

Campylobacteriosis in returning travellers and potential secondary transmission of exotic strains

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SUMMARY

Multilocus sequence types (STs) were determined for 232 and 737 *Campylobacter jejuni/coli* isolates from Dutch travellers and domestically acquired cases, respectively. Putative risk factors for travel-related campylobacteriosis, and for domestically acquired campylobacteriosis caused by exotic STs (putatively carried by returning travellers), were investigated. Travelling to Asia, Africa, Latin America and the Caribbean, and Southern Europe significantly increased the risk of acquiring campylobacteriosis compared to travelling within Western Europe. Besides eating chicken, using antacids, and having chronic enteropathies, we identified eating vegetable salad outside Europe, drinking bottled water in high-risk destinations, and handling/eating undercooked pork as possible risk factors for travel-related campylobacteriosis. Factors associated with domestically acquired campylobacteriosis caused by exotic STs involved predominantly person-to-person contacts around popular holiday periods. We concluded that putative determinants of travel-related campylobacteriosis differ from those of domestically acquired infections and that returning travellers may carry several exotic strains that might subsequently spread to domestic populations even through limited person-to-person transmission.

Key words: Multilocus sequence typing, person-to-person transmission, risk factor, source attribution, travel-related campylobacteriosis.

INTRODUCTION

Diarrhoeal infections remain a major concern for travellers, especially for those bound for destinations

where relatively substandard hygienic conditions exist. A Dutch study showed that in a sample of 1202 individuals travelling to developing countries, 50% experienced one or more diarrhoeal episodes [1].

Campylobacteriosis is a leading cause of traveller's diarrhoea, particularly in travellers returning from Southern and South East Asia [2, 3]. In The Netherlands (16.5 million population), an estimated

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90 000 cases of campylobacteriosis occur annually, with ~12% estimated as international travel-related [4]. Moreover, fluoroquinolone-resistant *Campylobacter* infections occur significantly more often in travel-related cases [3, 5].

Travellers are particularly prone to experiencing (symptomatic) *Campylobacter* infections [2, 6, 7], and susceptibility to disease is associated with duration of foreign stay. For instance, duration of residence of expatriates in Nepal was linearly correlated with protection from diarrhoeal infection [8], and travellers experiencing multiple diarrhoeal episodes had a shorter duration of symptoms after the first episode [1]. Similarly, in expatriates in Thailand, campylobacteriosis occurred significantly more often in those living there for <1 year [9]. A documented case of acquired immunity in developed countries concerned people professionally exposed. Newly employed poultry abattoir workers in Sweden have been shown to suffer more often from campylobacteriosis than their longer employed colleagues [10]. If partial immunity to (severe) disease is acquired over time with repeated exposure, then such protection should correlate with age. Indeed, campylobacteriosis incidence peaks in infancy worldwide and older age groups are significantly less prone to infection with common *Campylobacter* strains compared to the young [11]. Moreover, Swedish travellers to countries such as Germany, France, Belgium, The Netherlands, Austria, Luxembourg or Switzerland, all of which are developed countries with high hygienic standards, still have a 4.4–21 times higher risk of acquiring campylobacteriosis compared to those travelling to neighbouring Norway [12]. A recent study comparing *Campylobacter* multilocus sequence typing (MLST) datasets from different countries has further highlighted the importance of geographical distance in strain dissimilarity [13]. Moreover, serological studies of patients and volunteers infected with campylobacters (reviewed in [14]) have revealed an array of immunogenic components and elicited antibodies displayed little cross-reactivity, indicating considerable antigenic variation. Although core-genome (as described by MLST) is not necessarily related to antigens, and cross-protection is expected to develop for strains sharing similar antigenic properties, the higher risk of campylobacteriosis in travellers does not appear to be limited just to higher levels of exposure in developing countries, but also to the possible presence of ‘new’ *Campylobacter* strains, endemic in the different regions to which travellers have rarely been

exposed before [14]. It follows that, probabilistically, these new strains are more likely to be associated with regionally untested antigens than widespread strains, and acquired protection may be ineffective when exposed to uncommon strains, as evidenced by a Canadian study [15].

It is conceivable that the infected, but not necessarily symptomatic [14], returning travellers may introduce into the domestic population several ‘exotic’ strains with a higher probability of possessing antigens that are underrepresented in the local reservoirs, i.e. food-producing animals, pets and wildlife. These exotic strains may therefore constitute a distinctive, primarily human-restricted *Campylobacter* population that may have at least the potential to spread domestically via the person-to-person route. Although *Campylobacter* person-to-person transmission is believed to be uncommon and up to 66% and 21% of laboratory-confirmed cases in The Netherlands are attributable to chicken and cattle, respectively [16], it has been shown that campylobacteriosis household outbreaks are more common than believed [17] and up to 18% of putatively household outbreak-related cases are suggestive of secondary spread [18]. This raises the question to what extent exotically introduced strains may spill over into the domestic population and at first spread anthroponotically.

In this study, we investigated the MLST profiles of *Campylobacter* strains isolated from travellers returning to The Netherlands compared to those from domestically acquired cases. We also investigated putative determinants of travel-related campylobacteriosis by comparing the exposures of the returned travellers with those of travellers in the control population. Furthermore, we used a population genetics model for source attribution to estimate the probability that the domestically acquired infections were caused by exotic strains, putatively carried by returned travellers. Finally, factors associated with exotically introduced domestic campylobacteriosis were investigated and the percentage of domestically acquired cases due to person-to-person transmission of exotic strains was estimated.

