

Effect of inulin on the human gut microbiota: stimulation of *Bifidobacterium adolescentis* and *Faecalibacterium prausnitzii*

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Prebiotics are food ingredients that improve health by modulating the colonic microbiota. The bifidogenic effect of the prebiotic inulin is well established; however, it remains unclear which species of *Bifidobacterium* are stimulated *in vivo* and whether bacterial groups other than lactic acid bacteria are affected by inulin consumption. Changes in the faecal microbiota composition were examined by real-time PCR in twelve human volunteers after ingestion of inulin (10 g/d) for a 16-d period in comparison with a control period without any supplement intake. The prevalence of most bacterial groups examined did not change after inulin intake, although the low G + C % Gram-positive species *Faecalibacterium prausnitzii* exhibited a significant increase (10.3 % for control period v. 14.5 % during inulin intake, $P=0.019$). The composition of the genus *Bifidobacterium* was studied in four of the volunteers by clone library analysis. Between three and five *Bifidobacterium* spp. were found in each volunteer. *Bifidobacterium adolescentis* and *Bifidobacterium longum* were present in all volunteers, and *Bifidobacterium pseudocatenulatum*, *Bifidobacterium animalis*, *Bifidobacterium bifidum* and *Bifidobacterium dentium* were also detected. Real-time PCR was employed to quantify the four most prevalent *Bifidobacterium* spp., *B. adolescentis*, *B. longum*, *B. pseudocatenulatum* and *B. bifidum*, in ten volunteers carrying detectable levels of bifidobacteria. *B. adolescentis* showed the strongest response to inulin consumption, increasing from 0.89 to 3.9 % of the total microbiota ($P=0.001$). *B. bifidum* was increased from 0.22 to 0.63 % ($P<0.001$) for the five volunteers for whom this species was present.

Prebiotic: Inulin: Microbiota: *Bifidobacterium*

The microbial community resident in the human colon is a highly complex consortium of many different bacterial species. The application of molecular tools targeting the 16S rRNA gene has revealed that the numerically dominant groups are low G + C % Gram-positive bacteria and Gram-negative *Bacteroidetes*; however, many phylotypes remain uncultured and consequently poorly characterised⁽¹⁾. The microbiota plays an important role in host health through various mechanisms, such as protection against pathogenic bacteria and provision of nutrients, and its composition can be modulated by dietary means, which can lead to either more health-promoting or more detrimental consequences for the host⁽¹⁾. One way of modulating the gut microbiota is through the consumption of prebiotics, non-digestible food ingredients that pass through the upper gut and are selectively fermented by colonic bacteria. This leads to specific changes in the composition and/or activity of the gut microbiota that confers benefits upon host well-being and health^(2,3). The dietary fructan inulin and its breakdown product fructo-oligosaccharide are particularly well-studied prebiotics, and evidence supporting their health-promoting effects, mostly in animal models, is accumulating rapidly^(3,4). A direct consequence

of inulin ingestion is stimulation of lactic acid bacteria (lactobacilli and bifidobacteria) within the gut microbiota⁽²⁾. While the bifidogenic effect of inulin is well demonstrated, it is less clear whether this is a characteristic of the whole genus, or whether certain *Bifidobacterium* spp. are selectively stimulated. Several *Bifidobacterium* spp. are commonly found in the adult human colon^(5–8). The degradation of fructo-oligosaccharides seems to be widespread among bifidobacterial strains in pure culture, while fewer strains are able to utilise inulin⁽⁹⁾. *In vivo*, however, cross-feeding of fructo-oligosaccharides and fructose from primary inulin degraders might lead to a stimulation of other bifidobacteria⁽⁹⁾.

Possible effects of inulin on other members of the gut microbiota are less well studied; however, it is increasingly recognised that prebiotics are likely to lead to further microbial changes other than bifidogenesis⁽¹⁰⁾. For example, an increase in the concentration of butyrate (butyrogenic effect) has been found in animal models⁽²⁾; however, lactic acid bacteria do not produce butyrate as a fermentation product. This butyrogenic effect could be due to several mechanisms. Several butyrate-producing bacteria can utilise inulin *in vitro*^(11,12) and are therefore likely to be stimulated directly in the gut. Metabolic

Abbreviations: Bact, *Bacteroides* spp.; Clep, clostridial cluster IV; CoAT, butyryl-CoA CoA-transferase gene; Ehal, *Eubacterium hallii*; Erec, clostridial cluster XIVa; Fprau, *Faecalibacterium prausnitzii*; Rrec, *Roseburia/E. rectale* group; Rum, cluster IV *Ruminococcus* spp.

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cross-feeding of the inulin breakdown product fructose and the fermentation product lactate from bifidobacteria to butyrate producers has been demonstrated in co-culture experiments^(13,14), and conversion of lactate into butyrate and propionate has been shown in fermentor studies⁽¹⁵⁾. A decrease in pH due to the increased formation of fermentation acids could also lead to changes in the microbiota, as demonstrated *in vitro*⁽¹⁶⁾. It is likely that all these mechanisms contribute to the butyrogenic effect of inulin *in vivo*, and they may also influence each other, as was demonstrated for the interdependence of pH, lactate production and lactate utilisation *in vitro*⁽¹⁷⁾.

Despite the composition of the human gut microbiota being quite similar between healthy individuals on a broad scale, there are individual differences at a species and strain level⁽¹⁸⁾. Consequently, metabolic capabilities of the microbiota might vary between people, as has been shown for the conversion of lactate to either butyrate or propionate *in vitro*⁽¹⁵⁾. The level of stimulation of bifidobacteria after ingestion of inulin as determined in faecal samples also shows a big variation between volunteers^(2,19), as do the baseline levels of this bacterial group. Numbers vary by several logs between individuals and in some people they are undetectable^(19,20). Therefore, other bacterial groups might replace bifidobacteria as the main consumers of inulin in those individuals.

The aim of the present study was to investigate changes in the major groups of the human gut microbiota in response to the intake of 10 g/d inulin over a 16-d period to improve the understanding of the effects of inulin on the gut community as a whole. In addition, the composition of the genus *Bifidobacterium* was investigated to establish which species were stimulated by inulin *in vivo*.

Experimental methods

Study design

The present study was designed to assess the influence of cabbage-processing methods and prebiotic ingestion on glucosinolate breakdown in human volunteers and the full details, including volunteer designations (A–L), have been published previously⁽¹⁹⁾. Briefly, twelve healthy adult volunteers were randomly split into two groups and either consumed 5 g inulin–oligofructose (Beneo™; DKSH/Orafti Great Britain Ltd, Kent, UK) twice daily for 21 d (inulin period) or did not consume a supplement for 21 d (control period) in a balanced crossover study design. Faecal samples were taken at the outset of the present study and at day 16 of both periods. DNA was extracted from the faecal samples as described before⁽¹⁹⁾. The present study was approved by the NHS Trust Grampian Research Ethics Committee.

Faecal fermentation product and pH analysis

For fermentation product analysis, approximately 0.5 g of faeces were mixed with 1.5 ml of sterile demineralised water and centrifuged at 13 000 g for 10 min. One millilitre of the supernatant was analysed by GC following conversion to *t*-butyldimethylsilyl derivatives⁽²¹⁾. The lower limit of reliable detection of each product was taken as 0.2 mM. For the determination of faecal pH, approximately 0.5 g of faeces was centrifuged for 10 min at 17 000 g and the pH was measured in the supernatant with a microelectrode.

