Response to foot-and-mouth disease vaccines in newborn calves. Influence of age, colostral antibodies and adjuvants*

By A. M. SADIR, A. A. SCHUDEL, O. LAPORTE

Department of Virology, Centro de Investigaciones en Ciencias Veterinarias, Instituto Nacional de Tecnología Agropecuaria, Castelar

M. BRAUN

Cátedra de Inmunologia, Facultad de Ciencias Veterinarias, Universidad de Buenos Aires, Argentina

AND R. A. MARGNI

Cátedra de Inmunología, Facultad de Farmacia y Bioquímica, Universidad de Buenos Aires, Argentina

(Accepted 19 May 1987)

SUMMARY

Oil-emulsified (OE) and aqueous (Aq) vaccines were prepared with the same batch of inactivated A₂₄ 8345 foot and mouth disease virus (FMDV). Calves born to vaccinated dams did not respond to the Aq vaccine 30 or 90 days post partum. When the OE vaccine was used on a similar group of calves, no responses were elicited up to 21 days post partum. However, calves 30 or more days old responded like adult cattle to the OE vaccine. When the OE vaccine was used in colostral antibody-free calves 3–30 days old, all animals showed good antibody responses but, in calves vaccinated 3 or 7 days post partum, antibodies were detectable only after a considerable period of time. Our results show that both passively acquired colostral antibodies and age are important in the response of very young calves to FMDV oil vaccines. From a practical point of view, in endemic areas where adult cattle are periodically vaccinated, vaccination of calves between 30 and 60 days post partum with OE vaccines would lead to high levels of herd protection.

INTRODUCTION

Foot and mouth disease virus (FMDV) is the aetiological agent of an acute febrile disease that causes enormous economic losses in many countries of the world. In endemic areas inactivated aqueous (Aq) vaccines with aluminium hydroxide and saponin adjuvants are often used but these vaccines induce protection for very short periods only, and repeated vaccinations at 4-month intervals are necessary (INTA & PIADC, 1977; Rivenson et al. 1982). One of the

* This work was partly presented at the Winter Meeting of the Sociedad Argentina de Inmunología, Rosario, Argentina, June 1985; abstract in Acta Physiologica Pharmacologica Latinoamericana 36, 183 (1986).

principal problems in mass immunization against FMDV is inducing protection of young calves, since it has been shown that newborn calves with maternal antibodies give very poor or no response to aqueous (Aq) FMDV vaccines (Nicholls *et al.* 1984), and that epidemic waves start in many countries with infection of these unprotected young calves (COSALFA, 1981).

The lack of response of newborn calves may be due to interference by passively acquired colostral antibodies (CAb) (Brun et al. 1974; Graves, 1963; Nicholls et al. 1984). Several workers have studied the immune response of young calves against FMDV and have shown that, using Aq-FMDV vaccines on the offspring from immunized dams, the latter not only fail to respond but their serum antibody titres drop after vaccination. Hence vaccination with Aq-FMDV has no positive effect on protection up to 5 months of age (Graves, 1963; Nicholls et al. 1984). Although Nicholls et al. (1985) have shown that, in the absence of CAb 1-week-old calves responded to FMDV vaccination as well as adult cattle, preliminary experiments in our Institute (Sadir, A. M. et al. unpublished results) showed very poor responses (log₁₀ neutralizing titres ≤ 1.67) to a commercial FMDV trivalent Aq vaccine in calves of less than 30 days of age, even if they were born to nonvaccinated dams. Recently, oil-emulsion (OE) vaccines have been introduced and it has been demonstrated, both in laboratory and in controlled field conditions, that they confer a much longer lasting protection in cattle (INTA & PIADC, 1977; Rivenson et al. 1982). Furthermore, it has been shown that, in other species, vaccination of newborn animals with OE-FMDV vaccines induces immune responses even in the presence of colostral antibody (CAb) (Cunliffe & Graves, 1970; Morgan & McKercher, 1979), although some contradictory evidence has recently appeared (Francis & Black, 1986).

