Assessment of trophic segregation amongst gentoo penguin (*Pygoscelis papua*) individuals in Antarctica using a non-invasive methodology

LUCÍA RABINOVICH-LARRECHEA ¹, DANIEL E. NAYA^{1,2}, MARIANA COSSE³, NADIA BOU³ and VALENTINA FRANCO-TRECU¹

¹Department of Ecology and Evolution, Faculty of Sciences, University of the Republic, Montevideo, Uruguay ²Museum of Vertebrate Zoology, University of California, Berkeley, CA, USA ³Department of Biodiversity and Genetics, Clemente Estable Biological Research Institute (IIBCE-MEC), Montevideo, Uruguay Irabinovich@fcien.edu.uy

Abstract: Individual trophic specialization (ITS) refers to the trophic diversification amongst individuals within a population. The gentoo penguin (*Pygoscelis papua*) is considered a trophic generalist at the population level, but little is known about its individual trophic differentiation. We assessed the degree of ITS at one of its main breeding colonies: Ardley Island, South Shetland Islands. We used skin from 19 dead individuals to determine species and sex by molecular methods and a nail for stable isotope analysis of δ^{15} N and δ^{13} C. Isotopic niche metrics and ITS were estimated for the population and for each sex. We found a moderately high degree of ITS associated with the trophic position of the resources consumed (δ^{15} N) for the population and both sexes, as well as a moderate degree of ITS in the foraging habitat (δ^{13} C) for the population and females. Females showed a higher exclusive niche area, suggesting that they use resources and foraging areas that males do not, probably related to reproductive energy demands. Given the high population density of this species, ITS could function as a mechanism to decrease intraspecific competition. This combination of genetic and isotopic tools allowed us to provide relevant information on the trophic ecology of the gentoo penguin without manipulating animals or using invasive methods.

Received 14 April 2023, accepted 26 December 2023

Keywords: intrapopulation variation, penguins, stable isotope analysis, trophic ecology

Introduction

Studying the trophic ecology of wild populations is relevant given that feeding habits determine trophic web connections, and therefore such trophic ecology could affect community structure to a significant extent (Araújo et al. 2011, Dall et al. 2012). In this sense, a generalist population may be composed of generalist individuals who use all of the resources consumed by the population, or it may be the sum of groups of individuals with specialized trophic strategies (Bolnick et al. 2002, 2007, Dall et al. 2012). The latter is called individual trophic specialization (ITS). ITS refers to intra-population variation in the use of resources not attributable to sex, age or discrete morphs of individuals (Bolnick et al. 2003, Araújo et al. 2011). One proposed ecological cause for ITS is intraspecific competition: if individuals share a preferred (set of) resource(s), increased competition can lead them to diversify their trophic habits by consuming different secondary prey (Bolnick et al. 2003, 2007, Araújo et al. 2011, Dall et al. 2012). ITS can have important ecological and evolutionary consequences by affecting population and community dynamics (Araújo et al. 2011, Dall et al. 2012). In addition, the degree of individual specialization can affect the strength of competitive interactions between sympatric species and, consequently, the probability of persistence of a species in a given environment (e.g. Costa-Pereira *et al.* 2018).

The degree of ITS in a population can be estimated as the proportion of the population niche that is explained by the inter-individual variance component (Roughgarden 1974, Bolnick *et al.* 2002, 2007). The total niche width (TNW) of a population can be divided into two components: the within-individual variance component (WIC), which reflects the average diversity of resources used by each individual; and the betweenindividual variance component (BIC), which reflects the average variation in resource use amongst individuals in the population (TNW = WIC + BIC; Roughgarden 1972). Accordingly, the ITS index (BIC/TNW) ranges from 0, when the population is composed of generalist individuals, to 1, when all of the individuals are specialists (Bolnick *et al.* 2002).

The ITS in a population is determined by studying the temporal consistency in the trophic habits of its individuals (Costa-Pereira & Araújo 2022). In recent times, one of the main methodologies used to study this consistency is stable isotope analysis (SIA) of δ^{15} N and δ^{13} C. δ^{15} N mainly allows us to infer the trophic position of an individual (Post 2002) and, secondarily, to discriminate between feeding zones (McMahon *et al.* 2013, Brault *et al.* 2018). δ^{13} C provides information on the origin of primary productivity, enabling the foraging habitat to be inferred (France 1995). For instance, in the marine environment, δ^{13} C distinguishes between the use of benthic *vs* pelagic prey (France 1995).

Individual specialization could be assessed using SIA in three different ways: 1) sequential sampling of the same tissue in the same individuals, 2) simultaneous sampling of different tissues (i.e. a multi-tissue approach) and 3) sampling of different portions of a metabolically inert tissue that undergoes continuous growth (Nava & Franco-Trecu 2019 and references therein). Continuous growing metabolically inert tissues (e.g. vibrissae, nails, feathers) maintain an unchanged isotopic signal once synthesized, and hence repeated measurements of the trophic habits of an individual can be obtained over periods of months and even years (i.e. Silva et al. 2015). SIA of these tissues allows researchers to evaluate the variation in the trophic habits of the individuals relative to the total population variation and therefore the degree of ITS (Newsome et al. 2009, Vander Zanden et al. 2010). Whilst these tissues have been used to study trophic habits (not necessarily ITS), they are often sampled invasively as they involve the manipulation of living individuals (e.g. use of vibrissae from sea lions and sea otters (Newsome et al. 2009, de Lima et al. 2022) and feathers in birds (Jaeger et al. 2009)). Moreover, the tips of nails are cut off when using the nails in living animals, and this only allows us to obtain isotopic data from a short and sometimes unknown period of time (Cherel et al. 2007). Therefore, the use of metabolically inert tissues obtained from dead individuals is a good alternative to avoid sampling living animals, thereby constituting a valuable non-invasive methodology (Ainley et al. 2003, Vasil et al. 2012). Moreover, in the case of nails, using dead animals allows us to sample the entire nail and thus to obtain the isotopic data regarding the whole of the period of time it integrates.

The gentoo penguin (*Pygoscelis papua*) is a mesopredator species with a circumpolar distribution (Clucas *et al.* 2014), and it is considered a generalist species with a diverse and flexible diet at the population level (Polito *et al.* 2015, Negrete *et al.* 2016). Although some populations consume fish and squid in addition to Antarctic krill (*Euphausia superba*; Karnovsky 1997, Polito *et al.* 2011, Gorman 2015), in others Antarctic krill remains their main prey, constituting more than 90% of the diet (Pickett *et al.* 2018).

