

Whole body and muscle energy metabolism in preruminant calves: effects of nutrient synchrony and physical activity

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The effects of asynchronous availability of amino acids and glucose on muscle composition and enzyme activities in skeletal muscle were studied in preruminant calves. It was hypothesized that decreased oxidative enzyme activities in muscle would explain a decreased whole body heat production with decreasing nutrient synchrony. Preruminant calves were assigned to one of six degrees of nutrient synchrony, step-wise separating the intake of protein and lactose over the two daily meals. Calves at the most synchronous treatment received two identical meals daily. At the most asynchronous treatment, 85 % of the daily protein and 20 % of the daily lactose supply were fed in one meal and the remainder in the other meal. Daily intakes of all dietary ingredients were identical for all treatments. Oxidative enzyme activities and fat content increased with decreasing nutrient synchrony in *M. Rectus Abdominis* (RA), but not in *M. Semitendinosus*. Cytochrome-*c*-oxidase activity was positively correlated with fat content in RA (r 0.49; $P < 0.01$). Oxidative enzyme activities in both muscles were not correlated with average daily heat production, but citrate synthase activity in RA was positively correlated ($P < 0.01$) with the circadian amplitude (r 0.53) and maximum (r 0.61) of heat production associated with physical activity. In conclusion, this study indicates that muscle energy stores are regulated by nutrient synchrony. The lack of correlation between muscle oxidative enzyme activities and average daily heat production was in contrast with findings in human subjects. Therefore, oxidative enzyme activity in muscle should not be used as an indicator for whole body heat production in growing calves.

Energy metabolism: Muscle: Nutrient synchrony: Physical activity

Daily nutrient intake has been correlated with muscle enzyme activities in animals (Cassar-Malek *et al.* 2004) and man (Helge & Kiens, 1997), and with intramuscular fat and glycogen contents in animals (Pethick & Rowe, 1996; Gondret *et al.* 2000). Regulation of muscle oxidative enzyme activities and muscle composition also depends on the supply of individual macronutrients (Hocquette *et al.* 1998; Geelen *et al.* 2001; Gondret & Leuret, 2002). Apart from variation in daily nutrient supply, within-day variation in the supply of different nutrients (i.e. synchrony) may induce these effects. Asynchronous absorption patterns can be induced by either a separated intake of protein and carbohydrates in time (e.g. dissociated diets) or by supplying ingredients with different kinetics of digestion and absorption. In preruminants, for example, asynchronous absorption of glucose and amino acids may result from differences in passage behaviour of clotting *v.* non-clotting dietary ingredients (Guilloteau *et al.* 1986; Verdonk *et al.* 1999). Applying a theoretical approach, we separated the intake of glucose and amino acids within the day (i.e. across meals) in heavy preruminant calves (Van den Borne *et al.*

2006b). Surprisingly, and in contrast to similar investigations in pigs (JJGC Van den Borne, JW Schrama, MJW Heetkamp, MWA Verstegen and WJJ Gerrits, unpublished results), increased separation of the amino acid and glucose availability within a day decreased whole body heat production (–9%), and increased whole body fat deposition (+59%), in heavy preruminant calves (Van den Borne *et al.* 2006b). Muscle energy metabolism may contribute to these changes as it accounts for about 20 % of the daily heat production in growing farm animals (Ortigue *et al.* 1995). Therefore, muscle energy metabolism may help to spare energy in case of an asynchronous supply of protein and lactose. Effects of nutrient synchrony on the energy metabolism and composition of muscles may also have implications for muscle function, meat quality and development of metabolic disorders in calves.

The regulation of muscle energy metabolism is well documented (e.g. Hocquette *et al.* 1998). In non-growing man, oxidative enzyme activities in muscle are positively correlated with whole body heat production (Zurlo *et al.* 1994; Doucet

Abbreviations: BW, body weight; COX, cytochrome-*c*-oxidase; CS, citrate synthase; H_{act} , heat production associated with physical activity; H_{cor} , heat production corrected for physical activity; H_{tot} , total heat production; LDH, lactate dehydrogenase; ME_m , metabolizable energy requirements for maintenance; RA, *M. Rectus Abdominis*; SEQ, meal sequence; ST, *M. Semitendinosus*; SYN, degree of nutrient synchrony.

