

Nutrition Society Scottish Section Meeting held at The Royal College of Physicians, Edinburgh on 21-22 March 2016

Conference on 'Phytochemicals and health: new perspectives on plant-based nutrition' Symposium 3: Phytochemicals for healthier foods

Alterations in vitamin A/retinoic acid homeostasis in diet-induced obesity and insulin resistance

Nimesh Mody

University of Aberdeen, Institute of Medical Sciences, School of Medicine, Medical Sciences and Nutrition, Foresterhill Health Campus, Aberdeen, Scotland AB25 2ZD, UK

Vitamin A is an essential micronutrient for life and the phytochemical β -carotene, also known as pro-vitamin A, is an important dietary source of this vitamin. Vitamin A (retinol) is the parent compound of all bioactive retinoids but it is retinoic acid (RA) that is the active metabolite of vitamin A. The plasma concentration of retinol is maintained in a narrow range and its normal biological activities strictly regulated since excessive intake can lead to toxicity and thus also be detrimental to life. The present review will give an overview of how vitamin A homeostasis is maintained and move on to focus on the link between circulating vitamin A and metabolic disease states. Finally, we will examine how pharmacological or genetic alterations in vitamin A homeostasis and RA-signalling can influence body fat and blood glucose levels including a novel link to the liver secreted hormone fibroblast growth factor 21, an important metabolic regulator.

Retinoids: Adiposity: Glucose homeostasis: Ceramide: Fibroblast growth factor 21

Vitamin A dietary intake and deficiency

A historical perspective of vitamin A tells us that the symptoms of vitamin A deficiency and the ability to treat night blindness with animal liver have been known and documented for thousands of years (Fig. 1). Animal liver is enriched in retinyl esters (particularly retinyl palmitate, the main storage form of vitamin A). It was about 1930, in experiments in rats by T. Moore that determined that the yellow phytochemical β -carotene (pro-vitamin A) was the precursor for the active vitamin A, which was stored in the liver. Separately P. Karrer first isolated and then determined the distinct chemical structures of β -carotene and retinol⁽¹⁻³⁾.

Vitamin A/retinoic acid (RA) have been shown to play essential roles in the preservation of immune function, continued promotion of good vision and the development, growth and maintenance of multiple body tissues. Animals do not have the capability to generate vitamin A via *de novo* synthesis and daily dietary vitamin A intake

can vary. Important dietary sources of vitamin A are from richly-coloured fruit and vegetables and also fish and animal liver⁽⁴⁾. More generally, richly-coloured fruit and vegetables are abundant sources of phytochemicals (such as carotenoids), which are not essential, but do have potential health benefits. Carotenoids form a large class of colourful plant pigments, which include carotenes such as β-carotene (pro-vitamin A) and lycopene and also xanthophylls such as zeaxanthin and lutein (which unlike β -carotene are not metabolised to vitamin A). Dietary and nutritional surveys performed in the USA and Europe indicate that most people's dietary intake of β -carotene is 1–2 mg/d, which is sufficient if the intake of preformed vitamin A (from animal sources) is also adequate⁽⁴⁾. Potential health benefits from frequent consumption of fruit and vegetables are often ascribed to high concentrations of these phytochemicals and their relatively high antioxidant activities. However, the focus of this review is specifically how vitamin A and RA signalling can influence metabolic physiology.

Abbreviations: FGF, fibroblast growth factor; RA, retinoic acid; RBP, retinol binding protein. **Corresponding author:** N. Mody, email n.mody@abdn.ac.uk



598 N. Mody

Early history of vitamin A **Ancient Greece** Chinese Northern Europe Ancient Egypt (1825-1500 BC) (460-327 BC) (600-700 AD) (Middle Ages) **Dutch physician Jacob van Maerlandt** First connections between eye disease or night (1235-1299) wrote the poem: blindness and treatment with topical or nutritional Who does not at night see right application of raw liver of ox, goat or pig. Eats the liver of goat Hippocrates first described "nyktalopia" or night He will then see better at night. blindness. 19th - early 20th century history of vitamin A F. Magendie George Budd G. Lunin M. Mori Elmer V. McCollum (1907) and at the same time 1881 Lafayette Mendel and Thomas Burr Osborne 1817 1842 1904 First experimental studies linking not just general

Fig. 1. (Colour online) A historical perspective of vitamin A, known and documented for thousands of years, from ancient Egypt to the early 20th century discovery of an essential 'fat soluble factor A'⁽¹⁻³⁾.

