cambridge.org/jhl

Review Paper

Cite this article: Bhat AH, Tak H, Malik IM, Ganai BA, Zehbi N (2023). Trichostrongylosis: a zoonotic disease of small ruminants. *Journal of Helminthology* **97**, e26, 1–11. https://doi.org/ 10.1017/S0022149X2300007X

Received: 28 October 2022 Revised: 22 January 2023 Accepted: 23 January 2023

Keywords:

Zoonosis; *Trichostrongylus*; pathogenesis; immunology; Trichostrongylosis

Author for correspondence: A.H. Bhat, E-mail: bio.abid@gmail.com Trichostrongylosis: a zoonotic disease of small ruminants

A.H. Bhat¹ (10), H. Tak¹, I.M. Malik¹, B.A. Ganai² and N. Zehbi³

¹Department of Zoology, University of Kashmir, Hazratbal-Srinagar 190006, India; ²Centre of Research for Development, University of Kashmir, Hazratbal-Srinagar 190006, India and ³Department of Animal Sciences, Central University of Kashmir, Ganderbal, Jammu and Kashmir 191131, India

Abstract

In the present world a significant threat to human health is posed by zoonotic diseases. Helminth parasites of ruminants are one of the most common zoonotic organisms on the planet. Among them, trichostrongylid nematodes of ruminants, found worldwide, parasitize humans in different parts of the world with varying rates of incidence, particularly among rural and tribal communities with poor hygiene, pastoral livelihood and poor access to health services. In the Trichostrongyloidea superfamily, Haemonchus contortus, Teladorsagia circumcincta, Marshallagia marshalli, Nematodirus abnormalis and Trichostrongylus spp. are zoonotic in nature. Species of the genus Trichostrongylus are the most prevalent gastrointestinal nematode parasites of ruminants that transmit to humans. This parasite is prevalent in pastoral communities around the world and causes gastrointestinal complications with hypereosinophilia which is typically treated with anthelmintic therapy. The scientific literature from 1938 to 2022 revealed the occasional incidence of trichostrongylosis throughout the world with abdominal complications and hypereosinophilia as the predominant manifestation in humans. The primary means of transmission of Trichostrongylus to humans was found to be close contact with small ruminants and food contaminated by their faeces. Studies revealed that conventional stool examination methods such as formalin-ethyl acetate concentration or Willi's technique combined with polymerase chain reaction-based approaches are important for the accurate diagnosis of human trichostrongylosis. This review further found that interleukin 33, immunoglobulin E, immunoglobulin G1, immunoglobulin G2, immunoglobulin M, histamine, leukotriene C4, 6-keto prostaglandin $F_{1\alpha_2}$ and thromboxane B_2 are vital in the fight against Trichostrongylus infection with mast cells playing a key role. This review focuses on the prevalence, pathogenicity and immunological aspects of Trichostrongylus spp. in humans.

Introduction

Parasites that we encounter in nature may be species-specific or may have a wide range of hosts. The latter are strenuous to control because they can lie dormant in their reservoir hosts for long periods of time before infecting other hosts. Similarly, zoonotic parasites are difficult to control and pose a concern to human health due to their proclivity for residing in diverse hosts (Allen et al., 2017). Livestock helminths, among other zoonotic viruses, bacteria and other infections, are a cause of health concern for humans (Libera et al., 2022). Helminth parasites that infect the livestock, significantly affect their health and reproduction (Rehman & Abidi, 2022). Although *Haemonchus contortus* is considered as the notorious parasite of livestock because of its reproductive potential and blood sucking ability, Trichostrongylus remains one of the most frequent and extremely pathogenic parasites in cattle, and because of its zoonotic potential, *Trichostrongylus* is dangerous to human health (Getachew *et al.*, 2007). Human infection by *Trichostrongylus* is more prevalent in the pastoral communities who raise livestock or eat vegetables that are fecundated with faecal ordure. Humans become infected after consuming food or water contaminated with faeces of the definitive host (ruminant and humans). Gastrointestinal symptoms with disparate morbidity may develop in some patients; however, most patients are asymptomatic but have eosinophilia as the only symptom (Buonfrate et al., 2017). Till date, Trichostrongylus infections in humans have been reported only in a few parts of the world. The reason for such rarity in cases of human trichostrongylosis could most probably be the occurrence of asymptomatic infections and comparably less sensitivity of microscopic detection due to low egg-output (Buonfrate et al., 2017). As a result, there could be many undiagnosed cases of Trichostrongylus infection in the human populations worldwide (Wolfe, 1978). Trichostrongylus is similar to hook worms, particularly Ancylostoma duodenale and Necator americanus concerning its transmission, pathophysiology and morphology at certain life-cycle stages; as a result, very little is known about the population biology and epidemiology of Trichostrongylus spp. (Yong et al., 2007). There is lack of information regarding the

© The Author(s), 2023. Published by Cambridge University Press. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (http://creativecommons.org/licenses/ by/4.0), which permits unrestricted re-use, distribution and reproduction, provided the original article is properly cited.



global status of trichostrongylosis among humans. It is necessary to compile the pertinent scientific material and examine pastoral populations for the incidence of this parasite. Little to no work has been done on the pathogenicity and immune response to this worm in relation to humans. This review aims to compile literature about the: (a) pathogenesis of trichostrongylosis in humans; (b) immune response to *Trichostrongylus* invasion and its mechanism of action; and (c) epidemiology of this zoonotic parasite in relation to humans.

Life cycle and transmission

Using no intermediate host and exhibiting a direct life cycle, Trichostrongylus adults live in the gastrointestinal tract (GIT) at particular micro-niches depending on the species. Females lay eggs in the host's GIT which are then excreted with the faeces of the host. As soon as the eggs are in a favourable environment, they embryonate into the L1-larvae which moults two times and develops into the infective L3-larvae in about five days and may remain viable for about six months (Levine & Anderson, 1973). Throughout the summer months of June to August, most trichostrongylid larvae occur on the grass blades representing greater chances of infection in grazing animals (Crofton, 1948). Cattle become infected after consuming the L3-larvae while feeding on contaminated grasses. Once ingested, L3-larvae of Trichostrongylus spp. reach the predilection site, for example, the L3-larvae stage of Trichostrongylus axei inhabits the abomasum of ruminants where they complete their development and the adult worms then penetrate the lining of the abomasum. In some other species, the L3-larvae reach the small intestine and invade the crypts to complete their development into L4 and L5 larval stages. Depending on the species and the host, the prepatent period is usually three to four weeks (Janquera, 2017) but can extend up to two years (Ralph et al., 2006). Trichostrongylus colubriformis lives in mucoid passages on the surface of duodenal and intestinal villi (Shaw et al., 2003). Humans acquire trichostrongylosis when L3-larvae of Trichostrongylus spp. are ingested orally while consuming contaminated food. Application of night-soil (human excreta) or livestock faecal matter as manure and the resistant nature of the eggs gives rise to the propagation of this parasite in human populations (Sharma & Anand, 1997). Shady areas with high humidity and an abundance of grass are more favourable for the spread of Trichostrongylus (Watson, 1953).