Although several studies have assessed intra-individual differences in the trophic ecology of other *Pygoscelis*

species regarding environmental variability and population density (e.g. see Lescroël et al. 2009, 2014, 2020, Massaro et al. 2020 for studies in Pygoscelis adeliae), little is known about intra-population variation in P. papua. Until now, only two studies have explored the degree of ITS at the population level of this species (Herman 2016, Handley et al. 2017), without assessing potential differences between sexes. Given that ITS is affected by the diversity and abundance of local resources as well as by colony size, the degree of ITS in gentoo penguins varies not only between different islands (Herman 2016) but also between colonies of the same population (Handley et al. 2017). Moreover, the methodologies previously used have been invasive (i.e. stomach contents and SIA of feathers), involving the capture of individuals within the colony. Here, we aimed to assess a non-invasive methodology of using SIA of the nails of dead *P. papua* adults to estimate the degree of ITS in one of its main reproductive colonies: Ardley Island, South Shetland Islands. In addition, we tested differences in ITS between sexes. Considering the sustained growth in the population abundance of P. papua on Ardley Island (Roberts et al. 2017, Firla et al. 2019; see also 'Discussion' section below), which implies a decrease in the per capita abundance of local resources, we predicted that this population would have a high degree of ITS as a mechanism to decrease competition at the intra-population level (Araújo et al. 2011).

Materials and methods

Study area and sample collection

Ardley Island (62°13'S, 58°56'W) is located to the south-west of King George Island, which is part of the South Shetland Islands, Antarctica. It is considered a Site of Special Scientific Interest (SSSI No. 33) and is part of the Antarctic Specially Protected Areas (ASPA No. 150; ASPA 2009) given its relevance as a breeding and moulting site for various species of seabirds, such as *P. papua* (ASPA 2009). Ardley Island contains one of the largest reproductive colonies of *P. papua*, with more than 6000 breeding pairs (Roberts *et al.* 2017, Firla *et al.* 2019). There, *P. papua* inhabits in sympatry with chinstrap penguin (*Pygoscelis antarctica*) and Adélie penguin (*P. adeliae*), whose breeding colonies have ~50 and 300 breeding pairs, respectively (Roberts *et al.* 2017, Firla *et al.* 2019).

During January 2019, we made two visits to Ardley Island, in which feet from 19 carcasses belonging to adult individuals of the genus *Pygoscelis* were collected. The carcasses corresponded to recently predated individuals, so the feet were fresh. Both feet were collected from each carcass to ensure that the same

individual was not sampled twice during different sampling events. In the laboratory, the external portion of the central and longest nail was extracted. As nails are inert tissues, it is possible to obtain samples from recently dead individuals, constituting a non-invasive methodology (see Vasil *et al.* 2012). In addition, a skin sample was obtained from each foot and stored in Eppendorf tubes with 95% alcohol.

Sample analysis

DNA was isolated from tissue samples, following the procedure of González et al. (2015). The taxonomic identity of the samples was established by 500 bp amplification of the COI gene (mtDNA) using AWCF1 and AWCintR4 primers (Patel et al. 2010). Polymerase chain reactions (PCRs) were conducted in a final volume of 25 µl with 0.08 U Invitrogen[™] Platinum[™] Taq DNA Polymerase (Invitrogen Life Technologies, Carlsbad, CA, USA), 1X buffer, 2.5 mM, 1.25 µl bovine serum albumin (BSA), 0.75 µl of each primer and 100 ng of genomic DNA template. For the PCRs, an initial denaturing step at 94°C for 3 min was followed by 35 cycles of denaturation at 94°C for 45 s, annealing (55°C for 30 s) and extension at 72°C for 1 min, and this ended with a final extension of 72°C for 10 min. All PCR products were confirmed on 1% agarose gel and purified with Exonuclease I (Exo I) and FastAp following Werle et al. (1994). The purified products were sequenced on Macrogen Korea. Species was confirmed by a BLAST search against the GenBank nr database (http://blast. ncbi.nlm.nih.gov/Blast.cgi).

Sex determination was performed using molecular methods. PCR amplification of the CHD1 gene, located on birds' sex chromosomes, with P2 and P8 primers (Griffiths et al. 1998) produced fragments (CHD1W and CHD1Z) that differ in 18 bp length amongst the sex chromosomes of P. papua (Valenzuela-Guerra et al. 2013). Chromosome Z occurs in both sexes, whereas chromosome W is unique to females. Therefore, a single fragment size is amplified in males (ZZ) and two fragments of different sizes are amplified in females (ZW). PCR was carried out in a final volume of 20 µl with 1x of Platinum Multiplex Master Mix (Invitrogen Life Technologies, Carlsbad, CA, USA), 0.5 µl of each primer and 50 ng of genomic DNA template. We incorporated a fluorescent dye (FAM) on the P2 primer to analyse fragment sizes using capillary electrophoresis. The PCR profile consisted of an initial denaturing step at 94°C for 10 min, followed by 35 cycles of 94°C for 30 s, 47°C for 1 min and 72°C for 1 min, and then a final extension of 72°C for 10 min. Positive and negative controls were included in each PCR. Positive controls consisted of samples of Ara chloroptera of known sex. PCR amplification was confirmed by electrophoresis on 1% agarose gel, and the products were sent to the Institut Pasteur de Montevideo, Uruguay, for fragment analysis. Genotype assignment was carried out using *GeneMarker* 2.4.0 (Softgenetics LLC, State College, PA, USA). Males were homozygotes (370/370 pb) and females were heterozygotes (370/388 pb).

The collected nails were washed with distilled water to remove residues and contaminants and subsequently dried in an oven at 60°C for 24 h. Silva et al. (2015) estimated the nail growth rate of the Patagonian penguin (Spheniscus magellanicus) at 3.4 mm per month (assuming linear and constant growth). Since this is the closest phylogenetically available information, we used this growth rate to estimate the period that each portion of each nail integrated. The nail of each individual was measured and cut from the basal end in sections of \sim 3.5 mm (each portion integrated the trophic habits of \sim 1 month), allowing us to obtain five or six portions per individual. Each portion was washed and oven-dried again. Finally, each nail portion was cut into smaller fragments that were used for the SIA of $\delta^{15}N$ and $\delta^{13}C$. Samples were sent to the Center for Stable Isotopes, University of New Mexico (http://csi.unm.edu/) for analysis.

