

Original Article

*Undergraduate.

Cite this article: Opresko DM, Goldman SL, Johnson R, Parra K, Nuttall M, Schmahl GP, Brugler MR (2020). Morphological and molecular characterization of a new species of black coral from Elvers Bank, north-western Gulf of Mexico (Cnidaria: Anthozoa: Hexacorallia: Antipatharia: Aphanipathidae: Distichopathes). *Journal of the Marine Biological Association of the United Kingdom* **100**, 559–566. <https://doi.org/10.1017/S002531542000051X>

Received: 29 January 2020

Revised: 4 May 2020

Accepted: 27 May 2020

First published online: 8 July 2020

Key words:

Distichopathes; DNA barcoding; Flower Garden Banks National Marine Sanctuary; mesophotic coral ecosystem; scanning electron microscopy; taxonomy

Author for correspondence:


Mercer R. Brugler,

E-mail: mbrugler@amnh.org

© The Author(s), 2020. Published by Cambridge University Press on behalf of Marine Biological Association of the United Kingdom. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted re-use, distribution, and reproduction in any medium, provided the original work is properly cited.



Morphological and molecular characterization of a new species of black coral from Elvers Bank, north-western Gulf of Mexico (Cnidaria: Anthozoa: Hexacorallia: Antipatharia: Aphanipathidae: Distichopathes)

Dennis M. Opresko¹, Samantha L. Goldman^{2,*}, Raven Johnson^{3,*}, Katherine Parra^{3,*}, Marissa Nuttall^{4,5}, G.P. Schmahl⁴ and Mercer R. Brugler^{1,3,6} 

¹Department of Invertebrate Zoology, U.S. National Museum of Natural History, Smithsonian Institution, Washington, DC, USA; ²Department of Ecology and Evolutionary Biology, Cornell University, Ithaca, NY, USA; ³Biological Sciences Department, NYC College of Technology (City University of New York), Brooklyn, NY, USA; ⁴National Oceanic and Atmospheric Administration, Flower Garden Banks National Marine Sanctuary, Galveston, TX, USA; ⁵CPC, San Diego, CA, USA and ⁶Division of Invertebrate Zoology, American Museum of Natural History, New York, NY, USA

Abstract

The continental shelf edge of the NW Gulf of Mexico supports dozens of reefs and banks, including the West and East Flower Garden Banks (FGB) and Stetson Bank that comprise the Flower Garden Banks National Marine Sanctuary (FGBNMS). Discovered by fishermen in the early 1900s, the FGBs are named after the colourful corals, sponges and algae that dominate the region. The reefs and banks are the surface expression of underlying salt domes and provide important habitat for mesophotic coral ecosystems (MCE) and deep coral communities to 300 m depth. Since 2001, FGBNMS research teams have utilized remotely operated vehicles (e.g. ‘Phantom S2’, ‘Mohawk’, ‘Yogi’) to survey and characterize benthic habitats of this region. In 2016, a Draft Environmental Impact Statement proposed the expansion of the current sanctuary boundaries to incorporate an additional 15 reefs and banks, including Elvers Bank. Antipatharians (black corals) were collected within the proposed expansion sites and analysed using morphological and molecular methods. A new species, *Distichopathes hickersonae*, collected at 172 m depth on Elvers Bank, is described within the family Aphanipathidae. This brings the total number of black coral species in and around the sanctuary to 14.

Introduction

The Flower Garden Banks National Marine Sanctuary (FGBNMS) is located off the Texas and Louisiana coasts on the continental shelf margin in the north-western Gulf of Mexico, and encompasses just three of the dozens of reefs and banks that occur in this area. Antipatharians are relatively common in some areas of the FGBNMS and 13 species (representing ~5% of the 273 described species) have been recorded from the sanctuary (Opresko *et al.*, 2016), occurring at depths ranging from 50–150 m. The reefs and banks of the north-western Gulf of Mexico, including Elvers Bank, harbour significant mesophotic coral ecosystems and deeper coral communities (ONMS, 2016; Boland *et al.*, 2017; Semmler *et al.*, 2017). Elvers Bank is located 65 km east of the East FGB (Figure 1) at the extreme edge of the continental shelf, extending to a depth of 205 m. Due to its shelf-edge location, surveys during the FGBNMS expeditions suggest a biological community somewhat different from other banks in the region, including assemblages of differing varieties of invertebrates not observed in other locations in the north-western Gulf of Mexico study area. While light measurements at this location are not available, it may be assumed that these deeper environments and associated biota require less light than those at shallower depths. During a National Oceanic and Atmospheric Administration (NOAA) survey cruise (Northwestern Gulf of Mexico Cruise 2016; Cruise DFH-30; Dive 382) conducted in September 2016, a large green, pinnately branched antipatharian colony was photographed at a depth of 172 m at Elvers Bank (Figure 2); however, samples of the colony were not collected at that time due to a malfunction of the manipulator arm on the remotely operated vehicle (ROV). The *in-situ* photos suggested that the colony represented an undescribed species. In September 2017, during a follow-up cruise to the same area (NW GOMEX Expansion Sites I; Cruise DFH-32; Dive 524), two colonies of what appeared to be the same species were collected at 172 m depth (Figure 3) and preserved in 100% ethanol and RNA later for morphological and molecular analyses. These two specimens are the subject of this paper.