METHODS

Data

An earlier case-control study on risk factors for campylobacteriosis conducted in The Netherlands between April 2002 and March 2003, the ‘CaSa study’ [6], formed the basis of this study. Isolates of

3115 *Campylobacter jejuni/coli* cases identified by the Dutch Regional Public Health Laboratories (RPHL) through passive surveillance were sent to the Dutch Central Veterinary Institute (CVI) for molecular speciation [19, 20]. Controls were selected from RPHL population registries by frequency matching (aiming at two per case) according to age (0–4, 5–17, 18–29, 30–44, 45–59, ≥ 60 years), sex, urbanization degree (urban: >2500 addresses/km²; urbanized: 500–2500 addresses/km²; rural: <500 addresses/km²) and season (April–June 2002, July–September 2002, October–December 2002, January–April 2003) [6, 16, 21]. Cases and controls were asked to complete a questionnaire regarding foreign travel, food consumption, kitchen hygiene, contact with animals, contact with gastroenteritis cases, occupation, recreational activities, medication use and chronic disease history. Questions covered the 7 days prior to symptom onset (cases) or questionnaire completion (controls). Missing values were handled using multiple imputation [6, 21].

Cases/controls not returning the questionnaire and/or living abroad were excluded, leaving 1428 cases and 3363 controls enrolled in the study. Of these, 328 cases and 244 controls had travelled abroad with ≥ 1 overnight stay in the destination country. A total of 66 countries were visited, with 36 cases and 27 controls visiting >1 country during the same travel episode. For three cases and four controls the travel destination was unknown. Destination countries were grouped into travel regions by adapting the United Nations geoscheme (<http://unstats.un.org/unsd/methods/m49/m49regin.htm#europe>).

Isolates from 737 non-travellers (domestically acquired cases) and 232 travellers were typed using MLST [22]. Association of the travellers' five most frequent sequence types (STs) and clonal complexes (CCs) with travel regions was tested using χ^2 or Fisher's exact tests. Proportional Similarity Index (PSI) [13] was used to measure the (dis)similarity between ST frequency distributions of travellers and non-travellers. The PSI ranges between 0 (no common ST) and 1 (identical distributions). Simpson's index of diversity was calculated to define the ST diversity of travellers and non-travellers as the probability that two randomly selected individuals were infected with different STs [23].

Source attribution

The Asymmetric Island (AI) model, a Bayesian population genetics algorithm for modelling *Campylobacter*

evolution and transmission [24], was used to estimate the probability (Pr) of the 737 non-travellers being infected with exotically introduced STs or with STs originating from four putative animal reservoirs (chicken, cattle, sheep, pig) or from the environment (water, sand, wild birds), a proxy for other unidentified reservoirs putatively of wildlife origin [16]. This study was restricted to campylobacteriosis of probable exotic origin. Results regarding the other animal and environmental sources have been reported elsewhere [13, 16].

To run the AI model, *C. jejuni/coli* MLST data from the above-mentioned animal and environmental sources were supplied by the CVI and supplemented with other data [22, 25, 26] to provide a representative dataset for each source (Table 1). Supplementary data were selected from other published datasets (reported in [13]) using Smid's methodology, which allows for the selection of non-local (and/or non-recent) MLST datasets for *Campylobacter* source attribution while minimizing potential biases [13, 16]. Differences in Pr for exotic origin (Pr_e) were tested for the variables age, sex, and season using Kruskal–Wallis or Mann–Whitney tests.

Factors associated with travel-related campylobacteriosis

Logistic regression was used to investigate putative risk factors for travel-related campylobacteriosis. The 325 diseased travellers and the 238 healthy travellers with known travel destination were included as cases and controls, respectively. Analysis was performed in the same way as in previous studies [6, 16, 21]. A total of 131 factors were tested for association with *Campylobacter* infection in the preliminary single-variable analysis and those factors showing $P \leq 0.10$ were selected for inclusion in a multivariable model. A backward stepwise procedure was applied and variables with $P < 0.05$ were retained in the final model. Education level [16], travel region, and length of stay (days) were always included as covariates to control for confounding in addition to the frequency-matched variables. As travel regions were almost mutually exclusive, Western Europe (which The Netherlands belongs to) was made the base category against which the other regions were assessed.

To explore if the risk factors of the reduced model differed according to age, sex, education level, season, and travel region, we also tested for their two-way interactions. The final multivariable model was then

Table 1. *Campylobacter jejuni/coli* strains typed with MLST used to feed the Asymmetric Island model for source attribution

Country	Humans (travellers)	Chicken	Cattle	Sheep	Pig	Environment	Ref.
The Netherlands	232 ^a	236	0	9	0	106 (water)	Data ^b
United Kingdom	0	73	46	46	72	50 (sand)	[22]
Scotland	0	239	90	90	88	133 (wild birds)	[25]
Switzerland	0	77	23	23	0	0	[26]
Total	232	625	168	168	160	289	

^a From *Campylobacter jejuni/coli* infected travellers of the CaSa study [6].

