculture* **285**, 146–158.
3. National Research Council (2011) *Nutrients Requirements of Fish and Shrimp*, p. 376, Washington, D.C.: National Academies Press.
4. Panserat S, Hortopan GA, Plagnes-Juan E, *et al.* (2009) Differential gene expression after total replacement of dietary fish meal and fish oil by plant products in rainbow trout (*Oncorhynchus mykiss*) liver. *Aquaculture* **294**, 123–131.
5. De Francesco M, Parisi G, Médale F, *et al.* (2004) Effect of long-term feeding with a plant protein mixture based diet on growth and body/fillet quality traits of large rainbow trout. *Aquaculture* **236**, 413–429.
6. Lazzarotto V, Corraze G, Leprevost A, *et al.* (2015) Three-year breeding cycle of rainbow trout (*Oncorhynchus mykiss*) fed a plant-based diet, totally free of marine resources: Consequences for reproduction, fatty acid composition and progeny survival. *PLoS One* **10**, e0117609, doi:10.1371/journal.pone.0117609.
7. Geay F, Ferrareso S, Zambonino-Infante JL, *et al.* (2011) Effects of the total replacement of fish-based diet with plant-based diet on the hepatic transcriptome of two European sea bass (*Dicentrarchus labrax*) half-sibfamilies showing different growth rates with the plant-based diet. *BMC Genomics* **12**, 522.
8. Le Boucher R, Vandeputte M, Dupont-Nivet M, *et al.* (2013) Genotype by diet interactions in European sea bass (*Dicentrarchus labrax* L.): Nutritional challenge with totally plant-based diets. *J Anim Sci* **91**, 44–56.
9. Nasopoulou C & Zabetakis I (2012) Benefits of fish oil replacement by plant originated oils in compounded fish feeds. A review. *LWT – Food Sci Technol* **47**, 217–224.
10. Hardy RW (2010) Utilization of plant proteins in fish diets: Effects of global demand and supplies of fish meal. *Aquaculture Res* **41**, 770–776.
11. Yamamoto T, Unuma T & Akiyama T (2000) The influence of dietary protein and fat levels on tissue free amino acid levels of fingerling rainbow trout *Oncorhynchus mykiss*. *Aquaculture* **182**, 353–372.
12. Larsen BK, Dalsgaard J & Pedersen PB (2012) Effects of plant proteins on postprandial, free plasma amino acid concentrations in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* **326–329**, 90–98.



13. Schuhmacher A, Wax V & Gropp JM (1997) Plasma amino acids in rainbow trout (*Oncorhynchus mykiss*) fed intact protein or a crystalline amino acid diet. *Aquaculture* **151**, 15–28.
14. Ambardekar AA, Reigh RC & Williams MB (2009) Absorption of amino acids from intact dietary proteins and purified amino acid supplements follows different time-courses in channel catfish (*Ictalurus punctatus*). *Aquaculture* **291**, 179–187.
15. Jia S, Li X, Zheng S, *et al.* (2017) Amino acids are major energy substrates for tissues of hybrid striped bass and zebrafish. *Amino Acids* **49**, 2053–2063.
16. Bar NS & Radde N (2009) Long-term prediction of fish growth under varying ambient temperature using a multiscale dynamic model. *BMC Syst Biol* **3**, 107–125.
17. Panserat S (2009) Molecular regulation of intermediary metabolism focusing on utilization of dietary carbohydrates. In *Molecular Research in Aquaculture*, pp. 261–278 [K Overturf, editor]. Wiley-Blackwell.
18. Veron V, Panserat S, Le Boucher R, *et al.* (2016) Long-term feeding a plant-based diet devoid of marine ingredients strongly affects certain key metabolic enzymes in the rainbow trout liver. *Fish Physiol Biochem* **42**, 771–785.
19. Zduńczyk Z & Pareek CS (2009) Application of nutrigenomics tools in animal feeding and nutritional research. *J Animal Feed Sci* **18**, 3–16.
20. Primrose S, Draper J, Elsom R, *et al.* (2011) Metabolomics and human nutrition. *Br J Nutr* **105**, 1277–1283.
21. Claus SP & Swann JR (2013) Nutrimetabonomics: Applications for nutritional sciences, with specific reference to gut microbial interactions. *Annu Rev Food Sci Technol* **4**, 381–399.