Methodology

Databases such as Google Scholar, PubMed, Scopus, Web of Science and ScienceDirect were searched to collect and review the published scientific research articles related to trichostrongylosis among humans. Terms such as Trichostrongylus, trichostrongylosis/liasis, zoonosis/es/tic, human, transmission, case/s, report, diagnosis, pathogenicity/sis, immunological, immune response, etc. were used in multiple combinations to search for relevant research articles. Further, the bibliographic section of research articles was also searched to extract relevant references. In addition to these, any other relevant research articles and/or case reports from other sources were also reviewed during the study. Research articles with information regarding number of cases in humans, method of diagnosis, symptoms, country name and/or pathogenesis and immune response were included in this study. Research articles related to other hosts were excluded. We used Endnote to compile the articles and then thoroughly read the

papers to extract information such as year, country/region, number of cases, method of examination, mode of transmission, symptoms and any other important findings (table 1). Statistics from these articles are compiled in figs 3–5.

Pathogenicity

Human trichostrongylosis is typically a minor, subclinical condition as indicated by the fact that diagnosed cases have only been identified through screening. Nevertheless, heavier infection may result in emaciation, pain in the abdomen and diarrhoea along with slight anaemia and eosinophilia in adults while causing retardation of development in children (Hollo et al., 1970). Persons with infection intensity of 24-300 eggs per gram (EPG) of faeces are symptomatic while an infection intensity below 24 EPG shows no symptoms (Ghadirian & Arfaa, 1975; Wolfe, 1978). In small ruminants the symptoms of trichostrongylosis are more severe causing 'black scour disease' which is characterized by dark green to black diarrhoea, that covers the crutch, hocks and legs. There is extreme emaciation in heavily infected sheep with wasting of musculature and negligible amount of renal and omental fat (Edgar, 1933). Craig (2009) reported that trichostrongylosis may lead to moderate anaemia as the worms may feed on the host's blood from mucosa of the gut. Histopathological studies carried out by Barker (1975) showed that T. colubriformis causes severe villus atrophy and plasma loss in the sheep gut. Acutely infected mucosa becomes flat, has stunted epithelium, with projecting crypts often leaking eosinophilic material, exhibiting hyperplasia with highly inflammated lamina propria due to cell infiltration. The intestinal epithelial surface has erosions and necrosis which may be most probably caused by operational movement of adult nematodes. In the case of Trichostrongylus vitrinus, exsheathed L3-larvae burrow through intestinal villi and form submucosal tunnels and when adults emerge out of these tunnels, they cause considerable damage to the mucosal layer (Beveridge et al., 1989). Alterations in the intestinal morphology due to Trichostrongylus infection cause a decrease in the activity of enzymes such as alkaline phosphatase and leucine amino peptidase (Shayo & Benz, 1979). Though mildly pathogenic, Trichostrongylus may cause severe complications among young and weak livestock, sometimes proving fatal (fig. 1).

Immune response

The aim of the parasite is to establish itself successfully in/on the host without killing it. To do so, the parasite manipulates the immune response of the host, and either the parasite evades the host immune system by different mechanisms or it alters the immune response of host to make it ineffective. To counter this, the host tries to mount an effective immune response against the parasite to kill and expel it. Usually in helminth infection, T helper 2 or Type 2 response is initiated by the host. It includes expression of interleukin-4, interleukin-5, interleukin-9, interleukin-13, interleukin-21, interleukin-33 (IL-33) and proliferation and activation of plasma cells to secrete immunoglobulin E (IgE), ocytess and mast cells to secrete vasoactive amines. With respect to trichostrongylosis in humans, limited research has been conducted on immunological response. An in vitro study using a co-culture system of T. colubriformis and epithelial cell from humans showed that movement of T. colubriformis at the site of infection creates necrosis of intestinal epithelial cells. The necrosis

in turn induces the release of intracellular contents, including IL33 which is elemental in the commencement of appropriate host response to gastrointestinal nematodes (Andronicos et al., 2012). It has been found that after recurrent infections in natural conditions, sheep can develop immunity against T. colubriformis. In a natural foraging environment, the ability to withstand nematode establishment occurs after seven weeks of incessant infection (Dobson et al., 1990). When a sheep is fed with Trichostrongylus larvae, most of them are expelled in less than 24 h under a hypersensitivity response known as rapid rejection (Miller et al., 1985). The sheep shows resistance to Trichostrongylus by mounting an inflammatory response in gastro-intestinal mucosa which is evident by increase in mucosal mast cells and globule leucocytes (Miller et al., 1985; Douch et al., 1986). This inflammatory response seems to be genetically controlled and mast cells play a major role in resistance against Trichostrongylus (Gill, 1991). When treated with parasite antigen, mast cells from resistant sheep release around 39% of cellular mast cell protease (CMCP) as compared to less than 8% CMCP release by mast cells from sheep with primary infection of Trichostrongylus (Bendixsen et al., 1995). Jones & Emery (1991) demonstrated that sheep immunized with T. colubriformis release a number of inflammatory mediators such as histamine, leukotriene C4, 6-keto prostaglandin $F_{1\alpha}$ and thromboxane B_2 on secondary infection with leukotriene C4 being the most predominant inflammatory mediator in expulsion of nematodes from intestines. In guinea pigs immunized with irradiated T. colubriformis larvae, release of biological amines (histamine and 5-hydroxytryptamine) and enteric plasma was found to be involved in resistance to secondary infections of this parasite (Steel et al., 1990).

Serum IgE levels have been found to escalate following nematode infections (Shaw et al., 1999). Shaw et al. (2003) found T. colubriformis aspartyl inhibitor (Tco-API-1) as a strong allergen that produces an overwhelming IgE response in sheep, when produced endogenously by nematode; however, when administered separately, Tco-API-1 does not evoke IgE response. In response against T. colubriformis, serum immunoglobulin G1 (IgG1) and immunoglobulin M (IgM) titres elevate significantly by 35 days of infection with IgG1 being more persistent than IgM (Douch et al., 1994). IgG1 and immunoglobulin G2 (IgG2) levels in gut associated lymphoid tissue were observed to be greater in Merino sheep during T. colubriformis larval rejection (McClure et al., 1992). Treatment with the corticosteroid dexamethasone has been found to inhibit the progress of nematode resistance and reversibly reduce the expression of existing resistance in sheep, as indicated by higher faecal egg count and persistent weight loss. Dexamethasone functions by decreasing the production of leukotrienes and preventing the advent of mast cells in intestines (Douch et al., 1986, 1994) and eosinophils in serum (Buddle et al., 1992). Sheep also counter the nematodes by increased numbers of circulating antibodies and increased number of antibodies in the mucus of the intestine (Dawkins et al., 1988; Adams et al., 1989; McClure et al., 1992). Rabbit develops resistance against Trichostrongylus retortaeformis through three ways viz: self-cure; inhibition of larval development; and prevention of establishment of infective larval stages (Michel, 1952). All of these findings reveal that inflammatory response involving an interplay of different immune mediators including IL33, IgE, IgG1, IgG2, IgM, histamine, leukotriene C4, 6-keto prostaglandin $F_{1\alpha}$ and thromboxane B_2 is vital in the fight against Trichostrongylus infection with mast cells playing a key role (fig. 2).