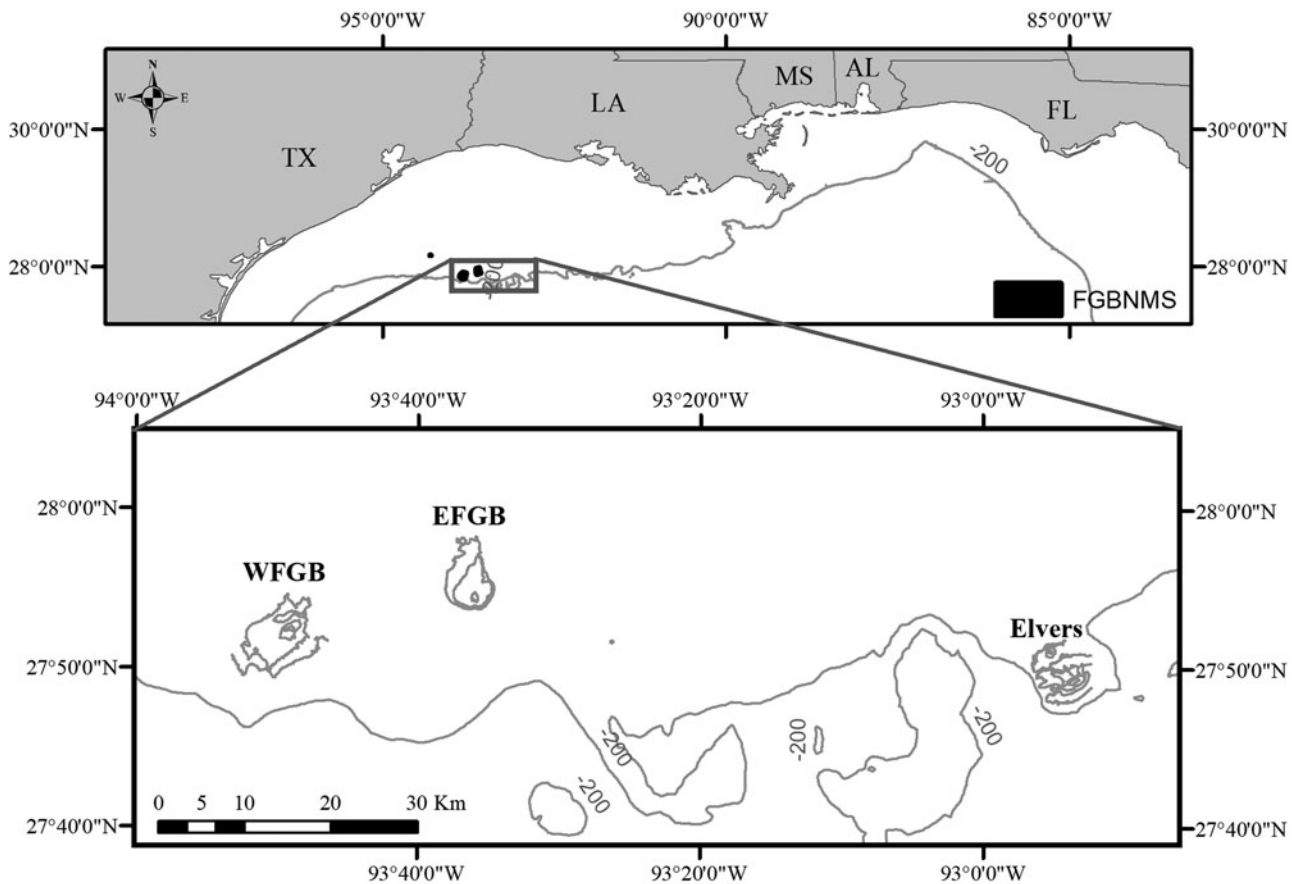


Fig. 1. The top panel shows the location of the Flower Garden Banks National Marine Sanctuary (FGBNMS), which is located off the Texas (TX) and Louisiana (LA) coasts on the continental shelf margin in the north-western Gulf of Mexico. The bottom panel shows that Elvers Bank is located ~65 km east of West and East FGB. The 200-m depth contour is shown in grey.

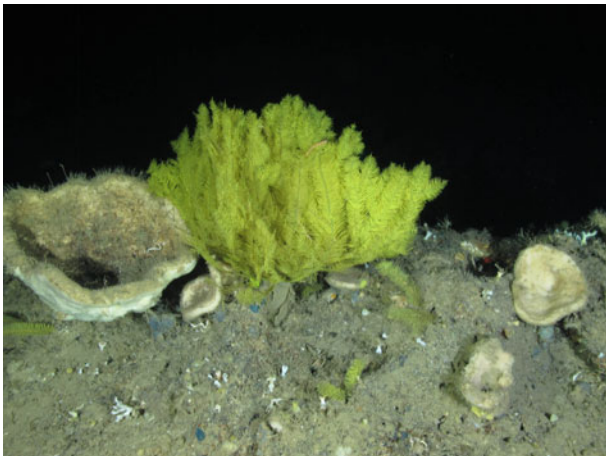


Fig. 2. *Distichopathes hickersonae* sp. nov., colony photographed *in situ*, at 172 m on Elvers Bank (specimen not collected).

Materials and methods

This report is based on specimens collected during expeditions aboard the RV 'Manta' to reefs and banks of the north-western Gulf of Mexico, conducted by NOAA staff of the FGBNMS as well as students and staff of NYC College of Technology (CUNY). The ROV 'Mohawk', maintained and operated by the Undersea Vehicle Program at the University of North Carolina at Wilmington, was used to collect the specimens after *in-situ*

photos were taken. Collected specimens were preserved in 100% EtOH and RNAlater and sent to the U.S. National Museum of Natural History (USNMNH), Smithsonian Institution, Washington, DC for further study. Photographs of the skeletal spines were made using a scanning electron microscope (SEM, Zeiss EVO MA 15) housed at the USNMNH. The specimens were coated with a 30–40 nm thick layer of 60% gold: 40% palladium. Analysis of the skeletal spines was conducted from direct examination of the material using a low-power binocular microscope or by examination of photographs taken with the SEM. The size of the polyps, referred to as the transverse diameter, was measured as the distance between the distal edge of the distal lateral tentacles and the proximal edge of the proximal lateral tentacles of the same polyp. The distance between spines was measured from the centre of the base of one spine to the centre of the base of an adjoining spine in the same axial row. The height of a spine was measured as the distance between the apex and the centre of the base of the same spine. Based on convention, the number of axial rows of spines was determined as the number of complete rows (those in which the bases of the spines are visible) that can be counted in one lateral view (also referred to as 'one aspect').

Subsamples of the specimens were subjected to molecular analysis which involved the sequencing of three mitochondrial gene regions (*trnW*-IGR-*nad2*, *cox3*-IGR-*cox1* and *nad5*-IGR-*nad1*; IGR = intergenic region) and three nuclear gene regions (ITS2, 28S and *SRP54*). DNA extraction, DNA quantification, PCR primers and reagents, PCR thermocycling profiles, PCR cleanup, cycle sequencing, cycle sequencing cleanup, traditional Sanger sequencing on an ABI-3730xL, multiple sequence alignment,

