

Heliport-associated Nosocomial Mucormycoses

To the Editor:

Nosocomial fungal infections have been associated with construction in or near hospitals¹⁻⁴ and contamination of ventilation systems,³ air conditioner filters,⁵ fireproofing material,⁶ and Elastoplast® dressings.⁷⁻⁸ We report the epidemiologic and microbiologic investigation of an apparent heliport-associated outbreak of nosocomial fungal infections.

Two cases of disseminated mucormycosis and one case of rhino cerebral mucormycosis were recognized in three newly diagnosed leukemia patients in a 162-bed pediatric teaching hospital between March and September 1985. The intervals between diagnosis of leukemia and onset of mucormycosis for the three cases were 12, 14, and 16 days. Each of the patients had been hospitalized for between six and nine days in the period between diagnosis of leukemia and onset of mucormycosis.

Review of microbiology, pathology, and nosocomial infection records from 1978 to 1985 revealed no cases of mucormycosis prior to March 1985. The three cases diagnosed in the remainder of 1985 represented a statistically significant aberration ($p = .004$, Poisson heterogeneity test). The number of cases of mucormycosis per newly diagnosed acute leukemia patient in 1985 was 3 per 60, versus 0 per 108 in the years 1982 to 1984 ($p = .08$, two-tailed Fisher's exact test). In the period of risk for nosocomial mucormycosis (January through August 1985), leukemia was newly diagnosed in 38 patients, compared with 30 patients in the same period of 1984 ($p = .33$, Poisson heterogeneity test).

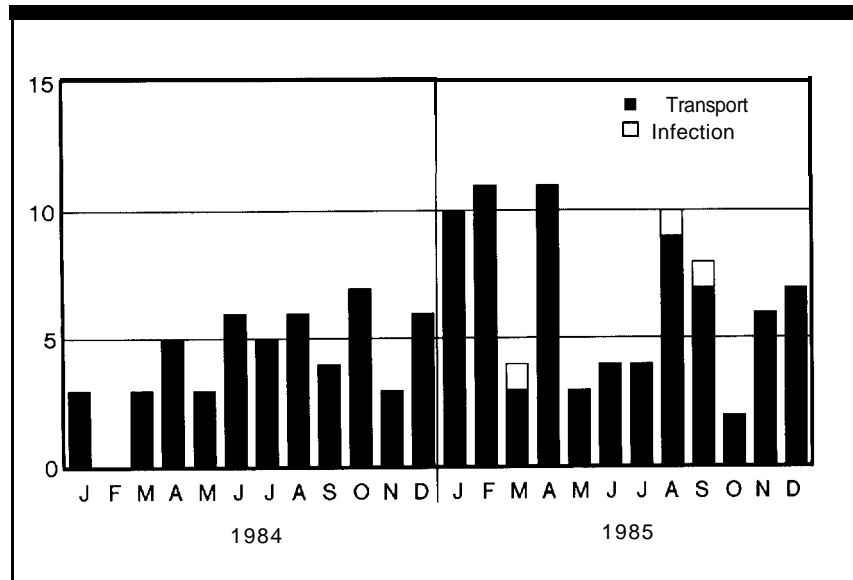


FIGURE. Helicopter transports and cases of mucormycosis infection in pediatric oncology patients, 1984 through 1985. There were 51 transports in 1984, compared with 77 in 1985. Infections occurred after periods of particularly heavy use of the heliport.

The ventilation pathway for the oncology unit was traced via hospital blueprints from outside the building to the patient rooms. The air that ventilates the oncology unit is transferred to the unit via an air handler whose intake vents are 20 yards from the hospital heliport. The heliport is elevated on columns and braces and sits 6.5 feet above the roof of the hospital's fifth floor. The roof area comprises 10,000 square feet, and at the time of the outbreak, was paved with a 1.5-inch-deep layer of gravel and sand. The air intake vents are 18 inches above the roof. When a helicopter was not present and the wind velocity was five or less miles per hour, air was "inhaled" into the air handler at a minimum speed of eight miles per hour (measured with an anemometer). When a helicopter landed or took off, air, along with dust and small particles from the roof, was blown into the air handler at speeds of 70 or more miles per hour. After the air enters the air handler through the intake vents, it passes through a prefilter, which filters approximately 40% of particulate matter, and then through a

bag filter, which traps approximately 90% of particles. The air is then delivered to the patient care units at a speed of one foot per second via air ducts that enter above the ceiling panels into patient rooms.

The heliport was constructed in mid-1983. In 1985 there were 77 helicopter transports, compared with 51 in 1984 ($p = .02$, Poisson heterogeneity test). During the period of risk (January through August 1985) there were 55 transports, compared with 31 during the same time period in 1984 ($p = .01$, Poisson heterogeneity test). The three cases of mucormycosis occurred during or following times of increased heliport use (Figure). No major hospital construction occurred in 1985.

Air sampling and culturing was performed with a centrifugal air sampler (Biotest Diagnostics, Fairfield, New Jersey) containing rose bengal agar. Thirty room air samples from nine patient rooms in the oncology unit obtained over five different days during a three-month period (October through December 1985) yielded a mean of 3.2 colony-forming units (CFU)

per m³ of zygomycetes. Eleven air samples from above the false ceiling panels in three patient rooms were similarly cultured on three days in October through November 1985, revealing a mean of 22 CFU per m³ of zygomycetes. Cultures obtained on two separate days (October and December, 1985) of the gravel that covered the roof near the air intake vents of the air handler that served the oncology unit had a mean of 1.3 CFU per 50 g. Cultures of the prefilters and bag filters inside the air handler yielded more than 50 CFU per cm² and 4 CFU per cm² of zygomycetes, respectively.

