

Non-nutritive bioactive components in maternal milk and offspring development: a scoping review

Review

Cite this article: Eisha S, Joarder I, Wijenayake S, and McGowan PO. (2022) Non-nutritive bioactive components in maternal milk and offspring development: a scoping review. *Journal of Developmental Origins of Health and Disease* **13**: 665–673. doi: [10.1017/S2040174422000149](https://doi.org/10.1017/S2040174422000149)

Received: 30 April 2021
Revised: 16 February 2022
Accepted: 9 March 2022
First published online: 7 April 2022

Keywords:

Mammalian milk; non-nutritive bioactive components of milk; maternal factors; offspring outcomes

Address for correspondence:

Patrick O. McGowan, Ph.D., University of Toronto, Scarborough Campus, 1265 Military Trail, Toronto, Ontario, Canada, M1C1A4.
E-mail: patrick.mcgowan@utoronto.ca

Shafinaz Eisha and Ishraq Joarder contributed equally to the manuscript.

Shafinaz Eisha^{1,2}, Ishraq Joarder¹ , Sanoji Wijenayake^{1,3} and Patrick O. McGowan^{1,2,4,5} 

¹Department of Biological Sciences, Center for Environmental Epigenetics and Development, University of Toronto Scarborough, Toronto, ON, Canada; ²Department of Cell and Systems Biology, University of Toronto, Toronto, ON, Canada; ³Department of Biology, Richardson College for the Environment and Science Complex, The University of Winnipeg, Winnipeg, MB, Canada; ⁴Department of Psychology, University of Toronto, Toronto, ON, Canada and ⁵Department of Physiology, University of Toronto, Toronto, ON, Canada

Abstract

Lactation is a critical time in mammalian development, where maternal factors shape offspring outcomes. In this scoping review, we discuss current literature concerning maternal factors that influence lactation biology and highlight important associations between changes in milk composition and offspring outcomes. Specifically, we explore maternal nutritional, psychosocial, and environmental exposures that influence non-nutritive bioactive components in milk and their links to offspring growth, development, metabolic, and behavioral outcomes. A comprehensive literature search was conducted using the Preferred Reporting Items for Systematic Reviews and Meta-Analysis Extension for Scoping Reviews (PRISMA-ScR) guidelines. Predetermined eligibility criteria were used to analyze 3,275 papers, and the final review included 40 primary research articles. Outcomes of this review identify maternal obesity to be a leading maternal factor influencing the non-nutritive bioactive composition of milk with notable links to offspring outcomes. Offspring growth and development are the most common modes of programming associated with changes in non-nutritive milk composition due to maternal factors in early life. In addition to discussing studies investigating these key associations, we also identify knowledge gaps in the current literature and suggest opportunities and considerations for future studies.

Background

Maternal milk is a heterogeneous biological fluid that is tailored to meet the developmental, digestive, and immune needs of offspring and is the sole source of nutrition for most newborn mammals.^{1–3} The World Health Organization (WHO) recommends that infants should be exclusively breastfed for a minimum of 6 months to receive the optimal benefits of maternal milk.⁴ Milk is rich in nutritive components, including carbohydrates, proteins, lipids, vitamins, and minerals, which have been discussed in relation to offspring development elsewhere.^{1,2,5–10} Milk also contains non-nutritive bioactive components, including growth and neurodevelopmental factors, hormones, active enzymes, peptides, a complex microbiota, immune factors, as well as non-coding RNA (microRNA – miRNA) encapsulated in milk-derived exosomes, which play roles in immune maturation, metabolism, overall health and developmental outcomes of offspring.^{11–18}

Maternal milk composition varies across lactation stages, milk fractions, circadian cycle, and even temporally within a single feed, exhibiting compositional differences between foremilk and hindmilk.^{1,2,19,20} The first fluid produced by mammary gland epithelial cells soon after delivery, known as colostrum, is rich in non-nutritive factors, including hormones, growth factors and immune components (e.g., lactoferrin, leukocytes, and immunoglobulins).^{2,8,21} In humans, colostrum is replaced by transition milk approximately 60 hours to 5 days post birth and lasts for about 10 days to 2 weeks post-partum, after which mature milk production is initiated.^{2,22,23}

Studies in human and animal models suggest that non-nutritive components in maternal milk contribute to early life developmental programming. Developmental programming refers to cellular and biochemical processes that lead to long-term functional changes as a result of a stimulus and/or insult during a critical period of development.²⁴ For example, maternal nutrition and psychosocial stress exposure during prenatal and/or early postnatal period can affect the overall growth and cognitive development of offspring, as well as increase the risk of disease later in life.^{25–27} Non-nutritive bioactive compounds in milk enhance offspring survival by promoting the healthy development of the immune system and intestinal microbial colonization.^{2,14,28–31} Emerging evidence also suggests that these bioactive components in milk may help

