Effects of a diet rich in sesame (Sesamum indicum) pericarp on the expression of oestrogen receptor α and oestrogen receptor β in rat prostate and uterus

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The expression of oestrogen receptors (ER α and ER β) in the prostate and uterus tissues of Wistar rats supplied for 8 weeks with a diet rich in sesame (Sesamum indicum) pericarp (30%) was monitored. Eight male rats, aged 6 weeks, were divided into a control group fed on a normal diet, and an experimental one, provided with the normal diet enriched with 30% sesame pericarp. A similar experiment was performed with female rats. At the end of the experiment, the prostate and uterus tissues were surgically removed and kept at -80° C for up to 2 months. Western blotting and quantitative real-time PCR (qRT-PCR) methods were used in order to investigate the levels of receptor proteins and mRNA. Significant increase in the expression of ER β in prostate and uterus was evident in both methods, while the magnitude of the observed alteration depended on the applied method. No statistically significant change was observed in the expression of ER α in uterus. In prostate, although the increase was more evident when investigated by means of qRT-PCR, the difference in expression of ER α was not statistically significant. In both tissues, a shift of the ratio of ER α :ER β in favour of ER β was evident, indicating, according to existing literature, a beneficial effect of the diet provided upon the health status of the organisms. It is suggested that this effect is attributed to the lignans present in the pericarp which exert phyto-oestrogenic activity.

Sesame: Sesame pericarp: Phyto-oestrogens: Oestrogen receptor α : Oestrogen receptor β

According to recent studies, several major complications could occur during and after long-term hormone replacement therapy, including the increased incidence of venous thromboembolism and breast cancer⁽¹⁾. Phyto-oestrogens, such as lignans, may constitute an alternative to hormone replacement therapy.

Sesame (Sesamum indicum, Pedaliaceae) is one of the most important oilseed crops with worldwide production reaching about 3.3 Mt annually and mainly cultivated in developing countries⁽²⁾. The interest in sesame oil arises from its high content of unsaturated fatty acids and antioxidant lignans, exemplified by sesamin, sesamolin and sesaminol, that are also classified as phyto-oestrogens (3). Usually, for the preparation of the various sesame products, the pericarp (hull) is removed. However, sesame oil from seeds with pericarp was more stable than that extracted from dehulled seeds⁽⁴⁾. This observation indicates that antioxidative components exist in the sesame perisperm, also confirmed by Chang et al. (5). Chemical analysis of the obtained extracts led to the identification of more than sixteen lignans⁽⁶⁾, while a polyphenol-rich extract, appearing to be a by-product of great importance due to its antioxidant and anti-mutational properties, has been obtained from the sesame perisperm⁽⁷⁾.

Oestrogens have diverse effects on many tissues in both males and females and the majority of these effects are mediated by oestrogen receptors that are ligand-dependent transcriptional activators belonging to the superfamily of nuclear receptors (8,9). The two subtypes of oestrogen receptors, oestrogen receptor α (ER α) and oestrogen receptor β (ERβ), contain certain structurally conserved functional domains such as the regions responsible for DNA binding, dimerisation, ligand binding and ligand-dependent transactivation of gene expression. The tissue distribution of the two subtypes is not completely coincident as some tissues (for example, uterus, vagina) express predominantly $ER\alpha$, whereas others (for example, lung, prostate, ovarian granulosa cells) express predominantly ERB. Other tissues, such as bone and the pituitary, express both ER α and ER $\beta^{(9)}$. The relative levels of ER α and ER β are important determinants of cellular sensitivity to oestrogens. Although, at normal concentrations of oestradiol, ERa is the strongest transcriptional activator of the two oestrogen receptor isoforms, co-expression of ERβ results in suppression of both the efficacy and the potency of hormone-stimulated responses $^{(10)}$.

