

Modulation of hypoxia-inducible factor-1 α /cyclo-oxygenase-2 pathway associated with attenuation of intestinal mucosa inflammatory damage by *Acanthopanax senticosus* polysaccharides in lipopolysaccharide-challenged piglets

Changyou Fan, Jie Han*, Xianjun Liu, Fei Zhang, Yingshan Long and Qitao Xie

Key Laboratory of Livestock Infectious Diseases in Northeast China of Ministry of Education, College of Animal Science and Veterinary Medicine, Shenyang Agricultural University, Shenyang 110866, Liaoning Province, People's Republic of China

(Submitted 22 February 2019 – Final revision received 17 May 2019 – Accepted 30 May 2019; First published online 10 June 2019)

Abstract

Intestinal barrier inflammatory damage is commonly accompanied by hypoxia. The hypothesis that dietary *Acanthopanax senticosus* polysaccharides (ASPS) might modulate the hypoxia-inducible factor-1 α (HIF-1 α) signalling pathway and contribute to attenuate intestinal injury was tested in lipopolysaccharide (LPS)-challenged piglets. Thirty-six weaned pigs were randomly allocated to one of the following three groups: (1) basal diet + saline challenge; (2) basal diet + LPS challenge; (3) basal diet with 800 mg/kg ASPS + LPS challenge. LPS was injected at 15, 18 and 21 d, and intestinal sections were sampled following blood collection at 21 d. The results showed ASPS reversed ($P < 0.05$) LPS-induced decrease in average daily feed intake and rise ($P < 0.05$) of diarrhoea incidence and index. Biochemical index reflecting gut barrier damage and function involving ileal pro-inflammatory cytokines (TNF- α and IL-1 β) and enzyme activity (diamine oxidase and lactase), as well as circulatory D-xylose, was normalised ($P < 0.05$) in LPS-challenged piglets receiving ASPS. ASPS also ameliorated intestinal morphological deterioration of LPS-challenged piglets, proved by elevated ileal villus height ($P < 0.05$) and improved appearance of epithelial villus and tight junction ultrastructure. Moreover, ASPS prevented LPS-induced amplification of inflammatory mediators, achieved by depressed ileal mRNA abundance of TNF- α , inducible NO synthase and IL-1 β concentration. Importantly, ileal protein expressions of HIF-1 α , cyclo-oxygenase-2 (COX-2) and NF κ B p65 were also suppressed with ASPS administration ($P < 0.05$). Collectively, these results suggest the improvement of mucosal inflammatory damage and diarrhoea in immune stress piglets is possibly associated with a novel finding where HIF-1 α /COX-2 pathway down-regulation is involved in NF κ B p65-inducible releasing of inflammatory cytokines by dietary ASPS.

Key words: *Acanthopanax senticosus* polysaccharides: Hypoxia: Inflammation: Piglets

A layer of intestinal epithelial cells lining the gastrointestinal tract forms a selective barrier to the harsh environment of intestinal lumen^(1,2). Disturbances in intestinal barrier, characterised by increased mucosal permeability, allow luminal bacteria, toxins and antigenic agents to 'leak' across the epithelium, resulting in inflammation, diarrhoea and potentially systemic disease^(3,4). Early life stress is a predisposing factor for the development of chronic intestinal barrier damage^(5,6). Immune stress in piglets resulting in sustained impairment in mucosal barrier is considered one of the major causes of diarrhoea and an economic loss case. The mechanism that immune stress-induced loss of intestinal barrier function is not fully understood but thought to be mediated by the release of pro-inflammatory

cytokines such as TNF- α as well as interleukins, which are central mediators of intestinal inflammatory diseases^(7,8).

Micro-environmental hypoxia has been identified to be a feature of sites of chronic inflammation. Hypoxia-inducible factor-1 α (HIF-1 α) is an oxygen-dependent subunit and masters transcription responses to hypoxia⁽⁹⁾. HIF-1 α activation plays a gut-injurious role associated with some conditions such as hypoxia and inflammation^(10,11). Many findings have converged to suggest HIF-1 α could be inducible in existence of pro-inflammatory cytokines, and the fact that its activation happens not only under hypoxia but also normoxic conditions^(12,13). Importantly, HIF-1 α promoter contains an active NF κ B binding in transcription start site⁽¹⁴⁾, and pro-inflammatory cytokines like

Abbreviations: ADFI, average daily feed intake; ADG, average daily gain; ASPS, *Acanthopanax senticosus* polysaccharides; BW, body weight; CONTR, basal diet + saline challenge (control); COX-2, cyclo-oxygenase-2; DAO, diamine oxidase; HIF-1 α , hypoxia-inducible factor-1 α ; iNOS, inducible NO synthase; LPS, lipopolysaccharide; TJ, tight junction; TLR4, toll-like receptor 4.

* **Corresponding author:** Dr J. Han, email hanjie1130@163.com

Table 1. Ingredient composition and nutrient contents of the basal diet (%; as-fed basis)

Items	
Ingredients	
Maize	22
Rice	10
Flour	10
Soya bean meal	15
Soya bean powder	8
Fish meal	3
Milk powder	15
Citric acid	2
Glucose	2
White granulated sugar	2
Whey powder	5
Calcium lactate	1.3
Dibasic calcium phosphate	1
Salt	0.2
Emulsified fat powder	1.5
Vitamin and mineral premix*	2
Nutrient content†	
Digestible energy (kcal/kg)‡	3460
Crude protein (%)	19.1
Ca (%)	0.68
Total P	0.58
Lysine (%)	1.42
Methionine (%)	0.44
Threonine (%)	0.92
Tryptophan (%)	0.34

* Premix provided per kg of diet: vitamin A 3.6 mg; vitamin D₃ 0.82 mg; vitamin E 40 mg; vitamin K₃ 4.7 mg; thiamine 4.4 mg; riboflavin 10.2 mg; niacin 40 mg; pantothenic acid 32 mg; pyridoxine 3.2 mg; vitamin B₁₂ 0.01 mg; folic acid 1.2 mg; biotin 0.14 mg; choline 500 mg; Cu (copper sulphate) 8 mg; Zn (zinc sulphate) 84 mg; Fe (ferrous sulphate) 84 mg; Mn (manganese sulphate) 33 mg; iodine (calcium iodate) 0.6 mg; Se (sodium selenite) 0.3 mg.

† Crude protein, Ca and total P were analysed values. The digestible energy and amino acid contents were calculated values.

‡ To convert energy in kcal to kJ, multiply by 4.184.

TNF- α ⁽¹⁵⁾ and IL-6⁽¹⁶⁾ are among the target gene identified for hypoxia-induced NF κ B. Theoretically, co-regulation of gastrointestinal NF κ B and HIF-1 α pathways by nutrition means and subsequent down-regulation of the pro-inflammatory cytokines genes may exert beneficial effects on the intestinal injury.

Natural plant-derived polysaccharides have been safely used mainly in nutritional and medical areas for a long period in many Oriental countries. *Acanthopanax senticosus* polysaccharides (ASPS) are the major active ingredients extracted from traditional herbal medicine *Acanthopanax senticosus*, and have been characterised by the bioactivity of immune regulation^(17,18). In our previous works^(19,21), following the elucidation of down-regulation of the genes expressions of pro-inflammatory cytokines under inflammatory conditions with ASPS application, subsequent study identified the important role of ASPS on suppressing the toll-like receptor 4 (TLR4)/NF κ B/myosin light chain kinase (MLCK) signalling pathway to sustain intestinal integrity in lipopolysaccharide (LPS)-challenged mice model. Given the intimate molecular link between NF κ B and HIF-1 α ⁽²²⁾, further identification of the role of dietary ASPS on the HIF-1 α signalling pathway under inflammation condition is the point of interest.