Quantitative real-time PCR

Primers for quantification of specific bacterial groups and the butyryl-CoA CoA-transferase gene are given in Table 1. Primers designed in the present study for *Roseburia* spp. and *Eubacterium hallii* were initially selected based on a sequence alignment of twenty-nine clostridial cluster XIVa (Erec) bacteria, two cluster IV and three cluster XVI strains. The specificity of the primers was checked with the Probe Match function of the Ribosome Database Project II⁽²²⁾. Primer and probe sequences taken from the literature were checked with Probe Match and sequence alignments and modified accordingly. Group-specific primer sets were validated by testing their amplification of either genomic DNA or the amplified 16S rRNA gene of a panel of fifteen cluster XIVa, seven cluster IV, one cluster XVI, three *Bacteroides* strains and *Bifidobacterium adolescentis* as shown in Table 2, using the real-time PCR conditions described in the next paragraph. Primers for clostridial cluster XIVa, *E. hallii* (Ehal), clostridial cluster IV (Clep), *Faecalibacterium prausnitzii* (Ffrau) and *Bacteroides* spp. (Bact) amplified all the strains in their target groups and none of the strains in other groups. Two different primer sets for *Roseburia* spp./*Eubacterium rectale* were used, as we could not identify any region of the 16S rRNA gene that was specific for all known species of the *Roseburia/E. rectale* cluster. Primer set Rrec1 targets the same species as fluorescent *in situ* hybridisation (FISH) probe Rrec584⁽²³⁾, and did not amplify *Roseburia inulinivorans* (Table 2), while primer set Rrec2 amplified all known *Roseburia* spp., *E. rectale* and *Butyrivibrio fibrisolvens* 16/4 and also led to low-level amplification of isolate A2-232 (Table 2). The primer set used for the amplification of cluster IV rumino-cocci (Rum) also amplified *Eubacterium siraeum* (Table 2). It should be noted that for primer sets Rrec2 and Rum, the group-specific primer was used together with a primer detecting a wider range of bacteria (i.e. clostridial cluster XIVa and IV, respectively); however, this did not seem to compromise the specificity or robustness of the assays. *Bifidobacterium* spp.-specific PCR conditions were validated with 16S rRNA gene amplicons of two strains of *Bifidobacterium adolescentis* (DSM 20083^T, L2-32⁽¹³⁾), two strains of *Bifidobacterium longum* (DSM 20219^T, DSM 20088), *Bifidobacterium pseudocatenulatum* DSM 20438^T, *Bifidobacterium bifidum* DSM 20456^T, *Bifidobacterium angulatum* DSM 20098^T and *Bifidobacterium breve* DSM 20213^T. For *B. adolescentis*, one of the primers used was specific for all bifidobacteria, as clone library analysis revealed that some *B. adolescentis* clones would not be recognised by species-specific primer Bi-ADO-1⁽⁵⁾.

Quantitative real-time PCR analysis of faecal DNA was performed as described previously⁽¹⁹⁾ with the following modifications: all samples and standards were examined in duplicate per PCR run. The annealing temperature was 60°C for all primer sets apart from Erec (55°C) and CoAT (53°C). For primer sets leading to amplicons over 150 bp, a 30-s extension step at 72°C was included. For CoA-transferase primers, data acquisition was performed at this step as described previously⁽²⁴⁾. For *B. bifidum*-specific primers, data acquisition was also performed at 72°C, as baseline fluorescence was high at 60°C presumably due to hairpin formation. Amplified 16S rRNA genes from the following bacteria served as standard templates: *Bacteroides thetaiotaomicron* DSM 2079^T for universal primers and *Bacteroides* spp. primers; *Roseburia*

Table 1. Primers used in the present study for real-time PCR analysis

Target group (abbreviation)	Primer name	Primer sequence	Amplicon*	Reference
All eubacteria	UniF	GTGSTGCAYGGYYGTCGTCA	147	Fuller <i>et al.</i> ⁽¹⁹⁾
	UniR	ACGTCRTCCMCNCCTTCCCTC		Fuller <i>et al.</i> ⁽¹⁹⁾
<i>Bacteroides</i> spp. (Bact)	Bac303F	GAAGGTCCCCACATTG	103	Bartosch <i>et al.</i> ⁽²⁷⁾
	Bfr-Fmrev	CGCKACTTGGCTGGTTCCAG		Liu <i>et al.</i> ⁽⁴⁶⁾ , modified
Clostridial cluster XIVa (Erec)	Erec482F	CGGTACCTGACTAAGAAGC	429	Rintillä <i>et al.</i> ⁽⁴⁷⁾
	Erec870R	AGTTTTYATTCTTGCGAACG		Rintillä <i>et al.</i> ⁽⁴⁷⁾
<i>Roseburia</i> spp. and <i>E. rectale</i> (Rrec1)	RrecF	GCGGTRCGGCAAGTCTGA	81	Walker <i>et al.</i> ⁽¹⁶⁾
	Rrec630mR	CCTCCGACACTCTAGTMCGAC		The present study
<i>Roseburia</i> spp. and <i>E. rectale</i> (Rrec2)	Rrec630F†	CGKACTAGAGTGTCCGGAGG	265	The present study
	RrecRi630F†	GTCATCTAGAGTGTCCGGAGG		The present study
	Erec870R‡	AGTTTTYATTCTTGCGAACG	278	Rintillä <i>et al.</i> ⁽⁴⁷⁾
<i>E. hallii</i> (Ehal)	EhalF	GCGTAGGTGGCAGTGCAA		Hold <i>et al.</i> ⁽⁴⁸⁾
	EhalR	GCACCGRAGCCTATACGG		The present study
Clostridial cluster IV (Clep)	Clep866mF	TTAACACAATAAGTWATCCACCTGG	314	Lay <i>et al.</i> ⁽⁴⁹⁾ , modified
	Clept1240mR	ACCTTCCTCCGTTTTGTCAAC		Sghir <i>et al.</i> ⁽⁵⁰⁾ , modified
<i>Faecalibacterium prausnitzii</i> (Fprau)	FPR-2F	GGAGGAAGAAGGTCTTCGG	248	Wang <i>et al.</i> ⁽⁵¹⁾
	Fprau645R	AATTCGCCTACCTCTGCACT		Suau <i>et al.</i> ⁽⁵²⁾ , modified
Cluster IV <i>Ruminococcus</i> spp. (Rum)	Rflbr730F	GGCGGCYTRCTGGGCTTT	157	Harmsen <i>et al.</i> ⁽⁵³⁾ , modified
	Clep866mR§	CCAGGTGGATWACTTATTGTGTTAA		Lay <i>et al.</i> ⁽⁴⁹⁾ , modified
Butyryl-CoA CoA-transferase gene (CoAT)	BCoATscrF	GCIGAICATTTACITGGAAYWSITGGCAYATG	557	Louis & Flint ⁽²⁴⁾
	BCoATscrR	CCTGCCTTTGCAATRTCIACRAANGC		Louis & Flint ⁽²⁴⁾
<i>Bifidobacterium adolescentis</i> (Bad)	Bif164F	GGGTGGTAATGCCGGATG	298	Bartosch <i>et al.</i> ⁽²⁷⁾
	BiADO-2	CGAAGGGCTTGCTCCCAGT		Matsuki <i>et al.</i> ⁽⁵⁾ , modified
<i>Bifidobacterium longum</i> (Blon)	BlonF	CAGTTGATCGCATGGTCTT	106	Malinen <i>et al.</i> ⁽⁵⁴⁾
	BlonR	TACCCGTGGAAGCCAC		Malinen <i>et al.</i> ⁽⁵⁴⁾
<i>Bifidobacterium catenulatum</i> group (Bcat)	BiCATg-1	CGGATGCTCCGACTCCT	285	Matsuki <i>et al.</i> ⁽⁵⁾
	BiCATg-2	CGAAGGCTTGCTCCCGAT		Matsuki <i>et al.</i> ⁽⁵⁾
<i>Bifidobacterium bifidum</i> (Bbif)	BiBIF-1	CCACATGATCGCATGTGATTG	278	Matsuki <i>et al.</i> ⁽⁵⁾
	BiBIF-2	CCGAAGGCTTGCTCCCAA		Matsuki <i>et al.</i> ⁽⁵⁾

*Product size (bp) is based on *Roseburia hominis* A2-183 for universal primers and on the bacterial strains used as standards for specific primers.

