



Conference on ‘The future of animal products in the human diet: health and environmental concerns’ Postgraduate Winner

Can milk proteins be a useful tool in the management of cardiometabolic health? An updated review of human intervention trials

Ágnes A. Fekete^{1,2*}, D. Ian Givens² and Julie A. Lovegrove¹

¹Department of Food and Nutritional Sciences, Faculty of Life Sciences, Hugh Sinclair Unit of Human Nutrition and Institute for Cardiovascular and Metabolic Research (ICMR), University of Reading, Reading RG6 6AR, UK

²Food Production and Quality Research Division, School of Agriculture, Policy and Development, Faculty of Life Sciences, University of Reading, Reading RG6 6AR, UK

The prevalence of cardiometabolic diseases is a significant public health burden worldwide. Emerging evidence supports the inverse association between greater dairy consumption and reduced risk of cardiometabolic diseases. Dairy proteins may have an important role in the favourable impact of dairy on human health such as blood pressure (BP), blood lipid and glucose control. The purpose of this review is to update and critically evaluate the evidence on the impacts of casein and whey protein in relation to metabolic function. Evidence from short-term clinical studies assessing postprandial responses to milk protein ingestion suggests benefits on vascular function independent of BP, as well as improvement in glycaemic homeostasis. Long-term interventions have been less conclusive, with some showing benefits and others indicating a lack of improvement in vascular function. During chronic consumption BP appears to be lowered and both dyslipidaemia and hyperglacemia seem to be controlled. Limited number of trials investigated the effects of dairy proteins on oxidative stress and inflammation. Although the underlying mechanisms of milk proteins on cardiometabolic homeostasis remains to be elucidated, the most likely mechanism is to improve insulin resistance. The incorporation of meals enriched with dairy protein in the habitual diet may result in the beneficial effects on cardiometabolic health. Nevertheless, future well-designed, controlled studies are needed to investigate the relative effects of both casein and whey protein on BP, vascular function, glucose homeostasis and inflammation.

Dairy protein: Metabolic health: Blood pressure: Vascular function

Milk and dairy products are widely consumed around the world on a daily basis. They are not only an important source of nutrients in the human diet, but they also represent important value in the food chain providing opportunities for farmers, food processors and retailers to contribute to increased food security and poverty alleviation⁽¹⁾. Therefore any change in milk and dairy consumption will have multiple impacts on human and animal health, environment, food security and

economics. Indeed, according to an OECD-FAO report, milk production is projected to increase by 180 million tonnes in the next decade, predominantly in developing countries⁽²⁾. Moreover, the inclusion of animal-derived products adds diversity to plant-based diets, providing an important source of many essential nutrients, the dietary requirements of which would be more difficult to meet by plant-based diets. However, the potential health impacts of animal-derived foods, and more specifically

Abbreviations: AA, amino acids; BCAA, branched-chain amino acids; BP, blood pressure; CRP, C-reactive protein; DBP, diastolic blood pressure; FMD, flow-mediated dilatation; GIP, gastric inhibitory peptide; GLP-1, glucagon-like peptide 1; LTP, lactotripeptides; RCT, randomised controlled trial; SBP, systolic blood pressure; WPL, whey protein and lycopene.

*Corresponding author: A. A. Fekete, email a.a.fekete@reading.ac.uk

milk and dairy consumption, have been questioned owing to their high saturated fat content (for review, see⁽³⁾). However, emerging epidemiological evidence supports the beneficial effects of milk and dairy consumption on health, particularly cardiometabolic health^(4–6).

Milk is a complex food, a unique package of many nutrients such as calcium, magnesium, iodine, phosphorus, vitamin B₁₂, pantothenic acid, riboflavin, high-quality protein, peptides and oligosaccharides. In the human body, these bioactive components may interact with each other and exert synergistic effects, making it difficult to assign the specific health effect of a single component. Bovine milk, which is widely consumed around the world, contains approximately 32–34 g/l protein of which 80 % (w/w) is casein and 20 % (w/w) is whey protein. Both milk proteins consist of smaller protein fractions such as casein: α -s1, α -s2, β and κ -casein; and whey: β -lactoglobulin, α -lactalbumin, lactoferrin, immunoglobulins, serum albumin, glycomacropeptide, enzymes and growth factors. Milk proteins are considered to be high-quality proteins. Whey protein is rich in branched-chain amino acids (AA; BCAA) such as leucine, isoleucine and valine, whilst casein contains more histidine, methionine, phenylalanine, proline, serine, tyrosine and valine. It is well established that casein and whey have differential effects on gastric emptying and kinetics of digestion and absorption⁽⁷⁾. Intact micellar casein clots in the stomach due to the low pH, and is, therefore, digested more slowly, which results in a prolonged and more sustained AA release. In contrast, intact whey (which is acid soluble) or hydrolysed whey and caseinate are absorbed more rapidly, with a faster AA release and half-life⁽⁷⁾. It is, however, of note that micellar casein is different from Ca or Na caseinate (micellar casein is acidified and neutralised with alkali e.g. NaOH or Ca(OH)₂ in order to form caseinate), as the latter are soluble and thus may show similarities to whey in terms of digestion rates^(8,9). As a result of their different inherent AA compositions leading to distinct absorption and kinetic behaviour, they may also have differential effects on human health.

The aim of this review is to update and critically evaluate the existing evidence on the effects of casein and whey on metabolic function, including blood pressure (BP), vascular function, glucose and lipid metabolism, and inflammation.

Comprehensive literature search

A comprehensive literature search was conducted using the electronic databases Medline, the Cochrane Library, EMBASE and Web of Science using the following terms: intervention, randomised controlled trials (RCT), clinical trials, high BP, hypertension, anti-hypert*, vascular function, endothelial function, vascular stiffness, milk protein, milk peptide*, casein, hydrolysate, human subjects, lipids, insulin, glucose, inflammation. Furthermore, hand-searching was performed on the reference lists of both studies and review articles. In addition, Google and Google Scholar were used to confirm

that the search was complete. The search period covered studies published until September 2015.

Blood pressure

CVD remain the leading cause of death in most countries worldwide. In the UK, there has been a significant decrease in death rates since 1961, and due to a combination of better healthcare and preventative strategies, in 2012 CVD became the second main cause of death (CVD caused 28 % of all death and cancer 29 %)⁽¹⁰⁾. Approximately seven million people live with CVD in the UK, which costs £19 billion each year (including premature death, lost productivity, hospital treatment and prescriptions) resulting in a significant economic burden⁽¹⁰⁾. Premature death from CVD can be prevented by improving modifiable risk factors. For example, it has been estimated that in the general population increasing physical activity, smoking cessation and dietary changes can lead to 50, 20–30 and 15–40 % mortality risk reduction, respectively⁽¹¹⁾.

High BP (hypertension) is the key modifiable risk factor of CVD and of stroke in particular. Nearly 30 % of adults in the UK have high BP; however, only half of them are aware of it and even less receive treatment⁽¹⁰⁾. High BP is present when systolic blood pressure (SBP) is ≥ 140 mmHg and/or diastolic blood pressure (DBP) is ≥ 90 mmHg⁽¹²⁾. It is important to treat hypertension and maintain BP in the normal range as elevated BP can cause irreversible damage to different organs such as kidneys, heart and eyes⁽¹²⁾.

Long-term studies on blood pressure

We have recently reviewed the evidence from RCT on the antihypertensive effects of milk proteins and peptides⁽¹³⁾. For that review we systematically searched and reviewed the literature until December 2012. There was an imbalance in the literature as more RCT were conducted using mainly one type of casein-derived peptides, called lactotripeptides (LTP). We, therefore conducted an updated meta-analysis on the impact of LTP on BP⁽¹⁴⁾, which included all available and relevant RCT and detailed subgroup and regression analyses, which were somewhat limited in previous meta-analyses in this area^(15–18). We found a small, but significant reduction in both SBP (-2.95 mmHg (95 % CI $-4.17, -1.73$; $P < 0.001$)) and DBP (-1.51 mmHg (95 % CI $-2.21, -0.80$; $P < 0.001$)) after 4 weeks of LTP supplementation in pre- and hypertensive populations. Since there was a statistically significant heterogeneity of treatment effects across studies, sub-group analyses were performed. These analyses suggested differences in countries where RCT were conducted: Japanese studies reported significantly greater BP-lowering effect of LTP (-5.54 mmHg for SBP; and -3.01 mmHg for DBP), compared with European studies (-1.36 mmHg for SBP; and -0.83 mmHg for DBP; $P = 0.002$ for SBP and < 0.001 for DBP). This was confirmed in a recent meta-analysis which focused on Asian RCT only. However, it only assessed SBP and

the authors reported a very similar reduction of -5.63 mmHg in SBP compared to our -5.54 mmHg⁽¹⁹⁾. There may be several explanations for this observation. Firstly Japanese diets contains less milk and dairy products than European diets, therefore consumption of milk proteins may have a greater overall impact when compared with populations that consume these proteins more regularly and in higher quantities⁽²⁰⁾. Furthermore, there are reported ethnic differences in the response to drug administration, BP lowering in particular⁽²¹⁾ which could impact on the response to these bioactive proteins. Finally differences in response may also have resulted from different spatial conformations (cis/trans) of LTP used in the studies, due to production processes⁽²²⁾. Intriguingly, we also found a small-study effect, and when all bias was considered it shifted the treatment effect towards a less significant SBP and non-significant DBP reduction in response to LTP supplementation. We concluded that with potential bias considered, LTP consumption may still be effective in lowering BP in mildly hypertensive or hypertensive groups⁽¹⁴⁾.

During our systematic literature search⁽¹³⁾ we found that there were very few studies investigating the BP-lowering effects of other casein-derived peptides in human subjects^(23–27). Furthermore these studies were limited, used different types of peptides and were often uncontrolled with poor methodological and study design. Due to these inconsistencies in the study design, it was impossible to compare these data and no firm conclusion could be drawn on the antihypertensive effects of casein-derived peptides. Similarly, we found a limited number of RCT conducted using intact whey or whey-derived peptides assessing their antihypertensive effects in human subjects^(28–33). These trials seem to be of higher quality than studies on casein-derived peptides; however, the findings of these studies were also inconsistent⁽¹³⁾.