Vitamin A homeostasis

and growth retardation.

malnutrition but an actual "vitamin A" deficiency

and causation of corneal defects, night-blindness

Vitamin A is essential for animals, but they cannot synthesise vitamin A and also daily dietary intake can vary widely, so animals have evolved an intricate pathway to maintain precise vitamin A homeostasis⁽⁵⁾. Firstly retinol is acquired from dietary sources in the form of preformed retinyl esters (animal sources) or the pro-vitamin β-carotene (fruit and vegetable sources), which can be enzymatically cleaved in the small intestine and converted into retinol. Secondly, to store vitamin A, retinol is coupled to fatty acids like palmitate in the liver to form retinvl esters and these are mobilised as required. Thirdly, the plasma concentration of retinol is strictly regulated and maintained at about 2-3 µm in human subjects (and about 1 µM in mice). Finally, retinol must be metabolised to RA within cells before it can enter the nucleus to regulate gene transcription via ligand induced activation of specific transcription factors.

Ingesting too much preformed vitamin A from foods (such as fish or animal liver), supplements, or prescription medications based on vitamin A can lead to hypervitaminosis A and toxicity of the liver. It may also lead to decreased bone mineral density⁽⁶⁾ and these compounds can induce teratogenic effects in the developing embryo, thus women of child bearing age are advised to avoid such foods and drugs around conception and the first trimester of pregnancy⁽⁴⁾. These problems are not an issue with intake of pro-vitamin A/ β -carotene, from fruit and vegetable sources, because of the lower conversion efficiency of β -carotene to retinol equivalents (compared with preformed vitamin A)⁽⁴⁾.

Retinoid transport, metabolism and signalling

In the molecular era, it is through the study of genetic knockouts of the key proteins that regulate vitamin A

storage and mobilisation that have elucidated how homeostasis is maintained⁽⁵⁾. Vitamin A is a lipophilic, fat soluble molecule and therefore requires specific binding proteins in order to be transported in the circulation and within the cell. Serum retinol binding protein (RBP; gene name RBP4) is essential for mobilisation of hepatic retinol into plasma, and for the delivery of retinol to the retina of the eye (where it is required for normal vision), especially when the availability of dietary retinoid is marginal. Approximately 80–90 % of all retinoids in the body are stored as retinyl esters in the liver. Here, cellular retinol binding protein-1 and lecithin:retinol acyltransferase serve to facilitate efficient retinyl ester synthesis, metabolism and mobilisation within cells⁽⁵⁾. Retinol derived from the hydrolysis of retinyl ester can bind to RBP and the retinol-RBP complex enters the bloodstream for transport to target tissues to meet the tissues retinoid requirements. Cellular retinol binding protein-1 is also proposed to play a role in regulating the oxidation of retinol in the two step conversion to $RA^{(5)}$.

Discovery of a "fat soluble factor A"

essential for growth and survival.

RA is the active metabolite of vitamin A, and is a critical signalling molecule involved in many biological processes via ligand induced activation of specific transcription factors, nuclear hormone receptors called RA-receptors. RA-receptors form heterodimers with retinoid-X receptors and bind to RA-response elements present in the promoters of target genes via the DNA-binding domain present within each receptor^(7,8). It is via this binding to DNA that enables RA to regulate gene transcription programmes. RA-signalling is well known to play essential roles in developmental biology. In contrast, within the retina of the eye, it is the metabolite 11-cis-retinal that drives photoreceptor function in the visual cycle⁽⁹⁾.