^b Provided by the Central Veterinary Institute (CVI) in Lelystad, The Netherlands.

expanded to include significant interactions. Overall model significance and goodness-of-fit were verified by likelihood ratio χ^2 and Hosmer–Lemeshow tests, respectively. The best-fitting model was identified using Akaike's Information Criterion (AIC). Bias-corrected bootstrap confidence intervals were also calculated (1000 iterations) and compared to the standard ones. As these did not differ significantly, the standard values were reported. Statistical analysis was performed using Stata v. 11.2 (StataCorp., USA).

Factors associated with exotically introduced domestic campylobacteriosis

To investigate putative risk factors for domestically acquired campylobacteriosis caused by STs of probable exotic origin, MLST data of travellers were included as an additional source in the AI model. Similar to previous studies [16, 27, 28], the Pr_e distribution was assessed and a cut-off was determined to optimize the number of domestic cases assigned to be exotically introduced and the confidence as to their correct assignment derived by the highest possible Pr_e . Logistic regression was then used to investigate risk factors for domestically acquired campylobacteriosis caused by STs with at least 77% probability (cut-off $Pr_e \geq 0.77$) of originating from abroad. This cut-off Pr_e resulted in the selection of 79 cases with a median Pr_e of 0.89 (mean 0.88, range 0.77–0.99) belonging to 35 different STs. The 3119 non-travelling controls were included in this analysis.

The effect of the assignment cut-off Pr_e on the risk factors was checked by sensitivity analysis, which was done by repeating the analysis for different cut-off Pr_e values from 0.5 to 0.9. Low numbers of cases did not allow for the construction of models based on a cut-off of $Pr_e > 0.9$. Finally, a case-case analysis

comparing exposures of domestic infections with exotic vs. non-exotic STs was performed.

Estimation of domestic cases due to person-to-person transmission of exotic strains

The percentage of domestic cases due to person-to-person transmission of exotic strains, denoted by γ , was estimated as:

$$\gamma = \frac{[1 - \prod_{i=1}^n (1 - \text{PAR}_i)] \times N_{\text{ED}}}{N_{\text{D}}} \times 100,$$

where PAR is the population attributable risk (calculated based on multivariable odds ratios and prevalence of exposure in cases) for factor i out of n factors entailing person-to-person transmission in the final multivariable model. N_{ED} is the estimated total number of domestic cases of probable exotic origin, given by $\text{Binomial}(N_{\text{D}}, Pr)$, where N_{D} is the total number of domestic cases (non-travellers) enrolled in the study and Pr is the estimated probability for a domestic case to be infected with an exotic ST, assumed to be $\text{Beta}(a, b)$ -distributed, with a being the 79 typed domestic cases with $Pr_e \geq 0.77$ and b being the total number of typed domestic cases ($n = 737$) minus 79. Estimation was performed using @RISK (Palisade Corp., USA) by setting 10000 iterations with the Latin Hypercube sampling technique and a seed of 1.

RESULTS

Genotypes

The 737 typed strains from non-travellers were assigned to 154 STs and 28 CCs, whereas those from the 232 travellers were assigned to 127 STs and 23 CCs. Twenty-eight STs from non-travellers and 23 STs from travellers were unassigned to previously identified CCs.

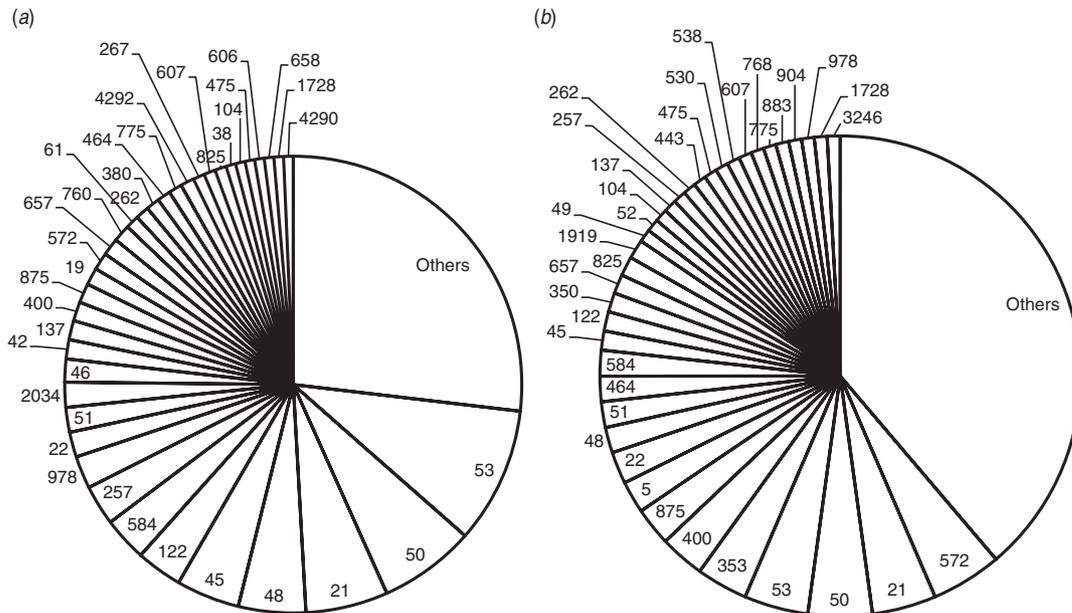


Fig. 1. Sequence types identified in *Campylobacter jejuni* isolates from (a) 737 non-travellers (infections acquired in The Netherlands) and (b) 232 travellers returning to The Netherlands. Category ‘others’ includes sequence types that occurred less than five times (non-travellers) and less than twice (travellers).