Therefore, in the present study, we employed *Escherichia coli* LPS-induced immune stress model in piglet to test the hypothesis that dietary supplemented with ASPS could modulate the HIF-1 α signalling pathway and concomitant amelioration of LPS-induced intestinal barrier dysfunction.

Methods

Preparation and analysis of *Acanthopanax senticosus* polysaccharides

ASPS were extracted from the root of *A. senticosus* by using a method of water extraction and alcohol precipitation with some modifications⁽²³⁾. Briefly, crushed roots were boiled to obtain filtrate, which was concentrated and precipitated with ethanol to collect precipitate by centrifugation. Following protein elimination from the precipitation by the Sevag method⁽²⁴⁾, recrystallisation and drying were employed to obtain a kind of tan powdery polysaccharide. About 96 % content of ASPS was determined by the phenol sulphuric acid method, and the composition analysis using HPLC with a diode-array-detector showed that it is a heteropolysaccharide composed of glucose, galactose, mannose, arabinose, glucuronic acid, rhamnose, galactosidonic acid and xylopyranose and fucose (online Supplementary Fig. S1).

Experimental design, feeding and lipopolysaccharide challenge

A total of thirty-six crossbred weaned piglets (Duroc \times Large White \times Landrace, weaned at 24–25 d of age) with an average initial body weight (BW) of 7.98 (SEM 0.04) kg were kept in a house equipped 2.0 \times 2.1 m² stainless steel pens (three piglets per pen). Each pen is equipped with plastic floor and a self-feeder and a nipple drinker to allow *ad libitum* access to feed and water. The temperature in the inner house was controlled at 24°C approximately. The piglets were randomly assigned according to birth weight and sex into three treatment groups as follows: (1) basal diet + saline challenge (CONTR); (2) basal diet + LPS challenge (LPS); (3) basal diet with 800 mg/kg ASPS + LPS challenge (ASPS + LPS). The maize–soya bean-meal basal diet was formulated to meet or exceed the nutrient requirements recommended by *Feeding Standard of Swine*⁽²⁵⁾ of 8–20 kg BW for all nutrients except for Ca (Table 1). The crude protein, Ca and total P contents in diet were analysed according to the method of *Association of Official Analytical Chemists*⁽²⁶⁾. The ASPS dose in the present work was determined on the basis of our published studies indicating that it ameliorated intestinal injury of LPS-challenged mice^(20,21). On days 15, 18 and 21 during the 21-d feeding trial, pigs in LPS and ASPS + LPS groups were challenged intraperitoneally with LPS (*E. coli* serotype 055:B5; Sigma Chemical Inc.) at 100 μ g/kg BW dissolved in sterile saline, and the pigs in CONTR were given an equivalent amount of sterile saline. At 21 d, pigs were orally given D-xylose at the dose of 500 mg/kg BW prepared as 0.5 g/ml solution in deionised water at 1 h post-challenge to assess intestinal absorptive function *in vivo*.

The institutional and national guidelines for the care and use of animals were followed, and all experimental procedures involving animals were approved by the Shenyang Agricultural University Institutional Animal Care and Use Committee.

Sample collection

On day 21, piglets were deprived of feed at 3 h pre-injection except for *ad libitum* access to water in order to avoid the potential effects on the characteristics of blood of LPS-induced feed

intake reduction. At 3 h after LPS challenge, whole blood samples were collected in pro-coagulant vacuum tubes by anterior vena cava. Plasma was then separated by centrifuging at 3500 *g* for 10 min, and stored at -20°C for further analysis of biochemical index. Following the blood collection, euthanasia and mid-line laparotomy were performed. After dissecting small intestine free of the mesenteric attachment, mid-ileum segments were then excised and flushed gently with ice-cold PBS and immediately immersed in 4 % chilled polyoxymethylene for histological evaluation and 2.5 % glutaraldehyde for electron microscopy identification, respectively. The remaining ileal mucosa was scratched with glass microscope slide and collected into sterile frozen tubes following longitudinal opening and ice-cold PBS rinsing, and then which was immediately snapped in liquid nitrogen and stored at -80°C until analysis.

Growth performance and faecal characteristics evaluation

Piglets were weighed individually at days 0, 14 and 21 of the trial to obtain the average BW of each pen and determine average daily gain (ADG). Surplus feed remaining in the feeder of each pen was cleared away and weighted to calculate average daily feed intake.

Faecal characteristics of piglets in each pen were assessed at 09.00 and 17.00 hours, and the degree of dehydration score was recorded twice daily during the entire LPS-challenged period (from 15 to 21 d). It was assigned a faecal score based on visual analysis of symptoms according to the following criteria^(27,29), 0, no stools; 1, normal, solid faeces; 2, pasty faeces; 3, droplets of watery faeces/diarrhoea; 4, moderate diarrhoea and 5, severe diarrhoea; and the criteria of 3, 4 and 5 were considered as diarrhoea condition. Diarrhoea incidence and diarrhoea index were calculated according to the following equations:

$$\text{Diarrhoea incidence (\%)} = \frac{\text{(number of diarrhoea pigs)}}{\text{(total number of pigs)} \times 100}$$

$$\text{Diarrhoea index} = \frac{\text{(total faecal score of pigs)}}{\text{(total number of pigs)}}$$

Intestinal barrier function parameters and inflammatory mediators in blood or intestinal mucosa

Intestinal function parameters, including enzymatic activity of intestinal DAO and lactase, invertase, maltase and the plasma concentration of D-xylose, as well as the levels of inflammatory cytokines involving TNF- α , IL-6 and IL-1 β in intestinal mucosa, were quantified using enzyme-linked immunosorbent assay kits (Omnimabs). All procedures were carried out strictly according to the manufacturer's protocols.

Intestinal mucosal histological assay

Villus height and crypt depth were measured on haematoxylin and eosin-stained and paraformaldehyde-fixed ileal histological slices. Detailed methods of these tests have been described previously⁽²⁰⁾. Ileal villus and microvillus morphology were further

researched by scanning electron microscope, and the processing was carried out as described elsewhere⁽³⁰⁾. Briefly, ileal sections measuring $3 \times 3 \times 3 \text{ mm}^3$ were cut and immediately transferred into 2.5 % glutaraldehyde for fixation. The segments were washed three times in phosphate buffer and then post-fixed in OsO_4 (1 % in phosphate buffer) for 30 min. After a series of dehydration processes in alcohol at the concentration of 30, 50, 70, 80, 90 and 100 %, and substitution in a series of tert-butyl alcohol at the concentration of 50, 75, 90 and 100 %, the segments were dried in freeze dryer (IXRF, VFD-30) and sputter-coated with a layer of platinum (Hitachi, MC1000), and finally examined under Regulus 8100 scanning microscope (Hitachi). The morphology of intestinal epithelial tight junction was evaluated by testing mid-ileal sections employing transmission electron microscope (HT-7700; Hitachi) as previously described⁽²¹⁾.