The aim of our work was to study the immune response of new-born calves, born to vaccinated and nonvaccinated dams, against FMDV vaccines in order to determine the best immunization programme in endemic areas. We here show that, although both age and the presence of CAb are important in the response to FMDV vaccine, when using OE vaccines on calves 30 or more days old, very good responses are obtained, even in the presence of CAb.

MATERIALS AND METHODS

Animals

Twenty-six calves of the Holando-Argentino breed, aged 1-90 days, born from dams vaccinated and nonvaccinated against FMDV were used. Offspring from nonvaccinated dams were born and kept during the whole experiment in a FMDV-free area (Chubut, Argentina).

Calves born to immunized cows were provided by the INTA-Castelar dairy farm. These dams had been revaccinated approximately 70 days before parturition with the OE-FMDV vaccine and had high anti-FMDV antibody titres (Sadir et al. 1984). Virus infection associated antibody (VIA) tests were negative in calves and dams during the entire experimental period (MeVicar & Sutmoller, 1870).

Vaccine

Vaccines were prepared from FMDV, strain A₂₄ 8345. The virus obtained from infected bovine tongue epithelium stocks was passaged seven times in primary

bovine kidney cultures (BK), once in BHK-21 clone 13 cells, and then grown in BHK-21 monolayers in roller bottles. The final viral titre was $10^{7\cdot2}$ 50% tissue culture infectious doses (TCID₅₀) per ml, its complement fixation titre was 16 after inactivation and the 146S concentration was $1\cdot8~\mu g/ml$.

The viral suspension was inactivated with binary 2-bromo-ethyleneimine as described by Bahnemann (1975). Tests for residual live virus in the antigen before emulsification were done in 100 suckling mice. The sterility was checked in anaerobic and Sabouraud culture media.

The OE vaccine was prepared according to the INTA formulation with Marcol 52 (42·5%), Arlacel 83 (6·55%) and Tween 40 (0·95%) using a Silverson type emulsifier (Rivenson *et al.* 1982). The Aq-FMDV control vaccine was produced by adsorption of the same viral suspension on to aluminium hydroxide (40% v/v and $2\cdot5\%$ Al₂O₃) with saponin and merthiclate added to a final concentration of 0·1% and 1/30000 respectively.

Immunization schedules

All calves were vaccinated by subcutaneous inoculation of 3 ml of vaccine per calf. All calves were first bled for serological tests and then vaccinated and revaccinated 30 days post partum (dpp) with 3 ml of OE vaccine. Calves born from nonimmunized dams received the same dose of OE vaccines at days 3, 7, 21 and 39 dpp; some were revaccinated as shown in Table 1.

Serology

Serum samples were obtained 7, 14, 21, 30 and 60 days postvaccination (dpv) and 7, 14, 21, 30, 60, 90, 120, 150 and 180 days post revaccination (dpr).

Neutralizing antibody titres were determined in each sample by the fixed virus-variable serum method, using $100~\mathrm{TCID}_{50}$ per tube. Results are expressed as the \log_{10} dilution of serum protecting half of the monolayers.

Decay rate of colostral antibodies

Three calves, born 60-70 days after the last vaccination of their dams, were studied. Blood samples were periodically collected from 5 to 180 dpp and their neutralizing antibodies measured. Individual slopes were studied by linear regression analysis, and the weighted mean slope was estimated (b = -0.014) in order to determine the half-life of CAb. The experimental error mean square (EEMS) was 0.46.

Challenge

Four calves born to immunized dams, vaccinated at 1, 30, 60 and 90 days of age with OE vaccines and revaccinated 60 days later, were challenged with 10000 50% suckling mouse doses 180 dpr.

RESULTS

Half-life of colostral antibodies

In order to be able to detect active immune responses in newborn calves, the half-life of CAb was first ascertained (Table 2). The mean slope from three