22. Gribbestad IS, Aursand M & Martínez I (2005) High-resolution 1H magnetic resonance spectroscopy of whole fish, fillets and extracts of farmed Atlantic salmon (*Salmo salar*) for quality assessment and compositional analyses. *Aquaculture* **250**, 445–457.
23. Mannina L, Sobolev AP, Capitani D, *et al.* (2008) NMR metabolic profiling of organic and aqueous sea bass extracts: Implications in the discrimination of wild and cultured sea bass. *Talanta* **77**, 433–444.
24. Dove ADM, Leisen J, Zhou M, *et al.* (2012) Biomarkers of whale shark health: A metabolomic approach. *PLoS One* **7**, e49379.
25. Lardon I, Nilsson GE, Stecyk JAW, *et al.* (2013) H-1-NMR study of the metabolome of an exceptionally anoxia tolerant vertebrate, the crucian carp (*Carassius carassius*). *Metabolomics* **9**, 311–323.
26. Kullgren A, Samuelsson LM & Larsson DGJ (2010) A metabolomics approach to elucidate effects of food deprivation in juvenile rainbow trout (*Oncorhynchus mykiss*). *Am J Physiol Regul Integr Comp Physiol* **299**, R1440–R1448.
27. Gil-Solsona R, Nacher-Mestr J, Lacalle-Bergeron L, *et al.* (2017) Untargeted metabolomics approach for unraveling robust biomarkers of nutritional status in fasted gilthead sea bream (*Sparus aurata*). *PeerJ* **5**, e2920; doi:10.7717/peerj.2920.
28. Picone G, Balling ES, Savorani F, *et al.* (2011) Metabolomics as a powerful tool for molecular quality assessment of the fish *Sparus aurata*. *Nutrients* **3**, 212–227.
29. Asakura T, Sakata K, Yoshida S, *et al.* (2014) Noninvasive analysis of metabolic changes following nutrient input into diverse fish species, as investigated by metabolic and microbial profiling approaches. *PeerJ* **2**, e550; doi:10.7717/peerj.550.<query for epub date>
30. Wagner L, Trattner S, Pickova J, *et al.* (2014) ¹H NMR-based metabolomics studies on the effect of sesamin in Atlantic salmon (*Salmo salar*). *Food Chem* **147**, 98–105.
31. Chauton MS, Galloway TF, Kjorsvik E, *et al.* (2015) ¹H-NMR metabolic profiling of cod (*Gadus morhua*) larvae: Potential effects of temperature and diet composition during early developmental stages. *Biol Open* **4**, 1671–1678, doi:10.1242/bio.014431.
32. Egea FCM, Toledo-Guedes K, Sanchez-Jerez P, *et al.* (2015) A metabolomic approach to detect effects of salmon farming on wild saithe (*Pollachius virens*) populations. *J Agric Food Chem* **63**, 10717–10726; doi:10.1021/acs.jafc.5b04765.
33. Schock TB, Newton S, Brenkert K, *et al.* (2012) An NMR-based metabolomic assessment of cultured cobia health in response to dietary manipulation. *Food Chem* **133**, 90–101.
34. Kullgren A, Jutfelt F, Fontanillas R, *et al.* (2013) The impact of temperature on the metabolome and endocrine metabolic signals in Atlantic salmon (*Salmo salar*). *Comp Biochem Physiol* **164**, 44–53.
35. Abro R, Moazzami AA, Lindberg JE, *et al.* (2014) Metabolic insights in Arctic charr (*Salvelinus alpinus*) fed with zygomycetes and fish meal diets as assessed in liver using nuclear magnetic resonance (NMR) spectroscopy. *Int Aquat Res* **6**, 1–11.
36. Wei Y, Liang M, Mai K, *et al.* (2016) The effect of ultrafiltered fish protein hydrolysate levels on the liver and muscle metabolic profile of juvenile turbot (*Scophthalmus maximus* L.) by 1H NMR-based metabolomics studies. *Aquac Res* **2016**, 1–13; doi:10.1111/are.13178.