mRNA expression analysis by real-time PCR

The procedures of mRNA expression analysis were done according to our previous description⁽²⁰⁾ with some modification. Total RNA of ileal mucosa was extracted using TRIzol reagent (Invitrogen) according to the manufacturer's instruction. The integrity of isolated RNA preparation was examined by electrophoresis on a 1 % agarose gel containing 0.5 mg/ml ethidium bromide, and the purity and concentration of RNA were quantified using a spectrophotometer (NanoDrop 2000, Thermo Fisher Scientific) according to the optical density at OD 260/280 readings. Total RNA was reverse-transcribed using the PrimeScript RT reagent kit (Takara Biotechnology). The primers for the quantitative RT-PCR of each gene transcript for TNF- α , HIF-1 α , inducible nitric oxide synthase (*iNOS*) and housekeeping gene were designed by Invitrogen and available in online Supplementary Table S1. The quantitative RT-PCR analysis was performed using the ABI StepOnePlus (Applied Biosystems), and the condition was set up as follows: 95°C for 30 s, followed by forty cycles at 95°C for 5 s, 60°C for 40 s; Gene-specific amplification was determined by melting curve analysis and agarose gel electrophoresis. The $2^{-\Delta\Delta C_t}$ method was used to analyse the relative changes in each target gene expression⁽³¹⁾ with glyceraldehyde 3-phosphate dehydrogenase (GAPDH) as the housekeeping gene. The change (Δ) in C_t values in each group was compared with the C_t value of GAPDH (ΔC_t), where $\Delta\Delta C_t = (C_t \text{ of the target gene} - C_t \text{ of GAPDH}) \text{ treatment} - (C_t \text{ of the target gene} - C_t \text{ of GAPDH}) \text{ CONTR}$. All the samples were analysed in triplicates.

Immunofluorescence microscopy

Immunofluorescence staining was carried out as per our previous description⁽²¹⁾ and Yue *et al.*⁽³²⁾. Ileal segments were fixed with 4 % paraformaldehyde and then cut into 3 μm thick slices. The slices were dewaxed and dehydrated with xylene and ethanol, respectively. The tissue samples were blocked with 5 % solution of bovine serum albumin (BSA) for 30 min at room temperature following incubation in 3 % hydrogen peroxide diluted by methanol, and then incubation with anti-HIF-1 α (1:500; Santa Cruz Biotechnology) and anti-cyclo-oxygenase-2 (COX-2) (1:500; LifeSpan Biosciences) antibodies diluted in PBS containing 5 % BSA at 4°C overnight. The resulting sections

Table 2. Effects of dietary *Acanthopanax senticosus* polysaccharides (ASPS) supplementation on growth performance and diarrhoea of lipopolysaccharide (LPS)-challenged piglets (Means of four pens with three pigs per pen with pooled standard errors)

Variable	CONTR	LPS	ASPS + LPS	SEM	P
Days 1–14 (before LPS challenge)					
Initial body weight (kg)	8.02	7.96	7.97	0.04	0.84
ADG (g/d)	259.82	258.39	268.57	5.92	0.79
ADFI (g/d)	461.31	466.67	472.92	8.88	0.89
Days 15–21 (during LPS challenge)					
ADG (g/d)	424.17 ^a	311.67 ^b	385 ^{a,b}	20.31	0.03
ADFI (g/d)	681.25 ^a	515.28 ^b	623.15 ^a	27.97	0.01
Diarrhoea incidence (%)	0 ^b	23.75 ^a	2.50 ^b	3.56	<0.01
Diarrhoea index	0.66 ^b	1.20 ^a	0.70 ^b	0.09	<0.01

CONTR, non-challenged piglets fed basal diet (control); LPS, LPS-challenged piglets fed basal diet; ASPS + LPS, LPS-challenged piglets fed basal diet supplemented with 800 mg/kg ASPS; ADG, average daily gain; ADFI, average daily feed intake.

^{a,b} Mean values within a row with unlike superscript letters are significantly different ($P < 0.05$).

were washed with PBS and incubated with Alexa fluor 488-conjugated secondary antibody for 2 h in dark conditions. After 4',6-diamidino-2-phenylindole (DAPI) treatment for 5 min and sealing, sample images were obtained using a biomicroscope (Axio Scope A1; Zeiss).

Western blot assay

The quantification of protein expression in ileal mucosa was done according to the procedures outlined by Han *et al.*⁽²⁰⁾ with some modification. In brief, an equal amount of protein extract was electrophoresed on 8 % reducing polyacrylamide gel and transferred onto polyvinylidene difluoride membranes. Immunoblots were blocked with 5 % BSA for 50 min at room temperature and incubated overnight at 4°C with specific primary antibodies involving rat anti-HIF-1 α and NF κ B p65 (1:1000; Santa Cruz Biotechnology) and rabbit anti-COX-2 (1:1000; LifeSpan biosciences) in Tris-buffered saline with Tween-20. Blots were washed and then incubated with the corresponding HRP-conjugated secondary antibodies for 40 min at room temperature. The relative abundance of each target protein was expressed as target protein: β -actin protein ratio. The protein expressions of all samples were expressed as fold changes, calculated relative to CONTR.

Statistical analyses

The sample size was calculated using software SAS (version 9.4; SAS Institute Inc.). A power test based on the data obtained from a recent study on gene expression and biochemical index of pigs⁽¹⁹⁾ showed that four replicates per treatment were needed to achieve 80 % power and $\alpha = 0.05$ according to Kononoff & Hanford⁽³³⁾, and statistical power of more than 80 % for a sample size of six could be expected in the present experiment, which enables the additional power to reject the null hypothesis (H_0), if H_0 was false ($P = 1 - \beta$).

The mean of three pigs in each pen/replicate was used to analyse growth performance and diarrhoea because each pen was regarded as an experimental unit, whereas individual pig was used as an experimental unit for analysis of the other parameters. All data sets were tested for normal distribution using the Shapiro–Wilk test, and then parametric data were analysed using one-way ANOVA with dietary as the main effect, and differences

among groups were compared using Duncan's test in the case where the significant main effect of diet was found.

The data of diarrhoea index were first performed with log-transformation due to lack of normal distribution, and then analysed using ANOVA. Statistical analyses were performed using IBM SPSS Statistics (version 22), and all the data were expressed as means with their standard errors. $P < 0.05$ was considered significant for all analyses.

Results

The effect of Acanthopanax senticosus polysaccharides on growth performance and diarrhoea of lipopolysaccharide-challenged piglets

As given in Table 2, ASPS did not affect growth performance including ADG and average daily feed intake prior to LPS challenge of weaned piglets (from day 1 to 14). LPS challenge (from day 15 to 21) resulted in suppressed ADG ($P < 0.05$) and average daily feed intake ($P < 0.05$) compared with the piglets in CONTR. In contrast, the values of ADG and average daily feed intake ($P < 0.05$) of LPS-challenged piglets were marked boosted by ASPS administration although the change of ADG was insignificant. In addition, the values of diarrhoea incidence ($P < 0.05$) and diarrhoea index ($P < 0.05$) in LPS-induced piglets were decreased by the inclusion of ASPS when compared with the LPS group, and their levels were comparable with the CONTR group.

Changes in biochemical parameters in blood or intestinal mucosa

The piglets in LPS group exhibited the elevated level of intestinal mucosal TNF- α ($P < 0.05$) and IL-1 β ($P < 0.05$), as well as decreased ($P < 0.05$) level in ileal enzyme activities of DAO ($P < 0.05$) and lactase ($P < 0.05$), and circulating D-xylose ($P < 0.05$) in LPS-challenged piglets when compared with CONTR (Table 3). Correspondingly, besides the significant normalising of aforementioned index ($P < 0.05$), the values of ileal activities of invertase and maltase, as well as ileal concentration of IL-6, were somewhere between CONTR and LPS groups although they were not significantly altered in the LPS-challenged piglets under ASPS application.