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calf. For calculating circadian means of heat production by the formula of Brouwer (1965), the nitrogen excretion was assumed to be constant throughout the day.

Measurement of muscle composition and enzyme activities

After five weeks on the experimental treatment, calves were transported to the slaughterhouse (20 min) and killed at 13.00 h (i.e. 7 h after the morning meal) by stunning and exsanguination. Within 15 min post-slaughtering, samples of the *M. Rectus Abdominis* (RA; oxido-glycolytic muscle) and *M. Semitendinosus* (ST; glycolytic muscle) were removed. The samples of skeletal muscle were immediately trimmed of visible fat and connective tissue. Samples were then cut into pieces, immediately frozen in liquid N₂ and stored at -80°C pending analyses. Samples for crude fat analysis (about 30 g for each muscle) were not frozen in liquid N₂, but ice-chilled and subsequently stored at -20°C pending analyses.

Protein (Bradford, 1976) and DNA (Labarca & Paigen, 1980) contents were measured in muscle homogenates. Activity of the oxidative enzyme citrate synthase (CS; EC 4.1.3.7), which is involved in the substrate flux through the tricarboxylic acid cycle, was determined in sonicated homogenates by measuring the rate of initial reaction at 412 nm by means of the DTNB (5,5'-dithiobis(2-nitrobenzoate)) method as described by Shepherd & Garland (1969). Activity of the oxidative enzyme cytochrome-*c*-oxidase (COX; EC 1.9.3.1), which is involved in the substrate flux through the respiratory chain, was determined in sonicated homogenates by measuring the oxidation of reduced cytochrome-*c* as described by Van Hinsberg *et al.* (1978). Activities of CS and COX were expressed in $\mu\text{mol CoA}$ liberated per min per g protein and in $\mu\text{mol cytochrome-}c$ oxidized per min per g protein at 25°C respectively. Activity of the glycolytic enzyme lactate dehydrogenase (LDH; EC 1.1.1.2.7), which catalyzes the formation of lactate from pyruvate, was determined as described by Ansay (1974). LDH activity was expressed in $\mu\text{mol NADH}$ oxidized per min per g protein. Glycogen content was measured by the method of Carroll *et al.* (1955). Briefly, glycogen was extracted from the tissue by homogenization with 5% trichloroacetic acid solution, precipitated from the extract by 95% ethanol and determined with the anthrone reagent in a colorimeter at 620 nm. Fat content was measured in muscle tissue after freeze-drying according to ISO 6492 (International Organization for Standardization, 1999). Ratios for CS:LDH, COX:LDH (relative enzyme activities), protein:DNA (indicator for muscle fibre size) and COX:CS (indicator for the biochemical properties of the mitochondria) were calculated.

Statistical analysis

The effects of nutrient synchrony and the interaction between nutrient synchrony and meal sequence on muscle traits and whole body energy metabolism traits were analysed by linear regression analysis, using the general linear model procedure of the SAS statistical software package version 9.1 (Statistical Analysis Systems Institute, Cary, NC, USA). The degree of nutrient synchrony was included as a regressor

(Default 1).

$$Y_{ij} = \mu + \beta_1 \times X_j + \beta_{2i} \times S_i X_j + \varepsilon_{ij}, \quad (1)$$

where Y_{ij} = dependent variable over the whole period, μ = average intercept, β_1 = effect of degree of nutrient synchrony, expressed as percentage of the daily protein intake in the high protein meal, β_{2i} = interaction between degree of nutrient synchrony and meal sequence i , S_i = fixed effect of meal sequence i , X_j = degree of nutrient synchrony (expressed as percentage of the daily protein intake in the high protein meal) for calf j , ε_{ij} = error term, $i = 1, 2$, and $j = 1, \dots, 18$.