Each sample was analysed in a continuous-flow mass spectrometer to determine the abundance of N and C isotopes with analytical precisions of $\pm 0.2\%$ for δ^{15} N and $\pm 0.04\%$ for δ^{13} C. Isotopic values are expressed in delta notation (d) in parts per thousand (‰) according to the equation: $dX = [(R \text{ sample}/R \text{ standard}) - 1] \times 1000$, where X corresponds to ¹³C or ¹⁵N, R sample is the ratio between the heavy and light isotope of the sample (¹⁵N/¹⁴N or ¹³C/¹²C) and R standard is the ratio between the heavy and light isotope of the reference standards. Standard values for ¹³C are determined with Pee Dee Belemnite (PDB), and for ¹⁵N atmospheric nitrogen (air) is used.

Given that five or six portions were obtained from each nail analysed and that their isotopic signatures ranged from August/September 2018 to January 2019, the information on the trophic habits of each individual reflected the end of the non-reproductive period and most of the reproductive period (Hinke *et al.* 2012).

Data analysis

Differences between sexes in the mean value of each isotope were evaluated using repeated-measures analysis of variance (ANOVA). The values of each isotope were modelled, including sex as a fixed effect and the identity of the individual (intercepts) and the portion of the nail (slopes) as random effects, using the *nlme* package (Pinheiro *et al.* 2021) of *R* (R Core Team 2020).

We estimated Layman metrics that allow for quantifying various aspects of the trophic habits of a population based on the distribution of the isotopic

Table I. Values of Layman metrics at the population level and for each sex. Isotopic values were obtained from the nails of 17 adult individuals of the population of *Pygoscelis papua* (8 females, 8 males, 1 could not be determined) from Ardley Island, South Shetland Islands.

Metrics	Population	Females	Males	
δ ¹⁵ N range	2.62	2.62	1.86	
δ ¹³ C range	2.10	1.80	1.55	
NND	0.22	0.29	0.28	
SDNND	0.12	0.21	0.09	

NND = mean nearest neighbour distance; SDNND = standard deviation of the nearest neighbour distance.



Fig. 1. Isotopic niche areas of males and females from the population of *Pygoscelis papua* from Ardley Island, South Shetland Islands. Standard ellipse areas were generated from the isotopic values (δ^{15} N and δ^{13} C) of nails from eight adult females (circles, red) and eight adult males (triangles, blue) using three percentages of the data (Stable Isotope Bayesian Ellipses in R): 25% (solid lines), 40% (dashed lines) and 99% (dotted lines).

signatures of the individuals in the isotopic space (biplot δ^{15} N- δ^{13} C; see Layman *et al.* 2007). The δ^{15} N range (NR) and the δ^{13} C range (CR) are the distances between

the highest and the lowest values of $\delta^{15}N$ and $\delta^{13}C$, respectively. Whilst the NR reflects the diversity of trophic levels exploited/used, the CR reflects the diversity of basal resources. The mean nearest neighbour distance (NND) is the average of the Euclidean distances of each individual to its nearest neighbour (individual) in the biplot δ^{15} N- δ^{13} C; the lower the values of NND, the greater the degree of trophic packing of the individuals. The standard deviation of the nearest neighbour distance (SDNND) indicates how uniform the distribution of the trophic niches of individuals is, being less influenced by sample size than NND. Whilst NR and CR are population metrics, NND and SDNND provide information about the relative position of the individuals in the population with respect to each other within the isotopic space (Layman et al. 2007). We calculated the Layman metrics at the population level and also separately for each sex.

Stable Isotope Bayesian Ellipses in R (SIBER; Jackson et al. 2011) were used to define the isotopic niche space at the population, sex and individual levels. This method is a Bayesian version of Layman metrics (Layman et al. 2007) that can incorporate uncertainties such as sampling biases and small sample sizes into niche metrics (Jackson et al. 2011). Based on Markov chain Monte Carlo (MCMC) simulation, this approach assigns measures of uncertainty to calculate parameters of ellipses in a way that is similar to a bootstrap procedure. Standard ellipse areas, corrected for small sample sizes (SEA_C), were used to estimate the width of the isotopic niche using Bayesian standard ellipse areas (SEA_B) . We calculated the SEA_B using three percentages of the data (25%, 40% and 99%) and 1000 replicates, from which we obtained the mean value and the 95% confidence interval (CI). The SEA_B calculated with: 1) 99% of the data represents the total area of the isotopic niche used, 2) 40% of the data represents the core of the isotopic niche and 3) 25% of the data represents the area of the isotopic niche that is most frequently used by the population, by individuals of the same sex or by each individual. We estimated the degree of SEA_C overlap between sexes. In addition, we estimated the mean and standard deviation (SD) of the SEA_C for all individuals and for the individuals of each sex using two percentages

Table II. Bayesian standard ellipse areas (SEA_B; number of replicates = 1000) at the population level and for each sex. Isotopic values were obtained from the nails of 17 adult individuals of the population of *Pygoscelis papua* (8 females, 8 males, 1 could not be determined), from Ardley Island, South Shetland Islands. SEA_B was calculated using three percentages of the data: 25%, 40% and 99% (Stable Isotope Bayesian Ellipses in R). The means are given, and 95% confidence intervals are shown in parentheses.

		Group	
	Population	Females	Males
SEA _B - 25%	0.216 (0.213–0.219)	0.312 (0.304–0.319)	0.174 (0.170-0.179)
SEA _B - 40%	0.385 (0.379-0.391)	0.553 (0.540-0.567)	0.309 (0.302-0.317)
SEA _B - 99%	3.475 (3.420–3.529)	4.987 (4.865–5.109)	2.790 (2.723–2.857)

Table III. Percentage of the corrected standard ellipse area (SEA_C) that each sex overlaps with the other. Isotopic values of δ^{15} N and δ^{13} C were obtained from the nails of eight adult females and eight adult males of the population of *Pygoscelis papua* from Ardley Island, South Shetland Islands. The area of overlap is shown using three percentages of the data: 25%, 40% and 99% (Stable Isotope Bayesian Ellipses in R).