† Primers Rrec630F and RrecRi630F are used together at 250 nm each.

‡ Primer is specific for cluster XIVa.

§ Primer is specific for cluster IV.

|| Primer is specific for bifidobacteria.

Table 2. Validation of 16S rRNA primers for real-time PCR analysis of gut microbiota*

Bacterial strain ^(reference)	Erec	Rrec1	Rrec2	Ehal	Clep	Fprau	Rum	Bact
Clostridial cluster XIVa								
<i>Eubacterium rectale</i> A1-86 ⁽⁵⁵⁾	+	+	+	-				
<i>Roseburia faecis</i> M72/1 (DSM 16840) ^{T(12)}	+	+	+	-				
<i>Roseburia hominis</i> A2-183 (DSM 16839) ^{T(12)}	+	+	+	-	-	-	-	-
<i>Roseburia intestinalis</i> L1-82 (DSM 14610) ^{T(12,42)}	+	+	+	-				
<i>Roseburia inulinivorans</i> A2-194 (DSM 16841) ^{T(12)}	+	+	+	-				
<i>Butyrivibrio fibrisolvens</i> 16/4 ⁽⁵⁶⁾	+	-	+	-				
Isolate A2-232 ⁽⁵⁵⁾	+	-	-†	-				
<i>Anaerostipes caccae</i> L1-92 (DSM 14662) ^{T(55)}	+	-	-	-				
<i>Coprococcus</i> sp. L2-50 ⁽⁵⁵⁾	+	-	-	-				
<i>Eubacterium hallii</i> L2-7 (DSM 17630) ⁽⁵⁵⁾	+	-	-	+				
<i>Eubacterium hallii</i> SM6/1 ⁽⁵⁶⁾	+	-	-	+				
Isolate SSC/2 ⁽⁵⁶⁾	+	-	-	-				
Isolate GM2/1 ⁽⁵⁶⁾	+	-	-	-				
Isolate M62/1 ⁽⁵⁶⁾	+	-	-	-				
<i>Ruminococcus obeum</i> A2-162 ⁽⁵⁷⁾	+	-	-	-				
Clostridial cluster IV								
<i>Faecalibacterium prausnitzii</i> A2-165 (DSM 17677) ⁽¹¹⁾	-	-	-	-	+	+	-	-
<i>F. prausnitzii</i> L2-6 ⁽⁵⁵⁾					+	+	-	
<i>F. prausnitzii</i> M21/2 ⁽⁵⁶⁾					+	+	-	
<i>Eubacterium siraeum</i> 70/3 ⁽⁵⁸⁾					+	-	+	
<i>Ruminococcus albus</i> SY3 ⁽⁵⁹⁾					+	-	+	
<i>Ruminococcus bromii</i> L2-63 ⁽⁵⁷⁾					+	-	+	
<i>Ruminococcus flavefaciens</i> N17 ⁽²⁵⁾					+	-	+	
Others								
<i>Eubacterium cylindroides</i> T2-87 ^(56,57)	-	-	-	-	-	-	-	-
<i>Bacteroides ovatus</i> V975 ⁽⁶⁰⁾								+
<i>Bacteroides thetaiotaomicron</i> VPI-5482 (DSM 2079) ^{T(61)}	-	-	-	-	-	-	-	+
<i>Bacteroides vulgatus</i> (DSM 1447) ^{T(61)}								+
<i>Bifidobacterium adolescentis</i> (DSM 20083) ⁽⁶²⁾	-	-	-	-	-	-	-	-

Bact, *Bacteroides* spp.; Clep, clostridial cluster IV; Ehal, *Eubacterium hallii*; Erec, clostridial cluster XIVa; Fprau *Faecalibacterium prausnitzii*; Rrec, *Roseburia/ E. rectale* group; Rum, cluster IV *Ruminococcus* spp.

* +, Real-time PCR quantification using standard curves, as described by Fuller *et al.* (19) of 1.5 ng genomic DNA or 10⁷ 16S rRNA copies of the respective strain resulted in over 10⁵ or 10⁶ gene copies, respectively; -, real-time PCR quantification resulted in less than 10² or 10³ gene copies, respectively.

† Amplification approximately 100-fold lower than that for positive strains.

hominis A2-183 (DSM 16839)^T for universal, cluster XIVa and *Roseburia* spp. primers; *E. hallii* L2-7 (DSM 17630) for *E. hallii* primers; *F. prausnitzii* A2-165 (DSM 17677) for cluster IV and *F. prausnitzii* primers; *Ruminococcus flavefaciens* N17⁽²⁵⁾ for cluster IV *Ruminococcus* spp. primers and *B. adolescentis* DSM 20083^T, *B. longum* DSM 20219^T, *B. pseudocatenulatum* DSM 20438^T and *B. bifidum* DSM 20456^T for *Bifidobacterium* spp.-specific assays. The percentage of 16S rRNA gene copy number of a specific group or species relative to gene copy numbers obtained with the universal primers was determined using the efficiency-corrected Δ Ct method⁽²⁶⁾. At least three independent standard curves were used to calculate the PCR efficiency $E = 10^{(-1/\text{slope})}$. Quantities for bacterial groups were calculated from mean Ct values of two independent PCR runs using the following formula: $E^{-\text{Ct}}$. As the two independent PCR runs led to highly reproducible results (CV of 10% or less), *Bifidobacterium* spp.-specific quantities were determined from single assays. The normalised value was obtained by dividing specific group quantities through universal quantities, which were multiplied by 100 to obtain the percentage of specific gene of universal gene copies. For *E. hallii*, CoA-transferase primers and *Bifidobacterium* spp., 5 ng per reaction was amplified, and for all other primer sets, 0.5 ng per reaction was amplified. Values for *E. hallii*, CoA-transferase and *Bifidobacterium* spp. were corrected to account for the different dilution of template DNA.