Table 3. The effects of *Acanthopanax senticosus* polysaccharides (ASPS) on intestinal damage and function parameters, and inflammatory mediators in blood or intestinal mucosa in lipopolysaccharide (LPS)-challenged piglets (Mean values with pooled standard errors; *n* 6 per group)

Variable	CONTR	LPS	ASPS + LPS	SEM	<i>P</i>
Plasma parameter (mmol/l)					
D-Xylose	2.73 ^a	2.16 ^b	2.38 ^{a,b}	0.09	0.03
Intestinal enzyme activity (U/ml)					
Diamine oxidase	5.14 ^a	3.72 ^b	4.95 ^a	0.26	0.04
Lactase	5.26 ^a	4.42 ^b	5.83 ^a	0.17	0.002
Invertase	32.23	28.62	29.82	0.63	0.08
Maltase	3.76	3.36	3.39	0.11	0.29
Intestinal inflammatory cytokines (pg/ml)					
TNF- α	138.5 ^b	201.64 ^a	126.64 ^b	11.73	0.02
IL-6	129.57	118.83	122.53	6.42	0.79
IL-1 β	70.21 ^b	86.00 ^a	62.74 ^b	3.95	0.007

CONTR, non-challenged piglets fed basal diet (control); LPS, LPS-challenged piglets fed basal diet; ASPS + LPS, LPS-challenged piglets fed basal diet supplemented with 800 mg/kg ASPS.

^{a,b} Mean values within a row with unlike superscript letters are significantly different ($P < 0.05$).

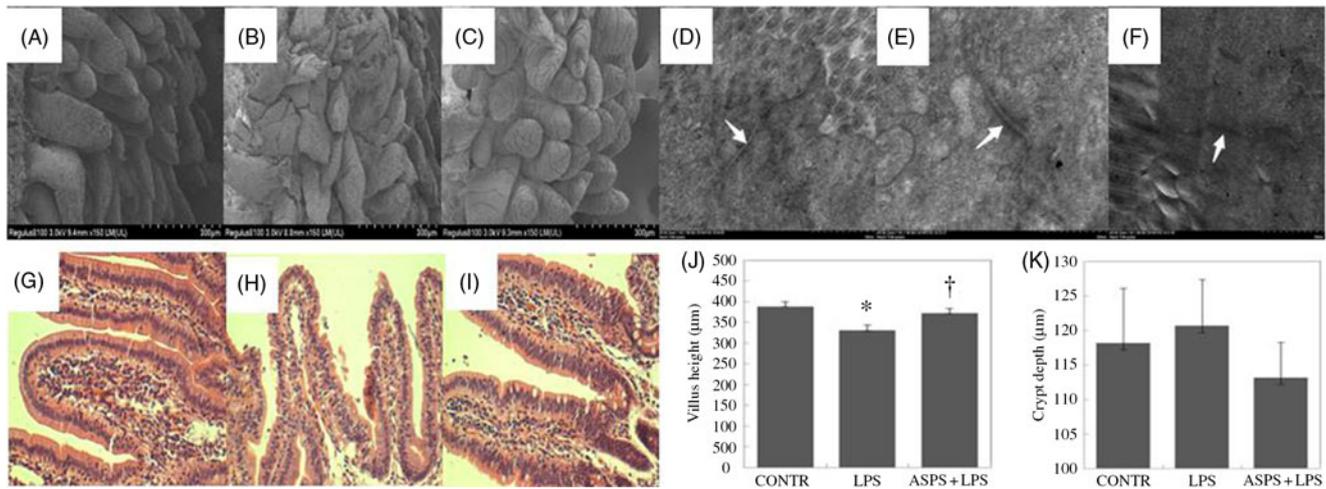


Fig. 1. Histological analysis of ileum influenced by *Acanthopanax senticosus* polysaccharides (ASPS) in lipopolysaccharide (LPS)-challenged piglets (*n* 6). (A–C) The representative photomicrographs of ileal villi from control (CONTR) (A), LPS (B) and ASPS + LPS (C) were observed by scanning electron microscope ($\times 150$). (D–F) The representative photomicrographs of ileal tight junctions from CONTR (D), LPS (E) and ASPS + LPS (F) were observed by transmission electron microscope ($\times 20\,000$). (G–I) The representative photomicrographs of ileal segments stained with haematoxylin and eosin ($\times 100$ magnification) of CONTR (G), LPS (H) and ASPS + LPS (I). (J) Ileal mucosal villus height; (K) ileal mucosal crypt depth. Values are means with standard errors represented by vertical bars. * $P < 0.05$ v. CONTR; † $P < 0.05$ v. LPS.

Intestinal histological observation

Compared with CONTR group (Fig. 1(A)), ileal histological observation using scanning electron microscope showed obvious destruction in intestinal epithelial morphology of villus and microvillus with irregular shape and arrangement in weaned piglets challenged with LPS (Fig. 1(B)). However, these pathologic changes were reversed drastically to form the normal intestinal epithelial appearance with the administration of ASPS (Fig. 1(C)), indicating that ASPS treatment prevented or repaired intestinal epithelial damage caused by LPS. Similarly, the evaluation for gut epithelial permeability using transmission electron microscope found that tight junction (TJ) ultra-structure characterised by intact structure and electron dense materials between the adjoining cells decreased following the LPS treatment (Fig. 1(E)) compared with CONTR group (Fig. 1(D)). As expected, ASPS inclusion significantly attenuated the negative changes induced by LPS challenge (Fig. 1(F)).

Ileal haematoxylin and eosin staining showed obvious damage characterised by atrophic villi with a discontinuous brush border and irregular epithelium in LPS-injected piglets without ASPS supplementation (Fig. 1(G) and (H)). Correspondingly, these negative histologic changes were significantly alleviated by pretreatment with ASPS (Fig. 1(I)). As expected, compared with LPS group, dietary ASPS significantly increased villus height in ileum ($P < 0.05$) (Fig. 1(J)). However, no effect was observed on crypt depth following ASPS supplementation (Fig. 1(K)).

Gene expression

The data from mRNA expressions of TNF- α , *i*NOS and HIF-1 α are shown in Fig. 2. Compared with the CONTR group, LPS challenge increased mRNA abundances of TNF- α ($P < 0.05$), HIF-1 α ($P < 0.05$) and *i*NOS ($P < 0.05$) in ileal mucosa. As expected, the altered mRNA expressions of aforementioned genes were reversed with ASPS administration ($P < 0.05$).