	% of SEA _C overlapped with the other sex for:		
	Females	Males	
SEA _C - 25%	33.26	61.39	
$SEA_{C} - 40\%$	40.09	74.01	
$SEA_C - 99\%$	52.19	96.33	

of the data: 25% and 99% (Jackson *et al.* 2011). In all of the cases, we used the mean value of the five or six nail portions of each individual to avoid redundancy of information associated with each individual.

We estimated the degree of ITS at the population level and for each sex using both a multidimensional approach (which incorporates the information of both isotopes, each constituting a niche axis; Ingram *et al.* 2018) and a one-dimensional approach (obtaining the degree of ITS for each isotope separately; Bolnick *et al.* 2007). For the multidimensional approach, we used a multiple-response generalized linear mixed model (MGLMM) to estimate intra- and inter-individual variance components, modelling the identity of the individual as a random effect (Ingram *et al.* 2018). We used the *MCMCglmm* package (Hadfield 2010), which employs a Bayesian MCMC approach, to estimate the variance components (WIC and BIC), TNW (as BIC + WIC) and ITS (as BIC/TNW) with a 95% CI. For the one-dimensional analysis, we used the *RInSp* package (Zaccarelli *et al.* 2013) to estimate the variance components (WIC and BIC), TNW and ITS, for both δ^{13} C and δ^{15} N, and also to evaluate the statistical significance of the ITS index against the null hypothesis that the population is composed of generalist individuals (see Zaccarelli *et al.* 2013). Finally, as an ITS proxy, we estimated the proportion of the population isotopic niche (SEA_C) represented by each single individual and by the individuals of each sex altogether, using 99% of the data. All of these analyses were performed in the free software *R* (R Core Team 2020), and the statistical significance was established at the 0.05 level.

Results

From 19 penguin samples, 17 were determined as *P. papua*, one corresponded to *P. adeliae* and other to *P. antarctica*. In our sample of 17 gentoo penguins, we identified eight females and eight males. Only in one case was it not possible to determine the sex, which was probably related to the low DNA quantity for amplifying nuclear markers.

The average length of the 17 nails analysed was 19.62 mm (SD = 1.55), allowing us to obtain five and six nail portions from 12 and five individuals, respectively. The average δ^{15} N and δ^{13} C values at the population level were 7.30‰ (0.52) and -24.24‰ (0.37), respectively. We did not find differences between sexes in either the δ^{15} N values (females = 7.21 ± 0.57‰, males = 7.42 ± 0.49‰, P = 0.26) or in the δ^{13} C values (females = -24.32 ± 0.39‰, males = -24.20 ± 0.36‰, P = 0.39).



Fig. 2. Isotopic niche areas of individuals of *Pygoscelis papua* from the population of Ardley Island, South Shetland Islands. Individual standard ellipse areas were generated from the isotopic values (δ^{15} N and δ^{13} C) of nails from 17 adult individuals using **a**. 99% and **b**. 25% of the data.

Table IV. Degree of multidimensional individual trophic specialization (ITS) for the population of *Pygoscelis papua* from Ardley Island, South Shetland Islands. Values of the within-individual variance component (WIC), between-individual variance component (BIC), total niche width (TNW) and multidimensional individual trophic specialization value (ITS = BIC/TNW) for the isotopic values (δ^{15} N and δ^{13} C) of the population of *P. papua* (*n* = 17 adults) from Ardley Island and of each sex (8 females, 8 males, 1 could not be determined) are calculated using 95% confidence intervals (*MCMCglmm* package in *R*). Isotopic values are shown.

	Population	Females	Males
WIC	0.002 (0.001-0.003)	0.002 (0.001-0.003)	0.002 (0.001-0.003)
BIC	0.004 (0.001-0.007)	0.006 (0.001-0.016)	0.005 (0.001-0.010)
TNW	0.006 (0.003-0.010)	0.008 (0.002-0.019)	0.007 (0.002-0.013)
ITS	0.645 (0.480-0.717)	0.732 (0.408–0.821)	0.711 (0.404–0.800)

Regarding isotopic niche metrics, both at the population level and for each sex, the range of δ^{15} N (NR) was greater than the range of δ^{13} C (CR). Females showed higher values than males for all Layman metrics. For example, the value of SDNND (uniformity in the distribution of the trophic niches of the individuals) was 0.21 for females and 0.09 for males (Table I).

Similarly, the mean SEA_B values (using 25%, 40% and 99% of the data) of females were always higher than those of males (Fig. 1 & Table II). In addition, the degree of overlap between the ellipses (SEA_C) of females and males showed that females have a greater exclusive area in all of the cases (Table III): whilst the exclusive area for males did not exceed 40%, the exclusive area for females exceeded 65%.

The individual mean (SD) of the SEA_C using 99% of the data was 1.87 (1.37), and there was high overlap between them (Fig. 2a). However, when we used 25% of the data, the ellipse area reduced to 0.12 (0.09), and some individuals overlapped little or not at all with others

Table V. Degree of individual trophic specialization (ITS) in δ^{15} N and δ^{13} C for the population of *Pygoscelis papua* from Ardley Island, South Shetland Islands. Values of the within-individual variance component (WIC), between-individual variance component (BIC), total niche width (TNW) and one-dimensional ITS value (ITS = BIC/TNW) are shown for the isotopic values (δ^{15} N and δ^{13} C) of 17 adult individuals at the population level and for each sex (8 females, 8 males, 1 could not be determined). Isotopic values were obtained from the nails. Statistical significance is shown regarding the ITS index against the null hypothesis that the population is made up of generalist individuals (*RInSp* package in *R*): ****P* ≤ 0.001.

	$\delta^{15}N$		$\delta^{13}C$			
	Population	Females	Males	Population	Females	Males
WIC	0.09	0.11	0.08	0.07	0.06	0.08
BIC	0.18	0.22	0.14	0.06	0.09	0.03
TNW	0.27	0.32	0.22	0.13	0.15	0.11
ITS	0.67***	0.67***	0.64***	0.49***	0.57***	0.31

(Fig. 2b). The mean SEA_C values of male individuals were higher than those observed for female individuals, both for 99% (males = 2.44 ± 1.79 , females = 1.36 ± 0.61) and for 25% of the data (males = 0.15 ± 0.11 , females = 0.08 ± 0.04).