Clone library construction and phylogenetic analysis

For the investigation of which bifidobacterial species were present, four volunteers, the two with the highest increase in *Bifidobacterium* spp. upon inulin consumption (volunteers I and L, both from group 2, treatment order control, inulin) and two volunteers with an intermediate increase (volunteers A and H, one from each group) were selected for clone library analysis. Faecal DNA was amplified with primers Bif164F⁽²⁷⁾ (GGGTGGTAATGCCGATG) and g-Bifid-R⁽²⁸⁾ (GGTGTTCTTCCCGATATCTACA) using BioTaq DNA polymerase (Bioline Ltd, London, UK). The amplification conditions were as follows: initial denaturation at 94°C for 2 min, followed by twenty cycles of denaturation (30 s, 94°C), annealing (30 s, 55°C) and extension (30 s, 72°C) and a final extension at 72°C for 10 min. PCR products were purified with a PCR clean-up kit (QIAquick; Qiagen Ltd, Crawley, UK or Wizard SV Gel and PCR Clean-Up System; Promega, Southampton, UK), cloned into vector pGEM[®]-T Easy (Promega) and transformed into XL1-Blue MRF['] Supercompetent cells (Stratagene, La Jolla, CA). Randomly chosen clones were amplified using vector primers and sequenced with primer g-Bifid-R on a capillary sequencer (CEQ8000 Genetic Analyser; Beckman Coulter, High Wycombe, UK).

Sequence analysis was performed on the in-house RRI/BioSS Beowulf cluster, running openMosix (<http://bioinformatics.rri.sari.ac.uk>). The sequences were base called with Phred^(29,30),

vector sequences trimmed off with Lucy⁽³¹⁾ and aligned with MUSCLE⁽³²⁾ and two iterations of Clustal W⁽³³⁾. Alignments were manually corrected using Jalview⁽³⁴⁾ and the sequences shorter than 400 nucleotides were deleted. A distance matrix was created with Phylip⁽³⁵⁾ and operational taxonomic units at 99% were obtained with Dotur⁽³⁶⁾. Any operational taxonomic units with less than four sequences were manually inspected for alignment errors and removed from the analysis if they were chimeras or of low sequence quality. The operational taxonomic units were assigned to different *Bifidobacterium* spp. by basic local alignment search tool (BLAST) analysis⁽³⁷⁾. The sequences have been deposited in GenBank under accession numbers EU421962–EU422224 and EU422226–EU422945.

Statistical analysis

The percentage of bacteria and SCFA concentrations from twelve volunteers and the percentage of *Bifidobacterium* spp. from ten volunteers were analysed with ANOVA, with volunteer as random effect and treatment (baseline, control, inulin), group (whether inulin was given before or after the control period) and their interaction as treatment effects. The effect of inulin was assessed by means of contrasts where the baseline and the control period were compared against the inulin period. Two of the *Bifidobacterium* spp. were detected in only five of the ten volunteers, and in addition to the parametric ANOVA described earlier, these strains were analysed by Friedman's non-parametric ANOVA. Results were similar for both approaches; so for consistency the ANOVA results are presented for these strains. For four volunteers, clone library data were obtained. The clone count per species was expressed as a percentage of the total clone count. When a strain was present in all four volunteers, these percentage data were analysed by ANOVA as described earlier, except that group effects were not assessed. Relationships between SCFA and bacteria were investigated using linear regression where a volunteer-specific intercept was allowed for. Data are presented as mean (SEM), where the latter is based on

variation between volunteers. All analyses were performed with GenStat 10th Edition Release 10.1 (VSN International, Hemel Hempstead, Hertfordshire, UK).

Results

Changes in microbiota composition, fermentation acids and pH upon inulin ingestion

It was shown previously for the samples analysed here that the proportion of bifidobacteria was significantly ($P < 0.001$) increased after inulin consumption (see Fuller *et al.*⁽¹⁹⁾ and Fig. 1); however, both the baseline abundance and the magnitude of the response to inulin were very different between individuals. We therefore investigated in the present study whether other bacterial groups were affected by ingestion of inulin. The real-time PCR primers for different phylogenetic groups of the faecal microbiota as well as primers against a gene involved in butyrate metabolism in the majority of known butyrate producers from the human gut⁽²⁴⁾ were used. Across all volunteers, a significant change upon inulin consumption was only found for *F. prausnitzii* ($P = 0.019$, Fig. 1); however, the response to inulin showed a tendency to be dependent on treatment order ($P = 0.057$). All the volunteers in group 2 (treatment order control, then inulin) showed an increase between the control and inulin periods, whereas in group 1, where inulin was given before the control period, *F. prausnitzii* had a tendency to remain high during the control period (data not shown).

Faecal SCFA and lactate were also determined, but there were no statistically significant effects of inulin (Fig. 2), except in the case of lactate, which was slightly increased during the inulin supplementation ($P = 0.041$). Similarly, the faecal pH did not differ significantly between treatment periods (baseline 6.96 (SEM 0.15), control 6.95 (SEM 0.13), inulin 6.91 (SEM 0.11)).

The percentage of *Roseburia/E. rectale* spp. showed a significant correlation with butyrate concentration ($P < 0.001$, linear regression, with 64% of the variance accounted), while the proportion of the *F. prausnitzii* group,

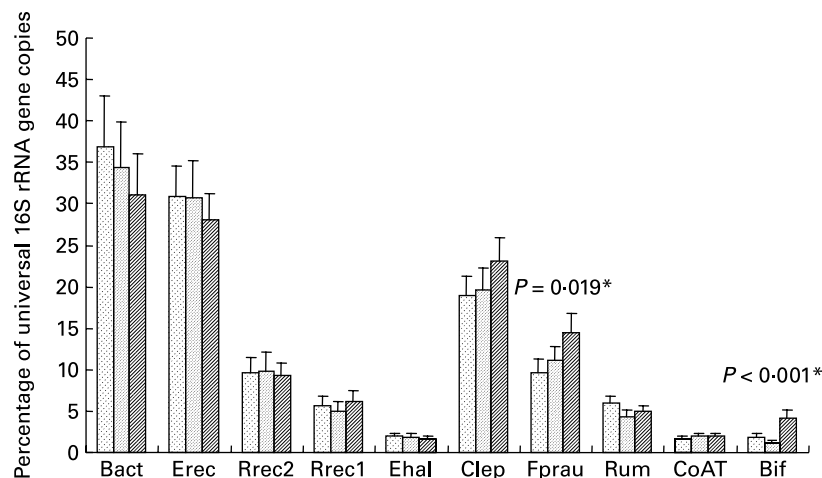


Fig. 1. Microbiota composition of faecal samples from twelve human volunteers before, (□; n 12) and after 16 d of control period (▤; n 12) or after 16 d of inulin supplementation (▨; n 11) based on real-time PCR. Details on bacterial groups detected are given in Tables 1 and 2. The data for *Bifidobacterium* spp. (Bif) were taken from Fuller *et al.*⁽¹⁹⁾. **P* values reflect the effect of inulin, obtained from comparing baseline and control periods against the inulin period using contrasts, from ANOVA with volunteer as random effect and treatment, treatment order and their interaction as fixed effects. Absence of *P* value means not significant ($P > 0.10$). Error bars reflect SEM (based on variation between volunteers).