Due to the maintenance of tight vitamin A homeostasis in adult animals, RA-mediated nuclear signalling was largely assumed that to be relatively less



important, especially in the regulation of metabolic physiology. Indeed much more attention has been paid to the role of other nuclear receptors in the regulation of metabolic processes including glucose and lipid metabolism, e.g. the RA-receptor heterodimer partner retinoid-X receptors, PPAR, liver-X-receptors and farnesoid-X-receptors (10). However, an increasing number of ground-breaking studies have now added to the largely ignored historical data that linked vitamin A levels with body mass regulation and type-2 diabetes (11–13).

Vitamin A levels and type-2 diabetes

Type-2 diabetes is a condition whereby blood glucose levels are elevated due to impaired insulin action (or insulin resistance) in muscle and liver and is commonly associated with increased fat mass and obesity. There are several reports associating increased circulating vitamin A levels or serum RBP4 (essential for transport of retinol in blood) with type-2 diabetes. Early reports of increased levels of serum and/or urine RBP4 in subjects with type-2 diabetes did not identify a cause or a down-stream effect^(11–13). In 2005, a role for elevated serum levels of RBP4 was first described from the laboratory of B.B. Kahn. They discovered that expression of RBP4 was elevated in adipose tissue of mice with impaired insulin action in muscle and liver in vivo⁽¹⁴⁾. Elevated levels of this adipokine (a term used to describe hormones or paracrine factors produced by adipose tissue) were found to associate with insulin resistance in multiple genetic and diet-induced models of obesity and type-2 diabetes. Several experimental models to alter circulating RBP4 levels, either genetically or pharmacologically, suggested strongly that elevated RBP4 levels might contribute to insulin resistance in muscle and liver and thus impaired glucose homeostasis and type-2 diabetes⁽¹⁴⁾.

However, human studies of obesity, insulin resistance or type-2 diabetes have been more controversial since not all of these have reported elevations in circulating RBP4 levels. Technical problems using enzyme-linked immunoassays may have led to inaccurate measurements of serum RBP4 concentrations⁽¹⁵⁾. Other confounding issues such as genetic background, age, sex, adipose distribution and kidney function should also be considered⁽¹⁵⁾. The exact mechanism by which elevated RBP4 levels contribute to insulin resistance and impaired glucose homeostasis remains unclear, but will be considered next. Serum RBP was first reported to induce hepatic expression of the key gluconeogenic enzyme phosphoenolpyruvate carboxykinase and thus increase circulating blood glucose levels via increased hepatic glucose production (14). This effect could be directly via increased RA-signalling to the RA-response elements in the gene promotor of phosphoenolpyruvate carboxykinase (also see next section). Elevated levels of serum RBP were also reported to impair insulin signalling in skeletal muscle by an unknown mechanism⁽¹⁴⁾.

The early findings, implicating elevated circulating levels of RBP4 with insulin resistance, had used the

available transgenic mouse models at the time, the whole body knockout of RBP4 and muscle-specific overexpression of RBP4^(14,16,17). Very recent advances in the field have been reported with the generation of mice with adipocyte-specific RBP4 overexpression⁽¹⁸⁾ and mice with hepatocyte-specific deletion of RBP4⁽¹⁹⁾. These complementary studies concluded that adipocyte RBP4 is not a significant source of circulating RBP4 since circulating RBP4 was undetectable in mice lacking hepatocyte RBP4⁽¹⁹⁾. However, in the setting of insulin resistance. adipocyte RBP4 may have a more important paracrine function that is confined within the adipose tissue compartment. Indeed, adipocyte-specific overexpression of RBP4 increased proinflammatory markers and genes encoding lipases associated with lipolysis in adipose tissue⁽¹⁸⁾. These alterations in adipose tissue were also associated with increased circulating NEFA and an increase in hepatic lipid accumulation without alterations in hepatic retinoid levels or genes.

In some tissues such as muscle and adipose, retinol-RBP4 is recognised by a plasma membrane receptor termed 'stimulated by retinoic acid 6' that transports retinol from the binding protein into cells⁽²⁰⁾. Stimulated by retinoic acid 6 also appears to signal via a phosphorylation cascade involving Janus kinases and their associated transcription factors called signal transducers and activators of transcription⁽²⁰⁾. These observations suggest more than one possible mechanism through which RBP4 may control insulin action but until a more complete network of signalling downstream of RBP4 and/or stimulated by retinoic acid 6 is determined, the role of RBP4 in metabolic disease will remain unclear.