Phyto-oestrogens are plant-derived compounds that promote oestrogenic actions mediated through the oestrogen receptors

ERα and ERβ in mammals⁽¹¹⁾. They are structurally similar to mammalian oestrogen 17β-oestradiol and are, therefore, considered to play an important role in the prevention of hormone-dependent diseases such as cancer, heart disease, menopausal symptoms and osteoporosis⁽¹²⁾. Due to interaction with oestrogen receptors, these compounds also have the potential to disrupt oestrogenic signalling, acting either oestrogenically as oestrogen agonists or anti-oestrogenically as antagonists and can also, therefore, be classified as selective oestrogen receptor modulators⁽¹³⁾.

According to our previous report⁽¹⁴⁾, the levels of mammalian lignans (enterodiol and enterolactone) increased by several-fold in the plasma of experimental animals provided for a time period of 8 weeks with a diet rich in sesame pericarp (30%), among other experimental diets. In the present study we attempted to further investigate the effect of this diet upon the expression of ER α and ER β in certain tissues of the experimental animals.

Materials and methods

Animals

All animal procedures were performed in accordance with the guidelines of the Greek Local Authority and Municipality of the Central Region of Macedonia.

Animals and diets

Special care was taken in order to eliminate any unwanted effects due to the provided diet. Commercial rat chow was used (type 510K; Hellenic Feed Company, Plati, Greece) containing 21% total protein from beetroots, 6·2% lipids, 4·5% fibres, 7·5% ash, 1·1% Ca, 0·9% P, 0·35% Na and 1·1% methionine. The pellets were grounded and one part (experimental diet) was mixed with the sesame pericarp (Haitoglou SA, Thessaloniki, Greece) at a final concentration of 30% (w/w), while the rest (control diet) was kept as it was. Subsequently, in both parts, the ground material was mixed with a minimum amount of tap water and by the use of a meat mincing machine, new pellets were created that were dried overnight at 45°C in a well-ventilated incubator.

Male Wistar rats were obtained from the Medical School of Aristotle University of Thessaloniki. The animal experimentation was conducted according to the Regional Veterinary Directorate (licence no. 13/9749/12-07-2006).

Animals, aged 6 weeks and of about 170 g body weight, were housed under controlled conditions of temperature (20–25°C) and lighting (12h light-dark cycle). Animals were fed on a commercial non-purified diet for 1 week after which they were fed on either the control diet or the experimental diet. At the end of the experiment, animals were fasted for 12h and killed after being anaesthetised using diethyl ether.

RNA extraction and cDNA synthesis

Total RNA was extracted from frozen tissues according to the method described by Chomczynski & Sacchi⁽¹⁵⁾. RNA was quantified spectrophotometrically at 260 nm and its integrity was verified also spectrophotometrically by measuring the

ratio of 260/280 nm and by electrophoresis in 1.5 % (w/v) agarose gel. A quantity of 1 µg total RNA was used to prepare cDNA. Synthesis of cDNA was performed with the iScript Select cDNA Synthesis Kit (170-8896; Bio-Rad Laboratories, Hercules, CA, USA) in a final volume of 20 µl. For reverse transcription, mixtures of RNA samples (1 µg total RNA), random primer and nuclease-free water (total volume 15 µl) were incubated at 65°C for 5 min and were then cooled on ice immediately. $5 \times$ iScript select reaction mix $(4 \mu l)$ and iScript reverse transcriptase (1 µl) were added to each reaction according to the manufacturer's instructions. The mixture was further incubated for 5 min at 25°C, following a 30 min incubation at 42°C and a final 5 min incubation at 85°C in order to heat-inactivate the reverse transcriptase. The finally derived cDNA products were stored at -20° C until further analysed by means of real-time PCR.

Quantitative real-time polymerase chain reaction

To verify changes in gene expression in the uterus and prostate tissues of the experimental animals due to the administration of sesame pericarp, quantitative real-time PCR (qRT-PCR) was performed using the iCycler iQ Real-Time PCR detection system (Bio-Rad Laboratories).