Since our review, published in 2013, three new studies which assessed the effects of milk proteins on BP as primary outcome were published. Petyaev *et al.*⁽³⁴⁾ examined the impacts of whey protein embedded in a protective lycopene matrix, a new proprietary formulation, the so-called whey protein lycosome, in a pilot study. Authors hypothesised that this formulation would protect whey protein from gastrointestinal degradation which would increase the bioavailability of the protein, and thus reduce the need for a high dose. They administered 70 mg whey protein along with 7 mg lycopene in the form of a capsule (WPL) and compared this with whey protein (70 mg) and lycopene (7 mg) separately (taken once daily for a month). A significant decrease in BP (-7 mmHg in SBP and -4 mmHg in DBP, $P < 0.05$) in the WPL group was reported compared with baseline only and no effect relative to the whey and lycopene given separately. Due to the nature of this pilot study, there was no information on blinding, the sample size was small (ten/treatment group) and due to the limited statistical analysis further investigation is needed to evaluate the potential antihypertensive effect of WPL. Another RCT was conducted in overweight and obese adolescents (aged 12–15 years), who were asked to

consume 1 litre/d of either water, skimmed milk, whey or casein (milk-based treatment drink contained 35 g/l protein) for 12 weeks⁽³⁵⁾. A decrease in brachial and central aortic DBP compared with baseline and control group (consuming water) was observed, whereas whey protein appeared to increase brachial and central aortic SBP, and central DBP. The authors acknowledged several limitations of the study, including difficulties in recruitment, changes in the research protocol after study commencement and not controlling for the extra energy intake that 1 litre/d treatment drinks provided, which led to an increase in weight in those in the treatment groups compared with a loss in the control group which consumed water. Therefore, due to these limitations it was difficult to draw firm conclusions from these data. A study of Figueroa *et al.*⁽³⁶⁾ examined the effects of both whey and casein on BP and vascular function combined with exercise training in obese, hypertensive women. In their 4-week trial, participants were assigned to consume 30 g casein, whey or 34 g maltodextrin (control) and perform resistance and endurance exercises 3 d/week under a qualified instructor's supervision. They reported significant reduction in both brachial and aortic SBP in both whey and casein groups compared with the control, although this was not observed for DBP. The exercise training did not have additional effects on BP or arterial function, owing the beneficial effect on the cardiovascular system to the milk proteins (Table 1).

In summary, emerging evidence suggest that milk protein consumption for at least 4 weeks may result in small BP lowering; however, further well-controlled studies involving 24-h ambulatory BP monitor should be performed for confirmation.

Short-term studies on blood pressure

According to a typical Western eating pattern, people spend up to 18 h/d in a postprandial state consuming three or more meals daily. Furthermore elevated postprandial lipemia, glycaemia and inflammation have been linked with increased risk for chronic disease development, including diabetes and CVD^(37–39). Therefore dietary strategies that attenuate the postprandial metabolic disturbance are urgently required.

To date only two studies have evaluated the acute (short-term) effects of milk proteins on BP. Pal and Ellis compared 45 g whey protein isolate, 45 g Na-caseinate with 45 g glucose in conjunction with a breakfast in normotensive overweight and obese women⁽³²⁾, but found no effect of treatment. A more recent study compared the postprandial effects of several dietary proteins (milk protein, pea protein and egg-white) and carbohydrate-rich meals on BP-related responses⁽⁴⁰⁾. Although the authors failed to specify the specific type of milk protein isolate used, its BP-lowering effect was not significantly different to pea protein, although both milk and pea protein were significantly lower than egg-white ($P \leq 0.01$; Table 1). The lack of evidence on the acute BP effects of milk proteins warrants further research.

Table 1. Impacts of milk proteins on blood pressure

| Reference | Subjects | Study design and duration | Treatment (g) | Comparison | Treatment effect |
|---|--------------------------------|---------------------------|---|--|---|
| Long-term | | | | | |
| Petyaev <i>et al.</i> ⁽³⁴⁾ | Prehypertensive (n 40) | Pilot, 4 weeks | Whey protein isolate (70 mg) embedded into lycopene micelles (7 mg) | Whey protein isolate, lycopene and placebo | ↓BP |
| Arnberg <i>et al.</i> ⁽³⁵⁾ | Overweight adolescents (n 193) | 12 weeks | Casein (35 g/l), whey protein (35 g/l) and skimmed milk (1 litre) | Water, pretest control group | ↓bBP and cDBP in casein group, ↑cDBP, bSBP and cSBP in whey group |
| Figuroa <i>et al.</i> ⁽³⁶⁾ | Obese women (n 33) | 4 weeks | Casein, whey protein | Carbohydrate | ↓bSBP and aSBP in casein and whey groups |
| Short-term | | | | | |
| Teunissen-Beekman <i>et al.</i> ⁽⁴⁰⁾ | Overweight or obese (n 48) | 240 min | Milk protein, pea protein, egg-white protein | Maltodextrin | ↓BP milk and pea protein groups compared to egg-white protein group |

↑, Increase; ↓, decrease; BP, blood pressure; bBP, brachial blood pressure; cBP, central blood pressure; DBP, diastolic blood pressure; SBP, systolic blood pressure.

Vascular function

Vascular dysfunction is often used as an umbrella term for abnormalities of the vascular system, such as endothelial dysfunction and arterial stiffness⁽⁴¹⁾. The endothelium, the inner layer of cells of the vasculature, plays a key regulatory role in the vascular system. Any disturbance in endothelial function, such as increased permeability, reduced vasodilation and activation of thrombotic and inflammatory pathways, can lead to atherosclerotic development⁽⁴²⁾. Due to the central role of the endothelium in the development of atherosclerosis, several non-invasive methods have been developed to assess endothelial dysfunction. Flow-mediated dilation (FMD) is considered to be the gold standard method of assessing endothelial function and may surpass the predictive value of traditional risk factors such as smoking, elevated cholesterol level in predicting cardiovascular events in patients with established CVD⁽⁴³⁾. However, it is of note that this technique requires extensive training and is operator dependent, which may limit its value.

Arterial stiffness is a measure of arterial elasticity which is the ability to expand and contract along with cardiac pulsation and relaxation. CVD risk factors such as ageing, hypertension, smoking and diet have been shown to have a detrimental effect on arterial distensibility, inducing an imbalance between the synthesis and degradation of elastin and types 1 and 3 collagen⁽⁴⁴⁾. Pulse wave velocity is considered to be the gold standard to measure arterial stiffness and has a substantial predictive value for CVD events⁽⁴⁵⁾.

Long-term studies on vascular function

Our previous review also evaluated the health effects of milk proteins and/or their peptides on vascular function⁽¹³⁾. In brief, we identified nine chronic

RCT^(33,46–53), of which eight used LTP^(46–54) and one trial used intact casein and whey⁽³³⁾. These studies were diverse in several aspects of methodologies such as design, length and dose of treatment, subject characteristics and measures of vascular function, and most importantly type of milk proteins used. Due to this heterogeneity, it is not possible to draw firm conclusions on the relative effects of milk proteins on the vascular function.

We have identified three further RCT: Petyaev *et al.*⁽³⁴⁾ examined the impacts of WPL not only on BP, but also on vascular reactivity, using FMD. They reported statistically significant improvements in FMD in the WPL group only (+2.6%, $P < 0.05$) compared with baseline. Arnberg *et al.* also evaluated the effects of intact whey, casein and semi-skimmed milk on arterial stiffness using pulse wave velocity, however, failed to show any changes in vascular function⁽³⁵⁾. Figuroa *et al.*⁽³⁶⁾ reported favourable changes in augmentation index (a measure of arterial stiffness) and brachial-pulse wave velocity in both whey and casein groups combined with exercise, compared with the control group. It is of note that the randomisation may not have been adequate as the baseline values for both BP and arterial stiffness were different in the treatment groups, which may have confounded the study (Table 2).

Short-term studies on vascular function

Only four RCT were conducted to evaluate the effects of milk proteins on vascular function in a postprandial setting^(32,54–56). Pal and Ellis failed to show any acute effects of whey and casein ingestion with a meal in normotensive obese postmenopausal women on arterial stiffness measured by pulse wave analysis⁽³²⁾. Likewise, Turpeinen *et al.*⁽⁵⁴⁾ also did not observe any statistically significant change in arterial stiffness measured by pulse wave velocity after acute ingestion of 25 mg LTP with 2 g plant

Table 2. Impacts of milk proteins on vascular function

| References | Subjects | Study design and duration | Treatment (g) | Comparison | Treatment effect |
|--|--------------------------------|---------------------------|---|---|------------------|
| Long-term | | | | | |
| Petyaev <i>et al.</i> ⁽³⁴⁾ | Prehypertensive (n 40) | Pilot, 4 weeks | Whey protein isolate (70 mg) embedded into lycopene micelles (7 mg) | Whey protein isolate, lycopene and placebo | ↑FMD |
| Arnberg <i>et al.</i> ⁽³⁵⁾ | Overweight adolescents (n 193) | 12 weeks | Casein (35 g/l), whey protein (35 g/l) and skimmed milk (1 litre) | Water, pretest control group | ↔ |
| Short-term | | | | | |
| Mariotti <i>et al.</i> ⁽⁵⁷⁾ | Overweight men (n 10) | 360 min | Casein | Whey protein isolate, α-lactalbumin-enriched whey protein | ↔ |

FMD, flow-mediated dilation, ↑, increase; ↔, no effect.

sterol ester mixed in a milk drink in mildly hypertensive subjects. However, Ballard *et al.* reported significant improvements in arterial reactivity assessed by FMD (+4.3 %) at 120 min after ingestion compared with placebo corresponding time point ($P < 0.05$) in mildly hypertensive, overweight individuals after whey hydrolysate (5 g NOP-47) ingestion with water⁽⁵⁵⁾. Mariotti *et al.* failed to report any significant effects of casein, whey or α-lactalbumin enriched whey protein on digital volume pulse (a measure of arterial stiffness)⁽⁵⁷⁾ (Table 2).

