# The effects of force-feeding on enzymes of the liver, kidney, pancreas and digestive tract of chicks

### By ZAFRIRA NITSAN\*

Division of Poultry Science, Agricultural Research Organization, The Volcani Center, Rehovot, Israel

## AND Y. DROR

Department of Agricultural Biochemistry, Faculty of Agriculture, Hebrew University of Jerusalem, Rehovot, Israel

## AND I. NIR AND NIVA SHAPIRA

Department of Animal Hygiene and Poultry Science, Faculty of Agriculture, Hebrew University of Jerusalem, Rehovot, Israel

(Received 2 August 1973 – Accepted 24 October 1973)

- 1. Force-feeding of young chicks for 15 d increased kidney arginase (EC 3.5.3.1) activity threefold. Fasting for 30 h decreased this activity by 50%.
- 2. Liver xanthine dehydrogenase was slightly increased after force-feeding and decreased following fasting.
- 3. The specific activities of two pentose-phosphate-cycle enzymes were not significantly affected by force-feeding, but glucose-6-phosphate dehydrogenase (EC 1.1.1.49) decreased following fasting.
- 4. The over-all secretion of digestive enzymes increased parallel to the increase in food consumption. Therefore, despite an increased absolute weight of the pancreas and intestinal chyme, specific activities were the same in the force-fed and *ad lib*.-fed groups, except for a higher activity in intestinal amylase.
  - 5. Fasting did not affect the pancreatic enzymic activities.

Force-feeding of young chicks increases growth rate (by 60%) and energy utilization. The increase in body-weight is due mainly to lean body-weight and partly to fat (Nir, Shapira, Nitsan & Dror, 1974). Force-feeding geese with maize is associated with an increase in the activities of liver xanthine dehydrogenase (XDH), kidney arginase (EC 3.5.3.1) and glucose-6-phosphate dehydrogenase (G6PD) (EC 1.1.1.43). The specific activities of digestive enzymes are reduced in the pancreas and intestinal tract (Nitsan, Nir, Dror & Bruckental, 1973), but the total amount of enzyme in the excreta is increased for the proteolytic enzymes and decreased for amylase (Nir, Nitsan & Vax, 1973). The activities of most of the enzymes, especially that of the amylase, are increased after supplementing the maize with soya-bean meal.

In the present study the enzymic adaptability of young chicks in digesting and metabolizing excessive amounts of nutrients was investigated. For this purpose we studied the effect of force-feeding and subsequent fasting on the activities of enzymes

<sup>\*</sup> Contribution No. 243-E, 1973 series.

involved in protein catabolism in liver and kidneys, enzymes of the pentose phosphate cycle in liver and enzymes in pancreas and digestive tract.

#### EXPERIMENTAL

The chicks were cross-bred male New Hampshire ×White Leghorn. Care of the chicks, diets, technique of force-feeding and of autopsy are described elsewhere (Nir et al. 1974). The birds were killed at 29 d of age. After autopsy, the livers were quickly rinsed with saline (9 g sodium chloride/l), blotted on filter paper, weighed and frozen at  $-22^{\circ}$ . The kidney, pancreas, intestinal and caecal contents were weighed and frozen  $(-22^{\circ})$  for enzymic determinations.

## Determination of enzyme activities

Renal arginase activity was determined as described by Dror & Gertler (1967) except that 4 mm-MnSO<sub>4</sub> was added to the medium. XDH activity in liver was determined according to Strittmatter (1965). The assay was performed with a model 2400 Gilford spectrophotometer (Gilford Instruments Ltd, Morden, Surrey). The extinction of each sample was measured for 2 min against its blank, in which xanthine was omitted. 6PGD and G6PD of liver were determined as described by Dror, Sassoon, Watson & Johnson (1970).