Oligonucleotide primer pairs for $ER\alpha$ were selected in accordance with previous references⁽¹⁶⁾. Primers for glyceral-dehyde 3-phosphate dehydrogenase (GAPDH) were designed from the published sequence of the rat GAPDH gene (GenBank reference AF106860). Primer pairs for $ER\beta$ were designed on the basis of the common sequence of seven isoforms according to the published sequence of the rat $ER\beta$ gene (ensembl.org: ENSRNOG00000005343). The primer pairs used were as follows:

ER α sense, 5-AAT TCT GAC AAT CGA CGC CAG-3; ER α antisense, 5-GTG CTT CAA CAT TCT CCC CCT C-3 with PCR product size (bp) 345.

ER β sense, 5-GCT CCT CTA TGC AGA ACC TCA AA-3; ER β antisense, 5-CAG AAG TGA GCA TCC CTC TTT G-3 with PCR product size (bp) 137.

GAPDH sense, 5-GCC ACTC AGA AGA CTG TGG A-3; GAPDH antisense, 5-AGG TGG AAG AAT GGG AGT TG-3 with PCR product size (bp) 337.

The position of the primers on cDNA is given below:

ER α sense, 681; ER α antisense, 1003.

ERβ 1, 2, 3, 6, 7 sense, 288; ERβ 1, 2, 3, 6, 7 antisense, 425; ERβ 4 sense, 568; ERβ 4 antisense, 705; ERβ 5 sense, 388; ERβ 5 antisense, 525.

These sets of primers were designed to span over intronic DNA sequences.

The synthesised cDNA were further amplified (1:10) in triplicate by PCR using SYBR green I as a fluorescent dye (SYBR Green Supermix, 170-8880; Bio-Rad Laboratories) in a final volume of $20 \,\mu l$.

The PCR cycling conditions were as follows: initial denaturation and enzyme activation at 95°C for 3 min, followed by thirty-five cycles of denaturation at 95°C for 30 s, annealing for 30 s, and extension at 72°C for 40 s. The specific annealing temperature of 60°C was the same for each target. Product purity was confirmed by dissociation curve (melting curve)

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followed by agarose gel electrophoresis. No template controls, as well as negative PCR controls without cDNA, were included in all assays, yielding no consistent amplification.

Calculation of relative expression levels of each target was conducted based on the cycle threshold (C_T) method according to the equation $2^{-\Delta\Delta CT(17)}$. Normalisation of the cDNA input was performed by using GAPDH gene expression as the internal control.

Gel electrophoresis and Western blot analysis

Western blot analysis was performed on uterus and prostate tissue samples. Tissues were removed from the animals and immediately immersed in liquid N2. The frozen tissues were pulverised in a mortar and homogenised further on ice in lysis buffer consisting of 50 mm-2-amino-2-hydroxymethylpropane-1,3-diol-HCl (pH 7.4), 1% (v/v) Nonidet P-40, 0.25 % (w/v) deoxycholic acid, 150 mm-NaCl, 1 mm-Na₃VO₄, 1 mm-EDTA, 1 mm-NaF and protease inhibitor cocktail from Sigma (P2714). The samples were kept on ice for 30 min and centrifuged further at 10 000 g for 10 min at 4°C to obtain the supernatant fraction. The protein concentration of samples was estimated by means of the Quick Start Bradford dye reagent (500-0205; Bio-Rad Laboratories). The samples were electrophoresed on 10 % SDS-PAGE. Prestained precision protein standards (161-0305; Bio-Rad Laboratories) were used as molecular weight markers.

After electrophoresis, the proteins were transferred onto a nitrocellulose membrane. Blots were probed with rabbit polyclonal antibody against ER α and against ER β . The anti-ER α antibody (ab34575) was purchased from Abcam (Cambridge, MA, USA) and raised by use of an immunising peptide corresponding to amino acid residues 21-32 from human ERα which is completely conserved between human, rat, rabbit, sheep, porcine and bovine species and cross-reacts with human and rat protein. The anti-ERβ antibody (ab786) was purchased from Abcam and it was raised in rabbit against a synthetic immunising peptide corresponding to amino acids 459-477 of human ERβ, and it cross-reacts with human, rat and baboon and possibly other species. As a secondary antibody goat anti-rabbit IgG conjugated with horseradish peroxidase was used (Goat Anti-Rabbit, 170-5046; Bio-Rad Laboratories), and was visualised by means of enhanced chemiluminescence (catalogue no. 2600; Chemicon International, Temecula, CA, USA). Signals were quantified by image analysis (Gel-Pro version 3.0; Media Cybernetics, Silver Spring, MD, USA). Protein loads were normalised by using the actin signal as the internal control.