In non-travellers, the top five STs (ST53, ST50, ST21, ST48, ST45) accounted for >25% of cases and the top five CCs (CC21, CC45, CC206, CC257, CC48) for >50% of cases (Fig. 1). In travellers, the top five STs (ST572, ST21, ST50, ST53, ST353) accounted for ~20% of cases and the top five CCs (CC21, CC353, CC828, CC206, CC52) for >50% of cases (Fig. 2). STs occurring once accounted for 46% and 70% of STs in non-travellers and travellers, respectively. PSI between travellers and non-travellers was 0.47 [95% confidence interval (CI) 0.34–0.59] while Simpson’s index was 0.972 (95% CI 0.968–0.976) in non-travellers and 0.988 (95% CI 0.984–0.992) in travellers.

There were 68 STs (74 cases) found only in travellers and absent in any of the considered sources and in non-travellers (Table 2). Most (73%) cases infected with these traveller-only STs had travelled to Asia or Africa compared to 46% of all travel-related cases returning from these continents (z test, $P < 0.001$). Conversely, 23% of cases infected with traveller-only STs had travelled within Europe compared to 53% of all travel-related cases travelling within Europe (z test, $P < 0.001$).

ST572 was significantly overrepresented (64%) in travellers from Western Europe ($P = 0.001$); ST50 in those from Western Asia (40%, $P = 0.014$); ST53 in those from Southern Europe (50%, $P = 0.035$); ST353, CC353 and CC828 in those from Northern

Africa (38%, 27%, and 46%; $P = 0.003$, $P < 0.001$, and $P < 0.001$, respectively); CC52 in those from Eastern Europe (23%, $P = 0.001$).

STs with $Pr_e \geq 0.77$ included in the risk factor analysis for exotically introduced domestic campylobacteriosis were ST1009, ST400, ST353, ST4307, ST5, ST2100, ST4308, ST2088, ST2151, ST1728, ST2844, ST508, ST403, ST443, ST350, ST3130, ST2156, ST878, ST657, ST4269, ST2882, ST2130, ST1080, ST4300, ST2187, ST586, ST696, ST587, ST2641, ST2553, ST3015, ST2123, ST4311, ST4278 and ST4284. There was a significant seasonal effect ($P = 0.036$) on Pr_e , which peaked in October–December and decreased in April–June. No significant age and sex effects on Pr_e were found.

Factors associated with travel-related campylobacteriosis

Compared to travelling within Western Europe, travelling to any region in Asia, Africa, Latin America and the Caribbean, and Southern Europe posed a higher risk of acquiring campylobacteriosis (Table 3), whereas the risk posed by Northern and Eastern Europe and Oceania, as well as the length of stay, were not significant ($P > 0.05$).

Factors positively associated with travel-related campylobacteriosis (Table 3) were: using proton-pump inhibitors, consuming vegetable salad when

Table 2. Sequence types found exclusively in travellers returning to The Netherlands but not in cases acquired in The Netherlands, nor in any of the sourced animal and environmental reservoirs

Sequence type	No. of cases	Travel destination
4291	1	Austria
474	1	Belgium
4274	1	Bolivia, Peru
3878, 4252	2	Czech Republic
830, 931, 2229, 2968	4	Egypt
3606	1	Ethiopia
892	1	France
1044	1	Germany
446, 451, 1042, 4288	4	India
530, 161, 2031, 2109, 2131, 2393, 2941, 4270, 4281, 4287, 4289, 4296	13	Indonesia
4298	1	Italy
4305	1	Jordan
3630	1	Jordan, Iraq
1380	1	Kenya
466	1	Luxemburg, France
4293	1	Malaysia
1039, 4309	2	Mali
2116, 3575, 4165, 4299	4	Morocco
986	1	Nepal, China
1233, 4053	2	Peru
2895	1	Philippines
614	1	Poland
4277	1	Portugal
904	2	Portugal, Spain
4275	1	Singapore
148, 1710, 4294, 4313, 4408	5	Spain
1919, 768, 407, 1953, 2083, 2315, 4303	10	Thailand
3246, 303, 305, 2066, 2184, 2275, 3142	8	Turkey
919	1	Vietnam, Malaysia

with STs that were undetected domestically had travelled predominantly to distant destinations in Asia and Africa, suggesting that differences in STs are related, to some extent, to the geographical distance of the travel region compared to The Netherlands, with STs from nearby European countries being generally more similar than those from farther destinations [13]. Another study [29] showed nearly identical *C. jejuni* ST distributions in two English populations, but clear differences were found compared to Finland, Australia and New Zealand. Yet, differentiation was not proportional to distance, with the English sample being more similar to the samples

from Australia and New Zealand than to the Finnish one, suggesting that similarities in STs between countries mainly reflect similarities of sources of infection rather than geographical proximity *per se*.