© The Author(s), 2022. Published by Cambridge University Press in association with International Society for Developmental Origins of Health and Disease. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted re-use, distribution, and reproduction in any medium, provided the original work is properly cited.

protect infants against acute infections, neuroinflammation and systemic inflammation, and metabolic diseases, including obesity, hypertension, and/or type I and II diabetes.^{1,2,12,32,33}

There is considerable evidence that perinatal (e.g., the combined period of prenatal and postnatal life) maternal overnutrition and psychosocial stress play a role in shaping metabolic and neurodevelopmental outcomes in the offspring.^{34,35} For example, in rodent models, a maternal diet high in saturated fats and complex sugars leads to increased offspring bodyweight,³⁴ elevated levels of inflammation in visceral adipocytes,³⁶ increased liver steatosis in males,³⁶ and increased anxiety-like behavior that persists into adulthood.^{37,38} In humans, maternal obesity is associated with an increased risk of childhood asthma, and higher infant weight-for-length and Body Mass Index (BMI) at birth.^{39,40} However, the milk of mothers with obesity was found to contain high levels of insulin and leptin, which may provide some benefit to the developing offspring,⁴¹ as increased levels of milk insulin and leptin reduce intestinal inflammation and increase intestinal barrier function.⁴²

Maternal anxiety during gestation as well as psychological and social stress are associated with a slower rate of cognitive development (lower scores on the mental development index of the Bayley Scales of Infant Development) in human infants over the first postnatal year.⁴³ In addition, maternal psychosocial stress is correlated with lower levels of milk bacterial diversity at 3 months postpartum,⁴⁴ which may contribute to a subsequent decrease in infant gut microbiome diversity.⁴⁵ While still debated,⁴⁶ there is evidence that this decreased complexity of the neonatal gut microbiome is associated with elevated risks of gastrointestinal, autoimmune, and metabolic disorders in adulthood.^{47,48} Although the compositional changes of non-nutritive components in milk due to maternal obesity and nutritional status is well-established,^{49–57} less is known about how maternal psychosocial stress during the pre-gestational/gestational/post-gestational periods impact milk composition.^{58,59} Also, while a majority of studies with human subjects has statistically controlled for anthropometric and demographic covariates, including maternal height, age and ethnicity, these measurements have not been investigated as independent variables that could potentially alter non-nutritive milk composition. This is particularly important because milk composition varies across demographics. For instance, maternal age and ethnicity are highly correlated with fat content in milk.^{60,61}

This scoping review was designed to provide a comprehensive overview of studies that have investigated non-nutritive bioactive components in milk associated with maternal factors and/or physiological and behavioral outcomes of offspring. In addition to a focus on the human literature, we did not exclude model organisms during the screening process to avoid missing relevant literature that fit within the scope of our study. Model systems that have been investigated in lactation biology research are highly heterogeneous and span non-human primates, rodents, bovine, and other mammals. We categorized the primary literature into different species to aid in elucidating knowledge gaps in lactation biology. We attempted to characterize emerging knowledge concerning the most studied maternal factors that play roles in altering the non-nutritive bioactive components of maternal milk, along with their associations with offspring outcomes during early childhood that can potentially persist into adulthood.

Methods

This scoping review was performed in accordance with the Preferred Reporting Items for Systematic Reviews and Meta-Analysis Extension for Scoping Reviews (PRISMA-ScR) guidelines.⁶²

Search strategy

A comprehensive literature search was performed using the databases MEDLINE, PubMed, EMBASE and the Web of Science, with “and/or” combinations of the following keywords: mammalian milk, lactation biology, maternal factors, and offspring developmental programming. The literature search was last updated on August 5th, 2020. All records were imported into Covidence systematic review software (Veritas Health Innovation, Melbourne, Australia) and duplicates were removed from the library.

Exclusion/eligibility criteria and article selection

Title and abstract screening were conducted by two, independent reviewers (IJ and SE), with disagreements resolved by a third independent reviewer (SW). During the abstract and title screening phase, articles were excluded if one or more of the following criteria were met: 1) Primary language is not English; 2) Not peer-reviewed; 3) Sole focus on metabolic, nutritional, and immune stress affecting maternal health as opposed to mother-offspring dyads or maternal factors contributing to changes in milk composition; 4) Do not link changes in milk composition with either maternal factors or offspring outcomes.

Remaining articles were subcategorized into four species-specific databases (human, rodent, bovine, and other mammals). We included studies that spanned human and non-human models, because many previously published studies used several non-human mammalian model systems to investigate different aspects of lactation biology that were relevant to our research objective. For instance, numerous studies have investigated the mechanistic and molecular mechanisms of milk derived exosome uptake and transfer using rodent, bovine, and marsupial milk. However, studies that have investigated non-nutritive milk components in relation to maternal factors and/or offspring outcomes mainly used human and/or rodent models. Further, differences in milk composition have been reported across mammals.⁶³ As such, we decided to include all relevant studies rather than to exclude non-human model systems, and used four subcategories to differentiate human studies from studies of rodents, bovines, and other mammals.