The multidimensional ITS (95% CI) was 0.65 (0.48–0.72) at the population level, 0.73 (0.41–0.82) for females and 0.71 (0.40–0.80) for males (Table IV). The one-dimensional ITS at the population level was higher for δ^{15} N (0.67) than for δ^{13} C (0.49; Table V). At the sex level, the one-dimensional ITS for δ^{15} N was similar between sexes (females = 0.67, males = 0.64). However, for δ^{13} C, the ITS of females was higher (0.57) than that of males (0.31). In all of the cases, the ITS index was significant (*P* < 0.001) with respect to the null model, except for the δ^{13} C index for males (*P* = 0.079; Table V).

The mean (SD) percentage of the population niche used by each individual (SEA_C - 99%) was 35.32% (19.49). Female individuals used 26.68% (8.69) of the total population niche and male individuals used 44.00%(24.92).

Discussion

In the present study, we estimated using a non-invasive methodology a moderate ITS for the gentoo penguin in Ardley Island, South Shetland Islands, and we also found some sexual differentiation. The degree of ITS found at the population level suggests that it could function as a mechanism to reduce potential intraspecific competition arising due to the sustained increase in population abundance (see below). As for the differences between sexes, the greater energy investment that females make at the beginning of the reproductive period could lead them to further minimize the number of conspecifics with which they must compete.

Isotopic niche metrics

The greater range reported for δ^{15} N in relation to δ^{13} C suggests that this population of *P. papua* shows a greater feeding diversity associated with the trophic position of the prey consumed rather than with the habitat used. In this sense, it should be noted that although differences in δ^{15} N could be related to differences in feeding zones, its variation in the West Antarctic Peninsula (i.e. the location of this study) occurs at a larger scale than that used by the *P. papua* individuals of this colony (Kokubun *et al.* 2010, McMahon *et al.* 2013, Brault *et al.* 2018). The values of NND and SDNND indicate that the individuals of the population are unevenly distributed in the isotopic space, so that groups of some individuals are closer to each other. The variation in the size of the individual isotopic niche as well as its

distribution in the isotopic space support the notion that this population includes both specialist and generalist individuals.

In addition, both sexes showed the same pattern as the overall population concerning the range of $\delta^{15}N$ and $\delta^{13}C$. Therefore, females and males show greater diversification regarding the trophic level of the prey consumed than regarding trophic habitat. A greater range of $\delta^{15}N$ and δ^{13} C and a greater area of the ellipse (percentage of data: 25%, 40% and 99%) in females suggest that they use a greater area of the isotopic space and therefore have greater diversity in their foraging habits than males. A higher NND, SDNND and exclusive isotopic niche area of females compared to males indicate that they have a lower degree of trophic redundancy (i.e. the relative position of females to each other within the δ^{15} N- δ^{13} C biplot differs more than that of males; Lavman et al. 2007) and that their distribution in the isotopic space is considerably less uniform. Similarly, this suggests that females use trophic resources and foraging areas that males do not. Since female individuals have on average smaller isotopic niche areas than male individuals, their trophic habits would be more limited to certain resources or foraging areas. The aforementioned result could be associated with the investment that this sex makes during the beginning of the reproductive period. Unlike males, females of *P. papua* fast for ~ 1 week during courtship, and a decrease in mass during the days before the first egg lay has been reported on King George Island (Trivelpiece & Trivelpiece 1990). In addition, females carry out the first incubation period (Black 2016). Each female individual could modify her foraging habits with respect to the other individuals in the population (of both sexes) and reduce their overlap, thus mitigating potential intraspecific competition for access to resources. Similarly, a greater isotopic niche area of females compared to males suggests that they are the sex that contributes to a greater extent to the diversification of the population isotopic niche.

Individual trophic specialization

At the population level, the estimation of ITS via the multidimensional and one-dimensional approaches indicated that the population of *P. papua* from Ardley Island presents a moderate to high degree of specialization, being composed mainly of specialist individuals. The one-dimensional analysis indicated that this population shows a higher degree of ITS associated more with the trophic position of the consumed resources (δ^{15} N) than with the foraging habitat (benthic/pelagic, δ^{13} C). Although the population degree of specialization in δ^{15} N is not affected by the foraging habits of individuals of a particular sex, at the level

of δ^{13} C the degree of population ITS is influenced more by the habits of females (see below). Studies on Livingston Island (near Ardley Island) reported a higher degree of ITS in $\delta^{15}N$ (0.53) than in $\delta^{13}C$ (0.44) during winter (e.g. Herman 2016). In this same population, greater inter-individual variation in $\delta^{15}N$ than in $\delta^{13}C$ was reported during the reproductive period (Polito et al. 2015). The greater specialization at $\delta^{15}N$, as found in this study and in Herman (2016), could be related to the fact that P. papua generally forages near the coast/ benthic zones (Kokubun et al. 2010, Polito et al. 2011), so the degree of differentiation in the isotopic values that individuals can achieve in the foraging habitat is less than the trophic segregation that can be achieved regarding the consumed prev. A degree of ITS close to 0.70 in $\delta^{15}N$ indicates that there is significant differentiation between individuals in the consumption of secondary resources, such as fish and squid. These results are supported by the ellipses analysis, given that on average each individual uses less than half of the total population niche, indicating that they are specialized in certain resources and/or foraging areas.

At the sex level, the multidimensional estimation of ITS indicated a moderate to high degree of specialization for both females and males. The degree of ITS of females was higher with respect to the trophic position of the consumed resources ($\delta^{15}N$) than to the foraging habitat $(\delta^{13}C)$. Males were specialized in the trophic position of the prey that they consume but without any degree of diversification in foraging habitat. The differences in the degree of specialization between females and males are consistent with the fact that female individuals use, on average, a lower percentage of the population niche compared to male individuals and show less variation between them. The fact that a moderate to high degree of multidimensional ITS was found in males whilst in the one-dimensional analysis this was reported for $\delta^{15}N$, but not for δ^{13} C, may be due to the fact that the multidimensional analysis considers how the paired values of $\delta^{15}N$ and $\delta^{13}C$ of each individual are distributed in the isotopic space.

