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TRAPPING SUBFEMTOLITER-SIZED VESICLES, AND ANALYZING THEM

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The chemical analysis of secretory vesicles has long been limited by the fact that many vesicles are analyzed together, resulting in an averaging of a population. Differences between vesicles within that population could not be detected. Wouldn't it be great if vesicles could be analyzed one at a time? You guessed it, this breakthrough has been recently achieved.

The technique of capillary electrophoresis (CE) was modified by Daniel Chiu, Sheri Lillard, Richard Scheller, Richard Zare, Sandra Rodriguez-Cruz, Evan Williams, Owe Orwar, Mats Sandberg, and Anders Lundqvist so that vesicles could be probed singly.² They pointed out that although we have been limited to analyzing populations of vesicles, there are examples in biology where the contents of a single vesicle is sufficient to elicit a response. But the problem remained of how to capture and analyze a vesicle in the femto- (10⁻¹⁶) to atto (10⁻¹⁶) liter size range.

It was demonstrated a few years ago that a single dye molecule could be detected by laser-induced fluorescence (LIF). However, detection is only the back half of the problem. The challenge that Chiu *et al.* addressed was the introduction and manipulation of an ultra-small sample. They chose the dense core vesicles from the atrial gland of the gastropod mollusk *Aplysia californica* as the test sample. These easily-isolated vesicles are known to contain bioactive peptides that are involved in the animal's reproductive behavior. Careful analysis of a (relatively speaking) bulk sample with several available methods showed that these vesicles contained NH₂-terminal peptides (NTPs), low-mass compounds (LMCs), and an unidentified peak (referred to as peak 4). Taurine was the main component of the LMCs, and they included leucine, isoleucine, and other modified or unusual amino acids.

To analyze a single vesicle, it was first optically trapped and a tapered capillary tip was moved into position next to it. The vesicle was about 0.5 microns in diameter, and the inside diameter of the tip was about ten times larger. A small voltage was applied which electrokinetically moved the vesicle into the tip. In video microscopic images published in their article,

one could literally see the vesicle streaking into the capillary tip. With a single vesicle inside the tip, the capillary inlet was transferred into a small droplet of a reaction mixture containing naphthalene-2,3-dicarboxaldehyde (NDA) and potassium cyanide in borate buffer. A small amount of this reaction mixture is electrokinetically injected into the tapered inlet to lyse the vesicle. The lysate is reacted inside the capillary column with NDA, separated electrophoretically, and the components detected with LIF. Ten vesicles were individually analyzed in this fashion. They were all found to contain NTPs in roughly equal quantities, and either taurine or peak 4; taurine and peak 4 were never present in the same vesicle. When two or more vesicles were analyzed together, both taurine and peak 4 were detected. It was suggested that there could be two distinct population of vesicles (one containing taurine, the other peak 4), or a single population of vesicles whose composition changes with maturity.

Chiu *et aL* have developed a method that can separate and probe the chemical messages within single vesicles of only attoliters of volume. Whereas these vesicles from *Aplysia* were rather large, we can logically expect that the method will be refined so that smaller vesicles can be examined in the future, even to the zepto (10⁻²¹) liter range. This will make it possible to address many previously unanswerable biologic questions. The heterogeneity of vesicle populations is no longer an insurmountable problem to determining the composition of a single quantum of intercellular communication.

1 The author gratefully acknowledges Dr. Richard Zare for reviewing this article. 2 Chiu, D.T., S.J. Lillard, R.H. Scheller, R.N. Zare, S.E. Rodriguez-Crnz, E.R. Williams, W. Onvar, M. Sandberg. and J.A. Lundqvist. Probing single secretory vesicles with capillary electrophoresis. *Science* **279**: 1190-1193. 1998.

Our next issue, combined for June and July, will be "special" in that it will be mailed well prior to, and handed out at, the MSA/MAS '98 Conference in Atlanta. The closing date for copy will be 19 June.

Front Page Image SEM Photograph of a Mosquito - Female (Aedes Aegypti)

Imaged on an ETEC SEM at 5 kV using the SEM Wideband Multi-Detector Color Synthesizer (designed, built and patented by David Scharf). Then acquired digitally at 2,048 X 1,536 pixels directly into a Macintosh Power PC at a TIFF file, using Digital Micrograph software and Digiscan hardware. Then output to a CELCO film recorder, using Ektachrome 100+ film, to produce a 4x5 transparency.

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