

Distribution of Calcium and Phosphorus in Leaves of the Proteaceae

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The overall aim of our research program is to discover the physiological basis of calcium toxicity in Proteaceae plant species that typically inhabit low-phosphorus, acidic soils and avoid calcareous (alkaline) soils. The phenomenon of Ca toxicity has been known for decades, but so far no clear mechanistic explanation is available. We propose that Ca toxicity in P-efficient Proteaceae is the result of a shift in cell type where P accumulates, from epidermal cells as seen in dicots generally to mesophyll cells in P-efficient Proteaceae species [1]. Conversely, we hypothesise that Ca toxicity is avoided in the few Proteaceae species that inhabit calcareous soils by shifting the accumulation of Ca from mesophyll cells, where dicots generally accumulate Ca, to epidermal cells so as to prevent interference with efficient P utilisation.

Leaves of several *Banksia* spp. growing in different soil environments in both Australia and Chile were dissected and rapid frozen in liquid nitrogen, before being subsequently prepared for cellular elemental analysis and mapping by EDS X-ray microanalysis. Frozen samples were either freeze-substituted in an anhydrous ether:acrolein mixture, resin embedded [2] and microplaned to produce transverse sections of leaf; or simply microplaned transversely while frozen and analysed in the fully frozen-hydrated state [3].

Samples were analysed at CMCA in a Zeiss Supra 55 field emission SEM fitted with a Leica cryostage and an Oxford X-Max80 SDD X-ray detector (80 mm²) interfaced to Oxford Instruments AZtecEnergy software. The microscope was operated at 15kV in high current mode. Immediately prior to each map acquisition, the instrument was calibrated and the beam current measured and recorded using a pure copper standard. Standards included polished mineral standards and microplaned frozen solutions of aqueous salts at various concentrations (e.g. CaCl₂). Elemental maps were acquired at a resolution of 1024 × 768 pixels, for > 400 frames with a dwell time of 50–100 μs per pixel. Drift correction and pulse-pile up correction were activated. Using the Oxford Instruments AZtecEnergy software, quantitative numerical data were subsequently extracted from regions of interest drawn on the element maps, with individual spectra from each pixel summed and processed to yield concentration data. Summed spectra from regions of interest were quantified using the AZtec XPP model for matrix corrections.

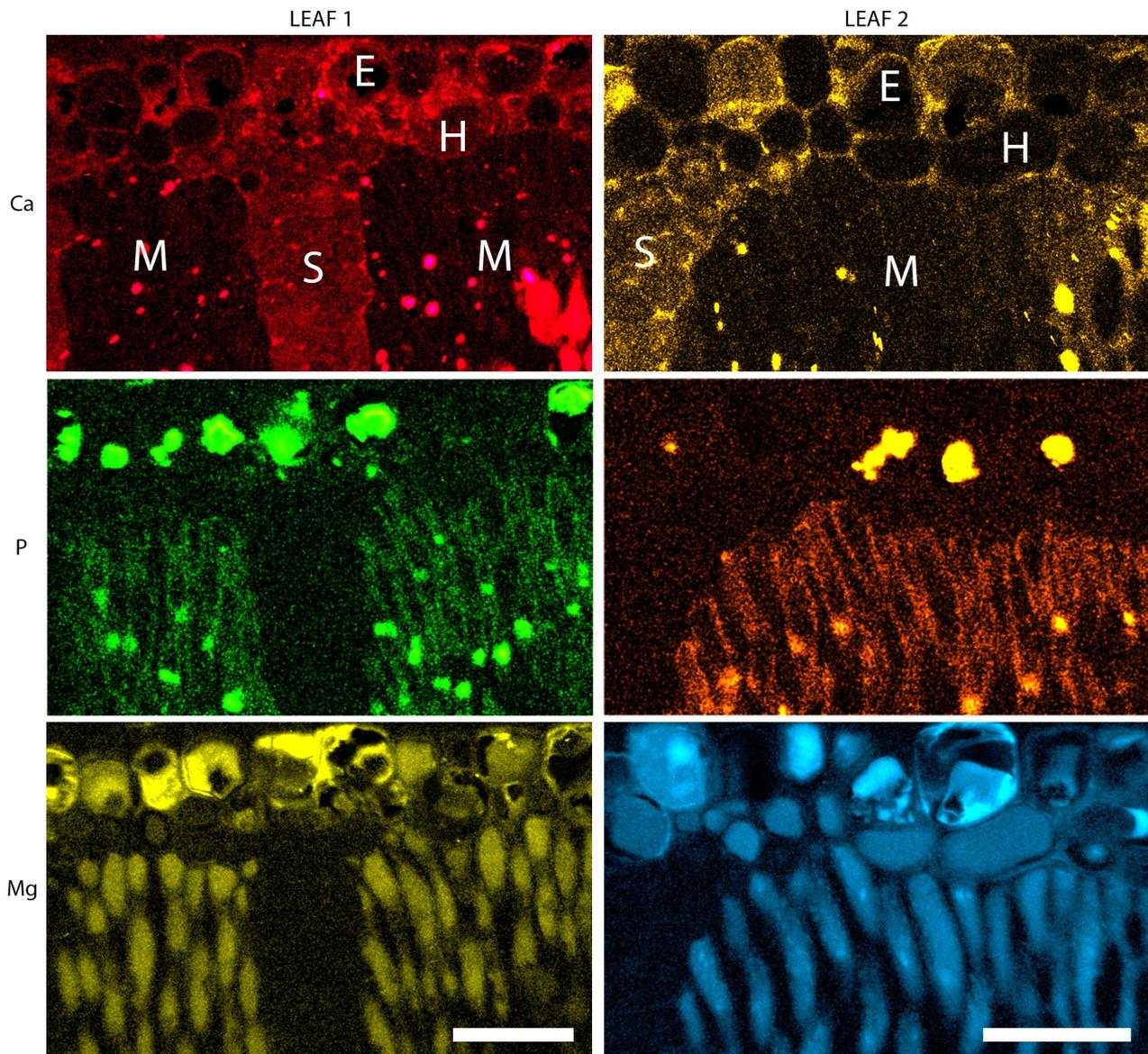
Elemental maps of leaf transverse sections are revealing the distribution and concentration of key elements in individual cell types, including the epidermis, hypodermis, sclerenchyma, palisade mesophyll and bundle sheath cells (Figure 1). Preliminary data suggest that P is concentrated within the mesophyll cells and Ca within the epidermal cells, as originally hypothesised [4].

References:

- [1] Shane, M. W. Tissue and cellular phosphorus storage during development of phosphorus toxicity in *Hakea prostrata* (Proteaceae). *Journal of Experimental Botany* 55, 1033–1044 (2004).
- [2] Marshall, A. Freeze-substitution as a preparation technique for biological X-ray microanalysis. *Scanning Electron Microscopy* 395–408 (1980).

[3] Marshall, A. T. & Clode, P. L. X-ray microanalysis of Rb^+ entry into cricket Malpighian tubule cells via putative K^+ channels. *Journal of Experimental Biology* 212, 2977–2982 (2009).

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Qualitative element maps of freeze-substituted *Banksia attenuata* leaves. E = epidermis; H = hypodermis; S = sclerenchyma; M = palisade mesophyll. Scale bars = 50 μ m.