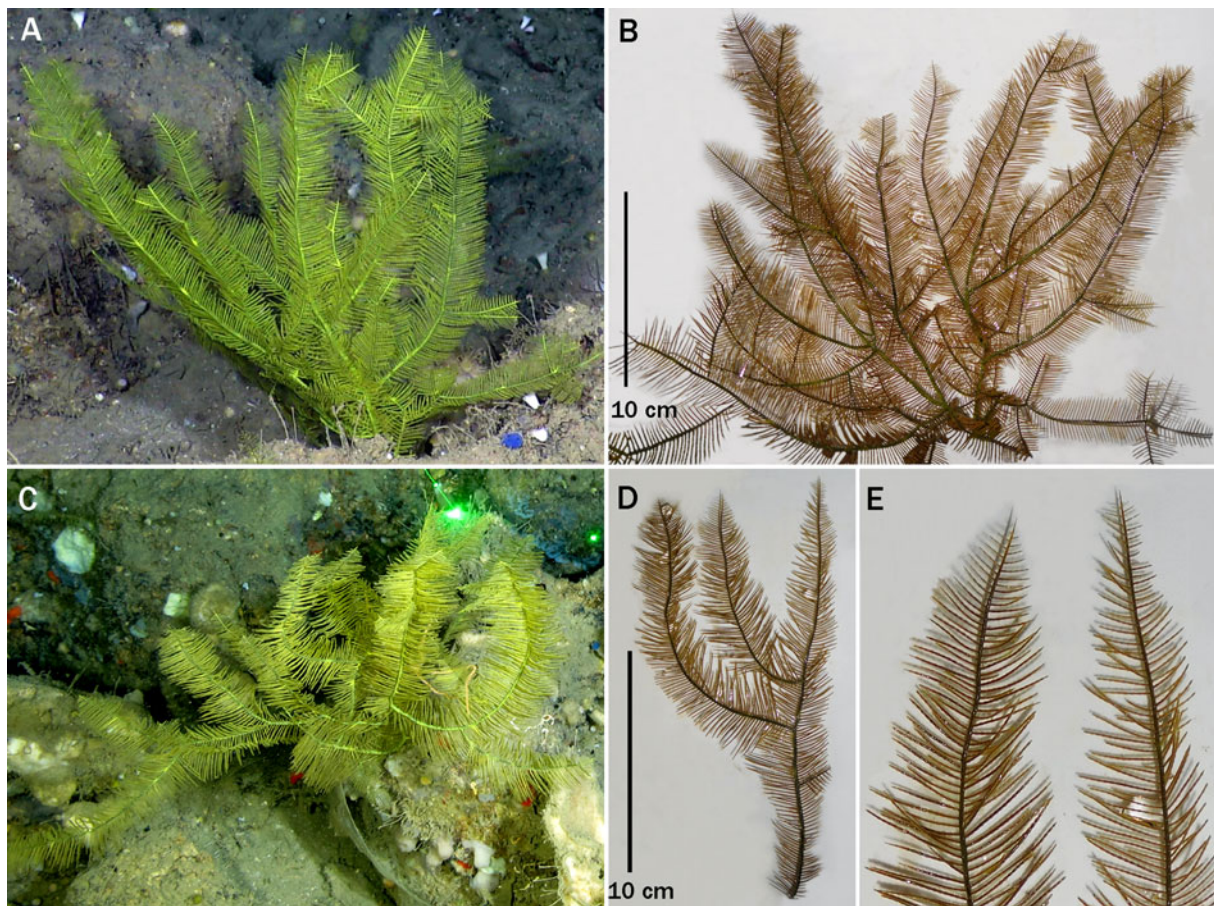


Fig. 3. (A) *In-situ* photo of holotype of *Distichopathes hickersonae* sp. nov. (USNM 1517703); (B) laboratory photo of holotype; (C) *In-situ* photo of paratype (USNM 1548274); (D) laboratory photo of paratype; (E) upper section of two branches of the paratype.

model selection, and Maximum likelihood-based tree building followed the protocol detailed in MacIsaac *et al.* (2013). Brugler *et al.* (2013) showed that the *nad5*-IGR-*nad1* region was the most variable region within the black coral mitogenome; however, the *cox3*-IGR-*cox1* region revealed the greatest number of unique haplotypes (i.e. unique species). Given that the phylogenetic reconstructions presented in Brugler *et al.* (2013) remain the most comprehensive to date, newly obtained sequence data (corresponding to *nad5*-IGR-*nad1* and *cox3*-IGR-*cox1* only) were added to the multiple sequence alignments from Brugler *et al.* (2013). Maximum likelihood-based phylogenies were built to infer the evolutionary relationship of *Distichopathes hickersonae* to known taxa within the Order Antipatharia. Currently, datasets for the three nuclear gene regions (ITS2, 28S and *SRP54*) are not as comprehensive (in terms of representative species) as the mitochondrial gene regions; thus, these regions were simply used to look for genetic differences between the holotype and paratype.

Results

Systematics

Order ANTIPATHARIA Milne-Edwards & Haime, 1857
Family APHANIPATHIDAE Opreško, 2004

Diagnosis. Corallum sparsely to densely branched (i.e. bramble-like, bushy, broom-like or fan-shaped), or pinnulated with simple pinnules arranged in two or more rows. Spines conical; usually covered to varying degrees with conical tubercles, but may be smooth; apex of spines simple (acute or rounded), occasionally bifurcated. Polyyps 0.6–3 mm in transverse diameter.

Remarks. The family was originally established with two subfamilies, the Aphanipathinae (including the genera *Aphanipathes*, *Phanopathes*, *Tetrapathes*, *Pteridopathes* and *Asteriopathes*) with subequal polypar spines, and the Acanthopathinae (including the genera *Elatopathes*, *Distichopathes*, *Rhipidipathes* and *Acanthopathes*) with very unequal (anisomorphic) polypar spines where the circumpolypar spines are very enlarged and the hypostomal spines are reduced or even absent (Opreško, 2004). Recent DNA sequencing studies (e.g. Brugler *et al.*, 2013) using mitochondrial markers (*cox3*-IGR-*cox1* and *nad5*-IGR-*nad1*) have not supported the recognition of these two subfamilies, and in fact the genera *Elatopathes* and *Distichopathes* have closer affinities to the family Myriopathidae than to other genera in the Aphanipathidae (Figures 4 and 5). In addition, the genus *Acanthopathes* grouped with several different families depending on the molecular marker or tree-building algorithm employed, and the genus *Rhipidipathes* grouped with genera in the Antipathidae (not shown). These results, if confirmed by more extensive molecular studies using higher resolution markers (i.e. Ultra-Conserved Elements; see Quattrini *et al.*, 2018), may necessitate a taxonomic revision of the family.

Genus *Distichopathes* Opreško, 2004

Antipathes, de Pourtalès, 1867: 112, 1871: 54, 1880: 118 (in part); van Pesch, 1914: 85 (in part, as subgenus *Aphanipathes*).

Aphanipathes Brook, 1889: 121 (in part); Opreško, 1972: 993 (in part).

Distichopathes Opreško, 2004: 235–237.