Pearson correlation coefficients were calculated for relationships between muscle energy metabolism traits within each muscle, for each trait between the two muscles and between muscle energy metabolism traits and whole body energy metabolism traits. Traits for the two muscles were compared by pairwise comparisons.

Results

Animal performance

Two calves were excluded from analysis, because of feed refusals or illness. The initial BW of the calves was 128 (SEM 1.7) kg and average daily gain during the 5 week experimental period was 885 (SEM 27.1) g. Average daily gain was lower ($P < 0.05$) for calves at SYN 5 and 6 (729 and 815 g respectively) than for calves at SYN 1–4. Calves at SYN 1–4 had a similar daily gain (on average 945 g). Feed intake did not differ between treatments and consequently the feed to gain ratio was higher ($P < 0.01$) for calves at SYN 5 and 6 (2.56 and 2.48 respectively) than for calves at the other treatments (on average 2.02). However, calves at the two most asynchronous treatments had a lower nutrient digestibility. The consequently lower digestible nutrient intake in calves at SYN 5 and 6 complicated interpretation of the results from SYN 1 to 6. Linear regression was therefore also performed for SYN 1 to 4 ($n = 22$) separately, because calves at those treatments had identical digestible nutrient intakes.

Muscle enzyme activities

In RA, activities of COX and CS increased ($P = 0.048$) and tended to increase ($P = 0.075$), respectively, with decreasing nutrient synchrony (SYN 1–4; Table 2). Ratios of enzyme activities were not affected by nutrient synchrony. In ST, enzyme activities were not affected by nutrient synchrony (Table 3). The sequence of the high protein and high carbohydrate meals did not affect enzyme activities.

Muscle composition

Intramuscular fat content increased in RA ($P = 0.037$) with decreasing nutrient synchrony at an identical digestible nutrient intake (Table 2), but nutrient synchrony did not affect intramuscular fat content in ST (Table 3). The fat content was substantially higher ($P < 0.001$) in RA (18.9 mg/g tissue) than in ST (6.5 mg/g tissue). The sequence of the high protein and high carbohydrate meals did not affect intramuscular fat content.

Table 2. Effects of nutrient synchrony on muscle enzyme activities and muscle composition in *M. Rectus Abdominis* of heavy preruminant calves (Values are means for treatments with their standard errors; *n* 5 per treatment for SYN 1 and 2, and *n* 6 per treatment for SYN 3–6)

Treatment (SYN)...	1	2	3	4	5	6	SYN 1–6			SYN 1–4		
	50	57	64	71	78	85	SEM	<i>b</i> *	<i>P</i> value†	SEM	<i>b</i>	<i>P</i> value
Enzyme activity ($\mu\text{mol/g}$ protein per min)												
LDH	4317	4269	4502	4447	4358	4076	286.2	-4.5×10^{-3}	0.673	344.4	8.7×10^{-3}	0.710
COX	119	97	119	154	121	99	13.7	0.2×10^{-4}	0.960	14.1	2.0×10^{-3}	0.048
CS	54	52	57	67	58	51	5.3	0.4×10^{-4}	0.850	5.0	0.7×10^{-3}	0.075
Relative enzyme activity												
COX:CS	2.22	1.87	2.16	2.70	2.33	2.00	0.321	2.3×10^{-3}	0.849	0.299	0.01	0.499
COX:LDH, $\times 10^{-3}$	27.8	24.6	27.7	35.3	28.4	24.2	3.92	2.1×10^{-6}	0.989	4.32	0.4×10^{-3}	0.191
CS:LDH, $\times 10^{-3}$	12.4	12.8	12.9	15.9	13.8	12.3	1.53	0.2×10^{-4}	0.728	1.48	0.2×10^{-3}	0.180
Muscle composition												
Glycogen(mg/g fresh tissue)‡	5.9	7.6	6.2	6.0	5.5	6.1	1.14	-0.02	0.630	1.25	-9.0×10^{-3}	0.907
Fat (mg/g fresh tissue)	16.3	13.5	21.9	23.9	16.3	13.7	2.46	-0.03	0.802	2.43	0.44	0.037
Protein (mg/g fresh tissue)	165	175	170	158	162	172	7.0	-0.07	0.764	7.7	-0.45	0.391
DNA ($\mu\text{g/g}$ fresh tissue)	1881	1972	2017	1996	1977	1989	55.1	1.95	0.309	47.9	5.75	0.088
Protein:DNA (mg/mg)	88.1	88.8	84.8	79.4	82.3	87.0	4.39	-0.1×10^{-3}	0.435	4.92	-0.5×10^{-3}	0.140