Intriguingly, BP-lowering effects of milk proteins were not associated with changes in vascular function in the reviewed RCT⁽¹³⁾ which is confirmed by emerging evidence on the relationship between BP and arterial stiffness. This suggests that the interaction between BP and arterial stiffness may be bi-directional^(58,59) via complex interactions between different pathways such as inflammatory^(60,61), hormonal (e.g. leptin and insulin)^(61–63) and disturbances in endothelial-derived mediators⁽⁵⁸⁾. Therefore it is important to determine the effects of milk proteins on other mediators of CVD risk that may indirectly affect BP.

Glycaemic control

Insulin has a range of biological actions within the human body⁽⁶⁴⁾, it not only has a key regulatory role in metabolic energy disposal and storage in tissues, but also it is responsible for cell growth and development⁽⁶⁵⁾, ion transport⁽⁶⁶⁾ and sympathetic nervous system activity⁽⁶⁷⁾. In addition, insulin has haemodynamic activities such as increasing blood flow and cardiac output, probably via increased NO production⁽⁶⁴⁾. Giugliano *et al.* demonstrated insulin release after an intravenous infusion of L-arginine resulted in improvements in FMD⁽⁶⁸⁾. However, Gates *et al.*⁽⁶⁹⁾ showed an insulin-independent vasodilation after L-arginine administration. Similarly, Ballard *et al.* reported an insulin-independent FMD improvement in response to the acute ingestion of a whey-derived peptide, NOP-47⁽⁵⁵⁾.

It is well established that food proteins and more specifically AA acutely stimulate insulin secretion⁽⁷⁰⁾ with several AA possessing direct insulinotropic effects^(71,72).

Both whey and casein appear to increase insulin secretion, however, to different extents⁽⁷³⁾. This may be due to their effect on gastric emptying, absorption and kinetics, since the insulin responses seemed to correlate with the increase in plasma AA concentration after protein ingestion⁽⁷⁴⁾. Likewise, hydrolysates appear to increase insulin production more than intact proteins⁽⁷⁵⁾.

It is not yet known how milk proteins exert their beneficial effects on glucose homeostasis; however, BCAA, in particular, leucine, isoleucine, valine, lysine and threonine are shown to act as insulin secretagogues (inducing insulin secretion from pancreatic β-cells), with leucine reportedly having the greatest insulinotropic effect acutely⁽⁷⁶⁾. This may be via the regulation of both ATP production (by metabolic oxidation and allosteric activation of glutamate dehydrogenase) and K_{ATP} activity⁽⁷⁷⁾. Similarly, BCAA and particularly leucine, have been reported to activate the mammalian rapamycin pathway resulting in a higher incretin hormone (insulin, glucagon-like peptide 1 (GLP-1) and gastric inhibitory peptide (GIP)) synthesis^(77,78). GIP is also known as glucose-dependent insulinotropic peptide, synthesised by K cells found in the mucosa of the duodenum and jejunum in response to food ingestion, which may subsequently further induce insulin production⁽⁷⁹⁾. While the effect of GIP appears to be more pronounced at normoglycaemic levels, GLP-1 is more active during hyperglycaemia⁽⁷⁹⁾. Jakubowicz and Froy showed that whey protein drink increased GIP response (+80 %) in healthy adults, yet a mixture of BCAA mimicking the supply of AA in whey protein, failed to exert the same effect⁽⁸⁰⁾. Therefore they suggested that certain bioactive peptides and/or AA deriving from whey protein during digestion may be responsible for this action⁽⁸⁰⁾. GLP-1 is a potent antihyperglycaemic hormone secreted by intestinal L cells⁽⁷⁹⁾. Interestingly, it has been shown to possess cardioprotective effects, which may be further complemented by natriuretic and antioxidative stress on the kidneys leading to beneficial impacts on BP and vasculature⁽⁸¹⁾. This warrants further consideration in future research when the effects of milk proteins on the cardiovascular system are assessed. Additionally, GLP-1 was more pronounced in healthy subjects after whey consumption



compared with casein or soya; however, after 2 h of ingestion the concentration of the hormone decreased, while it continued to increase after casein^(80,82,83). This may be explained by the different plasma kinetics of milk proteins. Two enzyme inhibitory peptides deriving from milk proteins have been associated with the beneficial effects on the glucose homeostasis: dipeptidyl peptidase-IV enzyme inhibitors and α -glucosidase enzyme inhibitors. Although dipeptidyl peptidase-IV plays several roles in different physiological processes, it has a distinct effect on glucose homeostasis by degrading incretin hormones GLP-1 and GIP⁽⁸⁴⁾. Whereas there is a definite lack of human studies examining the effects of dipeptidyl peptidase-IV inhibitory peptides deriving from milk proteins; some *in silico* (computer-aided), *in vitro* and limited animal studies suggest a potential role in controlling glucose metabolism. Lacroix and Li-Chan proposed that casein appears to be a better source of dipeptidyl peptidase-IV inhibitory peptides than whey protein⁽⁸⁵⁾. However, *in vitro* and *in vivo* studies suggest that whey protein may be equal or a better source of these inhibitory peptides (for review see⁽⁸⁶⁾). The α -glucosidase enzyme is found in the brush border of the enterocytes in the small intestine and is responsible for the synthesis and breakdown of carbohydrate by cleaving glycosidic bonds in complex carbohydrates to produce monosaccharides. A potential therapy in type 2 diabetic patients could be to reduce the absorption of glucose by carbohydrate hydrolysing enzymes such as α -glucosidase, which may also enhance and promote GLP-1 secretion⁽⁸⁷⁾. A very limited number of *in vitro* studies demonstrated that α -glucosidase inhibitory peptides may be derived from whey protein^(88,89). This clearly warrants further research.

Short-term studies on glycaemic control

Milk proteins have been extensively investigated for their insulinotropic and glucose-lowering effects in healthy subjects^(73,75,82,83,90–99) and to a limited extent in individuals with suboptimal glucose control^(100–106). The dose varied significantly between studies from as little as 10 g^(92,105,106)–51 g⁽⁹¹⁾. Milk proteins were administered on their own or with a meal or even served as pre-meals. Current evidence on the effects of whey protein on glucose control appears to be more promising than casein; furthermore it has been proposed that whey protein may be as effective at inducing insulin secretion as medication (sulfonylureas) prescribed for management of hyperglycaemia in type 2 diabetic patients^(80,107) (Table 3). Thus, providing a rationale for individuals with impaired glucose control or for patients with type 2 diabetes mellitus to consume whey protein prior to or with meals to control postprandial glucose metabolism. Future studies should examine the minimum dose at which whey protein exerts beneficial effects. Similarly due to the different time-frame by which milk proteins have an effect, longer postprandial trials (e.g. 24 h) may provide important information on how casein

could improve hyperglycaemia in individuals characterised by insulin resistance but with functional β -cells.

Long-term studies on glycaemic control

To the best of our knowledge, only three studies have investigated the chronic supplementation of milk proteins, rather than milk or dairy products, on glycaemic control. Pal *et al.* examined the effects of whey and casein (2×27 g/d for 12 weeks) in overweight and obese subjects⁽⁹⁶⁾. Most subjects had borderline impaired glucose tolerance at baseline, but at the end of the intervention a reduced fasting insulin concentration was observed in the whey protein group compared with the control group (glucose), although no change in fasting glucose was reported. In another study, a whey fermentation product (malleable protein matrix) decreased fasting plasma glucose concentration after 3 months supplementation compared with the control group, which was more pronounced in individuals with impaired fasting glucose at baseline⁽¹⁰⁸⁾. An acute-in-chronic study also reported a decrease in postprandial glucose response in whey group, which remained unchanged after the 4-week supplementation period⁽¹⁰²⁾ (Table 3).

Lipid metabolism

Short-term studies on lipids

Postprandial triacylglycerolaemia has been associated with markers of early atherosclerosis such as endothelial dysfunction and carotid media thickness^(109,110) and is strongly influenced by the composition of a meal, including the quality and quantity of fat^(111,112) and carbohydrate^(113,114). In theory due to the insulinogenic effects of milk proteins, their consumption would be predicted to attenuate postprandial lipaemia, as insulin has an inhibitory effect on hormone-sensitive lipase and hepatic release of free fatty acid and stimulatory effect on lipoprotein lipase which hydrolyses TAG for metabolism or storage. However, evidence from postprandial RCT is limited. Postprandial investigations reported decrease in TAG after both whey and casein ingestion in combination with a fat-rich meal in obese⁽⁹⁸⁾ and individuals with type 2 diabetes mellitus^(103,115), but showed no effect on TAG after acute consumption of whey protein^(99,104). Free fatty acid also decreased after whey and casein ingestion in obese⁽⁹⁹⁾ and type 2 diabetes mellitus patients⁽¹⁰⁴⁾. It is of note that parameters of lipid metabolism such as LDL and HDL and total cholesterol remain stable acutely^(116,117).

Recently an acute study reported that casein with a high-fat, high-energy meal, compared with whey protein and α -lactalbumin-enriched whey protein, significantly reduced postprandial TAG and had a marked effect of chylomicron kinetics⁽⁵⁷⁾. This could be due to the different physicochemical makeup of casein and whey protein, as casein forms a gel in the stomach influencing the rate of absorption and gastric emptying (Table 4).