Diagnosis. Corallum monopodial, unbranched, or sparsely to densely branched; tending to be planar with overlapping

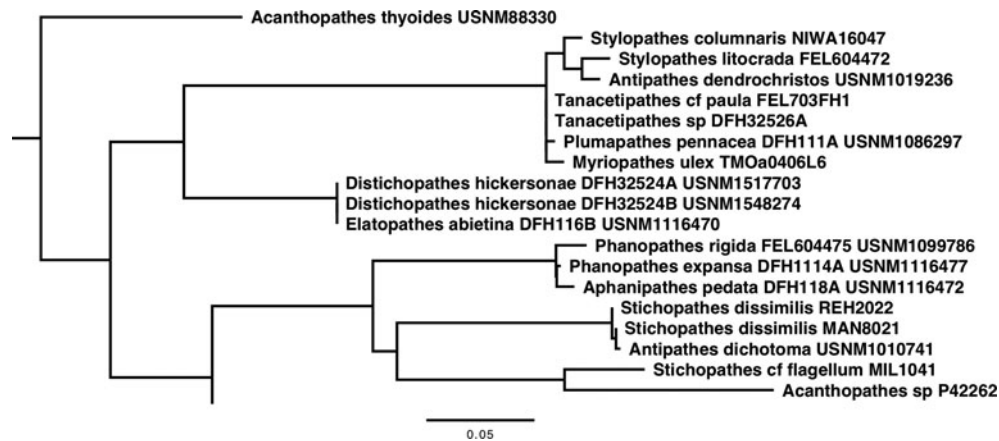
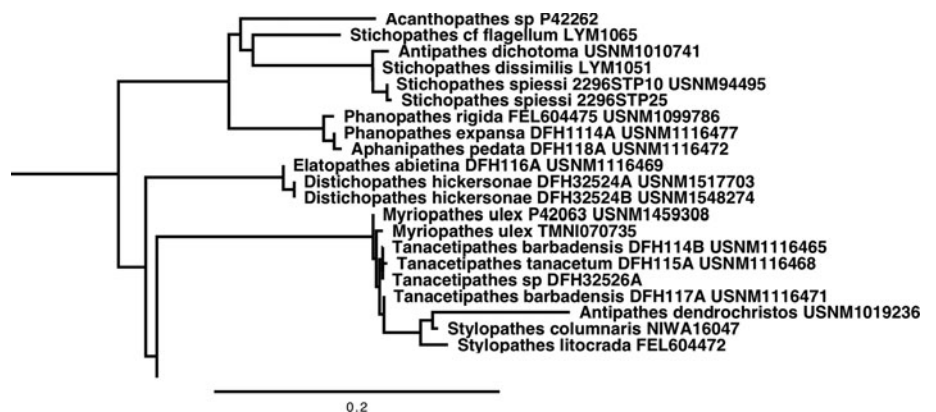


Fig. 4. Partial results of a Maximum likelihood-based phylogenetic reconstruction using mitochondrial *nad5*-IGR-*nad1* sequence data focusing specifically on the families Aphanipathidae and Myriopathidae. The MAFFT L-INS-i v7 based alignment consisted of 49 sequences and 686 sites. jModelTest v2.1.1 selected the TPM3uf + G model of sequence evolution. PhyML v3.1 utilized a BioNJ starting tree, best of NNI and SPR tree topology search options, and 1000 non-parametric bootstrap replicates. The tree was rooted internally to the Leiopathidae. Node support for *Distichopathes hickersonae* grouping with *Elatopathes abietina* is 99.9. *Distichopathes hickersonae* and *Elatopathes abietina* are genetically identical across 318 comparable base pairs. Node support for the clade grouping sister to *Distichopathes* + *Elatopathes* (which includes *Stylopathes*, *Antipathes*, *Tanacetipathes*, *Plumapathes* and *Myriopathes*) is 78.

Fig. 5. Partial results of a Maximum likelihood-based phylogenetic reconstruction using mitochondrial *cox3-cox1* sequence data focusing specifically on the families Aphanipathidae and Myriopathidae. The MAFFT L-INS-i v7 based alignment consisted of 59 sequences and 1111 sites. jModelTest v2.1.1 selected the GTR + I + G model of sequence evolution. PhyML v3.1 utilized a BioNJ starting tree, best of NNI and SPR tree topology search options, and 1000 non-parametric bootstrap replicates. The tree was rooted to sea anemones (*Metridium senile* and *Nematostella vectensis*) and zoanths (*Palythoa* sp. and *Savalia savaglia*). Node support for *Distichopathes hickersonae* grouping sister to *Elatopathes abietina* is 99.8. *Distichopathes hickersonae* and *Elatopathes abietina* can be genetically distinguished by a single base substitution (C/T) and a 38 bp insertion (in *Elatopathes*) across 826 comparable base pairs.



branches. Stem and branches pinnulate. Pinnules simple, not sub-pinnulate; arranged primarily in two lateral rows, but with simple short pinnules occurring very rarely on the abpolypar side of the axis. Pinnules also arranged alternately along the stem and branches.

Type species. *Distichopathes disticha* Opresko, 2004.

Remarks. The genus *Distichopathes* is distinguished from the other genera in the family Aphanipathidae by the simple, bilateral pinnules and distinctly anisomorphic spines. Species of the genus resemble those of *Pteridopathes* which also have two rows of simple bilateral pinnules; however, in *Pteridopathes* the polypar spines are subequal in size or, in some cases, the circumpolypar spines are slightly taller than the other polypar spines. The anisomorphic spines of *Distichopathes* are similar to those of *Elatopathes*, the colonies of which also have simple pinnules, but in the latter case the pinnules are arranged in four to six axial rows.

Species assigned to *Distichopathes*. Three species are currently assigned to the genus, *D. filix* (de Pourtalès, 1867), *D. disticha* Opresko, 2004, and *D. hickersonae* sp. nov.

Distribution. Species of *Distichopathes* are known only from the North-western Atlantic.

Distichopathes hickersonae sp. nov. Opresko & Brugler
(Figures 2, 3, 6, 7)

Aphanipathes filix, Opresko, 1974: 1003 (in part).

Aphanipathes filix de Pourtalès, 1867: 112 (in part)

?*Antipathes dissecta* Duchassaing & Michelotti, 1864: 48.

Type material

Holotype: USNM 1517703 (SEM 481). North-western Gulf of Mexico, Elvers Bank, 27.8434°N 92.9384°W, RV 'Manta', Stn DFH32-524A, Site E1-S, 172 m, NOAA, 25 Sept 2017.

Paratype: USNM 1548274, North-western Gulf of Mexico, Elvers Bank, 27.8433°N 92.9386°W, RV 'Manta', Stn DFH32-524B, Site E1-S, 172 m, NOAA, 25 Sept 2017.