Consistent with [30], travel-related isolates had a significantly higher ST diversity than domestic isolates, with a different spectrum of MLST genotypes representative of the different countries. The larger ST diversity in travellers combined with the association of some STs with specific destinations is also consistent with the presence of heterogeneously distributed clones that are endemic in the different regions but not so prevalent elsewhere in the world. Regionally endemic STs have been identified, e.g. in Australia [31], New Zealand [32] and Curaçao [33], and may emerge because of clonal expansion, niche adaptation, geographical isolation and host immune selection [34]. Although so far there has been no evidence of ST-specific immune responses, it can be hypothesized that the chance of being exposed to a ST with uncommon antigens is somewhat higher for STs that are rarely, rather than commonly, encountered. STs that are associated with strong regional clustering would therefore pose a higher risk to travellers because of limited, prior (repeated) exposure in addition to issues related to sanitation failure. The risk posed by uncommon STs is also substantiated by their age distribution [16, 35]. For instance, the three most common STs in non-travellers were mainly found in the young relative to other STs, decreasing steadily with age (data not shown). Conversely, rare STs (<5 isolates) occurred independently of age. According to interpretation of similar findings [11], it is likely that antigenic properties associated with the common STs are frequently encountered throughout life; thus, the young would be more susceptible because they have encountered these STs less often. In contrast, rare STs, more probably related to uncommon antigens, would have rarely been encountered by all age groups.

The original CaSa study was designed to investigate factors associated with domestic campylobacteriosis and not purposely with travel-related *Campylobacter* infection [6]. However, most risk exposures apply to some extent world-wide, and while the behaviour of travellers (and hence their exposure to campylobacters) may be different from the domestic population, it has been reported that most STs found in travellers returning to Scotland are still attributable to chicken in a similar proportion to that of domestic cases [29]. Although this study did not have the power

Table 3. Multivariable odds ratios (OR) and corresponding 95% confidence intervals (CI) of risk factors for *Campylobacter jejuni/coli* infection in travellers returning to The Netherlands

Risk factor (% of imputed missing values)	% cases exposed (n = 325)	% controls exposed (n = 238)	OR (95% CI) ^a
Duration of stay ^b (3)			1.0 (0.9–1.1) n.s.
Region of destination ^c			
Western Europe ^d	22.8	60.0	Reference
Northern Europe ^e	3.4	8.8	0.8 (0.3–2.2) n.s.
Eastern Europe ^f	4.3	5.0	1.1 (0.3–2.2) n.s.
Southern Europe ^g	22.8	24.8	1.7 (1.0–3.3)
Northern Africa ^h	8.6	1.3	10.6 (2.3–49.0)
Sub-Saharan Africa ⁱ	4.3	0.4	25.4 (2.7–310.7)
Western Asia ^j	13.8	2.1	10.6 (2.8–39.9)
South East Asia and China ^k	16.3	0.8	27.8 (4.5–170.9)
Southern Asia ^l	3.7	0.4	28.9 (2.4–265.1)
Oceania ^m	0.6	0.4	0.8 (0.0–43.040.2) n.s.
Latin America and the Caribbean ⁿ	4.9	0.4	20.8 (2.0–211.6)
Eating chicken vs. not eating chicken (2)	80.0	73.7	2.0 (1.1–3.5)
Eating yoghurt vs. not eating yoghurt (4)	62.5	88.1	0.4 (0.2–0.7)
Eating vegetable salad vs. not eating vegetable salad (3)			
When travelling within Europe	27.3	27.1	1.7 (0.9–3.1) n.s.
When travelling outside Europe	17.4	2.0	6.7 (2.1–40.2)
Drinking exclusively bottled water vs. drinking both bottled and non-bottled water (3)			
When travelling within Europe, excluding Southern Europe	6.0	8.2	1.5 (0.6–3.6) n.s.
When travelling to Southern Europe and outside Europe	33.3	7.2	2.3 (1.6–5.0)
Handling and/or eating raw or undercooked pork vs. not handling and/or eating raw or undercooked pork (5)	9.8	2.3	6.2 (1.3–29.3)
Using proton-pump inhibitors vs. not using proton-pump inhibitors	8.5	1.6	14.6 (3.0–82.0)
Having a chronic gastrointestinal disease vs. not having a chronic gastrointestinal disease (5)	6.2	3.5	2.9 (1.7–4.8)
Working in healthcare vs. not working in healthcare (1)	5.7	9.0	0.4 (0.1–0.9)

n.s., Not significant ($P > 0.05$).

^a Adjusted for age, sex, degree of urbanization, season and level of education.

^b Continuous variable, expressed as number of days stayed.

^c Adapted from the United Nations scheme of the composition of macro-geographical (continental) regions, geographical sub-regions, and selected economic and other groupings. (<http://unstats.un.org/unsd/methods/m49/m49regin.htm#europe>).

^d Includes travellers returning from Germany, France, Belgium, Austria, Luxemburg, and Switzerland.

^e Includes travellers returning from the UK, Ireland, Denmark, Sweden, Norway, and Finland.

^f Includes travellers returning from the Czech Republic, Hungary, Poland, Slovakia, and Romania.

^g Includes travellers returning from Spain, Italy, Portugal, Greece, Croatia, and Malta.

^h Includes travellers returning from Morocco, Egypt, and Tunisia.

ⁱ Includes travellers returning from Benin, Cameroon, Ethiopia, Ghana, Kenya, Mali, Nigeria, Rwanda, Tanzania, Botswana, Burkina Faso, South Africa and Namibia.

^j Includes travellers returning from Turkey, Jordan and Iraq.

^k Includes travellers returning from China, Indonesia, Malaysia, Singapore, Philippines, Thailand, and Vietnam.

^l Includes travellers returning from India, Nepal, and Bangladesh.

^m Includes travellers returning from Australia and Fiji Islands.