Prevalence of Trichostrongylus in humans

Few investigations on Trichostrongylus have revealed its widespread occurrence in human communities around the world. Watson (1953) reported that 48 million people were infected with Trichostrongylus spp. globally. However, researches on the epidemiology of Trichostrongylus proclaim a global distribution but low prevalence of infection in humans (Ghadirian & Arfaa, 1975; Cancrini et al., 1982; Millington et al., 1989; Boreham et al., 1995; John & Petri, 2006; Ralph et al., 2006; Yong et al., 2007). Ghadirian & Arfaa (1975) estimated 67%, 86% and 71% prevalence of trichostrongylosis among humans in Isfahan, Bakhtiari and Khuzestan regions of Iran, respectively, with primary species being Trichostrongylus orientalis and T. colubriformis. Watthanakulpanich et al. (2013) found 36.9% prevalence in Thakamrien Savannakhet, Laos. Trichostrongylus infections often become misreported due to the resemblance of their eggs with those of hookworms, for example, in Lahanam Laos, 2011, 93.5% of positive hookworm cases were of Trichostrongylus (Sato et al., 2011). Joe (1947) reported 36.42% prevalence in Java, Indonesia. Very low prevalence of 0.5% and 1.2% was reported in Chile and Brazil, respectively (Torres et al., 1972; Souza et al., 2013). Heydon & Green (1931) also reported a very low prevalence (0.3-0.4%) of trichostrongylosis in Queensland, Australia. Infection of T. orientalis has been reported in Armenia, China, Japan and Korea (John & Petri, 2006). Species such as T. axei, Trichostrongylus capricola, T. colubriformis, T. orientalis, Trichostrongylus probolurus, Trichostrongylus skrjabin and T. vitrinus have been found associated with infections in humans with T. axei, T. colubriformis and T. orientalis being the most common species which infect humans, mostly obtained via close contact with livestock (Ghadirian & Arfaa, 1975; Millington et al., 1989; John & Petri, 2006; Ralph et al., 2006; Yong et al., 2007). In Japan, the most predominant species among humans is T. orientalis which is also found in China and Korea (Miyazaki, 1991). Buonfrate et al. (2017) reported four clusters of Trichostrongylus infection in Italy. Multiple cases were reported from different regions of Hungary from time to time as mentioned by Hollo et al. (1970). El-Shazy et al. (2006) reported 2.6% prevalence of trichostrongylsis in Dakahlia, Egypt. Females of age group 41-50 have been found to be more susceptible to trichostrongylosis than males (Watthanakulpanich et al., 2013) (figs 3-5).

Discussion

Different studies on the incidence of *Trichostrongylus* prove that this parasite is prevalent among small ruminants worldwide and sporadically occurs in humans that live in close contact with these ruminants. This review, which included almost all cases of Trichostrongylus infection that had been reported and documented in the literature worldwide, showed that trichostrongylosis occur in people occasionally. A systematic review of Trichostrongylus infection in Iran by Rahimi-Esboei et al. (2022) showed its prevalence to be 0.01%. However, in rural communities the prevalence is found to be higher (3.13%) (Sharifdini et al., 2020). With ten species of Trichostrongylus reported in human beings, Iran has been a hotspot of human trichostrongylosis; globally, 11 species of Trichostrongylus have been linked to humans (Sharifdini et al., 2017b). The metrics indicate that T. colubriformis, followed by other species, is the predominant species infecting humans around the world (Lattes et al., 2011; Phosuk et al., 2013; Watthanakulpanich et al., 2013; Gholami

 Table 1. Reported cases of human trichostrongylosis globally in scientific publications from 1938 to 2022.

Species		Number of cases reported in	Tin line.	Defermen
number	Location	numans	Findings	Reference
1.	Louisiana, United States	01	Trichostrongylus colubriformis was found in the appendix of a girl suffering from recurrent pain	Chenken & Moss (1938)
2.	New South Wales, Australia	01	strongyle type eggs were collected from faecal sample of a four years old girl. Larvae from eggs were cultured and fed to rabbit which produced adult worms of <i>T.</i> <i>colubriformis</i>	Heydon & Bearup (1939)
3.	Democratic Republic of Congo	03	parasitological stool examination of three persons with abdominal symptoms and eosinophilia showed <i>Trichostrongylus</i> infection in them	O'teal & Magath (1947)
4.	Baghdad, Iraq	04	four persons with anaemia and epigastric pain were diagnosed with trichostrongylosis	Jawad (1952)
5.	United States	02	stool examination of two patients suffering from abdominal pain, diarrhoea and weakness, revealed <i>Trichostrongylus</i> infection with profound eosinophilia	Wallace <i>et al.</i> (1956)
6.	United Kingdom	03	three women aged 21, 23 and 27 years with epigastric pain, weakness, dizziness and anaemia reported positive for <i>Trichostrongylus</i> spp. All cases were introduced from other countries	Markell (1968)
7.	Israel	59	through stool analysis, 59 cases of trichostrongylosis were found between 1967 and 1971 in Israel. All of the cases, nevertheless, originated in Asia. Patients experienced nausea, abdominal pain and other associated symptoms	Cotin <i>et al.</i> (1972)
8.	Isfahan, Iran	40	<i>Trichostrongylus capricola</i> outbreak was reported in tribal population of Isfahan, Iran, with 22.5% prevalence among <i>Trichostrongylus</i> infected population	Ghadirian <i>et al.</i> (1974)
9.	Tunisia	01	54 years old woman complaining about abdominal cramps was diagnosed with trichostrongylosis by stool examination	Bouchekoua <i>et al.</i> (1977)
10.	Khuzestan, Iran	06	ether formalin stool examination of six persons showed <i>Trichostrongylus</i> infection. Oxantel-pyrantel was given to patients to resolve the infection	Farahmandian <i>et al.</i> (1977)
11.	Μοτοςςο	01	56 years old woman with asthenia, weight loss and pain in the lower limbs was diagnosed with <i>Entamoeba</i> and <i>Trichostrongylus</i> infection after stool examination. Species identified was <i>Trichostrongylus vitrinus</i>	Poirriez <i>et al</i> . (1984a)
12.	India	35	In 1981, ten and 1982, 25 cases of <i>T. colubriformis</i> were reported among tribal children using direct faecal smears, brine flotation and formol-ether concentration approaches	Poirriez <i>et al.</i> (1984b)
13.	Queensland, Australia	05	stool examination of five persons with high eosinophilia revealed <i>Trichostrongylus</i> eggs. Two of the persons suffered from abdominal pain and diarrhoea. Pyrantel embonate resolved the eosinophilia and infection	Boreham et al. (1995)
14.	Tasmania, Australia	01	seven years old refugee from Sudan was diagnosed with trichostrongylosis after formalin-ethyl acetate concentration (FEAC) stool examination	Bradbury (2006)
15.	Sydney, Australia,	02	stool microscopy of two persons with abdominal pain, nausea and diarrhoea revealed <i>Trichostrongylus</i> infection which showed mebendazole resistance. Colonoscopy of one person revealed mild terminal ileitis. Intestinal biopsies of both persons showed an inflammatory cell infiltrate, including profound eosinophils. Both of the persons had pet goats which were the source of infection	Ralph <i>et al</i> . (2006)
16.	France	01	FEAC stool examination of a 47 years old woman with stomach aches, abdominal bloating and occasional diarrhoea detected <i>Trichostrongylus</i> eggs, which were further identified to be of <i>T. colubriformis</i> by internal	Lattes et al. (2011)

Table 1. (Continued.)