Dietary, pharmacological or genetic manipulation of vitamin A/retinoic acid homeostasis

In contrast, dietary vitamin A deficiency or genetic intervention to decrease endogenous production of RA and thus reduce RA-signalling in mice leads to decreased fasting blood glucose levels and reduced hepatic gluconeogenesis^(21,22). Specifically, mice lacking the RA-synthesizing enzyme retinaldehyde dehydrogenase-1 have reduced expression of key gluconeogenic enzymes glucose-6-phosphatase and phosphoenolpyruvate carboxykinase, which is a *bona fide* RA-inducible target gene since the phosphoenolpyruvate carboxykinase gene promoter has been shown to contain a specific RA-receptor-binding site i.e. a RA-response element⁽²¹⁾.

Moreover, mice without retinaldehyde dehydrogenase-1 are protected from high-fat diet-induced obesity-insulin resistance. The mechanism of this protection was proposed to be from the accumulation of retinaldehyde (the sole precursor to RA). It was also suggested that retinaldehyde can increase the expression of mitochondrial uncoupling protein1 to drive uncoupled respiration and adaptive thermogenesis in white adipose tissue, essentially promoting a brown adipose phenotype to increase energy expenditure and inhibit weight gain. This effect is not unlike the well-characterised pharmacological



N. Mody

effects of RA to also induce uncoupling protein-1^(23–26). RA has also been shown to inhibit the cellular process of adipocyte differentiation and prevent obesity and insulin resistance in rodents^(27–30).

The overall beneficial metabolic effects of RA have also been reported to include changes in hepatic lipid metabolism leading to repartitioning of fatty acids away from TAG storage and towards oxidation⁽³¹⁾ However, liver toxicity with large doses and/or prolonged exposure, restricts the use of RA as a potential therapy for obesity and type-2 diabetes. Moreover, hypertriglyceridemia is a relatively frequent side effect of retinoid therapy (e.g. for dermatological disorders)(32). Retinoidinduced hypertriglyceridemia in human subjects has also been modelled in a number of rodent studies and has been reported to occur in response to high doses of vitamin A (as retinol or retinyl palmitate), RA isomers (including RA, 9-cis RA and 13-cis RA) and synthetic retinoid-X receptors-specific agonists (rexinoids)(33,34). These and other seemingly contradictory findings are reviewed in more detail elsewhere, and may depend on the balance between nuclear hormone heterodimers and the complex interplay with other transcription factors controlling adaptations to nutrient intake and metabolic needs⁽³⁵⁾.

Beneficial metabolic effect of the synthetic retinoid Fenretinide

We have shown that Fenretinide, a synthetic retinoid with decreased toxicological profile, can reduce obesity and prevent insulin-resistance in both muscle and liver in the high-fat diet-induced obese mouse model, via major effects on adipose and liver gene expression. Fenretinide was originally developed as a chemotherapeutic agent, has been widely studied in clinical trials of breast cancer chemoprevention⁽³⁶⁾. Fenretinide can strongly stimulate expression of classic RA-induced targets, decrease levels of serum retinol binding protein. inhibit adipocyte differentiation and decrease expression of the adipocyte secreted satiety hormone leptin (14,37-39). These effects are in contrast to the subtle effects of specifically altering RBP4 levels in the whole body or in tissue-specific manner, which as mentioned earlier, do not lead to alterations in hepatic retinoid levels or classic RA-induced target genes that are involved in the tight control of retinoid homeostasis.

Fenretinide also has effects independent of RA-signalling due to its chemical structure (addition of a bulky hydroxyphenyl group compared with RA). These effects have been attributed to the inhibition of ceramide biosynthesis, a sphingolipid that has been implicated in the pathogenesis of insulin resistance^(40,41). An excess supply of NEFA such as palmitate with a high-fat, high-sugar diet and/or obesity are thought to lead to increased ceramide production, which can impair mitochondrial function at the level of metabolic flux via fatty-acid β -oxidation and the tricarboxylic acid cycle^(42,43). This impairment is evident with an accumulation of acylcarnitines (fatty-acid β -oxidation intermediates) and tricarboxylic acid cycle intermediates. In mice, Fenretinide treatment completely normalised the accumulation of these intermediates in adipose tissue despite these mice continuing to feed a high-fat, high-sugar diet⁽⁴¹⁾.