Table 1. Experimental animals and immunization schedules

			Vaccination	Revaccination
Calf	Dam†	Adjuvant	(days post partum)	(days post partum)
1	Vacc.	OE*	6	No
2	Vacc.	OE	13	No
3	Vacc.	OE	13	No
4	Vacc.	OE	18	No
5	Vacc.	OE	4	61
6	Vacc.	OE	5	65
7	Vacc.	OE	5	65
8	Vacc.	OE	5	65
9	Vacc.	OE	10	70
10	Vacc.	OE	16	76
11	Vacc.	OE	21	81
12	Vacc.	\mathbf{OE}	30	90
13	Vacc.	OE	45	105
14	Vacc.	OE	60	120
15	Vacc.	OE	90	150
16	Nonvacc.	OE	7	No
17	Nonvacc.	OE	21	No
18	Nonvacc.	OE	30	No
19	Nonvacc.	OE	3	63
20	Nonvacc.	OE	7	63
21	Nonvacc.	OE	21	81
22	Nonvacc.	OE	30	90
Aq_{1-2}	Vacc.	\mathbf{Aq}	39	90
C_{1-2-3}	Vacc.	_	No	No

^{*} OE, oil-emulsified vaccine.

Table 2. Decline of maternally derived antibodies with time

Calf no.	Calf age (days)	NT*	Slope†
C 1	5	2.30	
	18	1.30	
	25	1.20	-0.028
	55	€0.70	
	178	€0.70	
C 2	11	3.10	
	48	2.10	-0.020
	90	1.50	
C 3	16	2.90	
	23	2.70	
	30	2.50	-0.012
	60	2.30	
	116	1.80	
	170	0.90	

^{*} NT, neutralizing titres (log₁₀).

[†] Vacc, Vaccinated; Nonvacc., Nonvaccinated. ‡ Aq; aqueous vaccine with hydroxysaponin adjuvant.

[†] The weighted mean slope was -0.014.

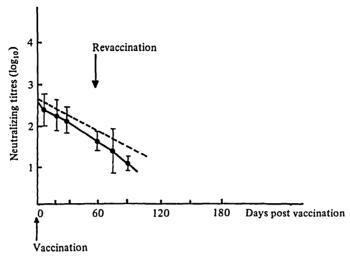


Fig. 1. Response to an aqueous foot-and-mouth disease vaccine in the presence of colostral antibodies. Calves were vaccinated when 30 days old and revaccinated 60 days later and their neutralizing antibodies tested as described in material and methods. (---): theoretical half-life slope of passively acquired antibodies; (---): actual titres in vaccinated calves.

non-immunized calves had a value of b = -0.140, giving a half-life value of 21.5 days, coincident with other data in the literature (Nicholls *et al.* 1984).

Effect of Aq-vaccine in calves born to vaccinated dams

As may be seen in Fig. 1, the antibody titres in calves with CAb were lower than expected from their theoretical half-life slope; furthermore, even when the calves were revaccinated 60 dpv, good responses could not be elicited.

Effect of OE-FMDV vaccines in calves born from vaccinated dams

Calves vaccinated 1 to 21 dpp had mean initial CAb \log_{10} titres of 3.18 ± 0.66 . At 60 dpv, their titres were similar or lower than expected from the theoretical half-life slope. In 4 calves which were not revaccinated no antibody rise could be detected up to 120 dpv (Fig. 2). In 5 out of 7 calves, revaccination induced a moderate antibody response (Fig. 2); the oldest calf of this group gave a good response, while one animal gave a very low and protracted response to revaccination.

Calves vaccinated 30 or more dpp had a mean prevaccination antibody titre of 1.67 ± 0.7 ; at 60 dpv their titre had risen to 2.17 ± 0.37 showing that vaccination at this age was effective; revaccination induced a further rise in antibody titres in all animals within this group (Fig. 3).

Effect of OE-FMDV vaccine on calves born from non-vaccinated dams

Calves vaccinated 21 and 30 dpp (Fig. 4, C and D) gave good antibody responses detectable 7 or 14 dpv. Calves vaccinated 3 and 7 dpp (Fig 4A and B) also gave good antibody responses, but with a lag of 30 to 60 dpv. Non-revaccinated calves

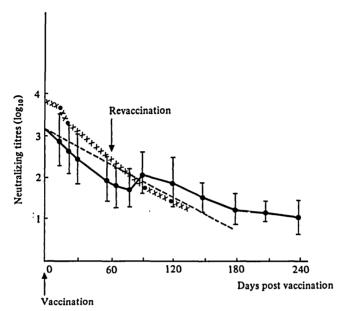


Fig. 2. Response to an oil-emulsified foot-and-mouth disease vaccine in calves immunized 1-21 days post partum in the presence of colostral antibodies. Calves born to vaccinated mothers were vaccinated 1-21 days post partum and revaccinated 60 days later, and their neutralizing antibodies tested as described in Materials and Methods. (---): theoretical half-life slope of passively acquired antibodies; (--): actual titres in vaccinated and revaccinated calves; (+++): actual titres in singly vaccinated calves.