Table 3. Impacts of milk proteins on glycaemic control

| Reference | Subjects | Study design and duration | Treatment (g) | Comparison | Treatment effect |
|---|-------------------------------------|---------------------------|----------------------------|---|--------------------------------------|
| Short-term | | | | | |
| Nilsson <i>et al.</i> ⁽⁷³⁾ | Healthy (<i>n</i> 12) | 120 min | WP (18.2 g) | White-wheat bread, milk, cod, cheese, gluten-low, gluten-high | ↑Insulin response, ↑GIP, ↔GLP-1 |
| Calbet <i>et al.</i> ⁽⁷⁵⁾ | Healthy (<i>n</i> 6) | 120 min | HC (36 g) | Intact casein | ↑GIP |
| Hall <i>et al.</i> ⁽⁸²⁾ | Healthy (<i>n</i> 9) | 180 min | WP (48 g) | Casein | ↑GLP-1 |
| Veldhorst <i>et al.</i> ⁽⁸³⁾ | Healthy (<i>n</i> 25) | 180 min | WP (10 and 25 %) | Casein, soya | ↑GLP-1 |
| Petersen <i>et al.</i> ⁽⁹⁰⁾ | Healthy (<i>n</i> 10) | 120 min | WP (20 g) | Glucose | ↓Glucose response |
| Pal and Ellis ⁽⁹¹⁾ | Healthy men (<i>n</i> 22) | 240 min | WP (50.8 g) | Turkey, egg, tuna | ↓Glucose response, ↑Insulin response |
| Akhavan <i>et al.</i> ⁽⁹²⁾ | Healthy (<i>n</i> 10) | 230 min | WP as pre-meal (10–20 g) | Glucose, water | ↓Glucose response, ↑GLP-1, ↑GIP |
| Akhavan <i>et al.</i> ⁽⁹³⁾ | Healthy (<i>n</i> 16/21) | 170 min | WP as pre-meal (10–40 g) | Water | ↓Glucose response |
| Acheson <i>et al.</i> ⁽⁹⁴⁾ | Healthy (<i>n</i> 23) | 330 min | WPI (50 % of diet) | Casein, soya, glucose | ↑Insulin response |
| Morifuji <i>et al.</i> ⁽⁹⁵⁾ | Healthy (<i>n</i> 10) | 120 min | WPH (86.9 %) | WP, soya, soya hydrolysate | ↑Insulin response |
| Nilsson <i>et al.</i> ⁽⁹⁷⁾ | Healthy (<i>n</i> 12) | 120 min | WP (18 g) | Glucose, amino acids | ↔GLP-1 |
| Holmer-Jensen <i>et al.</i> ⁽⁹⁸⁾ | Obese (<i>n</i> 11) | 480 min | WPI + fat-rich meal (45 g) | Casein and gluten | ↓GIP |
| Holmer-Jensen <i>et al.</i> ⁽⁹⁹⁾ | Obese (<i>n</i> 12) | 480 min | WPI + fat-rich meal (45 g) | WP specific fractions | ↔GLP-1 |
| Frid <i>et al.</i> ⁽¹⁰⁰⁾ | T2D (<i>n</i> 14) | 240 min | WP (27.6 g) | Ham (96 g) + lactose (5.3 g) | ↓Glucose response, ↑Insulin response |
| Ma <i>et al.</i> ⁽¹⁰¹⁾ | T2D (<i>n</i> 8) | 300 min | WP as pre-meal (55 g) | WP in main meal | ↑Insulin and incretin response |
| Ma <i>et al.</i> ⁽¹⁰²⁾ | T2D (<i>n</i> 7) | 240 min | WPI (25 g) | 'diet' drink | ↓Glucose response |
| Mortensen <i>et al.</i> ⁽¹⁰³⁾ | T2D (<i>n</i> 12) | 480 min | WPI + fat-rich meal (45 g) | Casein, gluten, cod | ↔GLP-1, ↓GIP |
| Mortensen <i>et al.</i> ⁽¹⁰⁴⁾ | T2D (<i>n</i> 12) | 480 min | WPI + fat-rich meal (45 g) | WP-specific fractions | ↔GLP-1 |
| Jonker <i>et al.</i> ⁽¹⁰⁵⁾ | T2D (<i>n</i> 13) | 250 min | CH (12 g) | CH (0 g) | ↑Insulin response |
| Geerts <i>et al.</i> ⁽¹⁰⁶⁾ | T2D (<i>n</i> 36) | 240 min | CH (12 g) | Intact casein | ↓Glucose response |
| Long-term | | | | | |
| Pal <i>et al.</i> ⁽⁹⁶⁾ | Overweight and obese (<i>n</i> 70) | 12 weeks | WPI (2 × 27 g/d) | Glucose | ↑Fasting insulin + HOMA-IR |
| Ma <i>et al.</i> ⁽¹⁰²⁾ | T2D (<i>n</i> 7) | 4 weeks | WPI (25 g) | 'diet' drink | ↓Glucose response |
| Gouni-Berthold <i>et al.</i> ⁽¹⁰⁸⁾ | MS (<i>n</i> 180) | 12 weeks | Whey MPM (15.3 g) | Placebo | ↓Glucose response |

↑, Increase; ↓, decrease; ↔, no effect; CH, casein hydrolysate; D, day; GIP, glucose-dependent insulinotropic polypeptide; GLP-1, glucagon-like peptide 1; HC, hydrolysed casein; HOMA-IR, homeostasis model assessment of insulin resistance; MS, metabolic syndrome; T2D, type-2 diabetes; Whey MPM, whey malleable protein matrix; WP, whey protein; WPH, whey protein hydrolysate; WPI, whey protein isolate.

Long-term studies on lipids

To date, five chronic RCT, which examined the lipid-lowering effects of milk proteins, have been identified. Three month supplementation of whey (2 × 25 g/d) and casein (2 × 25 g/d) during an *ad libitum* weight regain diet after substantial diet-induced weight loss in healthy obese subjects resulted in no change in plasma lipids⁽¹¹⁸⁾. However, whey protein isolate (2 × 27 g/d) significantly reduced fasting TAG, total cholesterol and LDL-cholesterol after 3 months in overweight, obese individuals⁽⁹⁶⁾. Another 3-month supplementation study with malleable protein matrix (15 g/d protein in two daily servings of 150 g yoghurt) reduced fasting TAG, which was more pronounced in subjects with elevated baseline TAG⁽¹⁰⁸⁾. In a 6-week study casein (35 g/d) also reduced total cholesterol in hypercholesterolaemic subjects⁽¹¹⁹⁾. Petyaev *et al.* reported a decrease in LDL-cholesterol, TAG and total cholesterol in their pilot study⁽³⁴⁾ (Table 4).

The limited evidence suggests that milk proteins have a beneficial impact on fasted lipids; however further studies are required. Although its possible mechanism of action is not clear, insulin may play a role. *In vitro* studies suggest that milk proteins and BCAA inhibit expression of genes involved in intestinal fatty acid and cholesterol absorption and synthesis⁽¹²⁰⁾. Whey has been shown to induce urinary excretion of tricarboxylic acid cycle compounds such as citric acid and succinic acid in rats, which are substrates for lipogenesis, suggesting an increased catabolic state (e.g. lipolysis) and reduced lipid accretion compared with casein⁽¹²¹⁾. This could be a possible mechanism of lipid reduction. Similarly, in another metabolic study conducted in human subjects, cheese (casein) appeared to induce lowering of urinary citrate⁽¹²²⁾, which suggests that cheese consumption affects the tricarboxylic acid cycle. Additionally, microbiota-related metabolite, hippuric acid was significantly higher in the cheese group, than in the milk, implying

Table 4. Impacts of milk proteins on lipid metabolism

| References | Subjects | Study design and duration | Treatment (g) | Comparison | Treatment effect |
|---|------------------------------|---------------------------|---|--|--|
| Short-term | | | | | |
| Brader <i>et al.</i> ⁽¹¹⁵⁾ | T2D (n 11) | 480 min | Casein combined with carbohydrates and a fat-rich meal (45 g) | Control meal, control meal + carbohydrate, control meal + casein | ↓TAG concentration in chylomicron-rich fraction |
| Holmer-Jensen <i>et al.</i> ⁽⁹⁸⁾ | Obese (n 11) | 480 min | WPI + fat-rich meal (45 g) | Cod and gluten | ↓TAG response, ↓TAG concentration in chylomicron-rich fraction, ↓FFA |
| Holmer-Jensen <i>et al.</i> ⁽⁹⁹⁾ | Obese (n 12) | 480 min | WPI + fat-rich meal (45 g) | WP specific fractions | ↔TAG response |
| Mortensen <i>et al.</i> ⁽¹⁰³⁾ | T2D (n 12) | 480 min | WPI + fat-rich meal (45 g) | Casein, gluten, cod | ↓TAG response, ↓FFA |
| Mortensen <i>et al.</i> ⁽¹⁰⁴⁾ | T2D (n 12) | 480 min | WPI + fat-rich meal (45 g) | WP specific fractions | ↔TAG response |
| Long-term | | | | | |
| Pal <i>et al.</i> ⁽⁹⁶⁾ | Overweight and obese (n 70) | 12 weeks | WPI (2 × 27 g/d) | Glucose | ↓Fasting TAG, ↓TC, ↓LDL-c |
| Weisse <i>et al.</i> ⁽¹¹⁹⁾ | Hyper-cholesterolemic (n 43) | 6 weeks | Casein (35 g/d) | Baseline | ↓TC |
| Claessens <i>et al.</i> ⁽¹¹⁸⁾ | Obese (n 48) | 12 weeks | WP (2 × 25 g/d) | Casein | ↔fasting lipids |
| Petyaev <i>et al.</i> ⁽³⁴⁾ | Prehypertensive (n 40) | Pilot, 4 weeks | Whey protein isolate (70 mg) embedded into lycopene micelles (7 mg) | Whey protein isolate, lycopene and placebo | ↓TC, ↓TAG, ↓LDL-c, ↑HDL |
| Gouni-Berthold <i>et al.</i> ⁽¹⁰⁸⁾ | MS (n 180) | 12 weeks | Whey MPM (15.3 g) | Placebo | ↓TAG |

↑, Increase; ↓, decrease; ↔, no effect; CH, casein hydrolysate; D, day; FFA, free fatty acids; HC; hydrolysed casein; HDL-c, HDL cholesterol; LDL-c, LDL cholesterol; MS; metabolic syndrome; T2D, type-2 diabetes; TC, total cholesterol; Whey MPM, whey malleable protein matrix; WP, whey protein; WPH, whey protein hydrolysate; WPI, whey protein isolate.

a stimulation of gut bacteria activity. The enhanced bacterial activity also resulted in higher SCFA⁽¹²²⁾, which have been proposed as key regulatory metabolites in lipid metabolism⁽¹²³⁾. This effect may be due to the cheese matrix rather than the casein *per se*. An *in vivo* study proposed another potential mechanism of action through decreased lipid infiltration into the liver in rats with non-alcoholic fatty liver⁽¹²⁴⁾. Another possible putative mechanism is increased fat oxidation. Lorenzen *et al.*⁽¹²⁵⁾ demonstrated an increased lipid oxidation after acute casein consumption compared with whey. They speculated that it may be due to lower insulin secretion after casein consumption relative to whey since insulin down-regulates lipid oxidation. However, insulin was not measured in the study and this mechanism could not be confirmed. The same research group examined the effects of dairy Ca on lipid metabolism in conjunction with a low- and high-fat diet for 10 d⁽¹²⁶⁾. They found that dairy Ca attenuated the increase in total and LDL-cholesterol, without affecting the rise in HDL-cholesterol. This observed phenomenon may be due to the formation of insoluble Ca-fatty acid soaps and/or the production of hydrophobe aggregation with bile and with other fatty acids^(126–128).