Ceramides can also inhibit insulin-signalling by directly increasing the phosphatase activity towards protein kinase B/Akt⁽⁴⁴⁾. Thus, probably through its effect on ceramides, Fenretinide can increase protein kinase B/Akt phosphorylation in adipocytes, even in the absence of insulin⁽³⁸⁾. Moreover, the primary oxidised catabolite of Fenretinide, which is a very poor inducer of RA-signalling, can also inhibit the enzyme responsible for the final step of ceramide synthesis and so similarly to Fenretinide, can also increase protein kinase B/Akt phosphorylation in adipocytes⁽⁴¹⁾.

Fibroblast growth factor 21: a new physiological target of vitamin A/retinoic acid

Most recently, Fenretinide and RA have been reported to repress the levels of the liver derived hormone fibroblast growth factor (FGF) 21⁽⁴⁵⁾. Hepatic Fgf21 expression and circulating levels of FGF21 are increased by stimuli such as fasting, protein/amino acid restriction and fructose ingestion via a range of transcription factors, including PPARα and carbohydrate response-element binding protein^(46,47). Thus, FGF21 has emerged as an important beneficial regulator of glucose and lipid homeostasis but its levels are also abnormally increased in insulinresistant states in rodents and human subjects. Interestingly, Fenretinide can normalise elevated levels of FGF21 in both high-fat diet-induced obese mice and in genetically obese-diabetic mice in association with the wide-range of beneficial effects on metabolic parameters reported previously and summarised earlier (Fig. 2)⁽⁴⁵⁾. Furthermore, diverse treatments to improve glycaemic control in human subjects can decrease elevated levels of FGF21 e.g. with metformin, rosiglitazone. insulin, insulinotropic agents, bariatric surgery, lifestyle modification, fish oil supplements or exercise (48-54). Since Fenretinide has been widely studied as a cancer therapeutic due to its favourable toxicological profile and is undergoing phase II clinical trials for treatment of insulin resistance in obese human subjects with hepatic steatosis, these new findings should be of considerable interest to the field of nutrition, obesity and diabetes research.

Conclusions

There is now considerable evidence to implicate vitamin A homeostasis in the regulation of body fat and blood glucose levels. It appears that altered levels of circulating retinol (bound to RBP) has a completely distinct role compared with RA, in regulating endocrine hormones and gene transcription programmes to alter lipid and glucose metabolism. Powerful new approaches combining the techniques of phenotyping of transgenic mice with



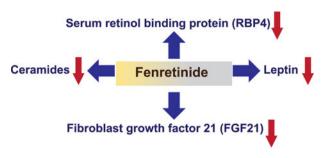


Fig. 2. (Colour online) Identified targets of synthetic retinoid Fenretinide via classic nuclear hormone signalling or via non-nuclear protein interaction. Fenretinide was originally found to decrease levels of serum retinol binding protein (RBP) 4 in cancer chemoprevention trials and in models of obesity and insulin resistance^(14,36,37). Fenretinide and retinoic acid (RA) can decrease expression of the adipocyte secreted satiety hormone leptin in cells and *in vivo*^(38,39) and repress levels of the liver derived hormone fibroblast growth factor (FGF) 21⁽⁴⁵⁾. Independently of RA-signalling, Fenretinide can also inhibit the enzyme responsible for the final step of ceramide synthesis^(40,41,45).

metabolomics, lipidomics and next-generation DNA sequencing to profile epigenomic changes and nuclear receptor occupancy across the genome will aid us to understand more about these interactions in the future. Moreover, these approaches are likely necessary to understand more about the potential of phytochemical and pharmaceutical strategies to treat obesity and type 2 diabetes in preclinical models and to be able to translate positive findings to human subjects.