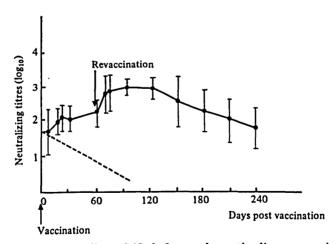


Fig. 3. Response to an oil-emulsified foot-and-mouth disease vaccine in calves immunized 30-90 days post partum, in the presence of colostral antibodies. Calves born to immunized dams were vaccinated 30 or more days post partum, revaccinated 60 days later and their neutralizing antibodies tested as described in Materials and Methods. (---): theoretical half-life slope of colostral antibodies. (---): actual titres of vaccinated calves.

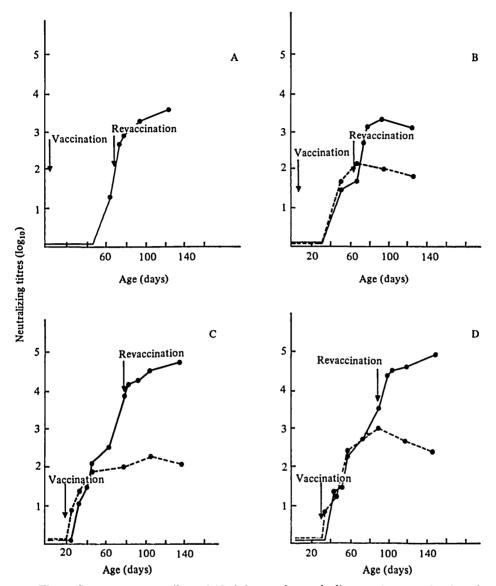


Fig. 4. Response to an oil-emulsified foot-and-mouth disease virus vaccine in calves born to nonvaccinated dams. Calves born in a disease-free area were vaccinated with an oil-emulsified vaccine at 3 (A), 7 (B), 21 (C) and 30 (D) days post partum; some of them (—) were revaccinated 60 days later; others (——) were not revaccinated.

showed persistent antibody titres up to 120 dpv; revaccination (Fig. 4) always led to enhanced responses.

Protection against viral challenge

Four calves were challenged 180 dpr (Table 3). The youngest developed local lesions, followed by production of virus infection-associated (VIA) antibodies but was protected against generalized infection. All the others were totally protected, and remained VIA-negative.

			Lesions				
Calf	Vaccination	Revaccination	Challenge	Titre*	VIA†	Local	Podal
e	1	61	241	1.2	+	+	_
1	30	90	270	1.90	_	-	_
n	60	120	300	3.30	_		_
o	90	150	330	1.70	_	-	-

Table 3. Protection against viral challenge

DISCUSSION

When studying the immune response of fetal and newborn calves against several antigens, Smith & Ingram (1965) showed that it depends both on the kind of antigen used and on the presence or absence of CAb. Husband & Lascelles (1975) showed that the most important cause of immunological unresponsiveness in neonates is not so much the immaturity of the lymphoid system as the effects of CAb, and the lack of response of young calves to FMDV vaccination has been attributed to the presence of CAb (Graves 1963; Nicholls *et al.* 1984).