Species number	Location	Number of cases reported in humans	Findings	Reference
			transcribed spacer 2 (ITS2) rDNA sequencing. The infection was transmitted from contaminated strawberries fertilized with sheep manure	
17.	New Zealand	03	three persons in Cornwall, United Kingdom, with abdominal pain, diarrhoea, weight loss and eosinophilia revealed <i>Trichostrongylus</i> sp. infection through parasitological stool examination. Albendazole resolved the eosinophilia and infection after six weeks. They had contracted the infection from a sheep farm in New Zealand	Wall <i>et al.</i> (2011)
18.	Thailand and Lao People's Democratic Republic (PDR)	08	five symptomatic persons from Thailand and three from Lao PDR showed <i>Trichostrongylus</i> infection upon stool examination by FEAC technique. Molecular identification of third stage larva obtained through agar plate culture technique revealed two species of <i>Trichostrongylus</i> , viz: <i>T.</i> <i>colubriformis</i> and <i>Trichostrongylus axei</i>	Phosuk <i>et al.</i> (2013)
19.	Guilan Province, Iran	08	eight persons in Guilan Province, Iran, with varying degree of weakness, backache, abdominal complications, diarrhoea, urticaria, dyspepsia and low-grade fever showed <i>T. colubriformis</i> and <i>Trichostrongylus vitrinus</i> eggs on stool examination	Ashrafi et al. (2015)
20.	Thailand	41	Srinagarind Hospital, Thailand, reported 41 cases of human trichostrongylosis from 2005–2012 through stool examinations using the standard FEAC technique. Most of the patients had abdominal symptoms with profound eosinophilia	Phosuk <i>et al.</i> (2015)
21.	Mazandaran Province, Iran	33	33 persons with medical complications were examined with FEAC technique and found positive for trichostrongylosis. Molecular identification by ITS2 rDNA sequencing revealed <i>T. colubriformis</i> (29 samples) and <i>T.</i> <i>axei</i> (four samples)	Gholami <i>et al.</i> (2015)
22.	Gakenke, Rwanda	159	conventional stool examination combined with the polymerase chain reaction (PCR) approach was used to diagnose 159 cases of <i>Trichostrongylus</i> spp. in children from a rural area of Rwanda	Irisarri-Gutiérrez <i>et al.</i> (2016)
23.	Mazandaran Province, Iran	07 51	five males and two females with clinical symptoms were examined and reported positive for trichostrongylosis. Molecular identification confirmed <i>T. colubriformis</i> infection. Out of 132 suspected cases, 51 persons were found positive to <i>Trichostrongylus</i> infection using the highly sensitive multiplex restriction enzyme-PCR technique. three species, <i>T. colubriformis</i> , <i>T. axei</i> and <i>Trichostrongylus vitrines</i> were reported	Sharifdini <i>et al.</i> (2017a) and Mizani <i>et al.</i> (2017)
24.	Northern Italy	11	from 2010 to 2015, Centre for Tropical Diseases of Negrar Verona, Italy, reported four family outbreaks of trichostrongylosis (total 11 cases). Most of the patients suffered from abdominal pain, diarrhoea and had profound eosinophilia. All four families had a history of eating vegetables contaminated with sheep faeces	Buonfrate <i>et al</i> . (2017)
25.	Guilan Province, Iran	41 60	 41 individuals were found to be positive for <i>Trichostrongylus</i> infection using formalin-ether approach. three species were reported in this study, <i>viz</i>: <i>T. colubriformis, T. vitrinus</i> and <i>Trichostrongylus longispicularis.</i> The latter species was reported for the first time in humans. 60 persons from Guilan Province, Iran, with abdominal complications were infected with <i>Trichostrongylus</i> sp. Identification of <i>Trichostrongylus</i> was done through FEAC technique and nutrient-agar plate culture 	Sharifdini et al. (2017b) and Ghanbarzadeh et al. (2019)

(Continued)

Table 1. (Continued.)

Species number	Location	Number of cases reported in humans	Findings	Reference
26.	Mazandaran province, Iran	12	12 persons with abdominal complications were diagnosed with trichostrongylosis by cellophane thick smear method	Babamahmoodi <i>et al.</i> (2020)
27.	Langroud district, Guilan Province, Iran	07	seven members of a family in Guilan Province, Iran, were diagnosed with trichostrongylosis after coproparasitological examination. Most of them experienced abdominal pain, diarrhoea, low grade fever, night rigors, allergic manifestations including urticaria and weight loss. ITS2 rDNA identification revealed <i>T.</i> <i>colubriformis</i> species	Ashrafi <i>et al.</i> (2020)
28.	Guilan Province, Iran	47	a cross-sectional study of 1500 individuals from 31 villages of Fouman region was done using different stool examination methods to estimate intestinal parasitic infections. <i>Trichostrongylus</i> spp. was found in 3.13% of individuals	Sharifdini <i>et al.</i> (2020)
29.	La Araucanía region, Chile	07	coproparasitological examination of 7 persons from rural La Araucanía region, Chile, proved positive to strongylid eggs. ITS2 rDNA sequencing further confirmed the <i>T.</i> <i>colubriformis</i> infection. Infected persons suffered from varying degree of diarrhoea, reflux, vomiting, meteorism and eosinophilia	Hidalgo <i>et al.</i> (2020)
30.	Valdivia, Chile	03	a person in Valdivia, Chile, reported with persistent diarrhoea for three weeks. Along with him, two other members of his family also proved positive for <i>T.</i> <i>colubriformis</i> infection on stool examination	Torres <i>et al.</i> (2021)
31.	China	01	a sheep herder from China presented with severe persistent diarrhoea, abdominal pain, weakness and hyper-eosinophilia. On stool examination, he was positive for <i>T. colubriformis</i> infection	Du et al. (2022)

et al., 2015; Hidalgo *et al.*, 2020; Torres *et al.*, 2021; Du *et al.*, 2022) with Iran reporting the highest number of cases.

Most cases of human trichostrongylosis around the world have been revealed following a parasitological stool examination particularly using the FEAC technique on persons with gastrointestinal complications (diarrhoea, abdominal pain, weakness and loss of appetite) and eosinophilia. In family outbreaks of trichostrongylosis, some family members with hypereosinophilia



Fig. 1. Activity and development of *Trichostrongylus* sp. in intestine of host; L3-larvae stages burrow through epithelium resulting in damage to intestinal villi and enteritis. Then adults emerge out through tunnels into the lumen of intestine and occasionally suck blood from the intestinal vasculature which leads to anaemia.