Acknowledgements

The author would like to thank M. Delibegovic and P. McCaffery (both University of Aberdeen) for their helpful comments and proof reading in preparation of this review.

Financial Support

Work in the author's laboratory was supported by the British Heart Foundation Intermediate Basic Research Fellowship FS/09/026, The Royal Society (of London) grant, Tenovus Scotland grants G10/04 and G14/14 to N. Mody, University of Aberdeen Centre for Genome Enabled Biology and Medicine (CGEBM) PhD studentship to Nicola Morrice and Biotechnology and Biological Sciences Research Council (BBSRC) studentship to George D. Mcilroy.

Conflict of Interest

None.

Authorship

The author was solely responsible for all aspects of preparation of the paper.

References

- Lanska DJ (2010) Chapter 29: historical aspects of the major neurological vitamin deficiency disorders: overview and fat-soluble vitamin A. Handb Clin Neurol 95, 435–444.
- Voss HE (1956) Nicolai I. Lunin, 1853–1937; a biographical assay. J Am Diet Assoc 32, 317–320.
- 3. Wolf G (1996) A history of vitamin A and retinoids. *FASEB J* **10**, 1102–1107.
- Grune T, Lietz G, Palou A et al. (2010) Beta-carotene is an important vitamin A source for humans. J Nutr 140, 2268S–2285S.
- 5. Shirakami Y, Lee S, Clugston RD *et al.* (2012) Hepatic metabolism of retinoids and disease associations. *Biochimica et Biophysica Acta (BBA) Mol Cell Biol Lipids* **1821**, 124–136.
- 6. Jackson HA & Sheehan AH (2005) Effect of vitamin A on fracture risk. *Ann Pharmacother* **39**, 2086–2090.
- McGrane MM (2007) Vitamin A regulation of gene expression: molecular mechanism of a prototype gene. *J Nutr Biochem* 18, 497–508.
- 8. Bastien J & Rochette-Egly C (2004) Nuclear retinoid receptors and the transcription of retinoid-target genes. *Gene* **328**, 1–16.
- 9. Kiser PD, Golczak M, Maeda A *et al.* (2012) Key enzymes of the retinoid (visual) cycle in vertebrate retina. *Biochimica et Biophysica Acta* (*BBA*) *Mol Cell Biol Lipids* **1821**, 137–151.
- 10. Evans RM & Mangelsdorf DJ (2014) Nuclear receptors, RXR, and the big bang. *Cell* **157**, 255–266.
- 11. Basualdo CG, Wein EE & Basu TK (1997) Vitamin A (retinol) status of first nation adults with non-insulindependent diabetes mellitus. *J Am Coll Nutr* **16**, 39–45.
- 12. Basu TK & Basualdo C (1997) Vitamin A homeostasis and diabetes mellitus. *Nutrition* **13**, 804–806.
- 13. Abahusain MA, Wright J, Dickerson JW *et al.* (1999) Retinol, alpha-tocopherol and carotenoids in diabetes. *Eur J Clin Nutr* **53**, 630–635.
- 14. Yang Q, Graham TE, Mody N *et al.* (2005) Serum retinol binding protein 4 contributes to insulin resistance in obesity and type 2 diabetes. *Nature* **436**, 356–362.
- 15. Kotnik P, Fischer-Posovszky P & Wabitsch M (2011) RBP4: a controversial adipokine. *Eur J Endocrinol* **165**, 703–711.
- Quadro L, Blaner WS, Salchow DJ et al. (1999) Impaired retinal function and vitamin A availability in mice lacking retinol-binding protein. Embo J 18, 4633–4644.
- 17. Quadro L, Blaner WS, Hamberger L *et al.* (2002) Muscle expression of human retinol-binding protein (RBP). Suppression of the visual defect of RBP knockout mice. *J Biol Chem* **277**, 30191–30197.
- 18. Lee SA, Yuen JJ, Jiang H *et al.* (2016) Adipocyte-specific overexpression of retinol-binding protein 4 causes hepatic steatosis in mice. *Hepatology* **64**, 1534–1546.
- 19. Thompson SJ, Sargsyan A, Lee SA *et al.* (2017) Hepatocytes are the principal source of circulating RBP4 in mice. *Diabetes* **66**, 58–63.
- 20. Noy N (2016) Vitamin A in regulation of insulin responsiveness: mini review. *Proc Nutr Soc* **75**, 212–215.
- 21. Shin DJ, Odom DP, Scribner KB *et al.* (2002) Retinoid regulation of the phosphoenolpyruvate carboxykinase gene in liver. *Mol Cell Endocrinol* **195**, 39–54.
- Kiefer FW, Orasanu G, Nallamshetty S et al. (2012) Retinaldehyde dehydrogenase 1 coordinates hepatic gluconeogenesis and lipid metabolism. Endocrinology 153, 3089– 3099.
- 23. Alvarez R, de Andres J, Yubero P et al. (1995) A novel regulatory pathway of brown fat thermogenesis. Retinoic