The pancreas was homogenized in 10 vol. cold distilled-water and centrifuged at 70000 g for 20 min. Lipase (EC 3.1.1.3) was determined in the homogenate by a modified procedure of Sarda & Desnuelle (1958); amylase by the method of Bernfeld (1955), with modification as described by Gertler & Nitsan (1970). Trypsin (EC 3.4.4.4) and chymotrypsin A (EC 3.4.4.5) were determined after activation of the pancreatic homogenate according to Gertler & Nitsan (1970). N-benzoyl-DL-argininep-nitroanilide-HCl (BANA) and N-acetyl-L-tyrosine-p-nitroanilide (ATNA) (E. Merck, Darmstadt, W. Germany), were used as substrates for trypsin and chymotrypsin, the final concentration being 1.25 mm in 3.7 mm-Tris buffer, pH 7.8, 0.6 mm-CaCl<sub>2</sub> and 25 mg/ml dimethylformamide. The reaction proceeded at 30° for 30 min and was stopped with acetic acid (300 ml/l). The colour developed was measured at 410 nm. The enzyme concentration was calculated using standard curves which were prepared with pure enzyme preparations (Sigma, St Louis, Missouri, USA) by the same procedure. Activity units were calculated as follows: for XDH, 6PGD and G6PD, μmol NAD or NADP reduced/min at 30°; for arginase and lipase, μmol product formed/min at 30°; for amylase, extinction at 550 nm (×10-2)/3 min, using 12.7 mm Bausch & Lomb test-tubes, at 37°.

Statistical analysis was done by the method of Snedecor & Cochran (1967) as detailed by Nir et al. (1974).

#### RESULTS

Activity of kidney arginase was markedly increased after force-feeding; the increase was threefold when expressed as units/g or about fivefold when expressed as units/kidney (Table 1). The activity was reduced by approximately 50% after fasting for

Table 1. Enzyme activities in the liver and kidney of chicks fed ad lib. or force-fed, before and after fasting

(Mean values for eight chicks/group)

	Treatment					Statistical significance of effects of			
	Ad libfed		Force-fed			or effects of			
Liver:	Fed	Starved	Fed	Starved	SEM	Force- feeding P <	Starving P <	Interac- tion P <	
Weight (g)	10.2	$7.4^d$	16.0a	$11.0_p$	0.2	0.01	0.01	0.01	
(g/kg body-wt)	$31.8_{p}$	27·9°	$40.8^a$	33.4 <sup>b</sup>	1.1	0.01	0.01	NS	
Enzyme activity (units*/g)									
XDH	0.63	$\circ \cdot 85^a$	0.81a	$0.89^a$	0.02	0.02	0.01	NS	
6PGD	0.88	0.99	1.05	0.98	0.04	NS	NS	NS	
G6PD	0.422	0.159	0.348	0.285	0.042	NS	0.01	0.02	
(units/liver)	•	•		•				-	
XDH	6·4°	6·3°	$13.7^{a}$	10.6p	0.6	0.01	0.02	0.01	
6PGD	8.90	7·3°	17·2a	11.6p	0.7	0.01	0.05	0.01	
G6PD	4.29b	1.18c	$5.87^a$	$3.38_{p}$	0.23	0.02	0.01	0.01	
Kidney:									
Weight (g)	$3.36^{b}$	$2.77^{b}$	$4.60^{a}$	$4.52^{a}$	0.26	0.01	NS	NS	
(g/kg body-wt)	10.2	10.2	11.5	12.8	0.7	NS	NS	NS	
Arginase activity					-				
(units*/g)	$34.8^{bc}$	20.30	121a	$54^{b}$		0.01	0.01	0.01	
se of mean	(3.5)		(12)						
(units/kidney)									
Arginase	117 <sup>c</sup>	$56^d$	$555^a$	242 <sup>b</sup>	_	0.01	0.01	0.01	
SE of mean	(16)		(54)						

NS, non-significant; XDH, xanthine dehydrogenase; 6PGD, 6-phosphogluconic dehydrogenase; G6PD, glucose-6-phosphate dehydrogenase.

Values followed by a common superscript do not differ significantly (P < 0.05).

about 30 h, and was reduced more in the force-fed group. This difference was highly significant ( $P < o \cdot o \cdot o \cdot$ ). The specific activity of liver XDH was increased by either force-feeding or fasting. Specific activities of liver 6PGD and G6PD were not affected by force-feeding, but the G6PD activity was reduced following fasting (Table 1). This effect was greater (P < 0.05) for the ad lib. group. Total activities of the liver enzymes increased concomitantly with the increase in liver weight of the force-fed chicks.