Statistical analysis

For statistical analysis of the results the Student's t test and ANOVA were applied.

Results

Immunoblotting studies were performed with total prostate and uterus protein from male and female rats provided daily with a sesame pericarp-enriched diet (experimental diet). After targeting the trans-blotted proteins with a specific antibody and visualising it by means of the enhanced chemiluminescence method, immuno-reactive full-length $ER\alpha$ and $ER\beta$ proteins of approximately $64-66\,kDa$ and $54-56\,kDa$ respectively were clearly observed in all the tissues studied except in the case of prostate, where $ER\alpha$ could not be detected under the described conditions.

The administration of sesame pericarp for a time period of 8 weeks resulted in a significant increase in the expression of ER β in prostate. It was evident that the surface area corresponding to prostate ER β protein from the treated animals was more dense and broad, as shown in Fig. 1. The histogram shows the mean values of the area corresponding to ER β for all the four experimental animals of each group quantified by image analysis (Gel-Pro version 3.0; Media Cybernetics). The mean value for the control group was set as 100 %. The difference among the mean values corresponding to each group was statistically significant as judged by Student's t test and ANOVA.

As far as the uterus is concerned, the effect of the provided experimental diet upon the expression of ER β in the experimental animals studied was similar (Fig. 2). The difference between the mean values corresponding to the visualised area for each group was statistically significant (P<0.01). The area corresponding to the ER β zone in the sesame pericarp-treated animals was increased by two-fold (Fig. 2).

Concerning the expression of ER α in prostate tissue homogenate, no specific protein was detected with this method while in the uterus, the observed changes were not statistically significant (P<0·1; Fig. 3) in spite of the fact that the visualised surface area corresponding to this receptor was broader in some of the treated animals.

Oestrogen receptor α and oestrogen receptor β mRNA in prostate and uterus of treated animals

By means of qRT-PCR, the level of transcription of the two receptors, ER α and ER β , was monitored.

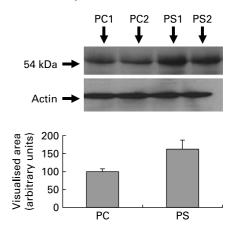


Fig. 1. Expression of oestrogen receptor β (ERβ) protein in the prostate of two control animals (PC1 and PC2; fed with the control diet) and two experimental animals (PS1 and PS2; fed with the experimental diet) as investigated by means of Western blotting. Actin was used as an internal control. In both cases, $100\,\mu g$ of total prostate protein was used. In the histogram is shown the visualised area (quantified by image analysis; Gel-Pro version 3.0; Media Cybernetics, Silver Spring, MD, USA) of all the animals from the control group (PC) and the sesame ($Sesamum\ indicum$) pericarp-fed group (PS). The area corresponding to the protein of ERβ in the prostate of the control group animals was taken as $100\,\%$. Values are means, with standard deviations represented by vertical bars.

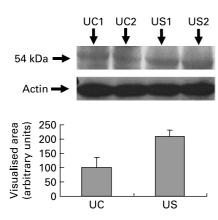


Fig. 2. Expression of oestrogen receptor β (ERβ) protein in the uterus of two control animals (UC1 and UC2; fed with the control diet) and two experimental animals (US1 and US2; fed with the experimental diet) as investigated by means of Western blotting. Actin was used as an internal control. In both cases, $100\,\mu g$ of total uterus protein was used. In the histogram is shown the visualised area (quantified by image analysis; Gel-Pro version 3.0; Media Cybernetics, Silver Spring, MD, USA) of all the animals from the control group (UC) and the sesame (*Sesamum indicum*) pericarp-fed group (US). The area corresponding to the protein of ERβ in the uterus of the control group animals was taken as $100\,\%$. Values are means, with standard deviations represented by vertical bars.