acid is a transcriptional activator of the mitochondrial uncoupling protein gene. *J Biol Chem* **270**, 5666–5673.

- 24. Puigserver P, Vazquez F, Bonet ML *et al.* (1996) *In vitro* and *in vivo* induction of brown adipocyte uncoupling protein (thermogenin) by retinoic acid. *Biochem J* **317**, Pt 3, 827–833.
- Kumar MV & Scarpace PJ (1998) Differential effects of retinoic acid on uncoupling protein-1 and leptin gene expression. J Endocrinol 157, 237–243.
- Kumar MV, Sunvold GD & Scarpace PJ (1999). Dietary vitamin A supplementation in rats: suppression of leptin and induction of UCP1 mRNA. J Lipid Res 40, 824–829.
- Murray T & Russell TR (1980) Inhibition of adipose conversion in 3T3-L2 cells by retinoic acid. *J Supramol Struct* 14, 255–266.
- Sato M, Hiragun A & Mitsui H (1980) Preadipocytes possess cellular retinoid binding proteins and their differentiation is inhibited by retinoids. *Biochem Biophys Res* Commun 95, 1839–1845.
- Xue JC, Schwarz EJ, Chawla A et al. (1996) Distinct stages in adipogenesis revealed by retinoid inhibition of differentiation after induction of PPARgamma. Mol Cell Biol 16, 1567–1575.
- Schwarz EJ, Reginato MJ, Shao D et al. (1997) Retinoic acid blocks adipogenesis by inhibiting C/EBPbetamediated transcription. Mol Cell Biol 17, 1552–1561.
- Berry DC & Noy N (2009) All-trans-retinoic acid represses obesity and insulin resistance by activating both peroxisome proliferation-activated receptor beta/delta and retinoic acid receptor. *Mol Cell Biol* 29, 3286–3296.
- 32. Bershad S, Rubinstein A, Paterniti JR *et al.* (1985) Changes in plasma lipids and lipoproteins during isotretinoin therapy for acne. *N Engl J Med* **313**, 981–985.
- Standeven AM, Beard RL, Johnson AT et al. (1996) Retinoid-induced hypertriglyceridemia in rats is mediated by retinoic acid receptors. Fundam Appl Toxicol 33, 264–271.
- 34. Standeven AM, Thacher SM, Yuan YD *et al.* (2001) Retinoid X receptor agonist elevation of serum triglycerides in rats by potentiation of retinoic acid receptor agonist induction or by action as single agents. *Biochem Pharmacol* **62**, 1501–1509.
- 35. Bonet ML, Ribot J & Palou A (2012) Lipid metabolism in mammalian tissues and its control by retinoic acid. *Biochimica et Biophysica Acta (BBA) Mol Cell Biol Lipids* 1821, 177–189.
- 36. Mody N & Mcilroy GD (2014). The mechanisms of Fenretinide-mediated anti-cancer activity and prevention of obesity and type-2 diabetes. *Biochem Pharmacol* 91, 277–286.
- 37. Preitner F, Mody N, Graham TE *et al.* (2009) Long-term Fenretinide treatment prevents high-fat diet-induced obesity, insulin resistance, and hepatic steatosis. *Am J Physiol Endocrinol Metab* **297**, E1420–E1429.
- 38. Mcilroy GD, Delibegovic M, Owen C *et al.* (2013) Fenretinide treatment prevents diet-induced obesity in association with major alterations in retinoid homeostatic gene expression in adipose, liver, and hypothalamus. *Diabetes* **62**, 825–836.