Fig. 2. Induction of Th2 immune response; Trichostrongylus colubriformis expressing allergen Tco-API-1, that is presented to Th2 cells by APC which releases cytokines (IL-4, IL-5, IL-9 and IL-13) that activate other cells including mast cell, eosinophil and B-cells. B-cells produce allergen specific IgE which bind to worm surface by Fab region and eosinophil by Fc region to induce their degranulation in order to kill the parasite. Allergen can also bind to mast cell bound IgE to cause their degranulation. In addition. damaged intestinal epithelial cells release IL-33 which binds to ST-2 receptor expressed by cells such as mast cell, basophil and eosinophil to cause their degranulation. Abbreviations: Tco-API-1, Trichostrongylus colubriformis sspartyl inhibitor; Th2 cell, T-helper 2 cell; APC, antigen presenting cell; IL, interleukin; Fab, fragment antigen-binding; Fc, fragment crystallizable; ST-2 is an IL-33 receptor belonging to the IL-1 family.



Fig. 3. Countries (coloured) with reported cases of *Trichostrongylus* infection in humans; Human infection by *Trichostrongylus* spp. is documented in several countries and is not limited to any one geographical area. Comprehensive inspection of hookworm patients may indicate otherwise in nations where *Trichostrongylus* has not yet been reported, as trichostrongyle eggs are frequently mistaken for hookworm eggs.

prove negative for *Trichostrongylus* eggs on stool examination which can be attributed to a long prepatent period of the parasite, that is, four months to two years (Wolfe, 1978; Boreham *et al.*, 1995; Ralph *et al.*, 2006). Saraei *et al.* (2019) showed a higher sensitivity of FEAC (95.8%) than the agar plate culture method (90.1%) in diagnosis of trichostrongylosis. However, a recent study showed that among conventional parasitological stool examination techniques, Willi's method is more sensitive (91.7%) followed by the agar plate culture method (52.8%), Harada–Mori culture (40.3%), FEAC (37.5%) and 5.6% for the wet mount technique (Pandi *et al.*, 2021). The same study showed

the polymerase chain reaction (PCR) assay to be highly sensitive (97.2%) and specific in diagnosing human trichostrongylosis. Recently, Mizani *et al.* (2017) the devised restriction enzyme-PCR assay for the detection of *Trichostrongylus* species, which has higher specificity to detect even 10 pg/ml of DNA from the sample. Arbabi *et al.* (2020) developed the PCR-high resolution melting assay for the diagnosis of *Trichostrongylus* species with higher specificity in differentiating *T. colubriformis*, *T. capricula*, *T. vitrinus* and *T. probblorous*. Most research to date has used the FEAC approach to diagnose human trichostrongylosis, which has lower sensitivity than Willi's method in



Fig. 4. Human trichostrongylosis case count by country. It is evident that Iran has reported the highest number of cases of *Trichostrongylus* among humans, which can be attributed to higher screening among human populations in part and the rest for pastoral livelihood of people in rural areas of Iran.

Fig. 5. Year wise human trichostrongylosis cases reported in the scientific literature from 1938 to 2022. Between the years 2015 and 2020, the number of cases is comparatively higher because of increased testing in rural human communities. Investigations in other pastoral communities of the world may detect more infections of trichostrongylosis.

conventional methods. Therefore, many more cases of trichostrongylosis can be discovered if human populations are screened using highly specific PCR along with the conventional Willi's and FEAC approaches.

Even though helminth zoonoses are declining globally, research has shown that warming caused by climate change, in particular, may eventually lead to a rise in helminth infections. The cause of this is that warming would lessen these helminths' larval arrest including Trichostrongylus, and so lengthen the time of year when the free-living larval stages would be active. This would result in a rise in the rates of inadvertent human and livestock ingestion of larval stages, as well as the rates of infection (Dobson & Hudson, 1992; Mas-Coma et al., 2008). This review indicates that contamination of food (vegetables) by livestock faeces is the main source of trichostrongylosis among humans which is further enhanced by rainfall due to dispersal of faecal material (Ghadirian & Arfaa, 1973; Ashrafi et al., 2020; Hidalgo et al., 2020). Along with close proximity to livestock, consumption of untreated wild aromatic plants, use of animal dung as fuel (common among rural residents), unhygienic eating practises, use of livestock manure as fertilizer, weakened immune system and advanced age are some other risk factors linked to the rising incidence of human trichostrongylosis (Ashrafi et al., 2020; Rahimi-Esboei et al., 2022). In light of many proven ill effects of chemical fertilizers, organic farming with use of animal dung as fertilizer is becoming common among rural areas which could further increase the incidence of human trichostrongylosis and other zoonotic diseases in the

near future (Ashrafi et al., 2020). Livestock being our most important source of meat and dairy must be monitored for helminth infections regularly through parasitological stool examination and be given periodic anthelmintic doses in order to prevent spillover of helminthiasis to humans. Proper hygiene decreases the rate of helminth infection greatly (Vaz Nery et al., 2019). Even though developed countries with advanced health care and proper hygiene have managed to curb the helminth infections, the 'hygiene hypothesis' and related studies have correlated the decrease in helminth infections with the increase of inflammatory and autoimmune diseases among human population as research proves that helminth infection in early stages of life are important contributors to the proper development of the immune system (Rook, 2012). With an increasing trend of anthelmintic resistance among different helminths, immunological studies among small ruminants in the direction of identifying potential antigenic proteins for the development of a vaccine against Trichostrongylus and other economically important nematodes in general are necessary. Pathogenicity has not been investigated in humans due to minimal number of infections. However, in sheep, Trichostrongylus causes high morbidity ranging from diarrhoea, severe degradation of intestinal wall to even mortality, which is more obvious in lambs. With the advent of new zoonotic diseases in the world, studies are needed to be conducted over these neglected mild pathogenic parasites to better understand their zoonotic potential and accordingly devise management strategies in the future.

Conclusion

The purpose of this review was to compile data regarding human trichostrongylosis in order to gain insights into the epidemiological, immunological and pathogenic aspects of Trichostrongylus species among humans. It was found that trichostrongylosis occurs among pastoral communities across the globe particularly in tropical countries. The majority of cases have been identified by traditional faecal examination techniques such as FEAC, however with the development of highly precise PCR-based approaches, it is important to combine these advanced techniques with traditional ones to accurately identify Trichostrongylus and other zoonotic helminths infecting humans. Most of the cases have been linked to the close association with the small ruminants or consumption of contaminated food. More investigations on human trichostrongylosis have led to an upsurge in instances being reported in recent years. With the recent finding of Trichostrongylus longispicularis among humans in Iran, the number of species associated with humans in this genus has increased to 11.

Further investigations are needed to be done to know the zoonotic potential and disease status of trichostrongylosis around the world. With meagre pathogenic studies and lack of information regarding the mechanism of immune response against *Trichostrongylus* spp., these two fields have remained unexplored; further research is needed in this direction to gather insights about the histological and physiological changes associated with human trichostrongylosis. Ruminants being an important source of food and other products, close association of humans with these domesticated animals is inevitable. In light of the emergence of new species of *Trichostrongylus* in humans, it is crucial to study their pathogenicity and zoonotic potential. Furthermore, it is recommended to devise proper management strategies in order to check the transmission of such zoonotic parasites in the future.