Pancreatic lipase, amylase, chymotrypsin and XDH total activities were significantly increased in the force-fed groups; lipase and XDH activities decreased after fasting. Trypsin activity was not affected significantly by either force-feeding or fasting (Table 2). Amylase activity (units/g) was increased in the small intestine contents of the force-fed group. No changes were observed in the activities of other enzymes in the small intestine when expressed as units/g. The total activities of these enzymes in the small intestine as units/g total chyme were increased in the forcefed group in parallel with the increase in the amount of the chyme, without a diluting effect. The small intestine content of the force-fed group was 63 % higher than that of the ad lib. group and 25 % higher when calculated as g/kg body-weight.

A general trend toward decrease was found in the enzyme activities of the caecum

<sup>\*</sup> See p. 242.

Table 2. Enzyme activities in the pancreas of chicks fed ad lib., or force-fed, before and after starving

(Mean values for eight chicks/group)

	Treatment					Statistical significance of effects of			
	Ad libfed		Force-fed						
Pancreas:	Fed	Starved	Fed	Starved	se of mean	Force- feeding P <	Starving P <	Interac- tion	
Weight (g)	1.418	1.09	$1.80^a$	1.64 <sup>ab</sup>	0.10	0.01	0.05	NS	
(g/kg body-wt) Enzyme activity (	4.4	4.1	4.2	4.6	0.3	NS	NS	NS	
Lipase	94ª	$66^{b}$	102a	$QI^a$	8	0.02	0.02	NS	
Amylase	94	105	103	107	9	NŠ	$\mathbf{N}\widetilde{\mathbf{S}}$	NS	
Trypsin	17.7	18.6	15.4	16.6	1.6	NS	NS	NS	
Chymotrypsin	84	82	97	99	II	NS	NS	NS	
XDH	0.237	0.512	0.289	0.228	0.045	NS	NS	NS	
(units*/pancreas	)								
Lipase	132b	71°	$193^{a}$	$149^{b}$	15	0.01	0.01	NS	
Amylase	$132^{b}$	114	195 <sup>a</sup>	175 <sup>a</sup>	16	0.01	NS	NS	
Trypsin	24.9	20.2	29.1	27.2	1.0	NS	NS	NS	
Chymotrypsin	1186	$89^b$	$183^{a}$	$162^a$	16	0.01	0.02	NS	
XDH	0.334	0.234	0.546	a 0.374 <sup>b</sup>	0.052	0.01	0.02	NS	

NS, non-significant; XDH, xanthine dehydrogenase.

Values followed by a common superscript do not differ significantly (P < 0.05).

Table 3. Enzyme activities in the small intestine and caecum of chicks fed ad lib. or force-fed

(Mean values for eight chicks/group)

	Treatment†						
	Ad lib fed	Force- fed	_	Ad lib fed	Force- fed		
			se of			se of	
a itt i it	(g/kg body-wt)		mean	(g)		mean	
Small intestine:			_				
Content	20.1 <sub>p</sub>	25.3a	1⋅8	$6.45^{b}$	10.50 <sup>a</sup>	0.72	
Enzyme activities	(unit	s*/g)		(units/g total chyme)			
Lipase	3.22	3.29	0.59	$22.8^{b}$	34.5 <sup>a</sup>	2.4	
Amylase	5.05 <sup>b</sup>	$8.71^{a}$	1.08	$32.6^{b}$	$91.5^{a}$	7.1	
Trypsin	3.84	3.37	0.10	24·8b	35·4 <sup>a</sup>	2 1	
Chymotrypsin	11.70	9.10	o·96	75.5	95.6	7.7	
Caecum:	(g/kg b		(g)				
Content	4.9	5.1	0.4	1.56	2.13	0.10	
Enzyme activities	(unit	s*/g)		(units/g total chyme)			
Lipase	4.68	4.26	0.35	7:30	9.07	0.21	
Amylase	2.47	1.26	1.09	3.85	2.68	1.08	
Trypsin	7.03 <sup>a</sup>	$3.01_{p}$	0.74	10.97	8.33	0.92	
Chymotrypsin	14.1	12.1	3.0	22.0	25.8	3.2	

Values followed by a common superscript do not differ significantly (P < 0.05).