Inflammation and oxidative stress

Inflammation and oxidative stress are chronic conditions which contribute to many diseases such as obesity⁽¹²⁹⁾,

type 2 diabetes mellitus⁽¹³⁰⁾ and CVD⁽¹³¹⁾. Different dietary components have an impact on low-grade inflammation⁽¹³²⁾; however, there is a lack of RCT evaluating the acute and chronic consumption of milk proteins on inflammation or oxidative stress with inconsistent outcomes.

Long-term studies on inflammation and oxidative stress

A recent meta-analysis evaluated the effects of chronic consumption of whey protein and hydrolysate on C-reactive protein (CRP), a systemic inflammatory marker⁽¹³³⁾. Nine RCT were included which showed a small, non-significant reduction in CRP 0.42 mg/l (95 % CI −0.96, 0.13). Sub-group analyses suggested that >20 g/d may be more effective, and the elevated baseline CRP level (≥3 mg/l) could be more responsive to whey or whey peptides consumption⁽¹³³⁾. Similarly, Arnberg *et al.*⁽³⁵⁾ reported no change in CRP in adolescence after whey, casein or skimmed milk consumption for 12 weeks.

IL-6, IL-8 and TNF- α are also recognised inflammatory markers, which induce CRP. Pal and Ellis failed to observe significant changes in these inflammatory markers (2 × 27 g whey or casein or glucose for 12 weeks) in overweight individuals⁽³³⁾. However, Sugawara *et al.*⁽¹³⁴⁾ reported decreased level of IL-6, IL-8 and TNF- α in patients with chronic obstructive pulmonary disease after whey intervention compared with the control group. Likewise, IL-6 and TNF- α were decreased after lactoferrin consumption for 6 months in postmenopausal women⁽¹³⁵⁾.

Table 5. Impacts of milk proteins on inflammation and oxidative stress

| Reference | Subjects | Study design and duration | Treatment (g) | Comparison | Treatment effect |
|--|--|---------------------------|--|--|----------------------------|
| Long-term | | | | | |
| Sugawara <i>et al.</i> ⁽¹³⁴⁾ | COPD (<i>n</i> 36) | 12 weeks | WP (20 g) | 0 g WP | ↓CRP, ↓IL-6, ↓IL-8, ↓TNF-α |
| Bharadwaj <i>et al.</i> ⁽¹³⁵⁾ | Post-menopausal women (<i>n</i> 38) | 24 weeks | Ribonuclease-enriched lactoferrin (2 × 125 mg/d) | Placebo | ↓IL-6, ↓TNF-α |
| Arnberg <i>et al.</i> ⁽³⁵⁾ | Overweight adolescents (<i>n</i> 193) | 12 weeks | Casein (35 g/l), whey protein (35 g/l) | Water, pretest control group | ↑CRP |
| Pal and Ellis ⁽³³⁾ | Overweight (<i>n</i> 70) | 12 weeks | WPI (54 g), Casein (54 g) | Glucose | ↔CRP, ↔IL-6, ↔TNF-α |
| Hirota <i>et al.</i> ⁽⁴⁶⁾ | Mild hyper-tensives (<i>n</i> 25) | 1 week | VPP (3.42 mg), IPP (3.87 mg) | Baseline | ↔CRP, ↓TNF-α |
| Short-term | | | | | |
| Pal and Ellis ⁽³²⁾ | Overweight postmenopausal women (<i>n</i> 20) | 480 min | WPI (45 g), Casein (45 g) | Glucose | ↔CRP, ↔IL-6, ↔TNF-α |
| Ballard <i>et al.</i> ⁽⁵⁶⁾ | Healthy (<i>n</i> 20) | 120 min | Whey-derived peptide (NOP-47, 5 g) | Placebo | ↔CRP, ↔IL-6, ↔IL-8, ↔TNF-α |
| Kerasiotti <i>et al.</i> ⁽¹³⁶⁾ | Healthy men (<i>n</i> 9) | 48 h | WP (0.26 g protein/kg BW/h) | Placebo | ↓CRP, ↓IL-6, ↑IL-10 |
| Holmer-Jensen <i>et al.</i> ⁽¹³⁷⁾ | Obese (<i>n</i> 11) | 240 min | WP + high-fat meal | Casein, cod and gluten + high-fat meal | ↓CCL5/RANTES, ↑MCP-1 |

↑, Increase; ↓, decrease; ↔, no effect; BW, body weight; CCL5, CC chemokine ligand-5; CH, casein hydrolysate; COPD, chronic obstructive pulmonary disease; CRP, C-reactive protein; IPP, isoleucine-proline-proline; MCP-1, monocyte chemoattractant protein-1; VPP, valine-proline-proline; Whey MPM, whey malleable protein matrix; WP, whey protein; WPH, whey protein hydrolysate; WPI, whey protein isolate.

Similarly Hirota *et al.*⁽⁴⁶⁾ reported decreased levels of TNF-α in mildly hypertensive subjects fed with the casein-derived LTP (Table 5).

Short-term studies on inflammation and oxidative stress

Pal and Ellis also reported no change in IL-6, IL-8 and TNF-α in a postprandial study investigating whey and casein⁽³²⁾. Likewise, a whey-derived peptide, NOP-47, also failed to change the level of serum cytokines (TNF-α, IL-6, IL-8, monocyte chemoattractant protein-1, vascular endothelial growth factor, soluble E-selectin, soluble vascular cell adhesion molecule-1) and chemokines⁽⁵⁶⁾. However, consumption of a cake containing whey protein after exhaustive cycling in nine subjects reported reduced levels of CRP and IL-6 by 46 and 50 %, respectively⁽¹³⁶⁾. Holmer-Jensen *et al.* assessed the postprandial effects of whey protein, casein, gluten and cod on low-grade inflammatory markers (monocyte chemoattractant protein-1, CC chemokine ligand-5/RANTES (Regulated on activation, normal T cell expressed and secreted)) in conjunction with a high fat meal⁽¹³⁷⁾. They reported that all meals increased CC chemokine ligand/RANTES; however, the smallest increase was observed after the whey protein meal. Monocyte chemoattractant protein-1 was initially suppressed after all meals, and the meal containing whey protein induced the smallest overall postprandial suppression⁽¹³⁷⁾ (Table 5).

The mechanism of action of milk proteins on oxidative stress and inflammation are unclear but Ca may suppresses the pro-inflammatory and reactive oxygen species

production *in vitro*⁽¹³⁸⁾. Interestingly, the milk protein-derived inhibitors of the angiotensin-I-converting enzyme may also be involved in the anti-inflammatory process⁽¹³⁹⁾.

Conclusion and implication for future studies

Taken together, there is a growing number of RCT which suggest that casein and whey protein may have a role in cardiometabolic health. Studies focused on reducing chronic disease risk factors such as hypertension and dysregulated lipid/glucose metabolism by non-pharmacological, dietary strategies will have significant implications not only for social and economic welfare, but also for the healthcare system.

Due to the different physicochemical makeup of casein and whey protein, they may exert differential effects in human subjects. Notably, manufacturing may play a significant role in the physiological effects of milk proteins; however, future studies should investigate which processing method results in more bioactive effects. There is inconclusive evidence on the relative impacts of milk proteins on diurnal BP and vascular function, yet there appears to be strong evidence on the insulinotropic impacts of dairy proteins, owing to the specific AA composition such as BCAA. They also appear to play a beneficial role in lipid homeostasis. Nevertheless the mechanism underlying the action of dairy proteins on the cardiometabolic health warrants further research.

The incorporation of a meal enriched with protein in the habitual diet may result in the improvement of cardiometabolic health as well as the prevention of developing



cardiometabolic diseases. Additionally, in contrast with pharmacological antihypertensive treatments, food-derived proteins have not been shown to cause any side-effects or hypotension, making them safe to consume by individuals with a variety of other disease conditions. After careful consideration of the available evidence and knowledge gaps, we have conducted two double-blind, controlled, cross-over studies (Whey2Go studies) aiming to compare the chronic ($n = 38$) and postprandial ($n = 27$) impacts of whey protein (2×28 g) and Ca-caseinate (2×28 g) with control (2×27 g, maltodextrin) on vascular function, BP, markers of insulin resistance, lipid metabolism and inflammatory status in men and women with mild hypertension ($\geq 120/80$ mmHg). These studies aim to provide valuable information on the relative effects of milk proteins on BP and on detailed aspects of vascular function compared with maltodextrin. These trials will further our knowledge of whether milk proteins have significant influences as health-promoting food components and whether the public as well as the food industry could benefit. The results from these studies are likely to be available in mid-2016.

Financial Support

This review received no specific grant from any funding agency in the public, commercial or not-for-profit sectors. A. A. F. is supported by a UK Biotechnology and Biological Sciences Research Council (BBSRC) studentship. The human trials were financially supported by BBSRC and Volac Int. Ltd.

Conflicts of Interest

J. A. L. and D. I. G. have previously received funding for research from AHDB Dairy. J. A. L. and D. I. G. have acted as advisors to the Dairy Council. J. A. L. and D. I. G. have received 'in kind' foods from Arla for an MRC funded study.