CON, contribution of the high protein meal to the daily protein supply (50% = two identical, balanced meals; 100% = all daily protein in one meal); SYN, degree of nutrient synchrony; LDH, lactate dehydrogenase; COX, cytochrome-c-oxidase; CS, citrate synthase.

* Regression coefficient ($y = a + b \times x$), representing the change in response parameter per % increase of the contribution of the high protein meal to the daily protein supply.

† Probability for test if the regression coefficient (*b*) equals 0. Effects of meal sequence and interactions between meal sequence and degree of nutrient synchrony were non-significant and were therefore excluded from the model.

‡ An interaction between CON and the meal sequence on muscle glycogen content was found ($P=0.023$).

Table 3. Effects of nutrient synchrony on muscle enzyme activities and muscle composition in *M. Semitendinosus* of heavy preruminant calves (Values are means for treatments with their standard errors; *n* 5 per treatment for SYN 1 and 2, and *n* 6 per treatment for SYN 3–6)

Treatment (SYN)...	1	2	3	4	5	6	SYN 1–6			SYN 1–4		
	50	57	64	71	78	85	SEM	<i>b</i> *	<i>P</i> value†	SEM	<i>b</i>	<i>P</i> value
Enzyme activity ($\mu\text{mol/g}$ protein per min)												
LDH	4167	4618	4500	4738	4381	4562	242.2	5.2×10^{-3}	0.543	272.9	0.2×10^{-4}	0.240
COX	113	94	78	103	90	96	12.7	-0.2×10^{-3}	0.544	12.8	-0.5×10^{-3}	0.525
CS	46	53	43	61	54	58	4.9	0.3×10^{-3}	0.095	5.4	0.5×10^{-3}	0.166
Relative enzyme activity												
COX:CS	2.51	1.80	2.04	1.72	1.68	1.62	0.264	-0.02	0.051	0.326	-0.03	0.199
COX:LDH, $\times 10^{-3}$	29.4	21.2	17.8	22.0	20.9	21.6	3.91	-0.1×10^{-3}	0.347	4.14	-0.3×10^{-3}	0.267
CS:LDH, $\times 10^{-3}$	11.5	11.8	9.7	13.1	12.4	12.8	1.37	0.1×10^{-3}	0.311	1.53	0.6×10^{-4}	0.533
Muscle composition												
Glycogen (mg/g fresh tissue)‡	6.7	6.6	8.2	6.4	6.8	6.8	1.06	-1.4×10^{-3}	0.969	1.14	-0.3×10^{-3}	0.881
Fat (mg/g fresh tissue)	5.6	5.4	7.4	7.6	6.1	6.0	0.98	0.01	0.717	1.19	0.12	0.166
Protein (mg/g fresh tissue)	177	171	171	166	172	175	5.5	-0.04	0.841	6.9	-0.42	0.283
DNA ($\mu\text{g/g}$ fresh tissue)	1964	1880	1908	1950	1944	1844	80.8	-1.57	0.594	84.9	-0.74	0.889
Protein:DNA (mg/mg)	92.7	93.1	89.6	85.9	90.2	94.8	5.69	0.1×10^{-4}	0.961	6.62	0.01	0.475

CON, contribution of the high protein meal to the daily protein supply (50% = two identical, balanced meals; 100% = all daily protein in one meal); SYN, degree of nutrient synchrony; LDH, lactate dehydrogenase; COX, cytochrome-c-oxidase; CS, citrate synthase.