Financial support. The authors acknowledge financial support for this study from the Human Resource Development Group Council of Scientific and Industrial Research (HRDGCSIR), India and University of Kashmir.

Conflicts of interest. None.

Ethical standards. We, as authors of this review article, have made every effort to conduct a thorough and unbiased evaluation of the literature on the topic at hand. We have taken care to ensure that all sources are properly cited. We have also strived to maintain objectivity and accuracy in our interpretation of the data, and have not allowed personal biases to influence our conclusions. It is our hope that this review article will provide readers with a fair and comprehensive overview of the current state of knowledge on the subject.

References

- Adams DB, Anderson BH and Windon RG (1989) Cross immunity between Haemonchus contortus and Trichostrongylus colubriformis in sheep. International Journal for Parasitology 19(7), 717–722.
- Allen T, Murray KA, Zambrana-Torrelio C, Morse SS, Rondinini C, Di Marco M, Breit N, Olival KJ and Daszak P (2017) Global hotspots and correlates of emerging zoonotic diseases. *Nature Communications* 8(1), 1–10.
- Andronicos NM, McNally J, Kotze AC, Hunt PW and Ingham A (2012) *Trichostrongylus colubriformis* larvae induce necrosis and release of IL33 from intestinal epithelial cells *in vitro*: implications for gastrointestinal nematode vaccine design. *International Journal for Parasitology* **42**(3), 295–304.

- Arbabi M, Hooshyar H, Lotfinia M and Bakhshi MA (2020) Molecular detection of *Trichostrongylus* species through PCR followed by high resolution melt analysis of ITS-2 rDNA sequences. *Molecular and Biochemical Parasitology* 236, 111260.
- Ashrafi K, Tahbaz A, Sharifdini M and Mas-Coma S (2015) Familial *Trichostrongylus* infection misdiagnosed as acute fascioliasis. *Emerging Infectious Diseases* 21(10), 1869–1870.
- Ashrafi K, Sharifdini M, Heidari Z, Rahmati B and Kia EB (2020) Zoonotic transmission of *Teladorsagia circumcincta* and *Trichostrongylus* species in Guilan province, northern Iran: molecular and morphological characterizations. *BMC Infectious Diseases* **20**(1), 1–9.
- Babamahmoodi F, Ahangarkani F, Bahrami Dounchali F and Nikbakht A (2020) Trichostronyliasis outbreak in North of Iran. *Tabari Biomedical Student Research Journal* 2(4). http://tbsrj.mazums.ac.ir/article-1-3720-en. html.
- Barker IK (1975) Intestinal pathology associated with *Trichostrongylus colubriformis* infection in sheep: histology. *Parasitology* 70(2), 165–171.
- Bendixsen T, Emery DL and Jones WO (1995) The sensitization of mucosal mast cells during infections with *Trichostrongylus colubriformis* or *Haemonchus contortus* in sheep. *International Journal for Parasitology* 25(6), 741–748.
- Beveridge I, Pullman AL, Phillips PH, Martin RR, Barelds A and Grimson R (1989) Comparison of the effects of infection with Trichostrongylus colubriformis, T. vitrinus and T. rugatus in Merino lambs. *Veterinary Parasitology* 32(2-3), 229–245.
- Boreham RE, McCowan MJ, Ryan AE, Allworth AM and Robson JM (1995) Human trichostrongylosis in Queensland. *Pathology* 27(2), 182–185.
- Bouchekoua N, Garbouri M, Ben R and Triki S (1977) A case of trichostrongyliasis. *Tunisie Médicale* 55(6), 405–406.
- Bradbury R (2006) An imported case of trichostrongylid infection in Tasmania and a review of human trichostrongylidiosis. Annals of the ACTM: An International Journal of Tropical and Travel Medicine 7(2), 25-28.
- Buddle BM, Jowett G, Green RS, Douch PGC and Risdon PL (1992) Association of blood eosinophilia with the expression of resistance of Romney lambs to nematodes. *International Journal for Parasitology* 22(7), 955–960.
- Buonfrate D, Angheben A, Gobbi F, Mistretta M, Degani M and Bisoffi Z (2017) Four clusters of *Trichostrongylus* infection diagnosed in a single center, in Italy. *Infection* **45**(2), 233–236.
- Cancrini G, Boemi G, Iori A and Corselli A (1982) Human infestations by *Trichostrongylus axei*, *T. capricola* and *T. vitrinus*: 1st report in Italy. *Parasitologia* 24(2-3), 145–149.
- Chenken JR and Moss ES (1938) Trichostrongylus colubriformis in the human appendix. Report of a case in Louisiana. Journal of Laboratory and Clinical Medicine 24(1), 15–17.
- Cotin M, Talis B and Wertheim G (1972) Clinical and therapeutic aspects of *Trichostrongylus colubriformis* (Nematoda–Trichostrongylidae) infection in man. *Harefuah* 83(1), 26–54.
- Craig TM (2009) Helminth parasites of the ruminant gastrointestinal tract. pp. 78–91 In Anderson DE and Rings DM (Eds) *Food animal practice*. 5th edn. Amsterdam, Elsevier.
- Crofton HD (1948) The ecology of immature phases of trrichostrongyle nematodes: I. the vertical distribution of infective larvae of *Trichostrongylus* retortaeformis in relation to their habitat. Parasitology 39(1-2), 17–25.
- Dawkins HJS, Windon RG, Outteridge PM and Dineen JK (1988) Cellular and humoral responses of sheep with different levels of resistance to *Trichostrongylus colubriformis. International Journal for Parasitology* 18 (4), 531–537.
- **Dobson AP and Hudson PJ** (1992) Regulation and stability of a free-living host-parasite system, *Trichostrongylus tenuis* in red grouse. II: population models. *Journal of Animal Ecology* **61**, 487–498.
- **Dobson RJ, Waller PJ and Donald AD** (1990) Population dynamics of *Trichostrongylus colubriformis* in sheep: the effect of infection rate on the establishment of infective larvae and parasite fecundity. *International Journal for Parasitology* **20(3)**, 347–352.
- Douch PGC, Harrison GBL, Elliott DC, Buchanan LL and Green RS (1986) Relationship of gastrointestinal histology and mucus antiparasite activity

with the development of resistance to trichostrongyle infections in sheep. *Veterinary Parasitology* **20(4)**, 315–321.