Given the sustained population increase of *P. papua* in Ardley Island (Roberts *et al.* 2017, Firla *et al.* 2019), the degree of ITS found in this study could constitute one of the mechanisms adopted by individuals to deal with intraspecific competition for trophic resources. In recent decades, gentoo penguins have increased their trophic level by consuming more fish and squid (McMahon *et al.* 2019). Although the percentage of consumption of these items can be highly variable amongst populations (e.g. Polito *et al.* 2011, Pickett *et al.* 2018), the dietary diversification found in this study may occur through differentiation in the proportion of krill *vs* highertrophic-level species in the diet. The increase in fish consumption could be favoured by both the recovery of marine mammal predators that also consume krill (McMahon *et al.* 2019) and the recovery of some fish species in King George Island (see Marschoff *et al.* 2012). Thus, the degree of specialization reported in this study could be due to multiple, non-mutually exclusive factors.

Furthermore, unlike other studies, we did not find differences between sexes in the trophic level of the prey consumed (Karnovsky 1997, Gorman 2015), but we did find a moderate to high degree of ITS in δ^{15} N for both sexes. This could also be explained by fluctuations in prey availability and interspecific competition. On the other hand, the differentiation between sexes regarding the degree of ITS in the foraging habitat suggests that the cost associated with the reproductive period could influence females to differentiate themselves even more than males from the rest of the individuals.

To our knowledge, this work is the first to estimate the degree of ITS of *P. papua* on Ardley Island at the population level, and it is also the first to assess ITS separately for each sex. We also highlight the non-invasive methodology used, which allows for the study of trophic habits in wild populations without manipulating individuals. Future research could incorporate the use of isotopic mixing models to address the specific contribution of each prey species to the diet of individuals. In this way, we could gain even more insights into the ways by which this population is dealing with intraspecific competition.

Acknowledgements

We thank the Ministry of Education and Culture of Uruguay (N° I/FVF/2017/076) for financing the stable isotope analysis and the Uruguayan Antarctic Institute for financing the trip and the logistical support in Antarctica. We thank David Ainley and an anonymous reviewer for their valuable comments on the manuscript. The present article is a contribution to Lucía Rabinovich-Larrechea's. graduate thesis, tutored by Valentina Franco-Trecu.

Author contributions

LR-L and VF-T conceived and designed the study; DEN and VF-T conducted the fieldwork; LR-L, MC, NB and VF-T conducted the laboratory work; LR-L and VF-T analysed the data; LR-L and VF-T wrote the manuscript; VF-T obtained the funding. All authors contributed to editing the manuscript and approved the final version.

Financial support

This work was supported by the Vaz Ferreira Funds -Ministry of Education and Culture, Uruguay (grant number I/FVF/2017/076).

Competing interests

The authors declare none.

Data availability

To access the database that supports the findings of this study, please contact the corresponding author, LR-L.

References

- AINLEY, D.G., BALLARD, G., BARTON, K.J., KARL, B.J., RAU, G.H., RIBIC, C.A. & WILSON, P.R. 2003. Spatial and temporal variation of diet within a presumed metapopulation of Adélie penguins. *The Condor*, **105**, 10.1093/condor/105.1.95.
- ARAÚJO, M.S., BOLNICK, D.I. & LAYMAN, C.A. 2011. The ecological causes of individual specialisation. *Ecology Letters*, 14, 10.1111/ j.1461-0248.2011.01662.x.
- ASPA. 2009. No. 150 Ardley Island, King George Island: Management Plan. Retrieved from https://www.env.go.jp/nature/nankyoku/ kankyohogo/database/jyouyaku/aspa/aspa_pdf_en/150.pdf
- BLACK, C.E. 2016. A comprehensive review of the phenology of *Pygoscelis* penguins. *Polar Biology*, **39**, 10.1007/s00300-015-1807-8.
- BOLNICK, D.I., SVANBÄCK, R., ARAÚJO, M.S. & PERSSON, L. 2007. Comparative support for the niche variation hypothesis that more generalized populations also are more heterogeneous. *Proceedings of the National Academy of Sciences of the United States of America*, 104, 10.1073/pnas.0703743104.
- BOLNICK, D.I., YANG, L.H., FORDYCE, J.A., DAVIS, J.M. & SVANBÄCK, R. 2002. Measuring individual-level resource specialization. *Ecology*, 83, 10.1890/0012-9658(2002)083[2936:MILRS]2.0.CO;2.
- BOLNICK, D.I., SVANBÄCK, R., FORDYCE, J.A., YANG, L.H., DAVIS, J.M., HULSEY, C.D. & FORISTER, M.L. 2003. The ecology of individuals: incidence and implications of individual specialization. *American Naturalist*, **161**, 10.1086/343878.
- BRAULT, E.K., KOCH, P.L., MCMAHON, K.W., BROACH, K.H., ROSENFIELD, A.P., SAUTHOFF, W., et al. 2018. Carbon and nitrogen zooplankton isoscapes in West Antarctica reflect oceanographic transitions. *Marine Ecology Progress Series*, 593, 10.3354/meps12524.
- CHEREL, Y., HOBSON, K.A., GUINET, C. & VANPE, C. 2007. Stable isotopes document seasonal changes in trophic niches and winter foraging individual specialization in diving predators from the Southern Ocean. *Journal of Animal Ecology*, **76**, 10.1111/j.1365-2656.2007.01238.x.
- CLUCAS, G. V., DUNN, M.J., DYKE, G., EMSLIE, S.D., NAVEEN, R., POLITO, M.J., et al. 2014. A reversal of fortunes: climate change 'winners' and 'losers' in Antarctic Peninsula penguins. Scientific Reports, 4, 10.1038/srep05024.
- Costa-PEREIRA, R. & ARAÚJO, M.S. 2022. Individual specialization. Encyclopedia of Biodiversity, 6, 10.1016/b978-0-12-822562-2.00068-2.
- COSTA-PEREIRA, R., RUDOLF, V.H.W., SOUZA, F.L. & ARAÚJO, M.S. 2018. Drivers of individual niche variation in coexisting species. *Journal of Animal Ecology*, 87, 10.1111/1365-2656.12879.
- DALL, S.R.X., BELL, A.M., BOLNICK, D.I. & RATNIEKS, F.L.W. 2012. An evolutionary ecology of individual differences. *Ecology Letters*, 15, 10.1111/j.1461-0248.2012.01846.x.
- DE LIMA, R.C., FRANCO-TRECU, V., CARRASCO, T.S., INCHAUSTI, P., SECCHI, E.R. & BOTTA, S. 2022. Segregation of diets by sex and individual in South American fur seals. *Aquatic Ecology*, 56, 10.1007/s10452-021-09915-9.
- FIRLA, M., MUSTAFA, O., PFEIFER, C., SENF, M. & HESE, S. 2019. Intraseasonal variability of guano stains in a remotely sensed penguin colony using UAV and satellite. *ISPRS Annals of the Photogrammetry, Remote Sensing and Spatial Information Sciences*, 4, 10.5194/isprs-annals-IV-2-W5-111-2019.