Authorship

A. A. F. conceived and wrote the manuscript. All authors critically reviewed and approved the final version of the manuscript.

References

- Muehlhoff EBA, McMahon D (2013) Milk and dairy products in human nutrition: Food and Agricultural Organization of the United Nations. <http://www.fao.org/docrep/018/i3396e/i3396e.pdf> (accessed August 2015).
- OECD-FAO Agricultural Outlook 2014–2023 (2014) OECD Publishing. https://www.embrapa.br/documents/1024963/1025740/OECD-FAO_Agricultural_Outlook_2014-2023/20082926-0f88-4159-970a-2a1c65795c47 (accessed August 2015).

- Markey O, Vasilopoulou D, Givens DI *et al.* (2014) Dairy and cardiovascular health: friend or foe? *Nutr Bull* **39**, 161–171.
- Elwood PC, Pickering JE, Givens DI *et al.* (2010) The consumption of milk and dairy foods and the incidence of vascular disease and diabetes: an overview of the evidence. *Lipids* **45**, 925–939.
- Ralston RA, Lee JH, Truby H *et al.* (2012) A systematic review and meta-analysis of elevated blood pressure and consumption of dairy foods. *J Hum Hypertens* **26**, 3–13.
- Aune D, Norat T, Romundstad P *et al.* (2013) Dairy products and the risk of type 2 diabetes: a systematic review and dose-response meta-analysis of cohort studies. *Am J Clin Nutr* **98**, 1066–1083.
- Boirie Y, Dangin M, Gachon P *et al.* (1997) Slow and fast dietary proteins differently modulate postprandial protein accretion. *Proc Natl Acad Sci USA* **94**, 14930–14935.
- Phillips SM (2011) A comparison of whey to caseinate. *Am J Physiol Endocrinol Metab* **300**, E610; author reply E1–2.
- Reitelseder S, Agergaard J, Doessing S *et al.* (2011) Whey and casein labeled with L-[1-¹³C]leucine and muscle protein synthesis: effect of resistance exercise and protein ingestion. *Am J Physiol Endocrinol Metab* **300**, E231–E242.
- Townsend NWJ, Bhatnagar P, Wickramasinghe K *et al.* (2014) *Cardiovascular Disease Statistics*. London: British Heart Foundation.
- Iestra JA, Kromhout D, van der Schouw YT *et al.* (2005) Effect size estimates of lifestyle and dietary changes on all-cause mortality in coronary artery disease patients: a systematic review. *Circulation* **112**, 924–934.
- Mancia G, De Backer G, Dominiczak A *et al.* (2007) Guidelines for the management of arterial hypertension: the Task Force for the Management of Arterial Hypertension of the European Society of Hypertension (ESH) and of the European Society of Cardiology (ESC). *Eur Heart J* **28**, 1462–1536.
- Fekete AA, Givens DI & Lovegrove JA (2013) The impact of milk proteins and peptides on blood pressure and vascular function: a review of evidence from human intervention studies. *Nutr Res Rev* **26**, 177–190.
- Fekete AA, Givens DI & Lovegrove JA (2015) Casein-derived lactotriptides reduce systolic and diastolic blood pressure in a meta-analysis of randomised clinical trials. *Nutrients* **7**, 659–681.
- Xu JY, Qin LQ, Wang PY *et al.* (2008) Effect of milk tripeptides on blood pressure: a meta-analysis of randomized controlled trials. *Nutrition* **24**, 933–940.
- Cicero AF, Gerocarni B, Laghi L *et al.* (2011) Blood pressure lowering effect of lactotriptides assumed as functional foods: a meta-analysis of current available clinical trials. *J Hum Hypertens* **25**, 425–436.
- Turpeinen AM, Jarvenpaa S, Kautiainen H *et al.* (2013) Antihypertensive effects of bioactive tripeptides—a random effects meta-analysis. *Ann Med* **45**, 51–56.
- Qin LQ, Xu JY, Dong JY *et al.* (2013) Lactotriptides intake and blood pressure management: a meta-analysis of randomised controlled clinical trials. *Nutr Metab Cardiovasc Dis* **23**, 395–402.
- Chanson-Rolle A, Aubin F, Braesco V *et al.* (2015) Influence of the lactotriptides isoleucine-proline-proline and valine-proline-proline on systolic blood pressure in Japanese subjects: a systematic review and meta-analysis of randomized controlled trials. *PLoS ONE* **10**, e0142235.
- Bulletin of the International Dairy Federation (2010) The World Dairy Situation 2010. http://www.milksa.co.za/sites/default/files/KORINL070_world_dairy_situation_2010.pdf.

21. Johnson JA (2008) Ethnic differences in cardiovascular drug response: potential contribution of pharmacogenetics. *Circulation* **118**, 1383–1393.
22. Siltari A, Viitanen R, Kukkurainen S *et al.* (2014) Does the cis/trans configuration of peptide bonds in bioactive tripeptides play a role in ACE-1 enzyme inhibition? *Biologics* **8**, 59–65.
23. Ashar MCR (2004) Fermented milk containing ACE-inhibitory peptides reduces blood pressure in middle aged hypertensive subjects. *Milchwissenschaft* **59**, 363–366.
24. Sekiya SKY, Kita E, Imamura Y *et al.* (1992) Antihypertensive effects of tryptic hydrolysate of casein on normotensive and hypertensive volunteers. *J Nutr Food Sci* **45**, 513–517.
25. Townsend RR, McFadden CB, Ford V *et al.* (2004) A randomized, double-blind, placebo-controlled trial of casein protein hydrolysate (C12 peptide) in human essential hypertension. *Am J Hypertens* **17**(11 Pt 1), 1056–1058.
26. Cadee JA, Chang CY, Chen CW *et al.* (2007) Bovine casein hydrolysate (c12 Peptide) reduces blood pressure in prehypertensive subjects. *Am J Hypertens* **20**, 1–5.
27. Sugai R (1998) ACE inhibitors and functional foods. *Bull IDF* **336**, 17–20.
28. Kawase M, Hashimoto H, Hosoda M *et al.* (2000) Effect of administration of fermented milk containing whey protein concentrate to rats and healthy men on serum lipids and blood pressure. *J Dairy Sci* **83**, 255–263.
29. Lee YM, Skurk T, Hennig M *et al.* (2007) Effect of a milk drink supplemented with whey peptides on blood pressure in patients with mild hypertension. *Eur J Nutr* **46**, 21–27.
30. Fluegel SM, Shultz TD, Powers JR *et al.* (2010) Whey beverages decrease blood pressure in prehypertensive and hypertensive young men and women. *Int Dairy J* **20**, 753–760.
31. Hodgson JM, Zhu K, Lewis JR *et al.* (2012) Long-term effects of a protein-enriched diet on blood pressure in older women. *Br J Nutr* **107**, 1664–1672.
32. Pal S & Ellis V (2011) Acute effects of whey protein isolate on blood pressure, vascular function and inflammatory markers in overweight postmenopausal women. *Br J Nutr* **105**, 1512–1519.
33. Pal S & Ellis V (2010) The chronic effects of whey proteins on blood pressure, vascular function, and inflammatory markers in overweight individuals. *Obesity (Silver Spring)* **18**, 1354–1359.
34. Petyaev IM, Dovgalevsky PY, Klochkov VA *et al.* (2012) Whey protein lysosome formulation improves vascular functions and plasma lipids with reduction of markers of inflammation and oxidative stress in prehypertension. *Sci World J* **2012**, 269476.
35. Arnberg K, Larnkjaer A, Michaelsen KF *et al.* (2013) Casein improves brachial and central aortic diastolic blood pressure in overweight adolescents: a randomised, controlled trial. *J Nutr Sci* **2**, e43.
36. Figueroa A, Wong A, Kinsey A *et al.* (2014) Effects of milk proteins and combined exercise training on aortic hemodynamics and arterial stiffness in young obese women with high blood pressure. *Am J Hypertens* **27**, 338–344.
37. Alipour A, Elte JW, van Zaanen HC *et al.* (2007) Postprandial inflammation and endothelial dysfunction. *Biochem Soc Trans* **35**(Pt 3), 466–469.
38. Klop B, Proctor SD, Mamo JC *et al.* (2012) Understanding postprandial inflammation and its relationship to lifestyle behaviour and metabolic diseases. *Int J Vasc Med* **2012**, 947417.
39. Lopez-Miranda J, Williams C & Lairon D (2007) Dietary, physiological, genetic and pathological influences on postprandial lipid metabolism. *Br J Nutr* **98**, 458–473.
40. Teunissen-Beekman KF, Dopheide J, Geleijnse JM *et al.* (2014) Differential effects of proteins and carbohydrates on postprandial blood pressure-related responses. *Br J Nutr* **112**, 600–608.
41. Schachinger V, Britten MB & Zeiher AM (2000) Prognostic impact of coronary vasodilator dysfunction on adverse long-term outcome of coronary heart disease. *Circulation* **101**, 1899–1906.
42. Verma S & Anderson TJ (2002) Fundamentals of endothelial function for the clinical cardiologist. *Circulation* **105**, 546–549.
43. Thijssen DH, Black MA, Pyke KE *et al.* (2011) Assessment of flow-mediated dilation in humans: a methodological and physiological guideline. *Am J Physiol Heart Circ Physiol* **300**, H2–12.
44. Bruno RM, Bianchini E, Faita F *et al.* (2014) Intima media thickness, pulse wave velocity, and flow mediated dilation. *Cardiovasc Ultrasound* **12**, 34.
45. Vlachopoulos C, Aznaouridis K & Stefanadis C (2010) Prediction of cardiovascular events and all-cause mortality with arterial stiffness: a systematic review and meta-analysis. *J Am Coll Cardiol* **55**, 1318–1327.
46. Hirota T, Ohki K, Kawagishi R *et al.* (2007) Casein hydrolysate containing the antihypertensive tripeptides Val-Pro-Pro and Ile-Pro-Pro improves vascular endothelial function independent of blood pressure-lowering effects: contribution of the inhibitory action of angiotensin-converting enzyme. *Hypertens Res* **30**, 489–496.
47. Jauhiainen T, Ronnback M, Vapaatalo H *et al.* (2010) Long-term intervention with *Lactobacillus helveticus* fermented milk reduces augmentation index in hypertensive subjects. *Eur J Clin Nutr* **64**, 424–431.
48. Turpeinen AM, Kumpu M, Rönback M *et al.* (2009) Antihypertensive and cholesterol-lowering effects of a spread containing bioactive peptides IPP and VPP and plant sterols. *J Funct Foods* **1**, 260–265.
49. Yoshizawa M, Maeda S, Miyaki A *et al.* (2009) Additive beneficial effects of lactotripeptides and aerobic exercise on arterial compliance in postmenopausal women. *Am J Physiol Heart Circ Physiol* **297**, H1899–H1903.
50. Yoshizawa M, Maeda S, Miyaki A *et al.* (2010) Additive beneficial effects of lactotripeptides intake with regular exercise on endothelium-dependent dilatation in postmenopausal women. *Am J Hypertens* **23**, 368–372.
51. Jauhiainen T, Rönback M, Vapaatalo H *et al.* (2007) *Lactobacillus helveticus* fermented milk reduces arterial stiffness in hypertensive subjects. *Int Dairy J* **17**, 1209–1211.
52. Cicero AF, Rosticci M, Gerocarni B *et al.* (2011) Lactotripeptides effect on office and 24-h ambulatory blood pressure, blood pressure stress response, pulse wave velocity and cardiac output in patients with high-normal blood pressure or first-degree hypertension: a randomized double-blind clinical trial. *Hypertens Res* **34**, 1035–1040.
53. Nakamura T, Mizutani J, Ohki K *et al.* (2011) Casein hydrolysate containing Val-Pro-Pro and Ile-Pro-Pro improves central blood pressure and arterial stiffness in hypertensive subjects: a randomized, double-blind, placebo-controlled trial. *Atherosclerosis* **219**, 298–303.
54. Turpeinen AM, Ehlers PI, Kivimaki AS *et al.* (2011) Ile-Pro-Pro and Val-Pro-Pro tripeptide-containing milk product has acute blood pressure lowering effects in mildly hypertensive subjects. *Clin Exp Hypertens* **33**, 388–396.
55. Ballard KD, Kupchak BR, Volk BM *et al.* (2013) Acute effects of ingestion of a novel whey-derived extract on vascular endothelial function in overweight, middle-aged men and women. *Br J Nutr* **109**, 882–893.