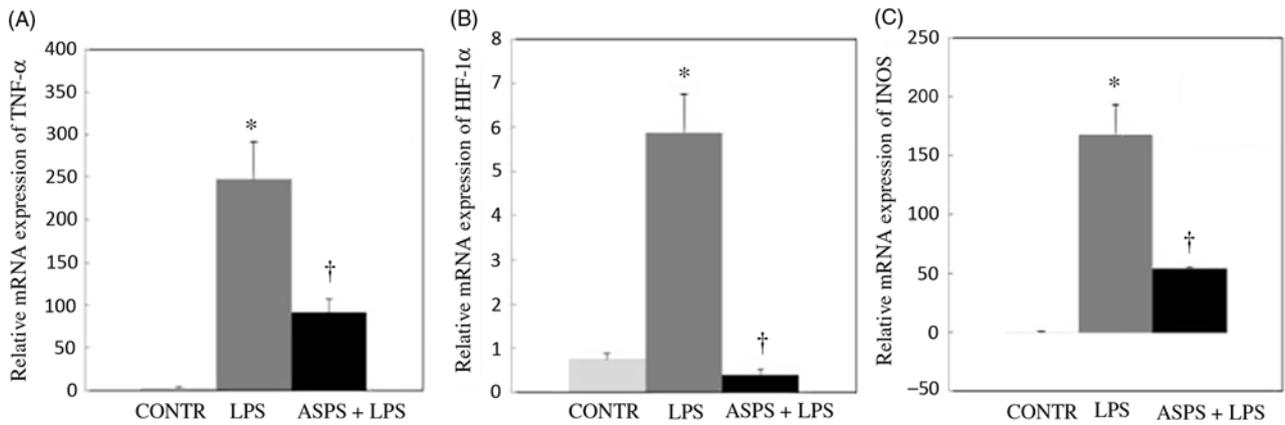


Fig. 2. Effects of dietary *Acanthopanax senticosus* polysaccharides (ASPS) supplementation on gene expression related to inflammation in lipopolysaccharide (LPS)-challenged piglets (n 6). Values are means of gene expression of TNF- α (A), hypoxia-inducible factor-1 α (HIF-1 α) (B) and inducible nitric oxide synthase (iNOS) (C), with standard errors represented by vertical bars. * $P < 0.05$ v. CONTR; † $P < 0.05$ v. LPS. CONTR, piglets receiving basal diet and injected with saline challenge (control); LPS, piglets receiving basal diet and injected with *Escherichia coli* LPS; ASPS + LPS, piglets receiving basal diet with 800 mg/kg ASPS and injected with LPS challenge.

Hypoxia-inducible factor-1 α , cyclo-oxygenase-2 and NF κ B p65 protein expressions in ileal mucosa by Western blotting and fluorescence microscopy

The localisation and expression of ileal HIF-1 α and COX-2 proteins are evaluated by immunofluorescence presented in Fig. 3. Piglets in LPS group exhibited more distribution of HIF-1 α and COX-2 staining with the presence of continuous bands along epithelial cell base compared with CONTR group (Fig. 3(A)). Correspondingly, ASPS pretreatment attenuated the HIF-1 α and COX-2 expressions. In addition, immuno-blotting was further employed to measure the expressions of the two proteins (Fig. 3(B)). Unanimously, the protein expressions of HIF-1 α , COX-2 and NF κ B p65 were amplified in ileal epithelium in LPS-challenged piglets (Fig. 3(C)) ($P < 0.05$). Inclusion of ASPS reversed LPS-induced elevation of HIF-1 α ($P < 0.05$), COX-2 ($P < 0.05$) and NF κ B p65 ($P < 0.05$).

Discussion

TLR recognise conserved pathogen-associated molecular patterns that are unique to micro-organisms⁽³⁴⁾. TLR4 is predominantly activated by LPS. The combination of LPS with TLR4 receptor can trigger downstream NF κ B and result in the activation of immune response⁽³⁵⁾. However, a high dose of LPS-inducible excessive activation of TLR4 signalling could lead to inflammatory damage in tissue⁽³⁶⁾. In the present study, we employed an *E. coli* LPS-challenged immune stress model for inducing gut barrier inflammatory damage in weaned piglets to determine whether ASPS supplementation might modulate HIF-1 α signalling response to hypoxia accompanied with inflammation and consequently contribute to improvement in loss of intestinal barrier. LPS as a toxin molecule present on the membrane of Gram-negative bacteria can induce systemic inflammatory and the various intestinal barrier disturbances induced by LPS injection in our current finding were coincident with many well-documented reports⁽³⁷⁾ and indicated an immune stress model built successfully in our study.

The present study validated that growth performance of piglets was similar among treatments despite the fact that ASPS was employed with lack of LPS challenge. Following LPS injection, ASPS treatment exhibited a positive role on feed intake elevation. However, the value of ADG of pigs fed ASPS was intermediate and no significance was obtained for ADG when comparing with the CONTR group. This finding was consistent with that in our previous report in weaned piglets⁽¹⁹⁾ and suggested that dietary ASPS preferred to alleviate the loss of growth performance under immunological stress than under normal condition. In addition, a noticeable decrease in diarrhoea incidence was observed in LPS-induced piglets with ASPS treatment and was tendentially similar to those in CONTR. Intestinal barrier disturbance is characterised by a high incidence of diarrhoea and decreased performance in immune stress piglets⁽³⁾. Improvement in gastrointestinal epithelial function may elaborate the cause of improved performance and decreased diarrhoea incidence due to ASPS application.

Deterioration in intestinal epithelial morphology and increased intestinal permeability are characterised by attenuation in absorption capacity of nutrients, fluids and electrolytes as a result of immune stress, which lead to fluid and electrolyte accumulation in the bowel and contribute to the development of diarrhoea⁽³⁸⁾. In the present study, dietary ASPS contributed to the irregular shape and arrangement of epithelial villus, as well as the elevation in villus height in ileum of LPS-challenged piglets in contrast to the challenged piglets without ASPS treatment. These signs of morphological improvement were further supported by biochemical parameters, including higher enzyme activity of ileal lactase and DAO. Lactase overlying the mucosal surface is responsible for decomposing disaccharides and facilitates mucosa maturation and digestive function⁽³⁹⁾. Likewise, DAO localised in the cytoplasm of apical villous cells of the small intestine with high activity should reflect the status of intestinal mucosa integrity^(40,41). Declined mucosal DAO activity as a biologic marker of boosted intestinal permeability under the condition of gut inflammation injury has been displayed in many research studies^(42,43). As expected, the activities of these two