Nicholls et al. (1985) reported that 1-week-old newborn calves responded as well as adult cattle to Aq-vaccines in the abscence of CAb. However, using a trivalent commercial Aq-vaccine, we have observed very poor or no responses in CAb-free calves from Patagonia up to 30 days of age (Sadir et al. to be published). Other authors have also reported that young cattle need higher doses than adults to produce comparable antibody responses (Brooksby, 1974). The reason for this difference in results is unknown but may be due to differences in the vaccine formulation or content, in the cattle population used or in other factors. Moreover, we here show that, although vaccination with OE-FMDV vaccines 3 and 7 dpp did elicit antibody responses, these were detectable only after a considerable period of time. In fact, the calves that were vaccinated 14 dpp had detectable antibodies before those that had been immunized 3 and 7 dpp. Although the number of animals is small, these and the data referred to above suggest that, at least in our conditions and during the first days of life, calves may be poor responders to FMDV vaccines in the absence of CAb; this unresponsiveness would have to be overcome before a positive reaction may be elicited. Using OE vaccines, the antigen is slowly released (Osebold, 1982) and may be presented to the immune system once it is able to respond. It should be noted that vaccination of 1-weekold pigs devoid of CAb with OE-FMDV vaccine led to poor immunity against challenge at 6 or 7 months of age (Francis & Black, 1986).

Although newborn calves are able to react to many antigens, they respond poorly to others such as polysaccharides, even if deprived of CAb (Smith & Ingram, 1965). In an experimental murine model, it has been reported that immune responses to FMDV infection may be elicited in the absence of T cells (Borca et al. 1986) and that in congenitally athymic (nu/nu) mice the course of infection is identical to that in their normal nu/+ littermates (Fernández et al.

^{*} Neutralizing antibodies on day of challenge

[†] VIA: virus infection-associated antigen. All animals were negative before challenge and were tested at 7, 21 and 30 days post challenge.

1986). This suggests that, in certain conditions, FMDV may induce non T-dependent reactions in mice, even if indirect evidence, for example the IgM to IgG switch, does point to helper T-cell involvement in anti-FMDV reactions. The delay in response to FMDV vaccines in newborn calves may also be due to an atypical response to these virus antigens in young cattle. It should be noted that functional studies at birth show that the stimulation of lymphocytes with mitogens is depressed (Fossum, 1985) – an effect which may be explained by the low numbers of T and B lymphocytes of very young calves. Finally, high perinatal levels of cortical hormones may have immunosuppressive effects in newborn calves (Roth, 1982) and different levels in these hormones may explain the differences between the results obtained by Nicholls et al. (1985) and ourselves.

Our data also show that CAb interfere with the immune response to FMDV vaccines; in fact calves born to vaccinated dams gave no detectable responses to OE vaccines until 21 dpp, but after 30 dpp, the responses to this vaccine are comparable to those of adult cattle. Nicholls et al. (1984) have shown that Aq vaccines should not be used before 5 months of age, since they do not induce antibody responses in calves with high CAb titres. Nevertheless, when very young calves from immunized dams were vaccinated and revaccinated with OE vaccine, good protection was observed, as shown by virus challenge 180 dpr. Once again, this protection may be due to a persistent stimulation of the immune system given by the slow release of antigen from the OE vaccine.

From a practical point of view, in endemic areas, where cattle are vaccinated periodically, CAb can confer a good protection to newborn calves for at least 60 days (Sadir *et al.* 1984). During this period, Aq-FMDV vaccines should not be used, since they lead to a reduction in CAb titre (Nicholls *et al.* 1984), while OE-FMDV vaccines are highly efficient from 30 dpp on, even in the presence of CAb. Vaccination of calves 30–60-days old with OE-FMDV vaccines will then lead to high levels of herd protection.

This work was supported by Grant no. 2109-4311/84 from the Comisión de Investigaciones Científicas de la Provincia de Buenos Aires. Thanks are due to Mrs A. M. Hernández and Mr A. Varone for the technical assistance and L. Marangunich for the statistical analysis. The typing of Mrs M. Sozzi is also acknowledged.

REFERENCES

Bahnemann, H. B. (1975). Binary ethyleneimine as an inactivant for foot-and-mouth disease virus and its application for vaccine production. *Archives of Virology* 47, 47-56.

Brooksby, J. B. (1974). Inmunización del animal joven contra la fiebre aftosa. Boletín del Centro Panamericano de Fiebre Aftosa 13-14, 1-5.

Borca, M. V., Fernández, F. M., Šadir, A. M., Braun, M. & Schudel, A. A. (1986). Immune response to foot-and-mouth diease virus in a murine experimental model: effective thymus-independent primary and secondary reaction. *Immunology* 59, 261-267.