Diagnosis

Corallum densely branched, branches tending to lie in one plane. Stem and branches with simple bilateral pinnules. Pinnules up to 2 cm long and 0.3 mm in diameter at their base; arranged in two very regular bilateral rows, with pinnules in each row alternating with those in opposite row. Small simple pinnules occurring very rarely on the abpolypar side of the axis. Spines anisomorphic: circumpolypar spines 0.26–0.35 mm tall, interpolypar spines 0.2–0.25 mm, the hypostomal spines 0.1–0.14 mm, and the abpolypar spines 0.1–0.18 mm. Polyps small, 0.65–0.8 mm in transverse

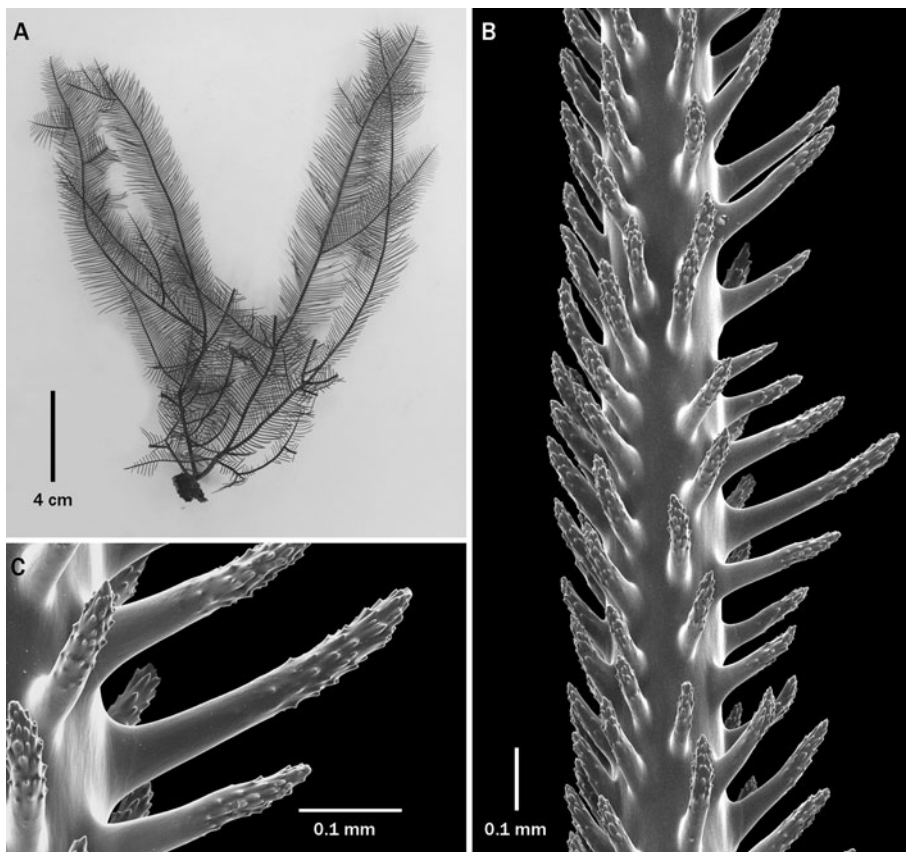


Fig. 6. *Distichopathes hickersonae* sp. nov., holotype, USNM 1517703: (A) corallum; (B) section of pinnule; (C) close-up view of polyp spines (B and C from SEM stub 481).

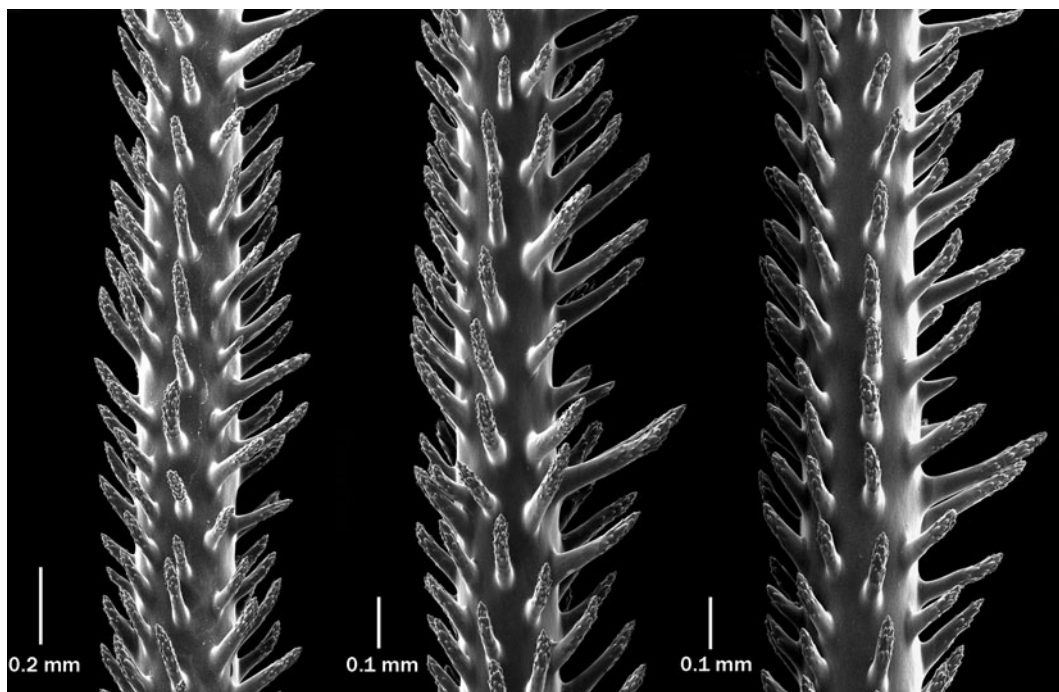


Fig. 7. *Distichopathes hickersonae* sp. nov., holotype, USNM 1517703: sections of pinnules (from SEM stub 481).

diameter and are placed in a single series on one side of the pinnules, with 8–10 polyps per cm.

Description of the holotype

The holotype (USNM 1517703) is a moderately sized complete colony about 21 cm tall and 17 cm wide (Figures 3A, B & 6A).