<sup>\*</sup> See p. 242.

<sup>\*</sup> See p. 242.

<sup>†</sup> No intestinal content was found in the fasted birds.

contents when expressed as units/g. Only the reduction of trypsin activity was statistically significant (P < 0.05). Similar activities were observed in the total amount of the caecal chyme of both groups (Table 3).

#### DISCUSSION

Kidney arginase activity. The specific activity of kidney arginase was increased threefold in the force-fed groups compared with that of the ad lib.-fed groups. This increase was much higher than the 50% increase in body-weight gain, the 70% increase in protein consumption, or the 20% decrease in protein utilization. In earlier work with chicks (Dror & Nir, 1971), a threefold increase in the activity was recorded when chicks were given a 400 g soya-bean protein/kg diet, compared with one of 200 g protein. An increase of two- to threefold in arginase activity was reported by Muramatsu & Nakagawa (1971), when starved rats were force-fed tryptophan, while a smaller increase was found when force-feeding with other essential amino acids. The increase in arginase activity in the force-fed chicks may also be due to the creation of imbalance in some essential amino acids by force-feeding (Austic & Nesheim, 1971).

A starvation period of about 30 h caused a decrease of about 50% in the arginase activity. The higher activity of the force-fed group was reduced to a greater extent than the activity of the *ad lib*.-fed group. According to this rate of decrease of arginase activity (50% in 30 h), the half-life of the enzyme may be estimated to be less than 24 h. This estimate is comparable with the half-life of rat liver arginase which was found to be 24 h by Szepesi & Freedland (1969) in a non-steady state during which the activity decreased.

XDH activity. In liver, XDH specific activity was increased either by force-feeding or by fasting (Table 1). The increase due to fasting confirms the result of Scholz & Featherston (1969), obtained when chicks given 250 g protein/kg diet were starved for 1 d. The latter increase is a result of protein catabolism during fasting. The increase was greater in the ad lib.-fed group, which contained 6% carcass fat compared with 14% in the force-fed group, and therefore used more protein as a source of energy than did the latter group, which utilized more fat during fasting (Nir et al. 1974).

A non-significant increase in pancreatic XDH specific activity was caused by the force-feeding. The response of XDH activities in the pancreas to the nutritional treatment was smaller than that found in the liver, but a small non-significant decrease was observed during fasting. Fisher, Curtis & Woodward (1967) showed that the pancreatic XDH activity in hatching chicks is developed only after they begin to eat and that fasting for 1 d resulted in a decrease in XDH activity of about 40 %. They suggested different control mechanisms for XDH activity in liver and pancreas. The liver XDH activity (Table 1) was increased after fasting and the pancreatic XDH (Table 2) was decreased. This diversity may be due to the different control mechanism of XDH activity in liver and pancreas, as suggested by Fisher et al. (1967).

Pentose phosphate cycle enzyme activities. Specific activities of 6PGD and G6PD were only slightly affected by force-feeding but G6PD activity decreased by about 60% following starvation. The lack of response of these activities to another extreme

nutritional treatment in chicks, such as a high-carbohydrate diet, was reported by Pearce (1972). Rats could differ from chicks in this respect. A marked increase in response to the same treatment was reported by Johnson & Sassoon (1968) in rats. A twofold increase in 6PGD activity and a 50 % decrease in G6PD activity in force-fed geese was reported by Nitsan *et al.* (1973). There could be some difference in the importance of pentose phosphate cycle enzymes between chicks and geese, but this could be due also to the higher fat content of the livers of the geese.