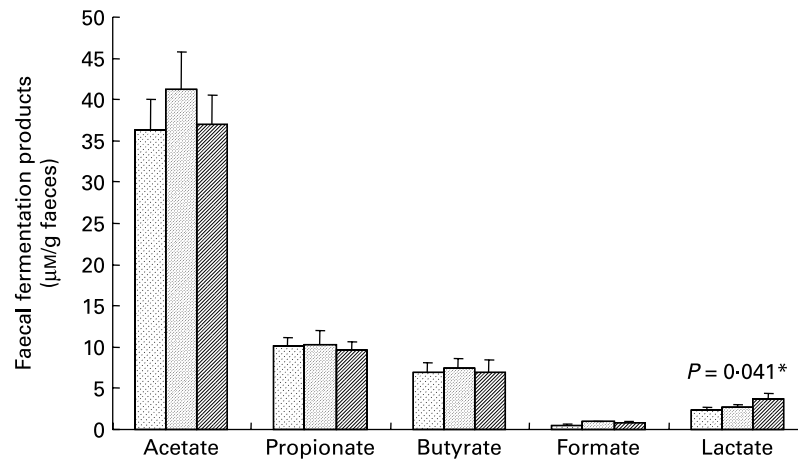


Fig. 2. Main bacterial fermentation products of faecal samples from twelve human volunteers before (□; *n* 12) and after 16 d of control period (▒; *n* 12) or after 16 d of inulin supplementation (▨; *n* 11). **P* value reflects the effect of inulin, obtained from comparing baseline and control periods against the inulin period using contrasts, from ANOVA with volunteer as random effect and treatment, treatment order and their interaction as fixed effects. Absence of *P* value means not significant (*P* > 0.10). Error bars reflect SEM (based on variation between volunteers).

which also produces butyrate, showed a trend towards a positive correlation with butyrate levels ($P=0.087$, linear regression, with 44 % of the variance accounted). The levels of the butyryl-CoA CoA-transferase gene significantly correlated with cluster XIVa bacteria (Erec, $P<0.001$, linear regression, with 70 % of the variance accounted; Rrec2, $P<0.001$, linear regression, with 59 % of the variance accounted), while no significant correlation was found for the *Roseburia/E. rectale* group Rrec1, *F. prausnitzii* or butyrate. Butyrate concentration, however, was negatively correlated with pH ($P=0.001$, linear regression with 60 % of the variance accounted).

Stimulation of *Bifidobacterium* spp. by inulin

Bifidobacterium spp. showed the strongest stimulation upon inulin ingestion in the present study. In order to identify the main species present, *Bifidobacterium*-specific clone library analysis was performed with faecal DNA of four donors. The effect of inulin on the bifidobacteria (as percentage of total bacteria) was significant in these four donors ($P=0.004^{(19)}$). This response was the strongest in donors I and L, with bifidobacteria in baseline and control samples forming less than 2 %, and in inulin samples over 8 % of all bacterial genes, while donors A and H showed a less dramatic inulin effect and had more variable levels of bifidobacteria between the baseline and control period (see Fuller *et al.* ⁽¹⁹⁾). Between sixty-four and ninety-one clones were analysed per sample. *B. adolescentis* and *B. longum* were detected in all four volunteers, *B. pseudocatenulatum* and *B. animalis* were present in three, *B. bifidum* in two and *B. dentium* in one. Each volunteer harboured between three and five different species. Of the strains that were detected in all four volunteers, the percentage of clones belonging to *B. adolescentis* was doubled during the inulin period (36 % at baseline and 27 % for the control period *v.* 66 % for the inulin treatment, $P=0.028$, based on ANOVA using contrasts). Clone numbers for *B. dentium* and *B. animalis* tended to be low (less than 10 % in all but one sample).

The most prevalent *Bifidobacterium* spp. based on the clone library results were quantified by the real-time PCR in all

volunteers apart from volunteers E and J, who carried very low or undetectable levels of bifidobacteria⁽¹⁹⁾.

All ten volunteers carried *B. longum* and nine of these also *B. adolescentis*. The *Bifidobacterium catenulatum* group was found in six and *B. bifidum* in five of the ten volunteers. For seven volunteers, the same species were found in all three samples (background, inulin and control). Ingestion of inulin led to a significant stimulation of *B. adolescentis* relative to the whole microbiota (Fig. 3) with six of the volunteers showing a clear increase (data not shown). *B. bifidum* also showed an increase (0.22–0.63 % ($P<0.001$) for the five volunteers for whom this species was present), while *B. longum* exhibited a tendency to increase on inulin (Fig. 3).

Discussion

The bifidogenic effect of the prebiotic inulin in man is well documented^(2,3). In the present study, we used the real-time PCR to analyse the effect of inulin ingestion on a wider range of human faecal bacteria. The human gut microbiota is dominated by low G+C % Gram-positive bacteria mainly belonging to clostridial clusters XIVa and IV⁽³⁸⁾ and Gram-negative bacteria related to *Bacteroidetes*, while other members such as lactic acid bacteria and proteobacteria are present in lower numbers⁽¹⁾. The real-time PCR primers used in the present study cover the three main bacterial groups as well as several subgroups within clostridial cluster XIVa and IV and a functional gene for butyrate synthesis that is shared between different groups⁽²⁴⁾. The data for all volunteers showed a significant increase for the cluster IV species *F. prausnitzii*. There was a tendency for a group effect, which may have been caused by a residual effect of inulin supplementation, although the faecal sample from the control period was collected 16 d after inulin supplementation was stopped. Furthermore, volunteers in group 2 consumed two cabbage test meals before the inulin treatment period⁽¹⁹⁾. Samples for microbiota analysis were taken 17 d after the test meals, however, and it is therefore unlikely that they had a significant effect on the levels of *F. prausnitzii*. Finally, the observed group effect may have arisen by

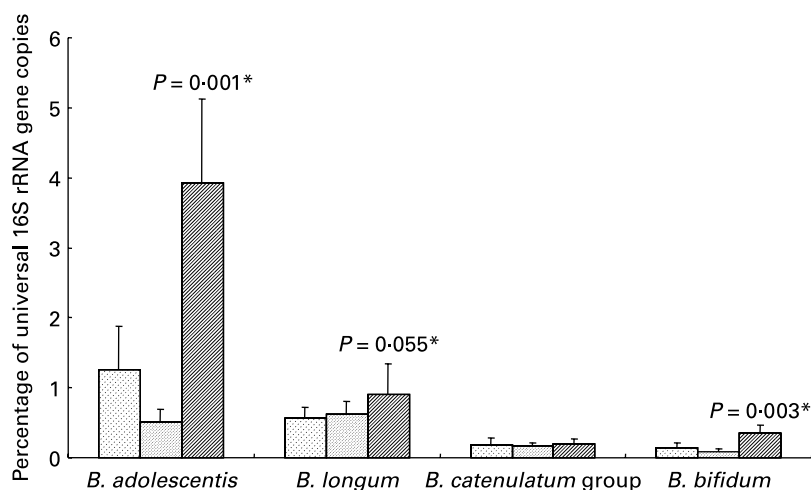


Fig. 3. Individual bifidobacterial species 16S rRNA genes as percentage of all bacterial 16S rRNA genes of faecal samples from ten human volunteers before (□; *n* 12) and after 16 d of control period (▒; *n* 12) or after 16 d of inulin supplementation (▨; *n* 11) based on real-time PCR. **P* values reflect the effect of inulin, obtained from comparing baseline and control periods against the inulin period using contrasts, from ANOVA with volunteer as random effect and treatment, treatment order and their interaction as fixed effects. Absence of *P* value means not significant (*P* > 0.10). Error bars reflect SEM (based on variation between volunteers).

