QUEBEC COOPERATIVE STUDY OF FRIEDREICH'S ATAXIA

Free Amino Acids and Calcium, Magnesium and Zinc Levels in Friedreich's Ataxia

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ABSTRACT: Free amino acid levels and zinc, magnesium and calcium content have been determined in autopsy samples of 9 areas of the brain, two skeletal muscles, and the right ventricle, left ventricle and septum of the heart of a Friedreich's ataxia subject.

RÉSUMÉ: Nous avons dosé, à l'autopsie, le niveau des acides aminés libres et celui du zinc, magnésium et calcium dans des échantillons de 9 régions du cerveau, deux muscles squelettaux, le ventricule droit, le ventricule gauche et le septum cardiaque d'un sujet souffrant de l'ataxie de Friedreich.

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Little information is available on free amino acid and metal ion content of tissues of Friedreich's ataxia patients. Free amino acid levels in the brains of two patients have been reported (Huxtable et al., 1979), but no report has yet appeared on metal ion content of the brain. Information of this nature is needed, however, to test certain hypotheses concerning biochemical defects in Friedreich's ataxia. Thus, it has been proposed that the cardiomyopathy invariably found associated with Friedreich's ataxia is due to a calcium overload defect (Huxtable, 1978), It has also been suggested that a defect in taurine regulation occurs in Friedreich's ataxia (Filla et al., 1978; Barbeau, 1980, 1982; Barbeau et al., 1982).

In this paper, we report values on free amino acid and metal ion concentrations in brain, heart, and muscle samples of a patient dying of Friedreich's ataxia.

METHODS

Clinical summary

Autopsy samples were obtained from a patient dying with Friedreich's ataxia. The patient was a 19-year-old white male who had been diagnosed as having Friedreich's ataxia at the age of 4 years. Poorly differentiated lymphoblastic lymphoma was diagnosed at age 18 years when he developed a superior venal caval syndrome. At that time he had severe scoliosis, and difficulty in walking and making fine movements with his hand because of ataxia. There was also slowness of speech. An echocardiogram showed left ventricular hypertrophy. Treatment included vincristine, adriamycin, and prednisone. Shortly after discharge following this therapy the patient developed a murmur with tachycardia and tachypnea that was thought to be an adriamycin-induced aggravation of his cardiomyopathy. Consequently, no further adriamycin was given. He had not received intrathecal methyltrexate because of his neurological problems; however, he returned 10 months later with headache when a lumbar puncture showed 600 blasts/mm³. He was treated then with intrathecal methyltrexate and systemic vincristine as well as 6-mercaptopurine, methyltrexate, L-asparaginase, and Septa (a mixture of trimethoprim and sulfamethoxazole). Subsequently he developed recurrent leptomeningeal involvement with loss of vision in the right eye, diminished vision in the left eye and a 6th cranial nerve palsy. A seizure disorder developed and he died 2 months later.

Pathological findings

At autopsy, lymphoma of a poorly differentiated lymphoblastic type was found throughout the entire body with massive diffuse lymphadenopathy and splenomegaly. Of particular note was alymphomatous infiltrate in the leptomeninges. In addition, there were multiple petechiae and ecchymoses suggesting a bleeding disorder.

Pathologic findings relating to Friedreich's ataxia included gliosis with tract degeneration in the cortical spinal, anterior and posterior spinal cerebellar and posterior column tracts as well as the gracile and cuneate nuclei. There was severe scoliosis. Also, there was marked hypertrophy of the left ventricle (1.8 cm).

Analyses

The amino acid analyses were performed as previously described (Huxtable et al., 1979). Ion analyses were performed

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by atomic absorption spectroscopy at the University of Arizona Analytical Center (Azari et al., 1980).

RESULTS AND DISCUSSION

A wealth of research has been performed on Friedreich's ataxia over the past ten years (summarized by Barbeau, 1980, 1982). Despite intensive investigation, basic biochemical indices are still difficult to establish for Friedreich's ataxia, due to the poor availability of biopsy or autopsy material. We feel, therefore, that the amino acid and metal determinations we report herein, although derived from only one case of Friedreich's ataxia, are of value in that they have rarely if ever been performed before (Huxtable et al., 1979).

Total tissue contents of calcium, magnesium and zinc for brain and muscle are reported in Tables 1 and 2. It has been suggested that the cardiomyopathy of Friedreich's ataxia may be secondary to a dysfunction in calcium regulation in the heart, whereby there is abnormally increased calcium flux into the myocyte (Huxtable, 1978). Data concerning the calcium content of Friedreich's ataxia hearts are sparse and inconsistent. One study reported areas of intracellular calcification in the heart from a Friedreich's ataxia patient at autopsy (Sanchez-Casis et al., 1976). A subsequent study of three hearts obtained from Friedreich's ataxia patients, however, reported finding no myocardial calcium deposits, as determined by histological analysis using a Von Kossa stain (Lamarche et al., 1980).