Digestive enzymes levels. In the pancreas and small intestine, the increase in the total activity of these enzymes paralleled the 70% increase in food consumption brought about by force-feeding compared with the ad lib.-fed group. The specific activities of the enzymes per g tissue or chyme were therefore the same in the two groups despite the bigger pancreas and increased amount of chyme in the intestines of the force-fed group. Force-feeding of maize in an amount two- to fourfold that of the ad lib. intake described in geese (Nitsan et al. 1973), caused a reduction in proteolytic and amylase activities in the enlarged digestive tract. However, the total activities of proteases in the excreta were increased, while that of amylase decreased (Nir et al. 1973). The different results obtained with the force-fed chicks and geese may be due either to the level of force-feeding or to the quality of the force-fed diet (maize v. starter mash). Increased food consumption level (170%) is accompanied by an increase in synthesis of digestive enzymes and secretion at the same rate, so that no dilution effect is seen despite the bigger pancreas and intestinal content. However, in force-fed geese (intake 200-400 % of ad lib. intake), although the enzyme synthesis was somewhat increased, it did not parallel the increase in the feeding level and therefore the specific activity/g chyme was reduced.

A good-quality diet supplies all the needed precursors for enzyme synthesis and therefore the force-fed chicks could produce higher amounts of digestive enzymes depending on the level of force-feeding. Maize, however, is probably deficient in some of the essential precursors for enzyme synthesis. A marked increase in the amylase secretion was observed in force-fed geese when the maize was supplemented by soya-bean meal (Nir et al. 1973). Amylase was found to be affected more than the proteolytic enzymes by the quality of the diet and responded more to methionine supplementation of the soya-bean diet (Nitsan & Gertler, 1972). In the present work, in which a good-quality mash was used, the amylase activity per g intestinal chyme was even greater in the force-fed than in the ad lib.-fed chicks (Table 3).

Digestibility of various nutrients is not changed by a wide range of food intake and digestive enzyme levels (Scott, Nesheim & Young, 1969; Nir et al. 1973; Dror, Shamgar & Budowski, unpublished results). The increased enzyme activity found in force-feeding is not necessarily needed for better digestion, but can be a result of mechanical and humoral stimulation caused by the passage of greater amounts of chyme through the digestive tract. It appears that the mechanism inducing the synthesis of enzymes of the digestive tract is not predominantly controlled by the amounts and concentrations of substrates or products in the gastrointestinal tract.

The authors acknowledge with thanks the generous contribution of Mr Joseph Taylor from London, England, which enabled them to carry out this work.

#### REFERENCES

Austic, R. E. & Nesheim, M. C. (1971). J. Nutr. 101, 1403.

Bernfeld, P. (1955). Meth. Enzym. I, 149.

Dror, Y. & Gertler, A. (1967). J. Nutr. 93, 401.

Dror, Y. & Nir, I. (1971). Nutr. Metab. 13, 75.

Dror, Y., Sassoon, H. F., Watson, J. J. & Johnson, B. C. (1970). Clinica chim. Acta 28, 291.

Fisher, J. R., Curtis, J. L. & Woodward, W. D. (1967). Devl Biol. 15, 289.

Gertler, A. & Nitsan, Z. (1970). Br. J. Nutr. 24, 893.

Johnson, B. C. & Sassoon, H. T. (1968). Proc. 7th int. Congr. Nutr. p. 496.

Muramatsu, K. & Nakagawa, T. (1971). Agric. biol. Chem. J. 35, 1594.

Nir, I., Nitsan, Z. & Vax, A. (1973). Annls Biol. anim. Biochim. Biophys. 13, 465.

Nir, I., Shapira, N., Nitsan, Z. & Dror, Y. (1974). Br. J. Nutr. 32, 229.

Nitsan, Z. & Gertler, A. (1972). Br. J. Nutr. 27, 337.

Nitsan, Z., Nir, I., Dror, Y. & Bruckental, I. (1973). Poult. Sci. 52, 474.

Pearce, J. (1972). Biochem. J. 130, 21P.

Sarda, L. & Desnuelle, P. (1958). Biochim. biophys. Acta 30, 513.

Scholz, R. W. & Featherston, W. R. (1969). J. Nutr. 98, 193.

Scott, M. L., Nesheim, M. C. & Young, R. J. (1969). Nutrition of the Chicken. Ithaca, New York: M. L. Scott and Associates.

Snedecor, G. W. & Cochran, W. G. (1967). Statistical Methods 6th ed. Ames, Iowa: Iowa State University.

Strittmatter, C. F. (1965). J. biol. Chem. 240, 2557.

Szepesi, B. & Freedland, R. A. (1969). Life Sci. II 8, 1067.