chance. Kleessen *et al.* (39) did not find a significant change in *F. prausnitzii* in human volunteers consuming bakery products containing inulin. Therefore, the effect seen in the present study will have to be confirmed in other human intervention studies to clarify whether the *F. prausnitzii* group is indeed stimulated by inulin; however, it is conceivable that this is the case, as several strains of *F. prausnitzii* have been shown to utilise inulin in pure culture (11). *F. prausnitzii* is a butyrate producer and inulin has been reported to be butyrogenic in animal models (2). An increase in butyrate production upon inulin ingestion could not, however, be detected in the present study. This was also true for other studies with similar levels of daily ingestion of inulin, and it must be recognised that SCFA changes in the upper colon may not be detected in the faeces as the majority of fermentation acids formed is taken up by the colon (40). Positive correlation between *F. prausnitzii* and butyrate levels was weak, whereas the levels of *Roseburia/E. rectale* bacteria showed a significant positive correlation with butyrate, as also observed in a recent study with obese volunteers (41). While this group did not increase upon inulin ingestion as a mean of all volunteers, two volunteers showed a strong increase (data not shown). This could have been caused by a fluctuation in the microbiota unrelated to inulin. It is possible, however, that only a subgroup of people reacts to inulin intake by an increase in *Roseburia/E. rectale* levels, possibly based on which strain/species of the *Roseburia/E. rectale* group are present. In pure culture, *Roseburia inulinivorans* has been shown to degrade inulin, whereas other *Roseburia/E. rectale* strains either show no or weaker inulin utilisation (12,42). Two primer sets were used for the *Roseburia/E. rectale* group that either exclude (Rrec1) or include *R. inulinivorans* (Rrec2). The high levels of this group after inulin intake in two volunteers seen with primer set Rrec1 were less pronounced with primer set Rrec2 (data not shown), indicating that *R. inulinivorans* is not the main species stimulated by inulin. Either other species within this group are directly stimulated by inulin, or this increase may be due to a cross-feeding effect from primary inulin degraders, as has been shown *in vitro* for *Roseburia intestinalis* and *R. hominis* (13,14).

A significant decrease in cluster XIVa (Erec) bacteria upon inulin ingestion was found in other studies (39,43) by fluorescent *in situ* hybridisation, but individual members of this group, such as *Roseburia/E. rectale* spp., were not investigated. We did not find a significant change in cluster XIVa bacteria in the present study. Interestingly, the butyryl-CoA CoA-transferase gene levels correlated significantly with cluster XIVa bacteria, but not with either the *Roseburia/E. rectale* group (Rrec1), *F. prausnitzii* or butyrate levels. This indicates that cluster XIVa contains many bacteria outside the *Roseburia/E. rectale* cluster carrying this pathway for butyrate formation that may be less metabolically active and thus may not contribute much to butyrate production in the colon.

It was observed that the spread within volunteers was similar to that between volunteers. This was about 1.8% (as percentage of total bacteria) for the bifidobacteria, 3.3% for Rrec1, 5.1% for *F. prausnitzii* and up to 13.4% for *Bacteroidetes*.

Bifidobacterium-specific clone libraries were constructed from the faecal samples of four volunteers to identify the main species present. Between three and five different *Bifidobacterium* spp. were found in each of the volunteers, similar to other studies based on multiplex PCR and PCR temperature gradient gel electrophoresis which found that a carriage of three to four species was most common in adults (5,8). The real-time PCR detection led to similar results, with the majority of volunteers carrying at least three of the four species analysed (data not shown). The bifidobacterial composition, examined by PCR-denaturing gradient gel electrophoresis, was found to be stable over a 4-week period (6). We also found that most species detected were present in each of the three samples for each volunteer with both the methodological approaches used here. Volunteer L carried five different *Bifidobacterium* species that were still detected even after a drastic increase in *B. adolescentis* after inulin consumption (data not shown). Volunteer H, on the other hand, had a less stable species composition, and the stability of the microbiota might therefore differ between individuals.

B. adolescentis and *B. longum* were the most prevalent species in the present study, followed by *B. pseudocatenulatum* and *B. bifidum*. These species were also found to be common in other studies utilising PCR-based methods^(5,7,8). *B. dentium*, detected in only one of the four volunteers using the clone library approach, was also found to be less prevalent in other investigations^(5,7,8). *B. animalis*, found here in three of the four volunteers, albeit at low numbers, is commonly used as a probiotic⁽⁴⁴⁾ and may therefore originate from the consumption of probiotic food products. Alternatively, it may form part of the indigenous microbiota as recently demonstrated in elderly subjects⁽⁴⁵⁾. Overall, the prevalence and species composition of bifidobacteria found in the present study are in good agreement with those from previous studies.

The ability of *Bifidobacterium* strains to grow on fructo-oligosaccharides and inulin has been investigated *in vitro*⁽⁹⁾. Of the fifty-five strains tested, all could grow on fructo-oligosaccharides but only eight on inulin. The study did not include all the species found here, but for *B. adolescentis* and *B. bifidum* both inulin degraders and non-degraders were found. Therefore, this trait does not seem to be unique to certain *Bifidobacterium* spp. and rather a strain-dependent characteristic. However, as growth on fructo-oligosaccharides was demonstrated *in vitro* for all strains tested, cross-feeding of oligosaccharides from the primary inulin degraders to other bifidobacteria is likely⁽⁹⁾. The present results indicate that *B. adolescentis* competes best for inulin *in vivo*, be it directly or by cross-feeding. This is particularly evident in volunteer L, who carried very low levels of five different *Bifidobacterium* spp. both in the baseline and control sample (data not shown), but showed a major boost of *B. adolescentis* after the inulin period. *B. pseudocatenulatum* has also been reported to be prevalent in man^(5,7). We did detect it in six volunteers, but it did not show an increase upon inulin ingestion.

In conclusion, the present study indicates that *B. adolescentis* plays a major role in the response to inulin *in vivo*. Furthermore, a significant stimulation was found for *F. prausnitzii*, confirming that ingestion of prebiotics is likely to lead to microbiota changes beyond the lactic acid bacteria⁽¹⁰⁾. Since inter-individual variation may have a major influence, studies involving larger numbers of volunteers together with more detailed analysis of the microbiota will be necessary to further define those bacteria that respond to inulin.

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of the data. K. S. analysed the faecal pH and SCFA concentration and contributed to optimising the real-time PCR quantification. Z. F. and A. D. designed the present study. G. H. performed the statistical analysis and contributed to the preparation of the manuscript. P. L. designed the present study and performed the real-time PCR quantification and clone library analysis and prepared the manuscript.