56. Ballard KD, Bruno RS, Seip RL *et al.* (2009) Acute ingestion of a novel whey-derived peptide improves vascular endothelial responses in healthy individuals: a randomized, placebo controlled trial. *Nutr J* **8**, 34.
57. Mariotti F, Valette M, Lopez C *et al.* (2015) Casein compared with whey proteins affects the organization of dietary fat during digestion and attenuates the postprandial triglyceride response to a mixed high-fat meal in healthy, overweight men. *J Nutr* **145**, 2657–2664.
58. Franklin SS (2005) Arterial stiffness and hypertension: a two-way street? *Hypertension* **45**, 349–351.
59. Dernelis J & Panaretou M (2005) Aortic stiffness is an independent predictor of progression to hypertension in non-hypertensive subjects. *Hypertension* **45**, 426–431.
60. Sesso HD, Buring JE, Rifai N *et al.* (2003) C-reactive protein and the risk of developing hypertension. *Jama* **290**, 2945–2951.
61. Tsai SS, Lin YS, Lin CP *et al.* (2015) Metabolic syndrome-associated risk factors and high-sensitivity c-reactive protein independently predict arterial stiffness in 9903 subjects with and without chronic kidney disease. *Medicine (Baltimore)* **94**, e1419.
62. Henry RM, Kostense PJ, Spijkerman AM *et al.* (2003) Arterial stiffness increases with deteriorating glucose tolerance status: the Hoorn Study. *Circulation* **107**, 2089–2095.
63. Singhal A, Farooqi IS, Cole TJ *et al.* (2002) Influence of leptin on arterial distensibility: a novel link between obesity and cardiovascular disease? *Circulation* **106**, 1919–1924.
64. Baron AD (1994) Hemodynamic actions of insulin. *Am J Physiol* **267**(2 Pt 1), E187–E202.
65. Rosen OM (1987) After insulin binds. *Science* **237**, 1452–1458.
66. Moore RD (1983) Effects of insulin upon ion transport. *Biochim Biophys Acta* **737**, 1–49.
67. Anderson EA, Hoffman RP, Balon TW *et al.* (1991) Hyperinsulinemia produces both sympathetic neural activation and vasodilation in normal humans. *J Clin Invest* **87**, 2246–2252.
68. Giugliano D, Marfella R, Verrazzo G *et al.* (1997) The vascular effects of L-arginine in humans. The role of endogenous insulin. *J Clin Invest* **99**, 433–438.
69. Gates PE, Boucher ML, Silver AE *et al.* (2007) Impaired flow-mediated dilation with age is not explained by L-arginine bioavailability or endothelial asymmetric dimethylarginine protein expression. *J Appl Physiol* **102**, 63–71.
70. Floyd JC Jr, Fajans SS, Conn JW *et al.* (1966) Insulin secretion in response to protein ingestion. *J Clin Invest* **45**, 1479–1486.
71. Schmid R, Schusdziarra V, Schulte-Frohlinde E *et al.* (1989) Role of amino acids in stimulation of postprandial insulin, glucagon, and pancreatic polypeptide in humans. *Pancreas* **4**, 305–314.
72. Schmid R, Schulte-Frohlinde E, Schusdziarra V *et al.* (1992) Contribution of postprandial amino acid levels to stimulation of insulin, glucagon, and pancreatic polypeptide in humans. *Pancreas* **7**, 698–704.
73. Nilsson M, Stenberg M, Frid AH *et al.* (2004) Glycemia and insulinemia in healthy subjects after lactose-equivalent meals of milk and other food proteins: the role of plasma amino acids and incretins. *Am J Clin Nutr* **80**, 1246–1253.
74. Calbet JA & MacLean DA (2002) Plasma glucagon and insulin responses depend on the rate of appearance of amino acids after ingestion of different protein solutions in humans. *J Nutr* **132**, 2174–2182.
75. Calbet JA & Holst JJ (2004) Gastric emptying, gastric secretion and enterogastrone response after administration of milk proteins or their peptide hydrolysates in humans. *Eur J Nutr* **43**, 127–139.
76. van Loon LJ, Saris WH, Verhagen H *et al.* (2000) Plasma insulin responses after ingestion of different amino acid or protein mixtures with carbohydrate. *Am J Clin Nutr* **72**, 96–105.
77. Yang J, Chi Y, Burkhardt BR *et al.* (2010) Leucine metabolism in regulation of insulin secretion from pancreatic beta cells. *Nutr Rev* **68**, 270–279.
78. Melnik BC (2012) Leucine signaling in the pathogenesis of type 2 diabetes and obesity. *World J Diabetes* **3**, 38–53.
79. Yabe D & Seino Y (2011) Two incretin hormones GLP-1 and GIP: comparison of their actions in insulin secretion and beta cell preservation. *Prog Biophys Mol Biol* **107**, 248–256.
80. Jakubowicz D & Froy O (2013) Biochemical and metabolic mechanisms by which dietary whey protein may combat obesity and type 2 diabetes. *J Nutr Biochem* **24**, 1–5.
81. Poudyal H (2015) Mechanisms for the cardiovascular effects of glucagon-like peptide-1. *Acta Physiol (Oxf)* **216**, 277–313.
82. Hall WL, Millward DJ, Long SJ *et al.* (2003) Casein and whey exert different effects on plasma amino acid profiles, gastrointestinal hormone secretion and appetite. *Br J Nutr* **89**, 239–248.
83. Veldhorst MA, Nieuwenhuizen AG, Hochstenbach-Waelen A *et al.* (2009) Dose-dependent satiating effect of whey relative to casein or soy. *Physiol Behav* **96**, 675–682.
84. Fan H, Yan S, Stehling S *et al.* (2003) Dipeptidyl peptidase IV/CD26 in T cell activation, cytokine secretion and immunoglobulin production. *Adv Exp Med Biol* **524**, 165–174.
85. Lacroix IME & Li-Chan ECY (2012) Evaluation of the potential of dietary proteins as precursors of dipeptidyl peptidase (DPP)-IV inhibitors by an *in silico* approach. *J Funct Foods* **4**, 403–422.
86. Patil P, Mandal S, Tomar SK *et al.* (2015) Food protein-derived bioactive peptides in management of type 2 diabetes. *Eur J Nutr* **54**, 863–880.
87. Slama G, Elgrably F, Sola A *et al.* (2006) Postprandial glycaemia: a plea for the frequent use of delta postprandial glycaemia in the treatment of diabetic patients. *Diab Metab* **32**, 187–192.
88. Lacroix IM & Li-Chan EC (2013) Inhibition of dipeptidyl peptidase (DPP)-IV and alpha-glucosidase activities by pepsin-treated whey proteins. *J Agric Food Chem* **61**, 7500–7506.
89. Konrad B, Anna D, Marek S *et al.* (2014) The evaluation of dipeptidyl peptidase (DPP)-IV, alpha-glucosidase and angiotensin converting enzyme (ACE) inhibitory activities of whey proteins hydrolyzed with serine protease isolated from Asian Pumpkin. *Int J Pept Res Ther* **20**, 483–491.
90. Petersen BL, Ward LS, Bastian ED *et al.* (2009) A whey protein supplement decreases post-prandial glycemia. *Nutr J* **8**, 47.
91. Pal S & Ellis V (2010) The acute effects of four protein meals on insulin, glucose, appetite and energy intake in lean men. *Br J Nutr* **104**, 1241–1248.
92. Akhavan T, Luhovyy BL, Panahi S *et al.* (2014) Mechanism of action of pre-meal consumption of whey protein on glycemic control in young adults. *J Nutr Biochem* **25**, 36–43.
93. Akhavan T, Luhovyy BL, Brown PH *et al.* (2010) Effect of premeal consumption of whey protein and its hydrolysate on food intake and postmeal glycemia and insulin responses in young adults. *Am J Clin Nutr* **91**, 966–975.