ⁿ Includes travellers returning from Bolivia, Ecuador, Peru, Venezuela, Chile, Costa Rica, Cuba, Dominican Republic, Haiti, Mexico, and Guatemala.

to conduct a stratified analysis by travel destination, it was possible to highlight some interactions between the exposures and the travel destinations.

Eight factors significantly associated with travel-related campylobacteriosis were identified. Consistent with evidence that poultry is the main reservoir

Table 4. Multivariable odds ratios (OR) and corresponding 95% confidence intervals (CI) of risk factors for *Campylobacter jejuni/coli* infection acquired in The Netherlands caused by strains of most likely exotic origin

Risk factor (% of imputed missing values)	% cases exposed (n = 79)	% controls exposed (n = 3119)	Multivariable OR (95% CI) ^a
Contact with people with gastroenteritis outside the household (3)	17.2	10.5	2.2 (1.9–4.6)
Recent use of proton-pump inhibitors	15.2	2.2	9.5 (4.4–20.6)
Eating yoghurt (2)	71.9	86.5	0.3 (0.2–0.6)
Not washing hands after toilet visit			
April–December 2002	1.3	0.6	6.8 (0.7–68.1) n.s.
January–April 2003	2.5	0.1	20.8 (1.9–233.4)
Being a school-attending child			
July–September 2002 and January–April 2003	6.3	6.5	1.4 (0.5–3.8) n.s.
April–June 2002	5.1	2.6	3.4 (1.0–11.4)
October–December 2002	7.6	2.0	4.0 (1.4–11.3)
Swimming in a public swimming pool (1)			
April–September 2002 and January–April 2003	10.1	11.8	1.0 (0.4–2.1) n.s.
October–December 2003	7.8	2.9	3.7 (1.3–11.0)

n.s., Not significant ($P > 0.05$).

^a Also adjusted for age, sex, degree of urbanization, season and level of education.

for campylobacters, most studies concerning risk factors for campylobacteriosis have identified an association with eating chicken [6, 7, 16, 36, 37], suggesting that this factor is not exclusive for acquiring infection abroad. This also applies to consuming antacids and having chronic enteropathies [6, 7, 16].

In The Netherlands, <1% of domestically acquired *Campylobacter* infections have been attributed to pigs [16]. Moreover, eating pork has been associated with a reduced risk for *C. coli* [6] and chicken-borne *C. jejuni/coli* [16] infections. Accordingly, Dutch retail pork has rarely been found contaminated with campylobacters [38]. The positive association with handling and/or eating raw or undercooked pork we found therefore suggests that pigs may be an important reservoir (and pork an important exposure) of campylobacteriosis outside The Netherlands.

In contrast to previous findings indicating that eating vegetable salad is negatively associated with the occurrence of (domestic) campylobacteriosis [6, 16], we observed that this factor was associated with increased odds of acquiring campylobacteriosis when travelling outside Europe. In Europe, extensive sampling of raw vegetables, including ready-to-eat salads, has generally found no, or very few, campylobacters [38], suggesting that contamination of such items during irrigation, harvesting and processing is unlikely and that salads may occasionally become

cross-contaminated during food preparation [36]. Conversely, exceptionally high *Campylobacter* isolation rates (~68%) in raw vegetables were reported from countries such as Malaysia [39], indicating that major problems can arise by consuming vegetables if hygiene practices are absent or break down.

Drinking bottled water exclusively was associated with increased odds of acquiring campylobacteriosis when travelling to high-risk destinations. In the UK, drinking bottled water has been found to be positively associated with campylobacteriosis [36], particularly *C. coli* infection [40], and ciprofloxacin-resistant *Campylobacter* infection acquired abroad [5], suggesting that bottled water could, given the right circumstances, provide a vehicle for campylobacters [36]. In fact, bottled water, unlike tap water, is not usually treated and testing for *Campylobacter* is rarely undertaken [36, 40]. Moreover, in the event of dual contamination of bottled water (campylobacters and organic matter), *C. jejuni* may survive for prolonged periods [41]. However, our association with bottled water was only significant when travelling to high-risk destinations, supporting the hypothesis that drinking bottled water acts as a proxy for local circumstances where there is a generally high risk for campylobacteriosis. Travellers are indeed usually advised to drink bottled water where there is any doubt about the local water quality. The use of bottled

water may help in preventing infection but there may be circumstances where the risk is higher than that which can be prevented by drinking bottled water. Moreover, our questionnaire did not distinguish between sparkling and still bottled water and did not ask whether it was consumed with or without ice. Therefore, further investigation is needed to assess if the advice of drinking bottled water merits any refinement.

Proportionally more controls than cases reported consuming yoghurt. It is believed that probiotic bacteria in yoghurt may alter the intestinal microflora in a way that prevents infection [37], while people working in healthcare might be particularly aware of the health risks (and ways to avoid them) when travelling.