- **Douch PGC, Risdon PL and Green RS** (1994) Antibody responses of sheep to challenge with *Trichostrongylus colubriformis* and the effect of dexamethasone treatment. *International Journal of Parasitology* **24**(7), 921–928.
- Du B, Zhang L, Dang W, et al. (2022) A sheepherder with a severe diarrhea caused by Trichostrongylus colubriformis. Travel Medicine and Infectious Disease 48, 102325.
- Edgar G (1933) Some observations on trichostrongylosis of young sheep. Australian Veterinary Journal 9(4), 149-154.
- El Shazly AM, Awad SE, Sultan DM, Sadek GS, Khalil HH and Morsy TA (2006) Intestinal parasites in Dakahlia governorate, with different techniques in diagnosing protozoa. *Journal of the Egyptian Society of Parasitology* **36**(3), 1023–1034.
- Farahmandian E, Arfaa F and Jalali H (1977) Evaluation of the effect of oxantel-pyrantel on various soil transmitted helminths in Iran. Iranian Journal of Public Health 6(2), 46–52.
- Getachew T, Dorchies P and Jacquiet P (2007) Trends and challenges in the effective and sustainable control of *Haemonchus contortus* infection in sheep. Review. *Parasite* 14(1), 3–14.
- Ghadirian E and Arfaa FNMN (1973) First report of human infection with Haemonchus contortus, Ostertagia ostertagi, and Marshallagia marshalli (family Trichostrongylidae) in Iran. Journal of Parasitology 59(6), 1144–1145.
- Ghadirian E and Arfaa F (1975) Present status of trichostrongyliasis in Iran. American Journal of Tropical Medicine and Hygiene 24(6), 935–941.
- Ghadirian E, Arfaa F and Sadighian A (1974) Human infection with Trichostrongylus capricola in Iran. American Journal of Tropical Medicine and Hygiene 23(5), 1002–1003.
- Ghanbarzadeh L, Saraei M, Kia EB, Amini F and Sharifdini M (2019) Clinical and haematological characteristics of human trichostrongyliasis. *Journal of Helminthology* **93**(2), 149–153.
- Gholami S, Babamehmoodi F, Abedian R, Sharif M, Shahbazi A, Pagheh A and Mehdi F (2015) *Trichostrongylus colubriformis*: possible most common cause of human infection in Mazandaran Province, North of Iran. *Iranian Journal of Parasitology* **10(1)**, 110–115.
- Gill HS (1991) Genetic control of acquired resistance to haemonchosis in Merino lambs. *Parasite Immunology* **13(6)**, 617–628.
- Heydon GAM and Bearup AJ (1939) A further case of human infection with Trichostrongylus colubriformis in New South Wales. Medical Journal of Australia 1(18), 694–695.
- Heydon GM and Green AK (1931) Some worm infestations of man in Australia. *Medical Journal of Australia* **21**(1), 619–628.
- Hidalgo A, Gacitúa P, Melo A, Oberg C, Herrera C and Fonseca-Salamanca F (2020) First molecular characterization of *Trichostrongylus colubriformis* infection in rural patients from Chile. *Acta Parasitologica* 65(3), 790–795.
- Hollo F, Rovo JT and Hidvegi Z (1970) A recent case of human *Trichostrongylus* infection in Hungary. *Parasitologia Hungarica* **3(1)**, 159–168.
- Irisarri-Gutiérrez MJ, Muñoz-Antolí C, Acosta L, Parker LA, Toledo R, Bornay-Llinares FJ and Esteban JG (2016) Hookworm-like eggs in children's faecal samples from a rural area of Rwanda. *African Health Sciences* 16(1), 83–88.
- Janquera P (2017) *Trichostrongylus* spp. parasitic roundworms of cattle, sheep, goats, pigs and horses: biology, prevention and control. Available at https://Parasitipedia.net/index.php?option = com_content&view = article&id= 2628&Itemid=2908 (accessed January 2023).
- Jawad H (1952) Four cases of trichostrongyliasis in Iraq. *Journal of the Faculty* of Medicine, Baghdad 16(2), 68–70.
- Joe LK (1947) *Trichostrongylus* infection in man and domestic animals in Java. *Journal of Parasitology* **33**(4), 359–362.
- John DT and Petri WA (2006) The intestinal nematodes. Markell and Voge's medical parasitology. pp. 266–267. 9th edn. St Louis, USA, Elsevier.
- Jones WO and Emery DL (1991) Demonstration of a range of inflammatory mediators released in trichostrongylosis of sheep. *International Journal for Parasitology* **21**(3), 361–363.
- Lattes S, Ferte S, Delaunay P, Depaquit J, Vassallo M, Vittier M, Kokcha S, Coulibaly E and Marty P (2011) Trichostrongylus colubriformis nematode infections in humans, France. Emerging Infectious Diseases 7(17), 1301–1302.

- Levine D and Anderson L (1973) Development and survival of *Trichostrongylus* colubriformis on pasture. Journal of Parasitology **59(1)**, 147–165.
- Libera K, Konieczny K, Grabska J, Szopka W, Augustyniak A and Pomorska-Mól M (2022) Selected livestock-associated zoonoses as a growing challenge for public health. *Infectious Disease Reports* 14(1), 63–81.
- Markell EK (1968) Pseudohookworm infection—trichostrongyliasis: treatment with thiabendazole. *New England Journal of Medicine* 278(15), 831–832.
- Mas-Coma S, Valero MA and Bargues MD (2008) Effects of climate change on animal and zoonotic helminthiases. *Revue Scientifique et Technique* 27 (2), 443–457.
- McClure SJ, Emery DL, Wagaland BM and Jones WO (1992) A serial study of rejection of *Trichosfrongylus colubriformis* by immune sheep. *International Journal for Parasitology* **22(2)**, 227–234.
- Michel JF (1952) Inhibition of development of *Trichostrongylus retortaeformis*. Nature 169(4309), 933–934.
- Miller HRP, Jackson F, Newlands GFJ and Huntley JF (1985) Rapid expulsion of gastrointestinal nematodes in the sheep: a role for immediate hypersensitivity reactions in the mucosa. pp. 460–479. In Morris B and Miyasaka M (Eds) *Immunology of the sheep*. Basel, Switzerland, Basel Institute for Immunology.
- Millington MA, Costa CH, Tavares AM, Dourado H, Reid WA and Macedo V (1989) Detection of helminthiasis with the Kato-Katz, Baermann-Moraes and Harada methods, in Tefé and various villages by the Japurá-Caquetá River, Amazonas. Revista da Sociedade Brasileira de Medicina Tropical 22(4), 217–218.
- Miyazaki I (1991) An illustrated book of helminthic zoonoses. Fukuoka, Japan, International Medical Foundation of Japan. Shukosha Printing.
- Mizani A, Gill P, Daryani A, Sarvi S, Amouei A, Katrimi AB and Sharif M (2017) A multiplex restriction enzyme-PCR for unequivocal identification and differentiation of *Trichostrongylus* species in human samples. *Acta Tropica* **173(1)**, 180–184.
- O'teal R and Magath TB (1947) *Trichostrongylus* infection of human beings: report of three cases. *Proceedings of Staff Meetings of the Mayo Clinic* 22 (10), 193–197.
- Pandi M, Sharifdini M, Ashrafi K, Atrkar Roushan Z, Rahmati B and Hajipour N (2021) Comparison of molecular and parasitological methods for diagnosis of human trichostrongylosis. *Frontiers in Cellular and Infection Microbiology* 11(75), 93–96.
- Phosuk I, Intapan PM, Sanpool O, Janwan P, Thanchomnang T, Sawanyawisuth K, Morakote N and Maleewong W (2013) Short report: molecular evidence of *Trichostrongylus colubriformis* and *Trichostrongylus* axei infections in humans from Thailand and Lao PDR. American Journal of Tropical Medicine and Hygiene 89(2), 376–379.
- Phosuk I, Intapan MP, Prasongdee KT, Changtrakul Y, Sanpool O, Janwan P and Maleewong W (2015) Human trichostrongylosis: a hospital case series. Southeast Asian Journal of Tropical Medicine and Public Health 46(2), 191–197.
- Poirriez J, Dei-Cas E, Guevart E, Abdellatifi M, Giard P and Vernes A (1984a) Human infestation by *Trichostrongylus vitrinus* in Morocco. *Annales de Parasitologie Humaine et Comparée* **59**(6), 636–638.
- Poirriez J, Pais G, Pais-Rajamma C and Vernes A (1984b) Human trichostrongyliasis in India. Transactions of the Royal Society of Tropical Medicine and Hygiene 78(3), 425–426.
- Rahimi-Esboei B, Pourhajibagher M and Bahador A (2022) Prevalence of human trichostrongyliasis in Iran: a systematic review and meta-analysis. *Reviews in Medical Microbiology* 33(1), 16–22.
- Ralph A, O'Sullivan MV, Sangster NC and Walker JC (2006) Abdominal pain and eosinophilia in suburban goat keepers – trichostrongylosis. *Medical Journal of Australia* 184(9), 467–469.
- Rehman A and Abidi SMA (2022) Livestock health: current status of helminth infections and their control for sustainable development. In Sobti RC (Ed.) Advances in animal experimentation and modeling. pp. 365– 378. Aligarh, India, Academic Press.
- Rook GA (2012) Hygiene hypothesis and autoimmune diseases. Clinical Reviews in Allergy & Immunology 42(1), 5–15.
- Saraei M, Ghanbarzadeh L, Hajialilo E, Barghandan T, Amini F and Sharifdini M (2019) Comparison of nutrient agar plate culture and