* Regression coefficient ($y = a + b \times x$), representing the change in response parameter per % increase of the contribution of the high protein meal to the daily protein supply.

† Probability for test if the regression coefficient (*b*) equals 0. Effects of meal sequence and interactions between meal sequence and degree of nutrient synchrony were non-significant and were therefore excluded from the model.

‡ An interaction between CON and the meal sequence on muscle glycogen content was found ($P=0.007$).

Glycogen content in both muscles was not affected by nutrient synchrony (Tables 2 and 3), but a positive correlation between glycogen content in RA and ST was found (r 0.72; $P < 0.001$). Glycogen content averaged 6.24 mg/g in RA and 7.01 mg/g in ST. An interaction between SEQ and SYN was found for intramuscular glycogen content in both muscles ($P < 0.05$), indicating that glycogen content was higher after feeding the high protein diet than after the high lactose diet. The difference in glycogen content between meal sequences was more pronounced at SYN 2 (3.08 mg/g in RA and 6.08 mg/g in ST) and SYN 3 (4.63 mg/g in RA and 4.96 mg/g in ST) than at the other treatments.

Whole body heat production

Average heat production and H_{cor} decreased with decreasing nutrient synchrony ($P < 0.05$), while the RQ increased ($P = 0.028$; Table 4). A detailed description of whole body energy partitioning and protein deposition is presented and discussed elsewhere (Van den Borne *et al.* 2006b). The maxima of the circadian patterns of H_{tot} and H_{cor} decreased with decreasing nutrient synchrony ($P < 0.05$), but the maximum of the circadian pattern of H_{act} was not affected by nutrient synchrony. The amplitudes of the circadian patterns of H_{tot} , H_{cor} and H_{act} were not affected by SYN. The sequence of the high protein and high carbohydrate meals did not affect any of the traits of whole body energy metabolism.

Correlations within muscle

In both muscles, CS and COX activities were positively correlated (r 0.35–0.36; $P < 0.05$). Intramuscular fat content showed a positive correlation (r 0.49; $P < 0.01$) with COX (and COX:LDH) in RA (data not shown).

Correlations between muscle energy metabolism and whole body energy metabolism

The decreased H_{tot} and H_{cor} with decreasing nutrient synchrony did not correspond with the increased (or unaffected) oxidative enzyme activities in muscle. In accordance, simple correlation analysis showed that average H_{tot} , H_{cor} and H_{act} were not correlated with enzyme activities in RA (Table 5) and ST (data not shown). Intramuscular fat content in RA was negatively correlated with average H_{tot} and H_{cor} .

The maximum of H_{act} and the amplitude of H_{tot} and H_{act} increased with increasing CS activity in RA in preruminant calves, resulting in positive correlations (Table 5). In ST, oxidative enzyme activities were not related to the maxima and amplitudes of heat production traits (data not shown).

Discussion

Animal performance

Feeding more than 71% of the daily protein in one meal and more than 68% of the daily lactose in the other one (i.e. SYN 5–6) induced diarrhoea, which was indicated by a lower faecal dry matter content, a lower faecal pH and a lower nutrient digestibility (Van den Borne *et al.* 2006b). The lower daily gain and higher feed to gain ratio for calves at SYN 5 and 6

were therefore likely due to a decreased digestible nutrient intake. Comparison of calves at SYN 1–4 allows studying the effects of nutrient synchrony at identical intakes of digestible nutrients. Therefore, this discussion focuses on treatments SYN 1–4.