The colony is branched to the fifth order, and the branching is planar with overlapping adjacent branches. The diameter at the base of the stem just above the holdfast is 3.7 mm. Several large first-order branches originate just above the holdfast, one of which extends to the top of the colony. The stem and branches possess simple, filiform pinnules up to about 2 cm long and 0.3

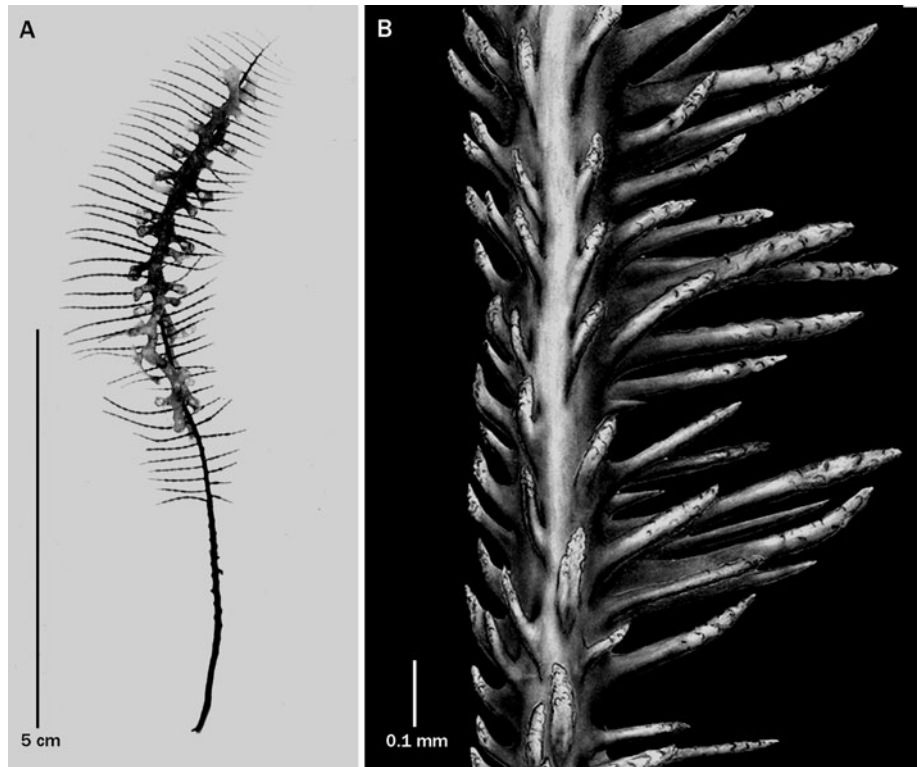


Fig. 8. *Distichopathes filix* Pourtales, specimen from R/V *Gerda* sta. 1125: (A) corallum; (B) section of pinnule.

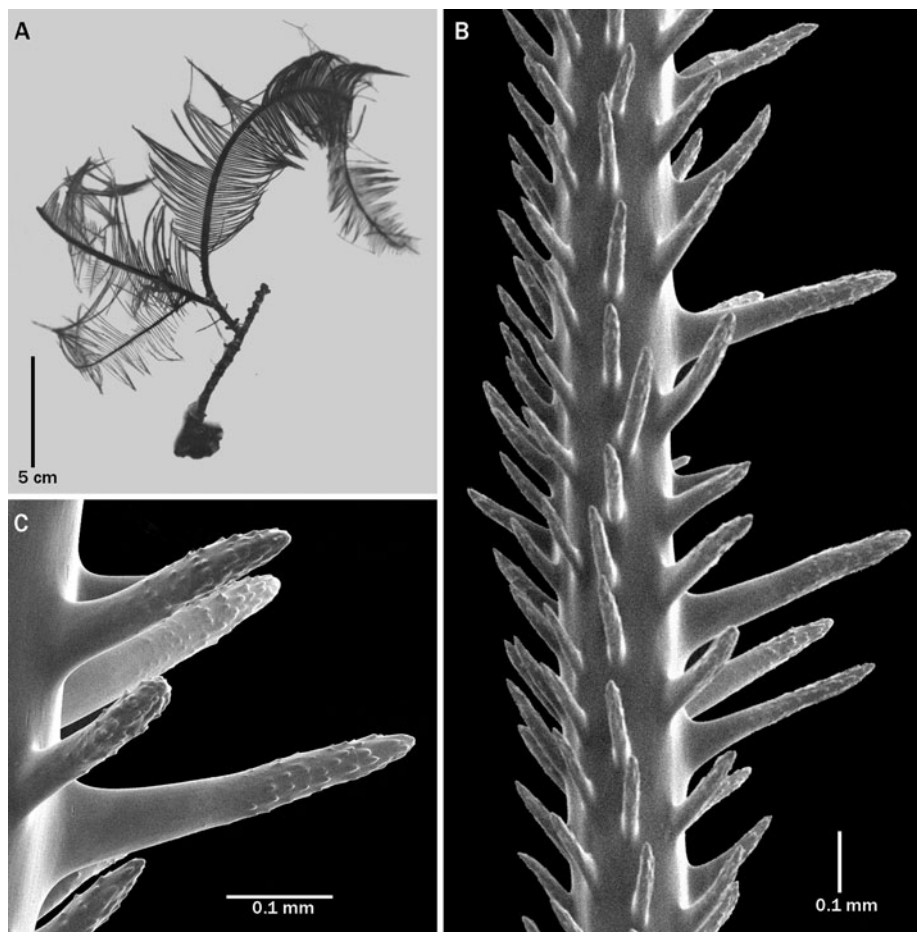


Fig. 9. *Distichopathes disticha*, holotype, MCZ 38476: (A) corallum; (B) section of pinnule; (C) close-up view of polyp spines.

mm in diameter near the base. The pinnules vary slightly in size from branch to branch, and can also be shorter on the lower parts of the branch compared with the more distal segment. Newly

forming pinnules at the branch tips gradually increase in size proximally until they reach a consistent length. The pinnules are arranged in two lateral rows with members of each row spaced

Table 1. Morphometric comparison of the three species of *Distichopathes*

Character	<i>D. filix</i>	<i>D. hickersonae</i>	<i>D. disticha</i>
Orders of branching	Unbranched	Up to five ^a	Up to three ^a
Length of pinnules (cm)	Usually <1	Mostly near 2 ^a	Mostly 3–7 (max. 12) ^a
Pinnular density (per cm)	14–16	16–18	16–18
Circumpolypar spines (mm)	0.55	0.26–0.35	≤0.4
Interpolypar spines (mm)	0.2	0.2–0.25	0.2
Hypostomal spines (mm)	0.1	0.1–0.14	≤0.1
Abpolypar spines (mm)	0.2	0.1–0.18	0.1
Polyps, transverse diameter (mm)	0.65–1.0	0.65–0.8	0.7
Polyp density (per cm)	7–9	8–10	8–9

^aBased on a limited number of specimens.

1–1.4 mm apart (from the middle of pinnule at the base to the middle of the base of adjacent pinnule in the same row), resulting in 16–18 pinnules per cm total for both rows. Most of the pinnules are inclined distally, but some extend out at nearly right angles to the axis of the stem or branch. The interior angle formed by the two rows of pinnules is close to 180°. Isolated pinnules occur on the abpolypar side of the stem and branches.