- FRANCE, R.L. 1995. Differentiation between littoral and pelagic food webs in lakes using stable carbon isotopes. *Limnology and Oceanography*, 40, 10.4319/lo.1995.40.7.1310.
- GONZÁLEZ, S., COSSE, M., DEL ROSARIO FRANCO, M., EMMONS, L., VYNNE, C., DUARTE, J.M.B., et al. 2015. Population structure of mtDNA variation due to Pleistocene fluctuations in the South American maned wolf (*Chrysocyon brachyurus*, Illiger, 1815): management units for conservation. *Journal of Heredity*, **106**, 10.1093/jhered/esv043.
- GORMAN, K.B. 2015. Integrative studies of Southern Ocean food-webs and Pygoscelis penguin demography: mechanisms of population response to environmental change. Doctor of Philosophy thesis. Burnaby, BC: Simon Fraser University, 311 pp.
- GRIFFITHS, R., DOUBLE, M.C., ORR, K. & DAWSON, R.J.G. 1998. A DNA test to sex most birds. *Molecular Ecology*, 7, 10.1046/ j.1365-294x.1998.00389.x.
- HADFIELD, J.D. 2010. MCMC methods for multi-response generalized linear mixed models: the MCMCglmm R package. Journal of Statistical Software, 33, 10.18637/jss.v033.i02.
- HANDLEY, J.M., CONNAN, M., BAYLIS, A.M.M., BRICKLE, P. & PISTORIUS, P. 2017. Jack of all prey, master of some: influence of habitat on the feeding ecology of a diving marine predator. *Marine Biology*, 164, 10.1007/s00227-017-3113-1.
- HERMAN, R.W. 2016. Investigating species and population level foraging variation and individual specialization in Pygoscelis penguins using stable isotope analysis. Thesis. Baton Rouge, LA: Louisiana State University, 73 pp.
- HINKE, J.T., POLITO, M.J., REISS, C.S., TRIVELPIECE, S.G. & TRIVELPIECE, W.Z. 2012. Flexible reproductive timing can buffer reproductive success of *Pygoscelis* spp. penguins in the Antarctic Peninsula region. *Marine Ecology Progress Series*, **454**, 10.3354/meps09633.
- INGRAM, T., COSTA-PEREIRA, R. & ARAÚJO, M.S. 2018. The dimensionality of individual niche variation. *Ecology*, 99, 10.1002/ ecy.2129.
- JACKSON, A.L., INGER, R., PARNELL, A.C. & BEARHOP, S. 2011. Comparing isotopic niche widths among and within communities: SIBER - Stable Isotope Bayesian Ellipses in R. *Journal of Animal Ecology*, 80, 10.1111/j.1365-2656.2011.01806.x.
- JAEGER, A., BLANCHARD, P., RICHARD, P. & CHEREL, Y. 2009. Using carbon and nitrogen isotopic values of body feathers to infer interand intra-individual variations of seabird feeding ecology during moult. *Marine Biology*, **156**, 10.1007/s00227-009-1165-6.
- KARNOVSKY, N.J. 1997. *The fish component of Pygoscelis penguin diets.* Master of Science in Biological Sciences thesis. Bozeman, MT: Montana State University, 76 pp.
- KOKUBUN, N., TAKAHASHI, A., MORI, Y., WATANABE, S. & SHIN, H.C. 2010. Comparison of diving behavior and foraging habitat use between chinstrap and gentoo penguins breeding in the South Shetland Islands, Antarctica. *Marine Biology*, **157**, 10.1007/s00227-009-1364-1.
- LAYMAN, C.A., ARRINGTON, D.A., MONTAÑA, C.G. & POST, D.M. 2007. Can stable isotope ratios provide for community-wide measures of trophic structure? *Ecology*, 88, 10.1890/08-0167.1.
- LESCROËL, A., DUGGER, K.M., BALLARD, G. & AINLEY, D.G. 2009. Effects of individual quality, reproductive success and environmental variability on survival of a long-lived seabird. *Journal of Animal Ecology*, 78, 10.1111/j.1365-2656.2009.01542.x.
- LESCROËL, A., BALLARD, G., GRÉMILLET, D., AUTHIER, M. & AINLEY, D.G. 2014. Antarctic climate change: extreme events disrupt plastic phenotypic response in Adélie penguins. *PLoS ONE*, **9**, 10.1371/ journal.pone.0085291.
- LESCROËL, A., LYVER, P.O., JONGSOMJIT, D., VELOZ, S., DUGGER, K.M., KAPPES, P., et al. 2020. Inter-individual differences in the foraging behavior of breeding Adélie penguins are driven by individual quality and sex. *Marine Ecology Progress Series*, 636, 10.3354/ meps13208.