Putative determinants of exotically introduced domestic campylobacteriosis were suggestive of anthroponotic transmission, namely contact with gastroenteritis cases outside the household (thus less likely to share the same exposure); not washing hands after toilet visit; being a school-attending child (usually having high frequencies of contacts); and attending public swimming pools (as recreational water has been proposed as a vehicle for *Campylobacter* transmission [16, 37]). Except for the first factor, the others were unidentified in previous analyses where the same cases were not split according to their estimated exoticism [6, 16]. Moreover, we found significant interactions with season, which is in accordance with the seasonal nature of travelling, as also shown by the finding that Pr_e varies seasonally. Periods most at risk were mainly those around popular holidays in The Netherlands, notably the autumn break in October, Christmas/New Year in December–January, and Easter in April–May. Moreover, people most at risk were school-attending children for which additional peaks in domestic campylobacteriosis have already been noted shortly after the end of school breaks, suggesting that these additional peaks are due to exposure to less common strains from less common foods consumed during the festivities and to the mixing of people that have not been in contact for a long time following on from the previous holidays [42]. It can therefore be hypothesized that travellers infected with strains possessing uncommon antigens might still be shedding them after returning home, most likely asymptotically [14]. As there is unlikely to be a high prevalence of acquired protection against these exotic strains domestically, there is at least the potential for them to spread even through limited person-to-person transmission, with an average of 3.2% domestic cases

being estimated to be due to anthroponotic transmission of exotically introduced strains. In conclusion, this study sought to provide an outline of *Campylobacter* transboundary epidemiology that may apply to other countries as well. This ranges from the identification of *C. jejuni/coli* MLST genotypes causing infection in international travellers to the risk factors potentially responsible for acquiring infection with such strains upon travelling, as well as those potentially responsible for their secondary spread to domestic populations. Besides the identification of high-risk travel destinations and universal correlates of campylobacteriosis, such as eating chicken, using antacids and having chronic enteropathies, this study also identified eating vegetable salad outside Europe, drinking bottled water in high-risk destinations and handling and/or eating raw or undercooked pork as factors associated specifically with travel-related campylobacteriosis. Moreover, suggestive evidence was provided that international travellers are particularly prone to infection with *Campylobacter* MLST genotypes that are endemic in the different travel regions, and that these exotic strains may be carried by the returning travellers and be spread to domestic populations through limited person-to-person transmission. As travellers have dynamic interactions with people, places, and microbes during their journeys, it is conceivable that they can be victims, carriers, and eventually transmitters of such agents to new regions and populations. Our understanding of campylobacteriosis may therefore depend on increased insight into regional risk differences, high-risk exposures and *Campylobacter* behaviour in response to newly available susceptible populations and changing environments.

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DECLARATION OF INTEREST

None.

REFERENCES

- Belderok SM, *et al.* Incidence, risk factors and treatment of diarrhoea among Dutch travellers: reasons not to routinely prescribe antibiotics. *BMC Infectious Diseases* 2011; **11**: 295.
- Ekdahl K, Andersson Y. Regional risks and seasonality in travel-associated campylobacteriosis. *BMC Infectious Diseases* 2004; **4**: 54.
- Hakanen A, *et al.* Fluoroquinolone resistance in *Campylobacter jejuni* isolates in travelers returning to Finland: association of ciprofloxacin resistance to travel destination. *Emerging Infectious Diseases* 2003; **9**: 267–70.
- Havelaar AH, *et al.* Disease burden of foodborne pathogens in the Netherlands, 2009. *International Journal of Food Microbiology* 2012; **156**: 231–238.
- Evans MR, *et al.* Risk factors for ciprofloxacin-resistant *Campylobacter* infection in Wales. *Journal of Antimicrobial Chemotherapy* 2009; **64**: 424–427.
- Doorduyn Y, *et al.* Risk factors for indigenous *Campylobacter jejuni* and *Campylobacter coli* infections in The Netherlands: a case-control study. *Epidemiology and Infection* 2010; **138**: 1391–1404.
- Tam CC, *et al.* Chicken consumption and use of acid-suppressing medications as risk factors for *Campylobacter* enteritis, England. *Emerging Infectious Diseases* 2009; **15**: 1402–1408.
- Hoge CW, *et al.* Epidemiology of diarrhea among expatriate residents living in a highly endemic environment. *Journal of the American Medical Association* 1996; **275**: 533–538.
- Gaudio PA, *et al.* Diarrhea among expatriate residents in Thailand: correlation between reduced *Campylobacter* prevalence and longer duration of stay. *Journal of Travel Medicine* 1996; **3**: 77–79.
- Cawthraw SA, *et al.* Antibodies, directed towards *Campylobacter jejuni* antigens, in sera from poultry abattoir workers. *Clinical and Experimental Immunology* 2000; **122**: 55–60.
- Miller G, *et al.* Does age acquired immunity confer selective protection to common serotypes of *Campylobacter jejuni*? *BMC Infectious Diseases* 2005; **5**: 66.
- Havelaar AH, *et al.* Estimating the true incidence of campylobacteriosis and salmonellosis in the European Union, 2009. *Epidemiology and Infection* 2013; **141**: 293–302.
- Smid JH, *et al.* Practicalities of using non-local or non-recent multilocus sequence typing data for source attribution in space and time of human campylobacteriosis. *PLoS ONE* 2013; **8**: e55029.
- Havelaar AH, *et al.* Immunity to *Campylobacter*: its role in risk assessment and epidemiology. *Critical Reviews in Microbiology* 2009; **35**: 1–22.
- Arsenault J, *et al.* Do patients with recurrent episodes of campylobacteriosis differ from those with a single disease event? *BMC Public Health* 2011; **11**: 32.
- Mughini Gras L, *et al.* Risk factors for campylobacteriosis of chicken, ruminant, and environmental origin: a combined case-control and source attribution analysis. *PLoS ONE* 2012; **7**: e42599.
- Ethelberg S, *et al.* Household outbreaks among culture-confirmed cases of bacterial gastrointestinal disease. *American Journal of Epidemiology* 2004; **159**: 406–412.
- Rotariu O, *et al.* Putative household outbreaks of campylobacteriosis typically comprise single MLST genotypes. *Epidemiology and Infection* 2010; **138**: 1744–1747.
- Fermér C, Engvall EO. Specific PCR identification and differentiation of the thermophilic campylobacters, *Campylobacter jejuni*, *C. coli*, *C. lari*, and *C. upsaliensis*. *Journal of Clinical Microbiology* 1999; **37**: 3370–3373.
- Marshall SM, *et al.* Rapid identification of *Campylobacter*, *Arcobacter*, and *Helicobacter* isolates by PCR-restriction fragment length polymorphism analysis of the 16S rRNA gene. *Journal of Clinical Microbiology* 1999; **37**: 4158–4160.
- Doorduyn Y, *et al.* Risk factors for *Salmonella* Enteritidis and Typhimurium (DT104 and non-DT104) infections in The Netherlands: predominant roles for raw eggs in Enteritidis and sandboxes in Typhimurium infections. *Epidemiology and Infection* 2006; **134**: 617–626.
- Dingle KE, *et al.* Multilocus sequence typing system for *Campylobacter jejuni*. *Journal of Clinical Microbiology* 2001; **39**: 14–23.
- Hunter PR, Gaston MA. Numerical index of the discriminatory ability of typing systems: an application of Simpson's index of diversity. *Journal of Clinical Microbiology* 1988; **26**: 2465–2466.
- Wilson DJ, *et al.* Tracing the source of campylobacteriosis. *PLoS Genetics* 2008; **4**: e1000203.
- Strachan NJC, *et al.* Attribution of *Campylobacter* infections in northeast Scotland to specific sources by use of multilocus sequence typing. *Journal of Infectious Diseases* 2009; **199**: 1205–1208.
- Korczak BM, *et al.* Multiplex strategy for multilocus sequence typing, *fla* typing, and genetic determination of antimicrobial resistance of *Campylobacter jejuni* and *Campylobacter coli* isolates collected in Switzerland. *Journal of Clinical Microbiology* 2009; **47**: 1996–2007.
- Bessell PR, *et al.* Using sequence data to identify alternative routes and risk of infection: a case-study of campylobacter in Scotland. *BMC Infectious Diseases* 2012; **12**: 80.
- Mullner P, *et al.* Molecular and spatial epidemiology of human campylobacteriosis: source association and genotype-related risk factors. *Epidemiology and Infection* 2010; **138**: 1372–1383.
- Strachan NJC, *et al.* Identifying the seasonal origins of human campylobacteriosis. *Epidemiology and Infection* 2013; **141**: 1267–1275.