The pinnular spines are distinctly anisomorphic (Figures 6B and 7); the circumpolypar spines are up to 0.35 mm tall, although most are 0.26–0.30 mm. The interpolypar spines are 0.2–0.25 mm, the hypostomal spines 0.1–0.14 mm, and the abpolypar spines 0.1–0.16 mm. The spines are arranged in axial rows, six rows of which can be seen in one lateral view. The density of the polypar spines is mostly 6–7 per mm in each row, whereas the abpolypar spines are much more crowded together with a density of 10–11 per mm. The polypar spines are slightly inclined distally (distal angle 60–80°), and the abpolypar spines are much more distally inclined with a distal angle of about 30°. The spines on the branches and stem are more acicular than those on the pinnules; they are mostly 0.16–0.2 mm in height. Almost all the pinnular spines, both polypar and abpolypar, are covered over the upper (distal) half of their surface with very distinct and very numerous conical tubercles (Figure 6C). The hypostomal spines, however, are either smooth or have only a few tubercles near the apex. In the samples examined, two hypostomal spines are reduced in size in each polyp area.

The polyps are 0.65–0.8 mm in transverse diameter and are placed in a single series on one side of the pinnules, with 8–10 polyps per cm. The distance between centres of adjacent polyps is 1–1.2 mm.

Intraspecific variation

The paratype (USNM 1548274, Figure 3C–E) agrees with the holotype in all skeletal and polypar characters. The pinnules are mostly 1.5 cm long, but can be as much as 2 cm. The density of the pinnules is 16–18 per cm total for both lateral rows. The pinnules at the tips of the branches increase in length proximally, but the rate of increase varies from branch to branch (Figure 3E). On sections of pinnules 0.20–0.24 mm in diameter, the circumpolypar spines range in size from 0.26–0.32 mm, the interpolypar spines are about 0.24 mm, the hypostomal spines 0.11–0.13 mm and the abpolypar spines 0.15–0.18 mm. The polyps are similar to those of the holotype in the transverse diameter, with a resulting similar density of 9 per cm.

Comparisons

The two other nominal species in the genus are *D. filix* (de Pourtalès, 1867) and *D. disticha* Opresko, 2004. Specimens

that conform to Pourtalès' original description of *D. filix* characteristically are monopodial, unbranched, and have very short pinnules 0.6–1 cm long (Figure 8A). Colonies of *D. disticha* are sparsely branched, with up to three orders of branching, and have pinnules that are mostly 3–7 cm long (maximum about 12 cm) (Figure 9A). In contrast, colonies of *D. hickersonae* can be densely branched (up to five orders of branching) and have pinnules that are up to 2 cm in length. The fact that the examined colonies of *D. disticha* and *D. hickersonae* are of comparable size would support the supposition that the differences in the length of pinnules and the density of the branching are not age-related intraspecific phenomena.

The pinnular spines of *D. hickersonae* are morphologically more similar to those of *D. disticha* (Figure 9B) than to those of *D. filix* (Figure 8B). In *D. filix* the spines usually have a clavate shape in that some are slightly swollen towards the apex, and the apolypar spines can be distinctly flattened and bent upwards at the apex. In contrast, the spines of *D. disticha* and *D. hickersonae* are more regularly cylindrical. In terms of the size of the spines, *D. hickersonae* is also more similar to *D. disticha*. The circumpolypar spines are mostly 0.26–0.30 mm tall in *D. hickersonae* (maximum about 0.35 mm); up to 0.4 mm in *D. disticha*, and up to 0.55 mm in *D. filix*. In the samples examined, the interpolypar spines, hypostomal spines and abpolypar spines in *D. hickersonae* are similar to or slightly larger than the corresponding ones in the other two species (Table 1). Although the number of observations is limited, one difference seen between *D. hickersonae* and *D. disticha* is in the number of hypostomal spines that are reduced in size; in *D. hickersonae* only two such hypostomal spines are found in each polyp area, whereas in *D. disticha* three are present.

The maximum height of the spines, especially the polypar spines, has in the past been a key character in separating closely related antipatharian species. The differences seen here in the size of the circumpolypar spines between *D. disticha* and *D. hickersonae*, although not great, are suggestive that as larger suites of specimens are examined, the population distribution of this character will be different for each of these species.

Remarks

As in most antipatharian species complexes, the morphological boundaries between the three nominal species of *Distichopathes* are likely to be difficult to clearly define because of the inherent variability in most taxonomic characters. Overlaps in characters such as the length of the pinnules and size of the spines between closely related species is to be expected. The information presented in Table 1 is based on small sample sizes, especially for *D. hickersonae* and *D. disticha*; therefore, the values shown,

Table 2. GenBank accession numbers for the holotype (USNM 1517703) and paratype (USNM 1548274) of *Distichopathes hickersonae* sp. nov.

Gene Region	Holotype	Paratype
Mitochondrial <i>cox3</i> -IGR- <i>cox1</i>	MT319029	MT319028
Mitochondrial <i>trnW</i> -IGR- <i>nad2</i>	MT330297	MT330296
Mitochondrial <i>nad5</i> -IGR- <i>nad1</i>	MT330294	MT330295
Nuclear ITS2	MT330303	MT330302
Nuclear 28S	MT330299	MT330298
Nuclear <i>SRP54</i>	MT330300	MT330301

such as the typical length of the pinnules and height of the spines will most likely change as more colonies are analysed. In addition, some specimens previously assigned to *D. filix* (see Table 4 in Opresko, 1972), particularly those that have intermediate pinnule lengths of 1–2 cm, will have to be re-evaluated to determine if the morphology of the spines provides a better indication of whether they are more closely related to *D. filix* or to *D. hickersonae*.

Molecular analysis

We compared the DNA sequence of the holotype (USNM 1517703) and paratype (USNM 1548274) using a combination of three mitochondrial and three nuclear gene regions. The holotype and paratype were genetically identical across all six gene regions (total number of base pairs compared: 2878 bp). We sequenced 1701 bp of mitochondrial DNA, which consisted of 492 bp from *trnW*-IGR-*nad2*, 880 bp from *cox3*-IGR-*cox1* and 329 bp from *nad5*-IGR-*nad1* (IGR: intergenic region). We also sequenced 1177 bp of nuclear DNA, which consisted of 603 bp from ITS2, 394 bp from 28S and 180 bp from *SRP54* (Table 2). We were unable to obtain tissue from representatives of *D. disticha* and *D. filix* and corresponding sequence data is not available on GenBank; thus, we could not include either congeneric in our molecular analysis.