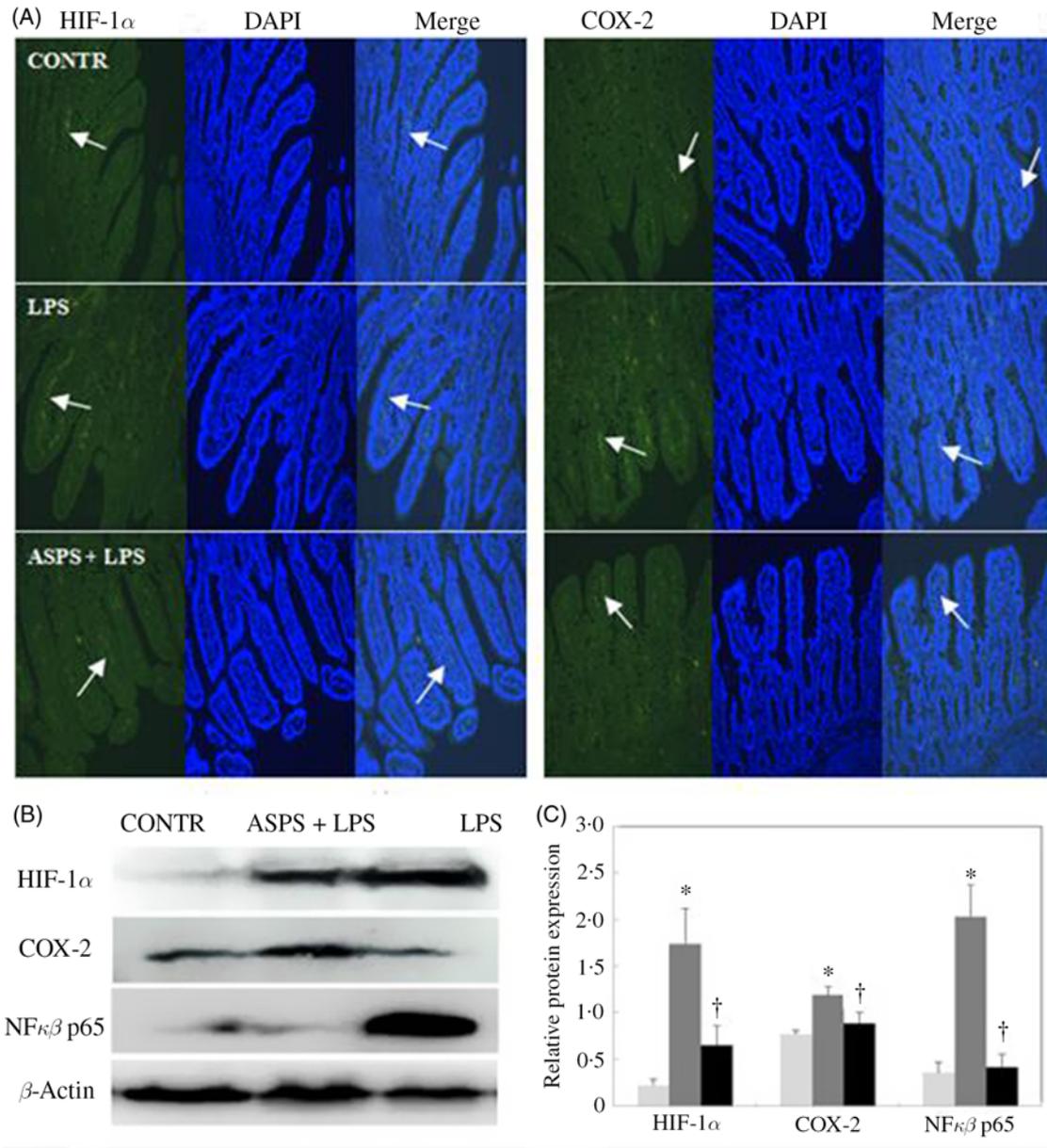


Fig. 3. Effects of *Acanthopanax senticosus* polysaccharides (ASPS) on the ileal expressions of hypoxia-inducible factor-1 α (HIF-1 α), cyclo-oxygenase-2 (COX-2) and NF κ B p65 in lipopolysaccharide (LPS)-challenged piglets (n 6). (A) The representative photomicrographs of ileal segments at a \times 100 magnification (green fluorescence) were observed by immunofluorescence. Nuclei were stained by 4',6-diamidino-2-phenylindole (DAPI) (blue fluorescence). Arrows indicate the location of HIF-1 α and COX-2 staining. (B) The bands are the representative Western blot images of HIF-1 α , COX-2, NF κ B p65 and β -actin in ileum. (C) Values are means of protein expression of HIF-1 α and COX-2, with standard errors represented by vertical bars. * $P < 0.05$ v. CONTR; † $P < 0.05$ v. LPS. CONTR, piglets receiving basal diet and injected with saline challenge (○); LPS, piglets receiving basal diet and injected with *Escherichia coli* LPS (■); ASPS + LPS, piglets receiving basal diet with 800 mg/kg ASPS and injected with LPS challenge (■).

enzymes in the present study were normalised with the treatment of ASPS under challenged condition. The circulatory concentration of D-xylose was measured to evaluate the intestinal absorption function, and the value of it had been increased in ASPS treatment and no significance was observed when comparing with the CONTR and LPS groups in the present study. Collectively, the preceding changes serving as a relatively stable marker of intestinal mucosal status confirmed the alleviated loss of mucosal absorptive capacity and intestinal permeability with dietary ASPS, which was also in agreement with lower diarrhoea incidence.

The facts that interaction of inflammation and hypoxia results in a compromised intestinal barrier function have been identified in a number of studies^(44,45). Hypoxia-inducible HIF-1 α is critical in activating the master regulator of inflammatory response of NF κ B and leads to NF κ B-induced inflammation⁽⁴⁶⁾, and pro-inflammatory cytokines like TNF- α ⁽¹⁵⁾, IL-6⁽¹⁶⁾ and iNOS⁽⁴⁷⁾ are among the target genes identified for hypoxia-inducible NF κ B. In turn, the existence of pro-inflammatory cytokines could induce HIF-1 α ⁽¹²⁾, and HIF-1 α also could be the direct target for NF κ B through binding the 197/188 location on the HIF-1 α promoter⁽⁴⁸⁾. To elucidate the mechanism where ASPS relieves

inflammatory intestinal injury, our previous investigations using LPS-challenged mice have identified that ASPS supplementation caused the inhibition of gut TLR4/NFκB signalling pathway and concomitant intestinal epithelial improvement^(20,21). However, the role of ASPS supplementation on HIF-1α regulation remains to be elucidated. Here, unanimous results of degressive HIF-1α expression were achieved in this work by the support of mRNA, protein and immune-fluorometric assay with ASPS treatment, and these changes were accompanied by a sharp decrease in intestinal mucosal expression and concentration of NFκB, TNF-α, iNOS, as well as IL-1β. Meanwhile, TNF-α is a central mediator in the predisposition and exacerbation of gastrointestinal inflammation⁽⁴⁹⁾, and it is thought to link with the inhibition of phosphorylation of the myosin light-chain mediated by the myosin light-chain kinase, leading to alternatively disrupted TJ stability and dysfunction of TJ protein expression⁽⁵⁰⁾. This observation can be extended to our study. Here, a drastic decline in the expression and concentration of ileal TNF-α in this work not only emphasised the important role of dietary ASPS in the regulation of TNF-α release and consequent improvement of gut epithelial barrier function of LPS-challenged piglets but also revealed that the down-regulation of TNF-α in ASPS-fed piglet is linked with HIF-1α change during immune stress. Our data were supported by our previous report in which the relief of releasing of TNF-α in the modulation NFκB/MLCK and the improvement of gut epithelial TJ by ASPS were observed in challenged mice⁽²¹⁾. Given the crosstalk between HIF-1α and NFκB, these emerging evidences in this work are particularly relevant to explain, at least in part, that HIF-1α expression involved in the modulation of the NFκB signalling pathway by dietary ASPS resulted in the reduction of the previous inflammatory mediators and led to improved intestinal barrier function of piglets under inflammation condition.

Cyclo-oxygenase (COX)-2, an inducible key enzyme in the production of inflammatory prostanoids, could be up-regulated as the direct target for HIF-1α due to its direct binding to a specific location at -506 on the COX-2 promoter, and this highlights the biological significance of COX-2 up-regulation during hypoxia⁽⁵¹⁾. COX-2 overexpression has been described in the destruction of intestinal barrier in rat peritonitis model⁽⁵²⁾. Therefore, identifying the regulatory role of ASPS in HIF-1α-inducible COX-2 up-regulation is crucial for further development of novel molecular target for diarrhoea prevention by ASPS. As expected, the decrease in gut mucosal HIF-1α expression accompanied with COX-2 activation by analysis detection of both protein and immunofluorescence following ASPS treatment in LPS-induced immune stress model of piglets was achieved in this work. In this regard, these emerging evidences suggested that up-regulation of COX-2 represented intestinal mucosa adaptive response to hypoxia, and the down-regulation of the HIF-1α/COX-2 signalling pathway may be a novel finding regarding the mechanism in which dietary ASPS relieved gut impair in LPS-challenged piglets. However, further research *in vitro* is needed to clarify whether the suppressed activation of the NFκB pathway with a sequential HIF-1α expression or modulation of HIF-1α-inducible NFκB is involved in ASPS work.