30. **McCarthy ND, et al.** Molecular epidemiology of human *Campylobacter jejuni* shows association between seasonal and international patterns of disease. *Epidemiology and Infection* 2012; **140**: 2247–55.
31. **Mickan L, et al.** Multilocus sequence typing of *Campylobacter jejuni* isolates from New South Wales, Australia. *Journal of Applied Microbiology* 2007; **102**: 144–152.
32. **McTavish SM, et al.** Wide geographical distribution of internationally rare *Campylobacter* clones within New Zealand. *Epidemiology and Infection* 2008; **136**: 1244–1252.
33. **Duim B, et al.** Molecular evidence for dissemination of unique *Campylobacter jejuni* clones in Curaçao, Netherlands Antilles. *Journal of Clinical Microbiology* 2003; **41**: 5593–5597.
34. **Gupta S, Maiden MC.** Exploring the evolution of diversity in pathogen populations. *Trends in Microbiology* 2001; **9**: 181–185.
35. **Kärenlampi R, et al.** Longitudinal study of Finnish *Campylobacter jejuni* and *C. coli* isolates from humans, using multilocus sequence typing, including comparison with epidemiological data and isolates from poultry and cattle. *Applied and Environmental Microbiology* 2007; **73**: 148–155.
36. **Evans MR, et al.** Hazards of healthy living: bottled water and salad vegetables as risk factors for *Campylobacter* infection. *Emerging Infectious Diseases* 2003; **9**: 1219–1225.
37. **Kapperud G, et al.** Factors associated with increased and decreased risk of *Campylobacter* infection: a prospective case-control study in Norway. *American Journal of Epidemiology* 2003; **158**: 234–242.
38. **European Food Safety Authority and European Centre for Disease Prevention and Control.** The European Union summary report on trends and sources of zoonoses, zoonotic agents and foodborne outbreaks in the European Union in 2010. *EFSA Journal* 2012; **10**(6): 2765.
39. **Chai LC, et al.** Thermophilic *Campylobacter* spp. in salad vegetables in Malaysia. *International Journal of Food Microbiology* 2007; **117**: 106–111.
40. **Gillespie IA, et al.** A case-case comparison of *Campylobacter coli* and *Campylobacter jejuni* infection: a tool for generating hypotheses. *Emerging Infectious Diseases* 2002; **8**: 937–942.
41. **Tatchou-Nyamsi-König J-A, et al.** Behaviour of *Campylobacter jejuni* in experimentally contaminated bottled natural mineral water. *Journal of Applied Microbiology* 2007; **103**: 280–288.
42. **van Pelt W, et al.** Similarities and differences in seasonality of *Campylobacter* in broilers and humans, 1998–2006, the Netherlands. *Zoonoses and Public Health* 2007; **54**: 51.