Etymology

Named in recognition of Emma L. Hickerson (NOAA's FGBNMS Research Coordinator). Since 2005, Emma has generously been inviting co-authors DMO and MRB, as well as underrepresented minority undergraduates from CUNY, to the FGBNMS to survey and collect black corals. These research cruises aboard the MV 'Spree' and RV 'Manta' have resulted in the discovery of a number of new species of antipatharian corals.

Distribution

Known only from the north-western Gulf of Mexico at 172 m depth.

Supplementary material. The supplementary material for this article can be found at <https://doi.org/10.1017/S002531542000051X>.

Data deposition information. All newly obtained sequence data have been uploaded to NCBI's GenBank (see Table 2 for individual accession numbers).

Acknowledgements. Special thanks to the shipboard crew of the RV 'Manta' and to the members of the scientific party (Nadia Alomari [CUNY], Erin Easton [UTRGV] and Robert McGuinn [DSCRTP]) for their efforts in helping collect the material on which this paper is based. Thanks to Sheila Moaleman (CUNY) and Nicole Bellaflora-Mejia (CUNY) for providing the first *in-situ* photos. MRB gratefully acknowledges Estefania Rodriguez (Curator of Marine Invertebrates at the AMNH) for allowing Team Black Coral to utilize her molecular lab. Specimens were collected using the National Marine Sanctuary Foundation's ROV 'Mohawk' operated by the Undersea Vehicle Program at the University of North Carolina at Wilmington, and the efforts of the ROV operators Lance Horn, Jason White and Eric Glidden are greatly appreciated. The specimens were examined in the SEM lab at the USNMNH,

and assistance was generously provided by S. Whitaker. DMO and MRB are Research Associates at the USNMNH and gratefully acknowledge that affiliation. MRB is also a Research Associate at the American Museum of Natural History and gratefully acknowledges that affiliation as well.

Author contribution. Conceptualization: DMO, MRB; Methodology: DMO, MRB; Formal analysis and investigation: DMO, SLG, RJ, KP, MN, MRB; Writing – original draft preparation: DMO, MRB; Writing – review and editing: DMO, SLG, RJ, KP, MN, GPS, MRB; Funding acquisition: DMO, GPS, MRB; Resources: DMO, MN, GPS, MRB; Supervision: MRB.

Financial support. Funding for cruises was provided, in part, by NOAA's Deep-Sea Coral Research and Technology Program to GPS. Funding for this project was provided in part by the City University of New York, the Flower Garden Banks National Marine Sanctuary, and a U.S. Department of Justice grant to the Smithsonian Institution.

References

- Boland G, Etnoyer PJ, Fisher C and Hickerson E (2017) State of deep-sea coral and sponge ecosystems in the Gulf of Mexico: Texas to the Florida straits. In Hourigan TF, Etnoyer PJ and Cairns SD (eds), *The State of Deep-Sea Coral and Sponge Ecosystems of the United States*. NOAA Technical Memorandum NMFS-OHC-4. Silver Spring, MD: NOAA, pp. 321–378.
- Brook G (1889) Report on the Antipatharia. *Report of the scientific results of the voyage of the H.M.S. Challenger*. *Zoology* 32, 222 pp.
- Brugler MR, Opresko DM and France SC (2013) The evolutionary history of the order Antipatharia (Cnidaria: Anthozoa: Hexacorallia) as inferred from mitochondrial and nuclear DNA: implications for black coral taxonomy and systematics. *Zoological Journal of the Linnean Society* 169, 312–361.
- de Pourtalès LF (1867) Contributions to the fauna of the Gulf Stream at great depths. *Bulletin of the Museum of Comparative Zoology, Harvard* 1, 103–120.
- de Pourtalès LF (1871) Deep-Sea Corals. III. *Catalogue of the Museum of Comparative Zoology, Harvard*. No. 4. 93 pp., 8 pls.
- de Pourtalès LF (1880) Zoological results of the Blake expedition to the Caribbean Sea. *Bulletin of the Museum of Comparative Zoology, Harvard* 6, 113–118.
- Duchassaing P and Michelotti J (1864) Supplément au mémoire sur les Coralliaires des Antilles. *Extrait des mémoires de l'Académie des Sciences de Turin* 23, 1–112.
- MacIsaac KG, Best M, Brugler MR, Kenchington ELR, Anstey LJ and Jordan T (2013) *Telopathes magna* gen. nov., spec. nov. (Cnidaria: Anthozoa: Antipatharia: Schizopathidae) from deep waters off Atlantic Canada and the first molecular phylogeny of the deep-sea family Schizopathidae. *Zootaxa* 3700, 237–258.
- Milne Edwards H. and Haime J. (1857). *Histoire Naturelle des Coralliaires ou Polypes Proprement Dits*. 360 pp., Paris.
- Office of National Marine Sanctuaries (2016) *Flower Garden Banks National Marine Sanctuary Expansion Draft Environmental Impact Statement*. Silver Spring, MD: U.S. Department of Commerce, National Oceanic and Atmospheric Administration, Office of National Marine Sanctuaries.
- Opresko DM (1972) Redesignations and reevaluations of the antipatharians described by LF de Pourtales. Biological results of the University of Miami Deep-Sea Expeditions. 97. *Bulletin of Marine Science* 22, 950–1017.
- Opresko DM (1974) *A Study of Classification of the Antipatharia (Coelenterata: Anthozoa) with Redesignations of Eleven Species* (Ph.D. thesis). Rosenstiel School of Marine and Atmospheric Science, University of Miami, Miami, Florida, USA.
- Opresko DM (2004) Revision of the Antipatharia (Cnidaria: Anthozoa). Part IV. Establishment of a new family, Aphanipathidae. *Zoologische Mededelingen, Leiden* 78, 209–240.
- Opresko DM, Nuttall MF and Hickerson EL (2016) Black corals of the Flower Garden Banks National Marine Sanctuary. *Gulf of Mexico Science* 33, 47–67.
- Quattrini AM, Faircloth BC, Duenas LF, Bridge TC, Brugler MR, Calixto-Botía IF, DeLeo DM, Foret S, Herrera S, Lee SM and Miller DJ (2018) Universal target-enrichment baits for anthozoan (Cnidaria) phylogenomics: new approaches to long-standing problems. *Molecular Ecology Resources* 18, 281–295.
- Semmler RF, Hoot WC and Reaka ML (2017) Are mesophotic coral ecosystems distinct communities and can they serve as refugia for shallow reefs? *Coral Reefs* 36, 433–444.
- van Pesch AJ (1914) The Antipatharia of the *Siboga* Expedition. *Siboga Expeditie Monographie* 17, 1–258.