Based on these investigations, the hospital implemented the following control measures at the end of December 1985: high efficiency particulate air (HEPA) filters were used in the oncology patient rooms; and the gravel under the helipad was removed and replaced with an impervious neoprene roofing material. Subsequent cultures of air samples from three oncology patient rooms on one day in February 1986 revealed no zygomycetes, and cultures of air samples from one patient room in April 1986 contained a mean of 0.5 CFU per m³ of zygomycetes. No further cases of confirmed mucormycosis in oncology patients have been reported as of December 1991.

The clustering of cases of mucormycosis within a limited period of time and the short interval between development of immunosuppression and occur-

rence of mucormycosis in all three patients suggested that our patients might have been exposed to a common environment contaminated unusually heavily with zygomycetes. Because all three patients had been hospitalized for periods of six to nine days during the two weeks between diagnosis of leukemia and onset of mucormycosis, the hospital was a likely site of exposure.

The findings of our investigation are consistent with the hypothesis that the gravel-paved roof beneath the heliport was a primary reservoir of zygomycetes, with the heavily colonized prefilters serving as a secondary source. Fungal spores were likely deposited from the gravel into the air handler by turbulence created by frequent helicopter landings and takeoffs. The high velocity winds also could have liberated spores from the heavily colonized air handler prefilters into the air supply of the oncology unit, thus exposing our patients to a large inoculum of zygomycetes. The increased use of the heliport during the time of the patients' hospitalizations supports this hypothesis. That zygomycetes were infrequently recovered from room air after the roof was paved and HEPA filters were used, despite continuing frequent use of the heliport, supports the efficacy of these interventions. It is also noteworthy that no new cases of mucormycoses have been recognized in our oncology patients.

We suggest that helicopter-created dust and proximity of heli-

ports to ventilation systems are potential environmental risk factors for nosocomial fungal infections.

Mark J. Abzug, MD

Sherry Gardner, RN, BA

Mary P. Glode, MD

Margrete Cymanski, MS (ASCP)

Martha H. Roe, MS (ASCP)

Lorrie F. Odom, MD

The University of Colorado

School of Medicine

Denver, Colorado

REFERENCES

1. Sarubbi FA Jr, Dopf HB, Wilson MB, et al. Increased recovery of *Aspergillus flavus* from respiratory specimens during hospital construction. *Am Rev Respir Dis.* 1982;125:33-38.
2. Arnow PM, Anderson RL, Mainous PHD, et al. Pulmonary *Aspergillus* during hospital renovation. *Am Rev Respir Dis.* 1978;118:49-53.
3. Lention JR, Rosenkranz MA, Michaels JA, et al. Nosocomial aspergillosis: a retrospective review of airborne disease secondary to road construction and contaminated air conditioners. *Am J Epidemiol.* 1982;116:430-437.
4. Grossman ME, Fithian EC, Behrens C, et al. Primary cutaneous aspergillosis in six leukemic children. *J Am Acad Dermatol.* 1985;12:313-318.
5. del Palacio Hernanz A, Feres J, Larregla Garraus S, et al. Nosocomial infection by *Rhizomucor pusillus* in a clinical haematology unit. *J Hosp Infect.* 1983;4:45-49.
6. Aisner J, Schimpff SC, Bennett JE, et al. *Aspergillus* infections in cancer patients: association with fireproofing materials in a new hospital. *JAMA.* 1976;235:411-412.
7. Mead JH, Lupton GP, Dillavon CL, et al. Cutaneous *Rhizopus* infection: occurrence as a postoperative complication associated with an elasticized adhesive dressing. *JAMA.* 1979;242:272-274.
8. Dennis JE, Rhodes KH, Cooney DR, Roberts GD. Nosocomial *Rhizopus* infection (zygomycosis) in children. *J Pediatr.* 1980;96:824-828.



The Society for Hospital Epidemiology of America

Position Search Announcement for

Chief Editor, Infection Control and Hospital Epidemiology
The Official Journal of the Society for Hospital Epidemiology of America

With the decision of founding Chief Editor, Dr. Richard Wenzel, to move on to other activities, the President and Board of the Society for Hospital Epidemiology of America announce the formation of a Search Committee to review candidates and make a recommendation to the Publications Committee and the Board for a successor. All members of SHEA who might be interested in the editorship are urged to submit the following:

- 1) a copy of your CV and bibliography
- 2) a letter of no more than two pages containing:
 - your proposal on how to further strengthen and improve the journal
 - the potential for local editorial board support and the recruitment of a managing editor to work under your direction
 - your experience to date in journal editing or reviewing journal articles.

An annual budget is provided to help you accomplish your work. Send your letter and CV to:

Chair, Chief Editor Infection Control and
Hospital Epidemiology Search Committee
SHEA National Office
875 Kings Highway
Suite 200
West Deptford NJ 08096



**This
publication is
available in
microform.**

University Microfilms
International reproduces this publication
in microform: microfiche and 16mm or 35mm film.
For information about this publication or any of the
more than 13,000 titles we offer, complete and mail
the coupon to: University Microfilms International,
300 N. Zeeb Road, Ann Arbor, MI 48106. Call us
toll-free for an immediate response: 800-521-3044.
Or call collect in Michigan, Alaska and Hawaii:
313-761-4700.

**University
Microfilms
International**

Please send information about these titles:

Name _____
Company/Institution _____
Address _____
City _____ State _____ Zip _____
Phone () _____