Effects of nutrient synchrony on muscle enzyme activities

Although the quantity (Brandstetter *et al.* 1998; Cassar-Malek *et al.* 2004) and the composition (Helge & Kiens, 1997; Geelen *et al.* 2001; Cuvelier *et al.* 2006) of the daily feed supply are known to affect oxidative and glycolytic enzyme activities in muscles, this study is one of the first to describe the effect of within-day distribution of nutrient availability on muscle metabolism. Ortigues-Marty *et al.* (2003) found increased oxidative enzyme activities in muscle (ST) of preruminant calves when skimmed milk protein was replaced by soluble wheat and whey proteins. Feeding soluble wheat and whey proteins resulted in larger circadian fluctuations of the amino acid availability compared with skimmed milk protein. Therefore, it was suggested that a transiently high amino acid availability may require an increased capacity of oxidative enzymes in muscle to provide energy for protein synthesis when amino acids are available (Ortigues-Marty *et al.* 2003). However, potential reasons for the affected oxidative capacity in the study of Ortigues-Marty *et al.* (2003) also include a lower intake of the skimmed milk diet than of the wheat and whey protein diet and the use of different dietary ingredients with different kinetics of fatty acid absorption (Petit *et al.* 1988; Cruywagen *et al.* 1990; Ortigues-Marty *et al.* 2003).

A decreased nutrient synchrony (i.e. increased circadian fluctuations in nutrient supply) also increased oxidative enzyme activities in RA in the current study (Table 2), but not in ST. Separating the availability of amino acids and glucose may have required temporarily high oxidative enzyme activities to provide energy for protein synthesis (see Ortigues-Marty *et al.* 2003) or to oxidize excessively available amino acids or glucose. As net protein utilization was unaffected and fat deposition was increased (Van den Borne *et al.* 2006b), temporarily increased rates of glucose and/or amino acid oxidation should be accompanied by decreased oxidation rates of oxidation during the remainder of the day.

It is noteworthy that nutrient synchrony only affected muscle enzyme activities in an oxidative muscle (RA), but not in a glycolytic muscle (ST). This corresponds with previous studies which demonstrated a higher response of oxidative (mitochondria-rich) than glycolytic muscles to nutrient supply (Cassar-Malek *et al.* 2004; Jurie *et al.* 2006).

Effects of nutrient synchrony on muscle composition

Intramuscular fat content was almost threefold higher in RA than in ST, which agrees with the generally described higher fat content in oxidative than in glycolytic muscles (Gondret *et al.* 1998; Hocquette *et al.* 2003). The increased intramuscular fat content in RA with decreasing nutrient synchrony corresponds to an increased whole body fat deposition in these calves (Van den Borne *et al.* 2006b) indicating that the extra fat is at least partly deposited in muscles. Intramuscular fat generally develops after abdominal, intermuscular and

Table 5. Pearson correlation coefficients between oxidative enzyme activities and muscle composition in *M. Rectus Abdominis* and whole body energy metabolism traits in heavy preruminant calves (*n* 22)

Trait	CS	COX	CS:LDH	COX:LDH	Fat	Glycogen
Average						
Heat production	-0.06	-0.33	-0.22	-0.42	-0.49*	-0.06
Activity related heat production	0.04	-0.41	0.06	-0.29	-0.43*	0.09
Activity corrected heat production	-0.09	-0.20	-0.28	-0.36	-0.37	-0.11
Maximum						
Heat production	0.28	-0.29	0.19	-0.27	-0.35	0.08
Activity related heat production	0.53**	-0.12	0.58**	-0.10	-0.22	0.26
Activity corrected heat production	-0.12	-0.26	-0.27	-0.39	-0.44*	-0.14
Amplitude						
Heat production	0.63**	0.08	0.66***	0.10	-0.08	0.16
Activity related heat production	0.61**	-0.14	0.58**	-0.11	-0.20	0.19
Activity corrected heat production	-0.04	-0.14	-0.05	-0.14	-0.21	-0.26

LDH, lactate dehydrogenase; COX, cytochrome-*c*-oxidase; CS, citrate synthase.

* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

was positively correlated with the circadian maximum and amplitude of H_{act} in growing calves. A similar correlation was found with H_{tot} , which can be explained by the high contribution of variation in H_{act} to variation in H_{tot} .

The high feeding level in preruminant calves may explain why oxidative enzyme activity in muscle showed a better relationship with within-day variation in H_{act} than with average H_{tot} . Apart from muscle, several other tissues contribute to whole body heat production. Feeding large amounts of nutrients results in a high diet-induced heat production predominantly caused by portal drained viscera and liver (Ortigue *et al.* 1995). This may also explain the different results in growing calves and non-growing man. The diet-induced heat production usually contributes for 30 to 35 % to H_{tot} in growing farm animals (Collin *et al.* 2001; Le Bellego *et al.* 2001; Van Milgen *et al.* 2001), but only for 5 to 15 % to H_{tot} in adult human subjects (Westerterp, 2004). Consequently, the overall contribution of H_{act} and muscle energy metabolism to average H_{tot} is likely to be higher in man than in calves. The individual housing conditions in the current study restricted the physical activity of calves, whereas calves under practical circumstances are housed in groups. Although group-housing only marginally increased H_{act} compared with individual housing as shown in pigs (Rijnen, 2003), this may have contributed to the lack of correlation between oxidative enzyme activity and H_{tot} in this study.

Oxidative enzyme activities in muscle were not correlated with the average H_{act} but only with the circadian fluctuations in H_{act} (maximum and amplitude) in preruminant calves. This may be explained by the measurement of enzyme activity, which truly reflects the maximum enzyme activity rather than the enzyme activity *in vivo*. Variation in fatty acid oxidation, for example, is not necessarily associated with variation in oxidative enzyme activities in muscle (Piot *et al.* 1998). Enzyme activities depend not only on the capacity of enzymes to convert substrates into new (co or end) products, but also on the availability of substrates. The availability of substrates for oxidative enzymes within a day is affected by nutrient supply, but also by particular energy requiring processes like physical activity. It can therefore be expected that the maximum oxidative enzyme activity

is regulated by the kinetics of substrate availability rather than by the daily average substrate availability. Although the major aim of this study was to determine effects of variation in nutrient supply on muscle energy metabolism, it appeared that variation in H_{act} was positively correlated with CS activity in RA.

The positive correlation between physical activity and CS activity in muscle corresponds with results obtained in man (Rimbert *et al.* 2004), rats (Spangenberg *et al.* 2005), pigs (Petersen *et al.* 1997) and steers (Jurie *et al.* 2006). Similarly, muscle CS activity was 23 % higher in group-housed calves than in individually housed calves (Ortigue-Marty *et al.* 2003) which likely relates to increased physical activity. Previous studies have, however, not related the muscle enzyme activity to *in vivo* energy expenditure. Therefore, the current study is one of the first to relate CS activity in muscle to heat production associated with physical activity (and hence to contractile activity in muscle tissue).

Conclusions

From the results of the present study it was concluded that a decreased nutrient synchrony increased oxidative enzyme activity and intramuscular fat content in the oxidative muscle RA of preruminant calves. This indicates that the within-day distribution of macronutrient availability can affect muscle composition and properties which may have consequences for muscle function and meat quality. However, the decrease in whole body heat production with increasing separation of protein and lactose intake could not be explained by a decrease in oxidative enzyme activities in muscle.

In human subjects, muscle enzyme activities relate to whole body energy expenditure. Consequently, enzyme activities can be used as predictors for energy expenditure. Our data indicate that this relationship is present only for the activity associated component of energy expenditure in growing preruminant calves. Therefore, this relationship loses significance when subjects are fed at higher intake levels. Consequently, muscle enzyme activities are probably useless as an indicator for whole body energy expenditure in rapidly growing animals.

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