It still remains unclear that how the polysaccharides isolated from *A. senticosus* possibly affect the TLR4/NFκB signalling

pathway. Due to share cell-type specificity of immune cell receptors with LPS, polysaccharide isolated from *A. senticosus* was reported to exert immunostimulation by the surface binding of the TLR4 and TLR2 receptors expressed on B cells and macrophages⁽¹⁷⁾, and natural plant-derived polysaccharides have been reported to be not connected with any tissue injuries at their biologically effective dose⁽⁵³⁾. In our relevant works, the possible reason why ASPS relieved the activation of TLR4/NFκB⁽²⁰⁾ and NFκB/HIF-1α/COX-2 pathway and intestinal barrier dysfunction following LPS challenge may be connected with competing with the binding of TLR4 receptor with LPS and resulting in the reverse in excessive activation of this signalling. Notably, activated kinds of immune cells and the acted receptors on the surface of immune cells are differences among different compositions of polysaccharides due to diversities and complexity in the sequence and spatial structure of the monosaccharides making up the polysaccharides^(54,55), which is likely connected with the difference in the extraction parts of plant, process and method. The ASPS used in the present research mainly consist of glucose residues (online Supplementary Fig. S1). Glucan, a polysaccharide consisting of glucose residues, is inferred to be likely the main active ingredient of ASPS to show the positive effect because glucan is also the main factor in functional additives of *Astragalus* polysaccharides⁽⁵⁶⁾ in China and many observations in previous reports suggesting the improved performance of pigs by dietary supplementation with β-glucan^(57,58). Nowadays, the herbal medicine of *A. senticosus* is widely used in China for modulating immune function although its polysaccharide products are under production. The further understanding on the action model of ASPS can help ASAP products to gain more attention.

In conclusion, the present study demonstrated that dietary supplementation with ASPS exerts alleviating role on inflammatory responses and intestinal barrier impairment and concomitant amelioration of diarrhoea incidence of immune stress piglets. These changes were possibly associated with a novel finding where down-regulation of HIF-1α/COX-2 is involved in NFκB-inducible release of inflammatory cytokines in immune stress piglets by ASPS supplementation.

Acknowledgements

The authors highly appreciate Mr Dongfeng Gao for assistance in the animal trial.

This study was jointly financially supported by the Natural Science Foundation of China (project no. 31702143; Beijing, P. R. China) and Key project of Liaoning Natural Science Foundation Plan (project no. 20170540796; Liaoning, P. R. China).

The authors contributed to this publication as follows: J. H. and X. J. L. designed the experiments; C. Y. F., F. Z., Y. S. L. and Q. T. X. participated in the animal experiment. J. H. and C. Y. F. discussed the results and wrote the paper. All authors read and approved the final manuscript.

The authors declare that they have no conflicts of interest.

Supplementary material

For supplementary material/s referred to in this article, please visit <https://doi.org/10.1017/S0007114519001363>

References

- Gewirtz AT, Liu Y, Sitaraman SV, *et al.* (2002) Intestinal epithelial pathobiology: past, present and future. *Best Pract Res Clin Gastroenterol* **16**, 851–867.
- Madara JL (1990) Warner-Lambert/Parke-Davis award lecture. Pathobiology of the intestinal epithelial barrier. *Am J Pathol* **137**, 1273–1281.
- Berkes J, Viswanathan VK, Savkovic SD, *et al.* (2003) Intestinal epithelial responses to enteric pathogens: effects on the tight junction barrier, ion transport, and inflammation. *Gut* **52**, 439–451.
- Livingston DH, Mosenthal AC & Deitch EA (1995) Sepsis and multiple organ dysfunction syndrome: a clinical-mechanistic overview. *New Horiz* **3**, 257–266.
- Farhadi A, Keshavarzian A, Van de Kar LD, *et al.* (2005) Heightened responses to stressors in patients with inflammatory bowel disease. *Am J Gastroenterol* **100**, 1796–1804.
- Bennett EJ, Tennant CC, Piesse C, *et al.* (1998) Level of chronic life stress predicts clinical outcome in irritable bowel syndrome. *Gut* **43**, 256–261.
- Suenaert P, Bulteel V, Lemmens L, *et al.* (2002) Antitumor necrosis factor treatment restores the gut barrier in Crohn's disease. *Am J Gastroenterol* **97**, 2000–2004.
- Prasad S, Mingrino R, Kaukinen K, *et al.* (2005) Inflammatory processes have differential effects on claudins 2, 3 and 4 in colonic epithelial cells. *Lab Invest* **85**, 1139–1162.
- Semenza GL (2000) HIF-1: mediator of physiological and pathophysiological responses to hypoxia. *J Appl Physiol* **88**, 1474–1480.
- Feinman R, Deitch EA, Watkins AC, *et al.* (2010) HIF-1 mediates pathogenic inflammatory responses to intestinal ischemia-reperfusion injury. *Am J Physiol Gastrointest Liver Physiol* **299**, G833–G843.
- Kannan KB, Colorado I, Reino D, *et al.* (2011) Hypoxia-inducible factor plays a gut-injurious role in intestinal ischemia reperfusion injury. *Am J Physiol Gastrointest Liver Physiol* **300**, G853–G861.
- Frede S, Freitag P, Otto T, *et al.* (2005) The proinflammatory cytokine interleukin 1 β and hypoxia cooperatively induce the expression of adrenomedullin in ovarian carcinoma cells through hypoxia inducible factor 1 activation. *Cancer Res* **65**, 4690–4697.
- Belaiba RS, Bonello S, Zahringer C, *et al.* (2007) Hypoxia up-regulates hypoxia-inducible factor-1 α transcription by involving phosphatidylinositol 3-kinase and nuclear factor κ B in pulmonary artery smooth muscle cells. *Mol Biol Cell* **18**, 4691–4697.
- van Uden P, Kenneth NS & Rocha S (2008) Regulation of hypoxia-inducible factor-1 α by NF κ B. *Biochem J* **412**, 477–484.
- Taylor CT, Dzus AL & Colgan SP (1998) Autocrine regulation of epithelial permeability by hypoxia: role for polarized release of tumor necrosis factor α . *Gastroenterology* **114**, 657–668.
- Matsui H, Ihara Y, Fujio Y, *et al.* (1999) Induction of interleukin (IL)-6 by hypoxia is mediated by nuclear factor (NF)- κ B and NF-IL6 in cardiac myocytes. *Cardiovasc Res* **42**, 104–112.
- Han SB, Yoon YD, Ahn HJ, *et al.* (2003) Toll-like receptor-mediated activation of B cells and macrophages by polysaccharide isolated from cell culture of *Acanthopanax senticosus*. *Int Immunopharmacol* **3**, 1301–1312.
- Chen RZ, Liu ZQ, Zhao JM, *et al.* (2011) Antioxidant and immunobiological activity of watersoluble polysaccharide fractions purified from *Acanthopanax senticosus*. *Food Chem* **127**, 434–440.
- Han J, Bian L, Liu X, *et al.* (2014) Effects of *Acanthopanax senticosus* polysaccharide supplementation on growth performance, immunity, blood parameters and expression of pro-inflammatory cytokines genes in challenged weaned piglets. *Asian-Australas J Anim Sci* **27**, 1035–1043.
- Han J, Liu LX, Yu N, *et al.* (2015) Polysaccharides from *Acanthopanax senticosus* enhances intestinal integrity through inhibiting TLR4/NF- κ B signaling pathways in lipopolysaccharide-challenged mice. *Anim Sci J* **87**, 1011–1018.
- Han J, Li JH, Bai G, *et al.* (2017) *Acanthopanax senticosus* polysaccharides-induced intestinal tight junction injury alleviation via inhibition of NF- κ B/MLCK pathway in a mouse endotoxemia model. *World J Gastroenterol* **23**, 2175–2184.
- Taylor CT (2008) Interdependent roles for hypoxia inducible factor and nuclear factor- κ B in hypoxic inflammation. *J Physiol* **586**, 4055–4059.
- Zhang LP, Chen LY & Zhang YS (1993) Study on water soluble polysaccharides from *Acanthopanax senticosus* fruits isolation and purification and structural investigating of AS-2. *Cbi J Biochem Mol Biol* **9**, 1–5.
- Staub AM (1965) Removal of protein – Sevag method. *Methods Carbohydrate Chem* **5**, 5–6.
- Ministry of Agriculture of People's Republic of China (2004) *Feeding Standard of Swine, NY-T 65–2004*. Beijing: China Agriculture Press.
- Association of Official Analytical Chemists (2002) *Official Methods of Analysis, Association of Official Analytical Chemists*, 17th ed. Washington, DC: AOAC.
- Andersen AD, Sangild PT, Munch SL, *et al.* (2016) Delayed growth, motor function and learning in preterm pigs during early postnatal life. *Am J Physiol Regul Integr Comp Physiol* **310**, 481–492.
- Kelly D, O'Brien JJ & McCracken KJ (1990) Effect of creep feeding on the incidence duration and severity of post-weaning diarrhea in pigs. *Res Vet Sci* **48**, 223–228.
- Zhang YC, Wang XB, Cui SQ, *et al.* (2014) Study on relationship between intestinal epithelium tight junction proteins mRNA expression and piglets weaning diarrhea. *Chin Anim Husb Vet Med* **41**, 221–224.
- Ringø E, Salinas I, Olsen RE, *et al.* (2007) Histological changes in Atlantic salmon (*Salmo salar* L.) intestine following in vitro exposure to pathogenic and probiotic bacterial strains. *Cell Tissue Res* **328**, 109–116.
- Livak KJ & Schmittgen TD (2001) Analysis of relative gene expression data using real-time quantitative PCR and the 2^{- $\Delta\Delta$ CT} method. *Methods* **25**, 402–408.
- Yue YY, Lin Y, Zhang Q, *et al.* (2016) Expression and clinical significance of the thymic stromal lymphopoietin in intestinal mucosa of patients with ulcerative colitis. *Prog Anat Sci* **22**, 289–291.
- Kononoff PJ & Hanford KJ (2006) Technical note: estimating statistical power of mixed models used in dairy nutrition experiments. *J Dairy Sci* **89**, 3968–3971.
- Medzhitov R & Janeway Jr CA (1997) Innate immunity: the virtues of a nonclonal system of recognition. *Cell* **91**, 295–298.
- Rehli M (2002) Of mice and men: species variations of Toll-like receptor expression. *Trends Immunol* **23**, 375–378.
- Modlin RL (2002) Mammalian toll-like receptors. *Ann Allergy Asthma Im* **88**, 543–547.
- Kruzel ML, Harari Y, Chen CY, *et al.* (2000) Lactoferrin protects gut mucosal integrity during endotoxemia induced by lipopolysaccharide in mice. *Inflammation* **24**, 33–44.
- O'Loughlin EV, Scott R & Gall D (1991) Pathophysiology of infectious diarrhea: changes in intestinal structure and function. *J Pediatr Gastroenterol Nutr* **12**, 5–20.
- Olsen WA, Li BUK, Loyd ML, *et al.* (1996) Heterogeneity of intestinal lactase activity in children: relationship to lactase-phlorizin hydrolase messenger RNA abundance. *Pediatr Res* **39**, 877–881.