formalin-ethyl acetate concentration methods in diagnosis of human trichostrongyliasis. *Journal of Ardabil University of Medical Sciences* **18**(4), 506–514.

- Sato M, Yoonuan T, Sanguankiat S, Nuamtanong S and Pongvongsa T (2011) Short report: human *Trichostrongylus colubriformis* infection in a Rural Village in Laos. *American Journal of Tropical Medicine and Hygiene* **84**(1), 52–54.
- Sharifdini M, Heidari Z, Hesari Z, Vatandoost S and Kia EB (2017a) Molecular phylogenetics of *Trichostrongylus* species (Nematoda: Trichostrongylidae) from humans of Mazandaran Province, Iran. *The Korean Journal of Parasitology* 55(3), 279–285.
- Sharifdini M, Derakhshani S, Alizadeh SA, Ghanbarzadeh L, Mirjalali H, Mobedi I and Saraei M (2017b) Molecular identification and phylogenetic analysis of human *Trichostrongylus* species from an endemic area of Iran. *Acta Tropica* 176(1), 293–299.
- Sharifdini M, Ghanbarzadeh L, Barikani A and Saraei M (2020) Prevalence of intestinal parasites among rural inhabitants of Fouman, Guilan Province, Northern Iran with emphasis on *Strongyloides stercoralis*. *Iranian Journal of Parasitology* 15(1), 91–100.
- Sharma S and Anand N (1997) Parasitic diseases: an overview. Pharmacochemistry Library 25(1), 1–45.
- Shaw RJ, Morris CA, Green RS, Wheeler M, Bisset SA, Vlassoff A and Douch PGC (1999) Genetic and phenotypic parameters for *Trichostrongylus colubriformis*-specific immunoglobulin E and its relationships with anti-*Trichostrongylus colubriformis* antibody, immunoglobulin G1, faecal egg count and body weight traits in grazing Romney lambs. *Livestock Production Science* 58(1), 25–32.
- Shaw RJ, McNeill MM, Maass DR, Hein WR, Barber TK, Wheeler M and Shoemaker CB (2003) Identification and characterisation of an aspartyl protease inhibitor homologue as a major allergen of *Trichostrongylus colu*briformis. International Journal for Parasitology 33(11), 1233–1243.
- Shayo ME and Benz GW (1979) Histopathologic and histochemic changes in the small intestine of calves infected with *Trichostrongylus colubriformis*. *Veterinary Parasitology* 5(4), 353–364.

- Souza RP, Souza JN, Menezes JF, Alcântara LM, Soares NM and Teixeira MCA (2013) Human infection by *Trichostrongylus* spp. in residents of urban areas of Salvador city, Bahia, Brazil. *Biomedica* 33(3), 439–445.
- Steel J, Jones W and Wagland B (1990) The response of immune sheep to challenge with *Trichostrongylus colubriformis*: enteric plasma loss and secretion of biogenic amines. *International Journal for Parasitology* 20(8), 1067–1073.
- Torres P, Figueroa L and Navarrete N (1972) Trichostrongylosis en la provincia de Valdivia, Chile [Trichostrongylosis in the province of Valdivia, Chile]. Boletín Chileno de Parasitología 27(1), 52–55. [In Spanish.]
- Torres HP, Arcos BA, Villa AE and Cerna DO (2021) Family outbreak caused by the nematode *Trichostrongylus colubriformis* in a rural area of the province of Valdivia: a rare occurrence zoonoses. *Revista Chilena de Infectología: Órgano Oficial de la Sociedad Chilena de Infectología* **38**(3), 455–460.
- Vaz Nery S, Pickering AJ, Abate E, Asmare A, Barrett L, Benjamin-Chung J and Brooker SJ (2019) The role of water, sanitation and hygiene interventions in reducing soil-transmitted helminths: interpreting the evidence and identifying next steps. *Parasites & Vectors* 12(1), 1–8.
- Wall EC, Bhatnagar N, Watson J and Doherty T (2011) An unusual case of hypereosinophilia and abdominal pain: an outbreak of *Trichostrongylus* imported from New Zealand. *Journal of Travel Medicine* 18(1), 59–60.
- Wallace L, Henkin R and Mathies AW (1956) Trichostrongylus infestation with profound eosinophilia. Annals of Internal Medicine 45(1), 146–150.
- Watson JM (1953) Human trichostrongylosis and its relationship to ancylostomiasis in southern Iraq, with comments on world incidence. *Parasitology* 43(1–2), 102–109.
- Watthanakulpanich D, Pongvongsa T, Sanguankiat S, et al. (2013) Prevalence and clinical aspects of human *Trichostrongylus colubriformis* infection in Lao PDR. Acta Tropica **126**(1), 37–42.
- Wolfe MS (1978) Oxyuris, Trichostrongylus and Trichuris. Clinics in Gastroenterology 7(1), 201–217.
- Yong TS, Lee JH, Sim S, et al. (2007) Differential diagnosis of *Trichostrongylus* and hookworm eggs via PCR using ITS-1 sequence. *Korean Journal of Parasitology* **45(1)**, 69–74.