Different authors have reported calcium levels in human hearts ranging from 40 to 155 μ g/g wet weight (summarized in Lyengar et al., 1978). Where values were originally reported

Table 1: Metal Ion Content of Heart and Skeletal Muscle in Fried-

per ash weight or per dry weight, they have been converted to per wet weight using the conversion factors of Lyengar et al. (1978). The median value found was 57 μ g/g wet weight. On the basis of this, it would appear that right ventricular calcium content in our patient is distinctly high, with the left ventricle having low to normal values. The mean of the averages for calcium content of skeletal muscle summarized by Lyengar et al. (1978) is 66 μ g/g wet weight. Information on variation from muscle to muscle, however, is not given. Of the two muscles we examined, the intercostal would appear to be abnormally high in calcium content, and the pectoralis major would appear to be abnormally low in calcium.

Average zinc values reported by various laboratories are $34 \pm 21 \ \mu g/g$ wet weight for the heart (standard deviation of means of 18 groups), and for muscle 54 ± 11 (11 groups). The values we report in Table 1, therefore, fall in the middle of the expected range. Magnesium levels, on the other hand, tend to be low. The average values reported for muscle are 224 ± 46 , and for heart 192 ± 44 (Lyengar et al., 1978).

We find considerable variation from region to region in the calcium content of the brain (Table 2). These values may be compared to the average and deviation of 9 sets of determinations for the brain of $128 \pm 19 \ \mu g/g$ (Lyengar et al., 1978). Corresponding values for magnesium and zinc are 173 ± 129 (mean of 19 sets) and 13 ± 4 (mean of 16 sets), respectively.

The free amino acid contents for the same regions are shown in Tables 3 and 4. The values for muscle may be compared to those of Bergstrom et al. (1974). Amino acid values in the brains of Friedreich's ataxia patients have been reported before (Huxtable et al., 1979). The major observation was that glutamate levels remain relatively invariant from region to region. As can

reich's Ataxia							
	µg/g wet weight of tissue						
		Mg	Zn				
Intercostal muscle	116	155	52				
Pectoralis major	19.4	180	52				
Left ventricle	40	188	29.6				
Septum	96	160	30.0				

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120

27.6

Table 2: Metal Ion Content of Brain in Friedreich's Ataxia						
	µg/g w	μg/g wet weight of tissue				
	Ca	Mg	Zn			
Anterior cerebellar vermis	16	140	16.0			
Dentate nucleus	96	204	26.8			
Posterior cerebellar vermis	29	136	14.8			
Cerebral cortex	22	128	15.2			
Cerebral hemisphere	112	188	23.2			
Precentral gyrus	160	190	47			
Inferior olivary nucleus	20	230	41			
Red nucleus	390	450	56			

Right ventricle

 Table 3: Free Amino Acid Content of Heart and Skeletal Muscle Regions in Friedreich's Ataxia

	μ mole/g wet weight							
	Intercostal muscle	Pectoralis major	left ventricle	septum	right ventricle			
Pser	0.11	0.09	0.12	0.14	0.11			
Tau	6.71	14.30	6.91	7.13	3.33			
Asp	0.18	0.26	0.33	0.27	0.26			
Thr	0.42	0.79	0.82	0.98	0.54			
Ser	0.52	1.06	1.45	1.65	0.89			
Asn	0.52	0.51	1.22	0.97	0.79			
Gln	5.72	8.57	13.43	14.83	6.68			
Pro	0.94	1.39	0.92	0.98	0.89			
Glu	1.22	1.74	15.74	20.08	8.89			
Gly	1.70	2.16	2.07	2.24	1.80			
Ala	3.04	3.76	12.73	12.79	7.50			
Val	0.67	0.92	0.90	1.11	0.67			
½ cys	0.14	0.15	0.23	0.26	0.21			
Met	0.13	0.24	0.38	0.43	0.16			
Isoleu	0.24	0.32	0.48	0.45	0.28			
Leu	0.52	0.69	1.34	1.45	0.75			
Tyr	0.26	0.34	0.46	0.48	0.29			
Phe	0.41	0.51	0.88	0.96	0.48			
ß-AIB	0.97	1.23	1.22	1.06	1.10			

be seen from Table 4, the same phenomenon is found in the patient reported on here. Thus, the mean variation for the glutamate values reported in Table 4 is 14%, compared with 36% for the taurine values, and 48% for the GABA values.

The ratios of concentrations of neuroactive amino acids may be of greater relevance than their absolute amounts (van Gelder, 1978). Certain ratios are reported in Table 5. The glutamate: glutamine ratio is significantly decreased in Friedreich's ataxia brains for all areas examined. Glutamine is largely contained within the glia, where one of its functions is to serve as a reservoir for maintaining synaptosomal glutamate content (van Gelder, 1978). The loss of the region to region variation in glutamate levels combined with the fall in glutamate:glutamine ratio suggests that Friedreich's ataxia brains suffer some anatomical or biochemical change in glutamate-containing synapses.