References

1. Flint HJ, Louis P, Scott KP & Duncan SH (2007) Commensal bacteria in health and disease. In *Virulence Mechanisms of Bacterial Pathogens*, pp. 101–114 [KA Brogden, editor]. Washington, DC: ASM Press.
2. Gibson GR, Probert HM, Van Loo J, Rastall RA & Roberfroid MB (2004) Dietary modulation of the human colonic microbiota: updating the concept of prebiotics. *Nutr Res Rev* **17**, 259–275.
3. Macfarlane S, Macfarlane GT & Cummings JH (2006) Review article: prebiotics in the gastrointestinal tract. *Aliment Pharmacol Ther* **24**, 701–714.
4. Guarner F (2005) Inulin and oligofructose: impact on intestinal diseases and disorders. *Br J Nutr* **93**, S61–S65.
5. Matsuki T, Watanabe K, Tanaka R, Fukuda M & Oyaizu H (1999) Distribution of bifidobacterial species in human intestinal microflora examined with 16S rRNA-gene-targeted species-specific primers. *Appl Environ Microbiol* **65**, 4506–4512.
6. Satokari RM, Vaughan EE, Akkermans AD, Saarela M & de Vos WM (2001) Bifidobacterial species diversity in human feces detected by genus-specific PCR and denaturing gradient gel electrophoresis. *Appl Environ Microbiol* **67**, 504–513.
7. Mullié C, Odou MF, Singer E, Romond MB & Izard D (2003) Multiplex PCR using 16S rRNA gene-targeted primers for the identification of bifidobacteria from human origin. *FEMS Microbiol Lett* **222**, 129–136.
8. Mangin I, Suau A, Magne F, Garrido D, Gotteland M, Neut C & Pochart P (2006) Characterization of human intestinal bifidobacteria using competitive PCR and PCR-TTGE. *FEMS Microbiol Ecol* **55**, 28–37.
9. Rossi M, Corradini C, Amaretti A, Nicolini M, Pompei A, Zannoni S & Matteuzzi D (2005) Fermentation of fructooligosaccharides and inulin by bifidobacteria: a comparative study of pure and fecal cultures. *Appl Environ Microbiol* **71**, 6150–6158.
10. Food Quality and Standards Service & Food and Agriculture Organization of the United Nations (FAO) (2007) FAO Technical Meeting on prebiotics, September 15–16
11. Duncan SH, Hold GL, Harmsen HJM, Stewart CS & Flint HJ (2002) Growth requirements and fermentation products of *Fusobacterium prausnitzii*, and a proposal to reclassify it as *Faecalibacterium prausnitzii* gen nov. comb. nov. *Int J Syst Evol Microbiol* **52**, 2141–2146.
12. Duncan SH, Aminov RI, Scott KP, Louis P, Stanton TB & Flint HJ (2006) Proposal of *Roseburia faecis* sp. nov. *Roseburia hominis* sp. nov. and *Roseburia inulinivorans* sp. nov., based on isolates from human faeces. *Int J Syst Evol Microbiol* **56**, 2437–2441.
13. Belenguer A, Duncan SH, Calder AG, Holtrop G, Louis P, Lobley GE & Flint HJ (2006) Two routes of metabolic cross-feeding between *Bifidobacterium adolescentis* and butyrate-producing anaerobes from the human gut. *Appl Environ Microbiol* **72**, 3593–3599.
14. Falony G, Vlachou A, Verbrugghe K & De Vuyst L (2006) Cross-feeding between *Bifidobacterium longum* BB536 and acetate-converting, butyrate-producing colon bacteria during growth on oligofructose. *Appl Environ Microbiol* **72**, 7835–7841.

15. Bourriaud C, Robins RJ, Martin L, Kozłowski F, Tenailleu E, Cherbut C & Michel C (2005) Lactate is mainly fermented to butyrate by human intestinal microfloras but inter-individual variation is evident. *J Appl Microbiol* **99**, 201–212.
16. Walker AW, Duncan SH, McWilliam Leitch EC, Child MW & Flint HJ (2005) pH and peptide supply can radically alter bacterial populations and short-chain fatty acid ratios within microbial communities from the human colon. *Appl Environ Microbiol* **71**, 3692–3700.
17. Belenguer A, Duncan SH, Holtrop G, Anderson SE, Lobley GE & Flint HJ (2007) Impact of pH on lactate formation and utilization by human fecal microbial communities. *Appl Environ Microbiol* **73**, 6526–6533.
18. Zoetendal EG, Collier CT, Koike S, Mackie RI & Gaskins HR (2004) Molecular ecological analysis of the gastrointestinal microbiota: a review. *J Nutr* **134**, 465–472.
19. Fuller Z, Louis P, Mihajlovski A, Rungapamestry V, Ratcliffe B & Duncan AJ (2007) Influence of cabbage processing methods and prebiotic manipulation of colonic microflora on glucosinolate breakdown in man. *Br J Nutr* **98**, 364–372.
20. Gueimonde M, Tölkö S, Korpimäki T & Salminen S (2004) New real-time quantitative PCR procedure for quantification of bifidobacteria in human fecal samples. *Appl Environ Microbiol* **70**, 4165–4169.
21. Richardson AJ, Calder GC, Stewart CS & Smith A (1989) Simultaneous determination of volatile and non-volatile fermentation products of anaerobes by capillary gas chromatography. *Lett Appl Microbiol* **9**, 5–8.
22. Cole JR, Chai B, Farris RJ, Wang Q, Kulam-Syed-Mohideen AS, McGarrell DM, Bandela AM, Cardenas E, Garrity GM & Tiedje JM (2007) The ribosomal database project (RDP-II): introducing myRDP space and quality controlled public data. *Nucleic Acids Res* **35**, D169–D172.
23. Aminov RI, Walker AW, Duncan SH, Harmsen HJ, Welling GW & Flint HJ (2006) Molecular diversity, cultivation, and improved detection by fluorescent *in situ* hybridisation of a dominant group of human gut bacteria related to *Roseburia* spp. or *Eubacterium rectale*. *Appl Environ Microbiol* **72**, 6371–6376.
24. Louis P & Flint HJ (2007) Development of a semiquantitative degenerate real-time PCR-based assay for estimation of numbers of butyryl-coenzyme A (CoA) CoA transferase genes in complex bacterial samples. *Appl Environ Microbiol* **73**, 2009–2012.
25. Flint HJ, McPherson CA & Bisset J (1989) Molecular cloning of genes from *Ruminococcus flavefaciens* encoding xylanase and beta(1-3,1-4)glucanase activities. *Appl Environ Microbiol* **55**, 1230–1233.
26. Bookout AL, Cummins CL, Kramer MF, Pesola JM & Mangelsdorf DJ (2006) High-throughput real-time quantitative reverse transcription PCR. In *Current Protocols in Molecular Biology*, pp. 158–1528 [FM Ausubel, R Brent, RE Kingston, DD Moore, JG Seidman, JA Smith and K Struhl, editors]. Hoboken, NJ: John Wiley & Sons, Inc.
27. Bartosch S, Fite A, Macfarlane GT & McMurdo ME (2004) Characterization of bacterial communities in feces from healthy elderly volunteers and hospitalized elderly patients by using real-time PCR and effects of antibiotic treatment on the fecal microbiota. *Appl Environ Microbiol* **70**, 3575–3581.
28. Matsuki T, Watanabe K, Fujimoto J, Miyamoto Y, Takada T, Matsumoto K, Oyaizu H & Tanaka R (2002) Development of 16S rRNA-gene-targeted group-specific primers for the detection and identification of predominant bacteria in human feces. *Appl Environ Microbiol* **68**, 5445–5451.
29. Ewing B & Green P (1998) Basecalling of automated sequencer traces using Phred. II. Error probabilities. *Genome Res* **8**, 186–194.
30. Ewing B, Hillier L, Wendl M & Green P (1998) Basecalling of automated sequencer traces using Phred. I. Accuracy assessment. *Genome Res* **8**, 175–185.
31. Chou HH & Holmes MH (2001) DNA sequence quality trimming and vector removal. *Bioinformatics* **17**, 1093–1104.
32. Edgar RC (2004) MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res* **32**, 1792–1797.
33. Chenna R, Sugawara H, Koike T, Lopez R, Gibson TJ, Higgins DG & Thompson JD (2003) Multiple sequence alignment with the clustal series of programs. *Nucleic Acids Res* **31**, 3497–3500.
34. Clamp M, Cuff J, Searle SM & Barton GJ (2004) The Jalview Java alignment editor. *Bioinformatics* **20**, 426–427.
35. Felsenstein J (2005) PHYLIP (PhylogenyInference Package) version 3.6. Distributed by the author. Department of Genome Sciences, University of Washington, Seattle, <http://evolution.genetics.washington.edu/phylip.html>
36. Schloss PD & Handelsman J (2005) Introducing DOTUR, a computer program for defining operational taxonomic units and estimating species richness. *Appl Environ Microbiol* **71**, 1501–1506.
37. Altschul SF, Gish W, Miller W, Myers EW & Lipman DJ (1990) Basic local alignment search tool. *J Mol Biol* **215**, 403–410.
38. Collins MD, Lawson PA, Willems A, Cordoba JJ, Fernandez-Garayzabal J, Garcia P, Cai J, Hippe H & Farrow JA (1994) The phylogeny of the genus *Clostridium*: proposal of five new genera and eleven new species combinations. *Int J Syst Bacteriol* **44**, 812–826.
39. Kleessen B, Schwarz S, Boehm A, Fuhrmann H, Richter A, Henle T & Krueger M (2007) Jerusalem artichoke and chicory inulin in bakery products affect faecal microbiota of healthy volunteers. *Br J Nutr* **98**, 540–549.
40. Nyman M (2002) Fermentation and bulking capacity of indigestible carbohydrates: the case of inulin and oligofructose. *Br J Nutr* **87**, S163–S168.
41. Duncan SH, Belenguer A, Holtrop G, Johnstone AM, Flint HJ & Lobley GE (2007) Reduced dietary intake of carbohydrates by obese subjects results in decreased concentrations of butyrate and butyrate-producing bacteria in feces. *Appl Environ Microbiol* **73**, 1073–1078.
42. Duncan SH, Hold GL, Barcenilla A, Stewart CS & Flint HJ (2002) *Roseburia intestinalis* sp. nov. a novel saccharolytic, butyrate-producing bacterium from human faeces. *Int J Syst Evol Microbiol* **52**, 1615–1620.
43. Harmsen HJM, Raangs GC, Franks AH, Wildeboer-Veloo ACM & Welling GW (2002) The effect of the prebiotic inulin and the probiotic *Bifidobacterium longum* on the fecal microflora of healthy volunteers measured by FISH and DGGE. *Microbial Ecol Health Dis* **14**, 211–219.
44. Masco L, Huys G, De Brandt E, Temmerman R & Swings J (2005) Culture-dependent and culture-independent qualitative analysis of probiotic products claimed to contain bifidobacteria. *Int J Food Microbiol* **102**, 221–230.
45. Ouwehand AC, Bergsma N, Parhiala R, Lahtinen S, Gueimonde M, Finne-Soveri H, Strandberg T, Pitkälä K & Salminen S (2008) *Bifidobacterium* microbiota and parameters of immune function in elderly subjects. *FEMS Immunol Med Microbiol* **53**, 18–25.
46. Liu C, Song Y, McTeague M, Vu AW, Wexler H & Finegold SM (2003) Rapid identification of the species of the *Bacteroides fragilis* group by multiplex PCR assays using group- and species-specific primers. *FEMS Microbiol Lett* **222**, 9–16.
47. Rinttilä T, Kassinen A, Malinen E, Kroggius L & Palva A (2004) Development of an extensive set of 16S rDNA-targeted primers for quantification of pathogenic and indigenous bacteria in faecal samples by real-time PCR. *J Appl Microbiol* **97**, 1166–1177.
48. Hold GL, Schwartz A, Aminov RI, Blaut M & Flint HJ (2003) Oligonucleotide probes that detect quantitatively significant