- MARSCHOFF, E.R., BARRERA-ORO, E.R., ALESCIO, N.S. & AINLEY, D.G. 2012. Slow recovery of previously depleted demersal fish at the South Shetland Islands, 1983–2010. *Fisheries Research*, **125–126**, 10.1016/ j.fishres.2012.02.017.
- MASSARO, M., AINLEY, D.G., SANTORA, J.A., QUILLFELDT, P., LESCROËL, A., WHITEHEAD, A., et al. 2020. Diet segregation in Adélie penguins: some individuals attempt to overcome colony-induced and annual foraging challenges. *Marine Ecology Progress Series*, 645, 10.3354/ meps13370.
- MCMAHON, K.W., HAMADY, L.L. & THORROLD, S.R. 2013. A review of ecogeochemistry approaches to estimating movements of marine animals. *Limnology and Oceanography*, 58, 10.4319/ lo.2013.58.2.0697.
- MCMAHON, K.W., MICHELSON, C.I., HART, T., MCCARTHY, M.D., PATTERSON, W.P. & POLITO, M.J. 2019. Divergent trophic responses of sympatric penguin species to historic anthropogenic exploitation and recent climate change. *Proceedings of the National Academy of Sciences of the United States of America*, **116**, 10.1073/ pnas.1913093116.
- NAYA, D.E. & FRANCO-TRECU, V. 2019. An unbiased method to estimate individual specialisation from multi-tissue isotopic data. *Freshwater Biology*, 64, 10.1111/fwb.13316.
- NEGRETE, P., SALLABERRY, M., BARCELÓ, G., MALDONADO, K., PERONA, F., MCGILL, R.A.R., *et al.* 2016. Temporal variation in isotopic composition of *Pygoscelis* penguins at Ardley Island, Antarctic: are foraging habits impacted by environmental change? *Polar Biology*, 40, 10.1007/s00300-016-2017-8.
- NEWSOME, S.D., TINKER, M.T., MONSON, D.H., OFTEDAL, O.T., RALLS, K., STAEDLER, M.M., *et al.* 2009. Using stable isotopes to investigate individual diet specialization in California sea otters (*Enhydra lutris nereis*). *Ecology*, **90**, 10.1890/07-1812.1.
- PATEL, S., WAUGH, J., MILLAR, C.D. & LAMBERT, D.M. 2010. Conserved primers for DNA barcoding historical and modern samples from New Zealand and Antarctic birds. *Molecular Ecology Resources*, 10, 10.1111/j.1755-0998.2009.02793.x.
- PICKETT, E.P., FRASER, W.R., PATTERSON-FRASER, D.L., CIMINO, M.A., TORRES, L.G. & FRIEDLAENDER, A.S. 2018. Spatial niche partitioning may promote coexistence of *Pygoscelis* penguins as climate-induced sympatry occurs. *Ecology and Evolution*, 8, 10.1002/ ece3.4445.
- PINHEIRO, J., BATES, D., DEBROY, S., SARKAR, D. & R CORE TEAM. 2021. *nlme*: linear and nonlinear mixed effects models. *R* package version 3.1-153 Retrieved from https://cran.r-project.org/package=nlme
- POLITO, M.J., LYNCH, H.J., NAVEEN, R. & EMSLIE, S.D. 2011. Stable isotopes reveal regional heterogeneity in the pre-breeding distribution and diets of sympatrically breeding *Pygoscelis* spp. penguins. *Marine Ecology Progress Series*, **421**, 10.3354/meps08863.
- POLITO, M.J., TRIVELPIECE, W.Z., PATTERSON, W.P., KARNOVSKY, N.J., REISS, C.S. & EMSLIE, S.D. 2015. Contrasting specialist and generalist patterns facilitate foraging niche partitioning in sympatric populations of *Pygoscelis* penguins. *Marine Ecology Progress Series*, 519, 10.3354/meps11095.
- Post, D.M. 2002. Using stable isotopes to estimate trophic position: models, methods, and assumptions. *Ecology*, 83, 10.1890/0012-9658 (2002)083[0703:USITET]2.0.CO;2.
- R CORE TEAM. 2020. R: a language and environment for statistical computing Retrieved from https://www.r-project.org/
- ROBERTS, S.J., MONIEN, P., FOSTER, L.C., LOFTFIELD, J., HOCKING, E.P., SCHNETGER, B., et al. 2017. Past penguin colony responses to explosive volcanism on the Antarctic Peninsula. *Nature Communications*, 8, 10.1038/ncomms14914.
- ROUGHGARDEN, J. 1972. Evolution of niche width. *The American Naturalist*, **106**, 10.1086/282892.
- ROUGHGARDEN, J. 1974. Niche width: biogeographic patterns among Anolis lizard populations. *The American Naturalist*, **108**, 429–442.

- SILVA, L.A., SILES, L., CARDONA, L., TAVARES, M., CRESPO, E. & GANDINI, P. 2015. Diferencias estacionales en la dieta de individuos juveniles del Pingüino Patagónico (*Spheniscus magellanicus*) reveladas en base al análisis de isótopos estables en uñas. [Seasonal differences in the diet of juvenile Patagonian penguins (*Spheniscus magellanicus*) revealed by stable isotope analysis of nails]. *Hornero*, **30**, 45–54.
- TRIVELPIECE, W.Z. & TRIVELPIECE, S.G. 1990. Courtship period of Adélie, gentoo, and chinstrap penguins. *In DAVIS*, L.S. & DARBY, J.T., *eds*, *Penguin biology*. Cambridge, MA: Academic Press, 113–127.
- VALENZUELA-GUERRA, P., MORALES-MORAGA, D., GONZÁLEZ-ACUÑA, D. & VIANNA, J.A. 2013. Geographic morphological variation of gentoo penguin (*Pygoscelis papua*) and sex identification: using morphometric characters and molecular markers. *Polar Biology*, 36, 10.1007/s00300-013-1389-2.
- VANDER ZANDEN, H.B., BJORNDAL, K.A., REICH, K.J. & BOLTEN, A.B. 2010. Individual specialists in a generalist population: results from a long-term stable isotope series. *Biology Letters*, 6, 10.1098/ rsbl.2010.0124.
- VASIL, C.A., POLITO, M.J., PATTERSON, W.P. & EMSLIE, S.D. 2012. Wanted: dead or alive? Isotopic analysis (δ¹³C and δ¹⁵N) of *Pygoscelis* penguin chick tissues supports opportunistic sampling. *Rapid Communications in Mass Spectrometry*, **26**, 10.1002/rcm.5340.
- WERLE, E., SCHNEIDER, C., RENNER, M., VOLKER, M. & FIEHN, W. 1994. Convenient single-step, one tube purification of PCR products for direct sequencing. *Nucleic Acids Research*, 22, 4354–4355.
- ZACCARELLI, N., BOLNICK, D.I. & MANCINELLI, G. 2013. *RInSp:* an *R* package for the analysis of individual specialization in resource use. *Methods in Ecology and Evolution*, **4**, 10.1111/2041-210X.12079.