mRNA in prostate. cDNA was synthesised from samples of eight prostate tissues by means of random primers and subjected to qRT-PCR amplifying ER α , ER β and GAPDH. As it is mentioned in the Materials and methods section, the primers were designed to detect either of the two receptors. Four of the samples were derived from control animals (aged 14 weeks) provided with the control diet, and four were from experimental animals (aged 14 weeks) provided with the experimental diet. The qRT-PCR reactions were set up for thirty-five cycles.

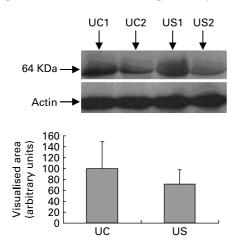


Fig. 3. Expression of oestrogen receptor α (ER α) protein in the uterus of two control animals (UC1 and UC2; fed with the control diet) and two experimental animals (US1 and US2; fed with the experimental diet) as investigated by means of Western blotting. Actin was used as an internal control. In both cases 100 μg of total uterus protein was used. In the histogram is shown the visualised area (quantified by image analysis; Gel-Pro version 3.0; Media Cybernetics, Silver Spring, MD, USA) of all the animals from the control group (UC) and the sesame (*Sesamum indicum*) pericarp-fed group (US). The area corresponding to the protein of ER α in the uterus of the control group animals was taken as 100%. Although the surface area corresponding to ER α protein in the uterus of the treated animals appears to be lower, the difference was not statistically significant (P<0.5). Values are means, with standard deviations represented by vertical bars.

As far as the ER α mRNA levels are concerned, although an increase by almost two-fold was observed, the difference was not statistically significant in prostate tissue (P<0·1; Fig. 4). However, a very dramatic increase in prostate tissue by more than four-fold of the levels of ER β mRNA was revealed after sesame pericarp administration (P<0·001; Fig. 4).

mRNA in uterus. The levels of mRNA were investigated in uterus tissue by following the same procedure and applying the same set of primers. In this course of experiments also, four of the samples were derived from control animals (aged 14 weeks) provided with the control diet, and four were from experimental animals (aged 14 weeks) provided with the experimental diet. The qRT-PCR reactions were set up for thirty-five cycles.

According to the results, both ER α and ER β mRNA are expressed in the uterus. Administration of sesame pericarp for 8 weeks caused induction of ER β mRNA by more than two-fold in uterus tissue (P < 0.005) while the levels of ER α mRNA were not statistically altered (P < 0.1) (Fig. 4).

Discussion

Despite the suggestion that the oestrogenic and/or anti-oestrogenic activity of phyto-oestrogens may reduce but also stimulate oestrogen-dependent tumour growth depending on dose and time of exposure⁽¹⁸⁾, reports providing information on the effects of a diet rich in plant products containing lignans upon the expression of oestrogen receptors in the organism are very rare. In the present paper we report on the effects of a diet rich in sesame pericarp that contains significant

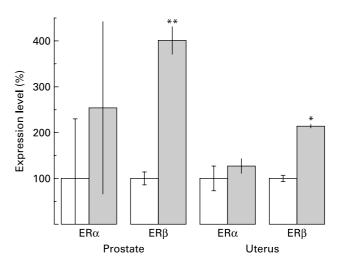


Fig. 4. Effect of sesame (*Sesamum indicum*) pericarp diet on the expression of oestrogen receptor α (ER α) and oestrogen receptor β (ER β) mRNA in the prostate and uterus as investigated by means of quantitative real-time PCR. Calculation of relative expression levels of each target was conducted based on the cycle threshold (C_T) method according to the equation $2^{-\Delta\Delta CT}$. Each one of the histograms represents the variation in expression levels of oestrogen receptors observed in the prostate and uterus of treated (\square) as related to untreated (\square) animals, that were set to 100%. The untreated animals were provided with rat chow while the experimental ones were provided with rat chow enriched with 30% sesame pericarp. Values are means, with standard deviations represented by vertical bars. Mean value was significantly different from that for the control animals: *P <0.005, *T <0.001. The observed difference between control and experimental animals was not statistically significant for ER α in the prostate and uterus tissues (P<0.1).