- groups of butyrate-producing bacteria in human feces. *Appl Environ Microbiol* **69**, 4320–4324.
49. Lay C, Sutren M, Rochet V, Saunier K, Doré J & Rigottier-Gois L (2005) Design and validation of 16S rRNA probes to enumerate members of the *Clostridium leptum* subgroup in human faecal microbiota. *Environ Microbiol* **7**, 933–946.
 50. Sghir A, Gramet G, Suau A, Rochet V, Pochart P & Doré J (2000) Quantification of bacterial groups within human fecal flora by oligonucleotide probe hybridization. *Appl Environ Microbiol* **66**, 2263–2266.
 51. Wang RF, Cao WW & Cerniglia CE (1996) PCR detection and quantification of predominant anaerobic bacteria in human and animal fecal samples. *Appl Environ Microbiol* **62**, 1242–1247.
 52. Suau A, Rochet V, Sghir A, Gramet G, Brewaeys S, Sutren M, Rigottier-Gois L & Doré J (2001) *Fusobacterium prausnitzii* and related species represent a dominant group within the human fecal flora. *Syst Appl Microbiol* **24**, 139–145.
 53. Harmsen HJ, Raangs GC, He T, Degener JE & Welling GW (2002) Extensive set of 16S rRNA-based probes for detection of bacteria in human feces. *Appl Environ Microbiol* **68**, 2982–2990.
 54. Malinen E, Rinttilä T, Kajander K, Mättö J, Kassinen A, Krogus L, Saarela M, Korpela R & Palva A (2005) Analysis of the fecal microbiota of irritable bowel syndrome patients and healthy controls with real-time PCR. *Am J Gastroenterol* **100**, 373–382.
 55. Barcenilla A, Pryde SE, Martin JC, Duncan SH, Stewart CS, Henderson C & Flint HJ (2000) Phylogenetic relationships of butyrate-producing bacteria from the human gut. *Appl Environ Microbiol* **66**, 1654–1661.
 56. Louis P, Duncan SH, McCrae SI, Millar J, Jackson MS & Flint HJ (2004) Restricted distribution of the butyrate kinase pathway among butyrate-producing bacteria from the human colon. *J Bacteriol* **186**, 2099–2106.
 57. Barcenilla A (1999) Diversity of the butyrate-producing microflora of the human gut. PhD Thesis. Robert Gordon University, Aberdeen, UK.
 58. Dabek M, McCrae SI, Stevens VJ, Duncan SH & Louis P (2008) Distribution of β -glucosidase and β -glucuronidase activity and of β -glucuronidase gene *gus* in human colonic bacteria. *FEMS Microbiol Ecol* (Epublication ahead of print version 4 June 2008).
 59. Varel VH & Dehority BA (1989) Cellulolytic bacteria and protozoa from bison, cattle-bison hybrids, and cattle fed three alfalfa-corn diets. *Appl Environ Microbiol* **55**, 148–153.
 60. Whitehead TR & Hespell RB (1990) The genes for three xylan-degrading activities from *Bacteroides ovatus* are clustered in a 3.8-kilobase region. *J Bacteriol* **172**, 2408–2412.
 61. Cato EP & Johnson JL (1976) Reinstatement of species rank for *Bacteroides fragilis*, *B. ovatus*, *B. distasonis*, *B. thetaiotaomicron*, and *B. vulgatus*: designation of neotype strains for *Bacteroides fragilis* (Veillon and Zuber) Castellani and Chalmers and *Bacteroides thetaiotaomicron* (Distaso) Castellani and Chalmers. *Int J Syst Bacteriol* **26**, 230–237.
 62. Reuter G (1963) Vergleichende Untersuchungen über die Bifidus-Flora des Säuglings- und Erwachsenenstuhl. *Zentralbl Bakteriol Parasitenkd Orig Abt I* **191**, 486–507.