37. Roques S, Deborde C, Richard N, *et al.* (2020) Metabolomics and fish nutrition: A review in the context of sustainable feed development. *Rev Aquac* **12**, 261–282.
38. Roques S, Debordes C, Richard N, *et al.* (2018) Characterizing alternative feeds for rainbow trout (*O. mykiss*) by ¹H NMR metabolomics. *Metabolomics* **14**, 155–167.
39. Urán P, Schrama JW, Jaafari S, *et al.* (2009) Variation in commercial sources of soybean meal influences the severity of enteritis in Atlantic salmon (*Salmo salar* L.). *Aquac Nut* **15**, 492–499.
40. Borey M, Panserat S, Surget A, *et al.* (2016) Postprandial kinetics of gene expression of proteins involved in the digestive process in rainbow trout (*O. mykiss*) and impact of diet composition. *Fish Physiol Biochem* **42**, 1187–1202.
41. Lazzarato V, Médale F, Larroquet L, *et al.* (2018) Long-term dietary replacement of fishmeal and fish oil in diets for rainbow trout (*Oncorhynchus mykiss*): Effects on growth, whole body fatty acids and intestinal and hepatic gene expression. *PLoS One* **13**, e0190730; doi.org/10.1371/journal.pone.0190730.
42. Moing A, Maucourt M, Renaud C, *et al.* (2004) Quantitative metabolic profiling by 1-dimensional ¹H-NMR analyses: Application to plant genetics and functional genomics. *Funct Plant Biol* **31**, 889–902.
43. Folch J, Lees M & Sloane-Stanley GH (1957) A simple method for the isolation and purification of total lipids from animal tissues. *J Biol Chem* **226**, 497–509.
44. Gatesoupe FJ, Fauconneau B, Deborde C, *et al.* (2018) Intestinal microbiota in rainbow trout, *Oncorhynchus mykiss*, fed diets with different levels of fish-based and plant ingredients: A correlative approach with some plasma metabolites. *Aquac Nutr* **24**, 1563–1576.
45. Jacob D, Deborde C, Lefebvre M, *et al.* (2017) NMRProcflow: A graphical and interactive tool dedicated to 1D spectra processing for NMR-based metabolomics. *Metabolomics* **13**, 36, doi:10.1007/s11306-017-1178-y.
46. Sumner LW, Amberg A, Barrett D, *et al.* (2007) Proposed minimum reporting standards for chemical analysis: Chemical analysis working group (CAWG) metabolomics standards initiative (MSI). *Metabolomics* **3**, 211–221, doi:10.1007/s11306-007-0082-2.
47. Tiril SU, Alagil F, Yagci FB, *et al.* (2008) Effects of betaine supplementation in plant protein based diets on feed intake and growth performance in rainbow trout (*Oncorhynchus mykiss*). *Isr J Aquacult – Bamid* **60**, 57–64.
48. Chowdhary S, Srivastava PP, Mishra S, *et al.* (2012) Synergistic effects of dietary glucosamine and plant/animal proteins on the growth performance of Asian catfish (*Clarias batrachus*) juveniles. *Online J Feed Anim Res* **2**, 50–57.
49. Shen G, Huang Y, Dong J, *et al.* (2018) Metabolic effect of dietary taurine supplementation on Nile tilapia (*Oreochromis niloticus*) evaluated by NMR-based metabolomics. *J Agric Food Chem* **66**, 368–377.
50. Obeid R (2013) The metabolic burden of methyl donor deficiency with focus on the betaine homocysteine methyltransferase pathway. *Nutrients* **5**, 3481–3495.
51. Salze GP & Davis DA (2015) Taurine: A critical nutrient for future fish feeds. *Aquaculture* **437**, 215–229.
52. Mannina L, Sobolev AP & Viel S (2012) Liquid state 1H high field NMR in food analysis. *Prog Nucl Magn Reson Spectrosc* **66**, 1–39.
53. Vidal NP, Manzanos MJ, Goicoechea E, *et al.* (2012) Quality of farmed and wild sea bass lipids studied by ¹H NMR: Usefulness of this technique for differentiation on a qualitative and a quantitative basis. *Food Chem* **35**, 1583–1591.