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amounts of lignans⁽⁶⁾ with phyto-oestrogenic activity upon the expression of ER α and ER $\beta^{(11,19)}$ in the prostate and uterus which express predominantly different classes of oestrogen receptors (ER β for prostate, ER α for uterus)^(20,21).

According to our previous report, the levels of enterolignans (i.e. enterolactone and enterodiole, which are the end metabolites of lignans in mammals and express strong phyto-oestrogenic activity) were increased dramatically in the plasma of Wistar rats receiving for 8 weeks the same, as in the present study, experimental diet. More specifically, the levels of enterodiole from zero reached 2663.4 mmol/l while those of enterolactone increased from 68.5 mmol/l to 2133-9 mmol/l⁽¹⁴⁾.

In the present study, sesame pericarp consumption led to a significant increase in the expression of ERB in the prostate. The increase was more than two-fold as judged by means of Western blotting and even higher, by four-fold, when the levels of mRNA were measured by means of qRT-PCR.

In the uterus tissue, expression of ERB was also elevated, although the observed difference was less prominent than in prostate (increase by two-fold), a finding that may be attributed to the variation in the natural levels of 17β-oestradiol due to the oestrus cycle of the experimental animals that were used.

No statistically significant difference was observed in the expressed levels of ER α in either tissue. The rather high differences observed in the band density of ERa protein from the uterus of animals deriving from the same experimental group are also attributed to the variation of the natural levels of 17β-oestradiol present in each animal due to the oestrus cycle. On the other hand, the very high variation of mRNA for ERa observed in prostate tissue is attributed to the very low levels that are naturally present in this tissue.

These findings are in agreement with previous reports that ERα and ERβ display marked differences in binding affinity and sensitivity to certain ligands and that phyto-oestrogens preferentially bind and activate $ER\beta^{(22)}$.

According to accumulated knowledge, the observed alterations in the expression of oestrogen receptors after administering an experimental diet are beneficial to the organism. Increased expression of ERB is associated with increased likelihood of response to endocrine therapy, while in tumour tissues v. normal ones in many cancers, including breast, ovary, colon and prostate, expression of ERB mRNA and ER β protein is decreased⁽²³⁾. Co-expression of ER α and ERB results also in the formation of receptor heterodimers (instead of homodimers) modifying, therefore, the ER α - and ERβ-mediated responses⁽²³⁾. Emerging data support different functions between ER β when it is expressed alone and ER β when it is co-expressed with ER $\alpha^{(24)}$. ER β seems to play a key role in the mitogenic action of oestrogen by providing protection against ERα-induced proliferation^(25,26)

The imbalance in ERa:ERB expression observed in oestrogen-dependent cancer opens up a new field in the hormone therapy of cancer. Targeted ERB therapies, including the development of Erß-specific ligands, may constitute a new therapeutic approach particularly for pre-invasive or proliferative lesions (27). According to the present results, it becomes evident that consumption of sesame pericarp that is rich in lignans and phenols⁽⁷⁾ is combined with increased levels of enterolignans in the plasma⁽¹⁴⁾ that induce the expression of ERB and alter the balance between the two types of oestrogen receptors. The balance of $ER\alpha$: $ER\beta$ is very important to the sensitivity of the cell to hormonal action and by shifting it to the benefit of ERB may help either to prevent cancer development on even to decrease its rate of development. The clinical value of ERB in cancer prognosis and its possible usefulness for prediction of the hormone response should be assessed in large-scale and prospective clinical studies. In general, these results offer additional information supporting the concept that consumption of vegetables and legumes rich in lignans and phenols is beneficial for the maintenance of good health.

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The authors state that there are not any conflicts of interest.

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