Brun, A., Chappuis, G., Favre, H., Roulet, C. & Terre, J. (1977). Utilisation chez les jeunes bovins du vaccin antiaphteux en adjuvant huileux. Developments in Biological Standardization 35, 117-122.

COSALFA (1981). Comisión Sudamericana para la Lucha Contra la Fiebre Aftosa (1981). Política y Estrategias del Combate de la Fiebre Aftosa en Sudamérica para la década 1981–1990. Río de Janeiro, Brasil: Centro Panamericano de Fiebre Aftosa.

- CUNLIFFE, H. F. & GRAVES, J. H. (1970). Immunologic response of lambs to emulsified foot-and-mouth disease vaccine. Canadian Journal of Comparative Medicine 27, 193-197.
- FERNÁNDEZ, F. M., BORCA, M. V., FONDEVILA, N., MAYO, J. & SCHUDEL, A. A. (1986). Foot-and-mouth disease virus (FMDV) experimental infection: susceptibility and immune response of adult mice. *Veterinary Microbiology* 12, 15-24.
- Fossum, C. (1985). Bovine lymphoid cells and monocytes. Identification and function. Thesis, Uppsala University.
- Francis, M. J. & Black, L. (1986). Response of young pigs to foot-and-mouth disease oil emulsion vaccination in the presence and absence of maternally derived neutralizing antibodies. Research in Veterinary Science 41, 31-39.
- Graves, H. J. (1963). Transfer of neutralizing antibody by colostrum to calves born of vaccinated dams. *Journal of Immunology* 91, 251-256.
- Husband, A. J. & Lascelles, A. K. (1975). Antibody responses to neonatal immunization in calves. Research in Veterinary Science 18, 201-207.
- INTA & PIADC (1977). Instituto Nacional de Tecnología Agropecuaria Plum Island Animal Disease Center. Foot-and-mouth disease: a vaccine study. Development in Biological Standardization 35, 123-133.
- McVicar, J. & Sutmoller, P. (1970). Foot-and-mouth disease: the agar gel diffusion precipitin test for antibody to virus-infection-associated (VIA) antigen as a tool for epizootiologic surveys. *American Journal of Epidemiology* 92, 273-278.
- MORGAN, D. O. & MCKERCHER, P. D. (1979). Immune response of neonatal swine to inactivated foot-and-mouth disease virus vaccine with oil adjuvant. I. Influence of colostral antibody. Presented at the 81st Annual Meeting of the United States Health Association, Minneapolis, Minnesota.
- NICHOLLS, M. J., BLACK, L., RWEYEMAMU, M. M., GENOVESE, J., FERRARI, R., HAMMANT, C. A., DA SILVA, E. & UMEHARA, O. (1984). The effect of maternally derived antibodies on the response of calves to vaccination against foot-and-mouth disease. *Journal of Hygiene* 92, 105–116.
- NICHOLLS, M. J., BLACK, L., RWEYEMAMU, M. M. & GRADWELL, D. V. (1985). Effect of age on response of cattle to vaccination against foot-and-mouth disease. *British Veterinary Journal* 141, 17-26.
- OSEBOLD, J. W. (1982). Mechanism of action by immunologic adjuvants. Journal of the American Veterinary Medical Association 181, 983-987.
- RIVENSON, S., SADIR, A. M., GAGGINO, O. P., MARCOVECCHIO, F. E., ZABAL, O. & LAPORTE, O. (1982). Estudio comparativo en bovinos de dos vacunas antiaftosas: oleosa e hidróxido saponinada. Revista de Medicina Veterinaria 63, 364-370.
- ROTH, J. A. (1982). Effect of glucocorticoids on the bovine immune system. Journal of the American Veterinary Medical Association 180, 894-901.
- Sadir, A. M., Schudel, A. A., Marcovecchio, F. E. & Margni, R. A. (1984). Inmunización de vacas preñadas con vacuna antiaftosa oleosa. Inmunidad pasiva. *Acta Physiologica Pharmacologica Latinoamericana* 36, 141-142.
- SMITH, A. N. & INGRAM, D. G. (1965). Immunological responses of young animals. II. Antibody production in calves. *Canadian Veterinary Journal* 6, 226–232.