Table 4: Fre	e Amino Acid	Content of Brain	n Regions in	Friedreich's	Ataxia mole/g wet weight

	Region								
	ACV	DN	PCV	<u></u>	CH	PG	ION	<u>RN</u>	SC
Pser	0.20	0.13	0.11	0.14	0.18	0.13	0.15	0.17	0.13
PEA	0.33	trace	0.33	0.25	trace	0.26	1.11	trace	0.15
Tau	3.67	1.82	3.47	1.54	4.12	1.41	2.70	2.49	2.62
Asp	1.71	1.37	1.68	2.63	1.92	2.29	1.44	3.59	2.38
Thr	0.71	0.41	0.67	0.57	0.95	0.48	0.99	0.56	2.09
Ser	1.25	0.92	1.21	1.13	1.37	1.00	1.88	1.24	3.91
Gln	11.47	18.25	11.73	11.41	11.42	11.65	10.30	16.89	13.60
Pro	0.14	trace	0.33	0.17	0.43	trace	1.03	0.32	1.32
Glu	9.99	7.40	9.90	10.79	9.56	9.13	7.69	10.21	10.51
Gly	2.72	2.29	2.30	1.79	2.69	1.45	4.40	2.89	6.89
Ala	3.45	3.43	3.27	3.00	3.17	2.64	4.74	4.98	6.07
Val	0.56	0.32	0.63	0.63	0.50	0.36	1.49	0.73	1.75
½ Cys	0.27	0.22	0.37	0.34	1.01	0.24	0.51	0.30	0.88
Cysta	1.60	5.35	2.32	2.22	1.91	3.36	1.44	3.12	1.33
Met	0.31	0.19	0.34	0.27	0.40	1.53	0.60	0.42	1.53
Isoleu	0.36	0.29	0.32	0.36	0.49	0.21	0.61	0.49	1.20
leu	1.20	0.90	1.14	1.00	1.29	0.69	1.68	1.43	3.06
Tyr	0.67	0.47	0.48	0.35	0.61	0.23	0.56	0.55	1.01
Phe	0.90	0.71	0.61	0.57	0.80	0.48	0.59	1.02	1.49
GABA	2.09	4.14	1.67	1.80	1.42	1.56	1.21	3.28	1.38

Abbreviations: ACV anterior cerebellar vermis; DN dentate nucleus; PCV posterior cerebellar vermis; CC cerebral cortex; CH cerebellar hemisphere; PG precentral gyrus; ION inferior olive nucleus; RN red nucleus; SC spinal cord.

Table 5: Selected Amino Acid Ratios						
Control	CH (4)	PCV (4)	ACV (4)	DN (3)		
Tau/Glu Tau/GABA GABA/Glu Glu/Gln	$\begin{array}{c} 0.24 \pm 0.14 \\ 1.57 \pm 0.63 \\ 0.14 \pm 0.04 \\ 1.94 \pm 0.55 \end{array}$	$\begin{array}{c} 0.18 \pm 0.11 \\ 1.00 \pm 0.52 \\ 0.16 \pm 0.04 \\ 2.32 \pm 1.23 \end{array}$	$\begin{array}{c} 0.37 \pm 0.42 \\ 1.05 \pm 0.50 \\ 0.32 \pm 0.30 \\ 2.20 \pm 1.05 \end{array}$	$\begin{array}{c} 0.23 \pm 0.15 \\ 0.51 \pm 0.36 \\ 0.45 \pm 0.04 \\ 1.22 \pm 0.38 \end{array}$		
Friedreich's Ataxia	(3)	(3)	(3)	(3)		
Tau/Glu Tau/GABA GABA/Glu Glu/Gln	$\begin{array}{c} 0.43 \pm 0.34 \\ 2.90 \pm 1.32 \\ 0.15 \pm 0.04 \\ 0.84 \pm 0.19^{a} \end{array}$	$\begin{array}{c} 0.35 \pm 0.23 \\ 2.08 \pm 1.02 \\ 0.17 \pm 0.04 \\ 0.84 \pm 0.08^{\rm b} \end{array}$	$\begin{array}{c} 0.37 \pm 0.18 \\ 1.76 \pm 0.54 \\ 0.21 \pm 0.02 \\ 0.87 \pm 0.17^{\rm b} \end{array}$	$\begin{array}{c} 0.25 \pm 0.43 \\ 0.44 \pm 4.24 \\ 0.56 \pm 0.35 \\ 0.41 \pm 0.44^{\mathrm{b}} \end{array}$		

Differences compared to control:

^a _p <.025; ^b _p <.05

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Abbreviations are as for Table 4. Control data are taken from Huxtable et al., 1979.

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