94. Acheson KJ, Blondel-Lubrano A, Oguey-Araymon S *et al.* (2011) Protein choices targeting thermogenesis and metabolism. *Am J Clin Nutr* **93**, 525–534.
95. Morifuji M, Ishizaka M, Baba S *et al.* (2010) Comparison of different sources and degrees of hydrolysis of dietary protein: effect on plasma amino acids, dipeptides, and insulin responses in human subjects. *J Agric Food Chem* **58**, 8788–8797.
96. Pal S, Ellis V & Dhaliwal S (2010) Effects of whey protein isolate on body composition, lipids, insulin and glucose in overweight and obese individuals. *Br J Nutr* **104**, 716–723.
97. Nilsson M, Holst JJ & Bjorck IM (2007) Metabolic effects of amino acid mixtures and whey protein in healthy subjects: studies using glucose-equivalent drinks. *Am J Clin Nutr* **85**, 996–1004.
98. Holmer-Jensen J, Mortensen LS, Astrup A *et al.* (2013) Acute differential effects of dietary protein quality on postprandial lipemia in obese non-diabetic subjects. *Nutr Res* **33**, 34–40.
99. Holmer-Jensen J, Hartvigsen ML, Mortensen LS *et al.* (2012) Acute differential effects of milk-derived dietary proteins on postprandial lipaemia in obese non-diabetic subjects. *Eur J Clin Nutr* **66**, 32–38.
100. Frid AH, Nilsson M, Holst JJ *et al.* (2005) Effect of whey on blood glucose and insulin responses to composite breakfast and lunch meals in type 2 diabetic subjects. *Am J Clin Nutr* **82**, 69–75.
101. Ma J, Stevens JE, Cukier K *et al.* (2009) Effects of a protein preload on gastric emptying, glycemia, and gut hormones after a carbohydrate meal in diet-controlled type 2 diabetes. *Diab Care* **32**, 1600–1602.
102. Ma J, Jesudason DR, Stevens JE *et al.* (2015) Sustained effects of a protein 'preload' on glycaemia and gastric emptying over 4 weeks in patients with type 2 diabetes: a randomized clinical trial. *Diab Res Clin Pract* **108**, e31–e34.
103. Mortensen LS, Hartvigsen ML, Brader LJ *et al.* (2009) Differential effects of protein quality on postprandial lipemia in response to a fat-rich meal in type 2 diabetes: comparison of whey, casein, gluten, and cod protein. *Am J Clin Nutr* **90**, 41–48.
104. Mortensen LS, Holmer-Jensen J, Hartvigsen ML *et al.* (2012) Effects of different fractions of whey protein on postprandial lipid and hormone responses in type 2 diabetes. *Eur J Clin Nutr* **66**, 799–805.
105. Jonker JT, Wijngaarden MA, Kloek J *et al.* (2011) Effects of low doses of casein hydrolysate on post-challenge glucose and insulin levels. *Eur J Intern Med* **22**, 245–248.
106. Geerts BF, van Dongen MG, Flameling B *et al.* (2011) Hydrolyzed casein decreases postprandial glucose concentrations in T2DM patients irrespective of leucine content. *J Diet Suppl* **8**, 280–292.
107. McGregor RA & Poppitt SD (2013) Milk protein for improved metabolic health: a review of the evidence. *Nutr Metab (Lond)* **10**, 46.
108. Gouni-Berthold I, Schulte DM, Krone W *et al.* (2012) The whey fermentation product malleable protein matrix decreases TAG concentrations in patients with the metabolic syndrome: a randomised placebo-controlled trial. *Br J Nutr* **107**, 1694–1706.
109. Teno S, Uto Y, Nagashima H *et al.* (200) Association of postprandial hypertriglyceridemia and carotid intima-media thickness in patients with type 2 diabetes. *Diab Care* **23**, 1401–1406.
110. Vigna GB, Delli Gatti C & Fellin R (2004) Endothelial function and postprandial lipemia. *Nutr Metab Cardiovasc Dis* **14**, 121–127.
111. Thomsen C, Storm H, Holst JJ *et al.* (2003) Differential effects of saturated and monounsaturated fats on postprandial lipemia and glucagon-like peptide 1 responses in patients with type 2 diabetes. *Am J Clin Nutr* **77**, 605–611.
112. Thomsen C, Rasmussen O, Lousen T *et al.* (1999) Differential effects of saturated and monounsaturated fatty acids on postprandial lipemia and incretin responses in healthy subjects. *Am J Clin Nutr* **69**, 1135–1143.
113. Cohen JC & Schall R (1988) Reassessing the effects of simple carbohydrates on the serum triglyceride responses to fat meals. *Am J Clin Nutr* **48**, 1031–1034.
114. Lairon D, Play B & Jourdeuil-Rahmani D (2007) Digestible and indigestible carbohydrates: interactions with postprandial lipid metabolism. *J Nutr Biochem* **18**, 217–227.
115. Brader L, Holm L, Mortensen L *et al.* (2010) Acute effects of casein on postprandial lipemia and incretin responses in type 2 diabetic subjects. *Nutr Metab Cardiovasc Dis* **20**, 101–109.
116. Olefsky JM, Crapo P & Reaven GM (1976) Postprandial plasma triglyceride and cholesterol responses to a low-fat meal. *Am J Clin Nutr* **29**, 535–539.
117. Roche HM, Zampelas A, Knapper JM *et al.* (1998) Effect of long-term olive oil dietary intervention on postprandial triacylglycerol and factor VII metabolism. *Am J Clin Nutr* **68**, 552–560.
118. Claessens M, van Baak MA, Monsheimer S *et al.* (2009) The effect of a low-fat, high-protein or high-carbohydrate ad libitum diet on weight loss maintenance and metabolic risk factors. *Int J Obes (Lond)* **33**, 296–304.
119. Weisse K, Brandsch C, Zernsdorf B *et al.* (2010) Lupin protein compared to casein lowers the LDL cholesterol: HDL cholesterol-ratio of hypercholesterolemic adults. *Eur J Nutr* **49**, 65–71.
120. Chen Q & Reimer RA (2009) Dairy protein and leucine alter GLP-1 release and mRNA of genes involved in intestinal lipid metabolism *in vitro*. *Nutrition* **25**, 340–349.
121. Lillefosse HH, Clausen MR, Yde CC *et al.* (2014) Urinary loss of tricarboxylic acid cycle intermediates as revealed by metabolomics studies: an underlying mechanism to reduce lipid accretion by whey protein ingestion? *J Proteome Res* **13**, 2560–2570.
122. Zheng H, Yde CC, Clausen MR *et al.* (2015) Metabolomics investigation to shed light on cheese as a possible piece in the French paradox puzzle. *J Agric Food Chem* **63**, 2830–2839.
123. Tremaroli V & Backhed F (2012) Functional interactions between the gut microbiota and host metabolism. *Nature* **489**, 242–249.
124. Hamad EM, Taha SH, Abou Dawood AG *et al.* (2011) Protective effect of whey proteins against nonalcoholic fatty liver in rats. *Lipids Health Dis* **10**, 57.
125. Lorenzen J, Frederiksen R, Hoppe C *et al.* (2012) The effect of milk proteins on appetite regulation and diet-induced thermogenesis. *Eur J Clin Nutr* **66**, 622–627.
126. Lorenzen JK & Astrup A (2011) Dairy calcium intake modifies responsiveness of fat metabolism and blood lipids to a high-fat diet. *Br J Nutr* **105**, 1823–1831.
127. Gacs G & Barltrop D (1977) Significance of Ca-soap formation for calcium absorption in the rat. *Gut* **18**, 64–68.
128. Govers MJ, Termont DS, Van Aken GA *et al.* (1994) Characterization of the adsorption of conjugated and unconjugated bile acids to insoluble, amorphous calcium phosphate. *J Lipid Res* **35**, 741–748.



129. Biro FM & Wien M (2010) Childhood obesity and adult morbidities. *Am J Clin Nutr* **91**, 1499s–1505s.
130. Conen D, Rexrode KM, Creager MA *et al.* (2009) Metabolic syndrome, inflammation, and risk of symptomatic peripheral artery disease in women: a prospective study. *Circulation* **120**, 1041–1047.
131. Libby P, Ridker PM & Hansson GK (2009) Inflammation in atherosclerosis: from pathophysiology to practice. *J Am Coll Cardiol* **54**, 2129–2138.
132. Galland L (2010) Diet and inflammation. *Nutr Clin Pract* **25**, 634–640.
133. Zhou LM, Xu JY, Rao CP *et al.* (2015) Effect of whey supplementation on circulating C-reactive protein: a meta-analysis of randomized controlled trials. *Nutrients* **7**, 1131–1143.
134. Sugawara K, Takahashi H, Kashiwagura T *et al.* (2012) Effect of anti-inflammatory supplementation with whey peptide and exercise therapy in patients with COPD. *Respir Med* **106**, 1526–1534.
135. Bharadwaj S, Naidu TA, Betageri GV *et al.* (2010) Inflammatory responses improve with milk ribonuclease-enriched lactoferrin supplementation in postmenopausal women. *Inflamm Res* **59**, 971–978.
136. Kerasiotti E, Stagos D, Jamurtas A *et al.* (2013) Anti-inflammatory effects of a special carbohydrate-whey protein cake after exhaustive cycling in humans. *Food Chem Toxicol* **61**, 42–46.
137. Holmer-Jensen J, Karhu T, Mortensen LS *et al.* (2011) Differential effects of dietary protein sources on postprandial low-grade inflammation after a single high fat meal in obese non-diabetic subjects. *Nutr J* **10**, 115.
138. Sun X & Zemel MB (2007) Calcium and 1,25-dihydroxyvitamin D3 regulation of adipokine expression. *Obesity (Silver Spring)* **15**, 340–348.
139. Kalupahana NS & Moustaid-Moussa N (2012) The renin-angiotensin system: a link between obesity, inflammation and insulin resistance. *Obes Rev* **13**, 136–149.