- 39. Shearer KD, Morrice N, Henderson C *et al.* (2015) Fenretinide prevents obesity in aged female mice in association with increased retinoid and estrogen signaling. *Obesity (Silver Spring)* **23**, 1655–1662.
- Bikman BT, Guan Y, Shui G et al. (2012) Fenretinide prevents lipid-induced insulin resistance by blocking ceramide biosynthesis. J Biol Chem 287, 17426–17437.
- Mcilroy GD, Tammireddy SR, Maskrey BH et al. (2016)
 Fenretinide mediated retinoic acid receptor signalling and
 inhibition of ceramide biosynthesis regulates adipogenesis,
 lipid accumulation, mitochondrial function and nutrient
 stress signalling in adipocytes and adipose tissue. Biochem
 Pharmacol 100, 86–97.
- Rodriguez-Cuenca S, Barbarroja N & Vidal-Puig A (2015)
 Dihydroceramide desaturase 1, the gatekeeper of ceramide induced lipotoxicity. *Biochim Biophys Acta* 1851, 40–50.
- 43. Chaurasia B & Summers SA (2015) Ceramides lipotoxic inducers of metabolic disorders. *Trends Endocrinol Metab* **26**, 538–550.
- 44. Stratford S, Hoehn KL, Liu F *et al.* (2004) Regulation of insulin action by ceramide: dual mechanisms linking ceramide accumulation to the inhibition of Akt/protein kinase B. *J Biol Chem* **279**, 36608–36615.
- 45. Morrice N, Mcilroy GD, Tammireddy SR *et al.* (2017) Elevated Fibroblast growth factor 21 (FGF21) in obese, insulin resistant states is normalised by the synthetic retinoid Fenretinide in mice. *Sci Rep* 7, 43782.
- Giralt M, Gavalda-Navarro A & Villarroya F (2015)
 Fibroblast growth factor-21, energy balance and obesity.
 Mol Cell Endocrinol 418, 66–73.
- 47. Fisher FM & Maratos-Flier E (2016) Understanding the physiology of FGF21. *Annu Rev Physiol* **78**, 223–241.
- 48. Fan H, Sun X, Zhang H *et al.* (2016) Effect of metformin on fibroblast growth factor-21 levels in patients with newly diagnosed type 2 diabetes. *Diabetes Technol Ther* **18**, 120–126.
- Srinivasa S, Wong K, Fitch KV et al. (2015) Effects of lifestyle modification and metformin on irisin and FGF21 among HIV-infected subjects with the metabolic syndrome. Clin Endocrinol (Oxf) 82, 678–685.
- Li K, Li L, Yang M et al. (2009) Effects of rosiglitazone on fasting plasma fibroblast growth factor-21 levels in patients with type 2 diabetes mellitus. Eur J Endocrinol 161, 391–395.
- 51. Yang M, Dong J, Liu H *et al.* (2011) Effects of short-term continuous subcutaneous insulin infusion on fasting plasma fibroblast growth factor-21 levels in patients with newly diagnosed type 2 diabetes mellitus. *PLoS ONE* **6**, e26359.
- 52. Haluzikova D, Lacinova Z, Kavalkova P *et al.* (2013) Laparoscopic sleeve gastrectomy differentially affects serum concentrations of FGF-19 and FGF-21 in morbidly obese subjects. *Obesity (Silver Spring)* **21**, 1335–1342.
- Taniguchi H, Tanisawa K, Sun X et al. (2016) Endurance exercise reduces hepatic fat content and serum fibroblast growth factor 21 levels in elderly men. J Clin Endocrinol Metab 101, 191–198.
- 54. Gomez-Ambrosi J, Gallego-Escuredo JM, Catalan V et al. (2017) FGF19 and FGF21 serum concentrations in human obesity and type 2 diabetes behave differently after diet-or surgically-induced weight loss. Clin Nutr 36, 861–868.