40. Thompson JS, Burnett DA, Markin RS, *et al.* (1988) Intestinal mucosa diamine oxidase activity reflects intestinal involvement in Crohn's disease. *Am J Gastroenterol* **83**, 756–760.
41. Schmidt WU, Sattler J, Hesterberg R, *et al.* (1990) Human intestinal diamine oxidase (DAO) activity in Crohn's disease: a new marker for disease assessment. *Agents Actions* **30**, 267–270.
42. Liu YL, Chen F, Odle J, *et al.* (2012) Fish oil enhances intestinal integrity and inhibits TLR4 and NOD2 signaling pathways in weaned pigs after LPS challenge. *J Nutr* **142**, 2017–2024.
43. Chen YP, Zhang H, Cheng YF, *et al.* (2018) Dietary L-threonine supplementation attenuates lipopolysaccharide-induced inflammatory responses and intestinal barrier damage of broiler chickens at an early age. *Br J Nutr* **119**, 1254–1262.
44. Yang S, Yu M, Sun L, *et al.* (2014) Interferon- γ -induced intestinal epithelial barrier dysfunction by NF- κ B/HIF-1 α pathway. *J Interferon Cytokine Res* **34**, 195–203.
45. Liu H, Li M, Wang P, *et al.* (2011) Blockade of hypoxia-inducible factor-1 α by YC-1 attenuates interferon- γ and tumor necrosis factor- α -induced intestinal epithelial barrier dysfunction. *Cytokine* **56**, 581–588.
46. Koong AC, Chen EY & Giaccia AJ (1994) Hypoxia causes the activation of nuclear factor κ B through the phosphorylation of I κ B α on tyrosine residues. *Cancer Res* **54**, 1425–1430.
47. Oliver KM, Taylor CT & Cummins EP (2009) Hypoxia. Regulation of NF κ B signalling during inflammation: the role of hydroxylases. *Arthritis Res Ther* **11**, 215.
48. Spirig R, Djafarzadeh S, Regueira T, *et al.* (2010) Effects of TLR agonists on the hypoxia-regulated transcription factor HIF-1 α and dendritic cell maturation under normoxic conditions. *PLoS ONE* **5**, e0010983.
49. Ye D, Ma I & Ma T (2006) Molecular mechanism of tumor necrosis factor- α modulation of intestinal epithelial tight junction barrier. *Am J Physiol Gastrointest Liver Physiol* **290**, G496–G504.
50. Clayburgh DR, Barrett TA, Tang Y, *et al.* (2005) Epithelial myosin light chain kinase-dependent barrier dysfunction mediates T cell activation-induced diarrhea *in vivo*. *J Clin Invest* **115**, 2702–2715.
51. Kaidi A, Qualtrough D, Williams AC, *et al.* (2006) Direct transcriptional up-regulation of cyclooxygenase-2 by hypoxia-inducible factor (HIF)-1 promotes colorectal tumor cell survival and enhances HIF-1 transcriptional activity during hypoxia. *Cancer Res* **66**, 6683–6691.
52. Short S, Wang J, Castle S, *et al.* (2013) Low doses of celecoxib attenuate gut barrier failure during experimental peritonitis. *Lab Invest* **93**, 119.
53. Mallick SK, Maiti S, Bhutia S, *et al.* (2010) Immunostimulatory properties of a polysaccharide isolated from *Astragalus hygrometricus*. *J Med Food* **13**, 665–672.
54. Shso BM, Xu W, Dai H, *et al.* (2004) A study on the immune receptors for polysaccharides from the roots of *Astragalus membranaceus*, a Chinese medicinal herb. *Biochem Biophys Res Commun* **320**, 1103–1111.
55. Shao BM, Dai H, Xu W, *et al.* (2004) Immune receptors for polysaccharides from *Ganoderma lucidum*. *Biochem Biophys Res Commun* **323**, 133–141.
56. Yuan SL, Piao XS, Li DF, *et al.* (2006) Effects of dietary *Astragalus* polysaccharide on growth performance and immune function in weaned pigs. *Anim Sci* **82**, 501–507.
57. Schoenherr WD, Pollmann DS & Coalson JA (1994) Titration of macrophages on growth performance of nursery pigs. *J Anim Sci* **72**, 57.
58. Dritz SS, Shi J, Kielian TL, *et al.* (1995) Influence of dietary β -glucan on growth performance, nonspecific immunity, and resistance to *Streptococcus suis* infection in weanling pigs. *J Anim Sci* **73**, 3341–3350.