54. Cheng K, Müllner E, Moazzami AA, *et al.* (2017) Metabolomics approach to evaluate a Baltic Sea sourced diet for cultured Arctic char (*Salvelinus alpinus* L.). *J Agric Food Chem* **65**, 5083–5090.
55. Gil-Solsona R, Caldach-Giner JA, Nacher-Mestre J, *et al.* (2019) Contributions of MS metabolomics to gilthead sea bream (*Sparus aurata*) nutrition. Serum fingerprinting of fish fed low fish meal and fish oil diets. *Aquaculture* **498**, 503–512.
56. Souci SW, Fachmann W & Kraut H (2008) *Food Composition and Nutrition Tables*, 7th ed., p. 1364. Germany: Medpharm Scientific Edition and CRC Press Book Taylor & Francis Group Stuttgart.
57. Wu TH & Bechtel PJ (2012) Screening for low molecular weight compounds in fish meal solubles by hydrophilic interaction liquid chromatography coupled to mass spectrometry. *Food Chem* **130**, 739–745.
58. Arvanitoyannis IS (2016) *Authenticity of Foods of Animal Origin. Food Biology Series*, p. 312. FL, USA: CRC Press Boca Raton.
59. Jasour MS, Wagner L, Sundekilde UK, *et al.* (2017) A comprehensive approach to assess feathermeal as an alternative protein source in aquafeed. *J Agri Food Chem* **65**, 10673–10684.
60. Sørensen M, Penn M, El-Mowafi A, *et al.* (2011) Effect of stachyose, raffinose and soya-saponins supplementation on nutrient digestibility, digestive enzymes, gut morphology and growth performance in Atlantic salmon (*Salmo salar*, L.). *Aquaculture* **314**, 145–152.
61. Basua P & Burgmayerb SJN (2011) Pterin chemistry and its relationship to the molybdenum cofactor. *Coord Chem Rev* **255**, 1016–1038.
62. Hemre GI, Lock EJ, Olsvik PA, *et al.* (2005) Atlantic salmon (*Salmo salar*) require increased dietary levels of B-vitamins when fed diets with high inclusion of plant based ingredients. *PeerJ* **4**, e2493; doi:10.7717/peerj.2493
63. Yamamoto T, Unuma T & Akiyama T (2000) The influence of dietary protein and fat levels on tissue free amino acid levels of fingerling rainbow trout *Oncorhynchus mykiss*. *Aquaculture* **182**, 353–372.
64. Larsen BK, Dalsgaard J & Pedersen PB (2012) Effects of plant proteins on postprandial, free plasma amino acid concentrations in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* **326–329**, 90–98.
65. Rolland M, Larsen BK, Holm J, *et al.* (2015) Effect of plant proteins and crystalline amino acid supplementation on postprandial plasma amino acid profiles and metabolic response in rainbow trout (*Oncorhynchus mykiss*). *Aquac Int* **23**, 1071–1087.
66. Dangin M, Boirie Y, Guillet C, *et al.* (2002) Influence of the protein digestion rate on protein turnover in young and elderly subjects. *J Nutr* **132**, 3228S–3233S.
67. Espe M, Zerrahn JE, Hølen E, *et al.* (2016) Choline supplementation to low methionine diets increase phospholipids in Atlantic salmon, while taurine supplementation had no effects on phospholipid status, but improved taurine status. *Aquac Nut* **22**, 776–785, doi:10.1111/anu.12298.
68. Cheng K, Wagner L, Moazzami AA, *et al.* (2016) Decontaminated fishmeal and fish oil from the Baltic Sea are promising feed sources for Arctic char (*Salvelinus alpinus* L.) – Studies of flesh lipid quality and metabolic profile. *Eur J Lipid Sci Technol* **118**, 862–873, doi:10.1002/ejlt.201500247.
69. Chao J, Huo TL, Cheng HY, *et al.* (2014) Gallic acid ameliorated impaired glucose and lipid homeostasis in high fat diet-induced NAFLD mice. *PLoS One* **9**, e96969, doi:10.1371/journal.pone.0096969.