Subsequently, the remaining articles were subjected to full-text screening. As per abstract and title screening, two independent reviewers (IJ and SE) conducted this analysis, and a third independent reviewer (SW) resolved disagreements. The following criteria were used to determine eligibility of the full-text articles towards the scoping review. Studies that contained the following criteria were included: 1) Primary literature; 2) A primary focus on non-nutritive bioactive components of milk as opposed to nutritive components; 3) Report experimentally validated physiological and psychological outcomes in offspring as opposed to conducting only *in vitro* experiments and/or *in-silico* predictions; 4) Research topics investigating non-nutritive bioactive components in milk and relating them to maternal factors and/or offspring outcomes (Table 1); 5) Experimental methodologies for milk exosome isolation and characterization adhering to the guidelines put forth by the International Society for Extracellular Vesicles (ISEV), where applicable.⁶⁴

Data collection and extraction

Data extraction was conducted for all articles that remained post full-text screening. A comprehensive and detailed excel inventory was created as a data charting form and the extractable variables were jointly developed by three reviewers (IJ, SE and SW).

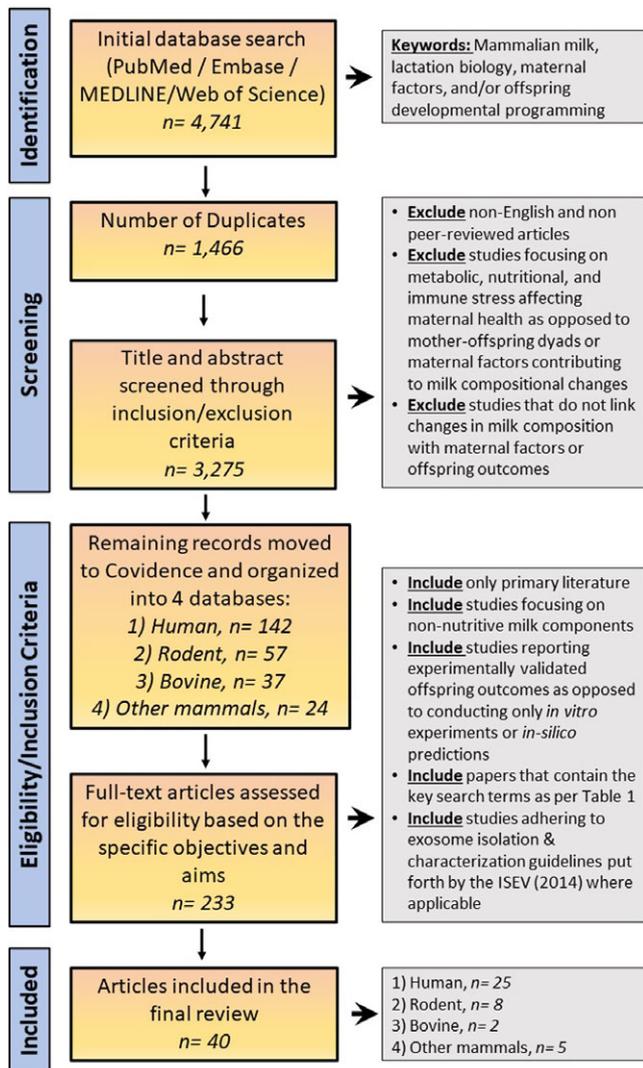


Fig. 1. PRISMA-ScR flow chart illustrating screening and eligibility criteria for article selection.

Extracted data included study characteristics (author, year of publication, country and doi), subtopic categorization based on overarching research objectives, maternal factors in relation to non-nutritive lactation biology and developmental outcomes of offspring. Two reviewers (IJ and SE) independently extracted and inputted data based on the distinct categorical variables developed.

Results

Study selection

A total of 4,741 journal articles were identified after the primary search. One thousand four hundred and sixty-six duplicates were removed using the Covidence reference manager and the 3,275 papers that remained were subjected to first round eligibility screening. Upon completion, 233 articles met the first eligibility criteria and were organized into four databases based on the species that was investigated, where the human database contained 142 entries, rodent contained 57, bovine contained 37, and the other category involving all other mammals

Table 1. Search terms and strategy

Non-nutritive bioactive components in maternal milk
Growth and neurodevelopmental factors
Cytokines
Chemokines
Hormones
Enzymes
Microbiota
Immune factors
Milk-derived exosomes
microRNA
Maternal factors
Nutrition
Psychosocial
Environmental
Anthropometry
Offspring outcomes
Growth
Neurodevelopment
Metabolism
Immunity
Microbiome
Behavior

Note: We searched for primary literature that included non-nutritive bioactive components in maternal milk AND linked that to maternal factors AND/OR offspring outcomes. We used multiple search terms (individually and in combination) for each category, and we included papers that contained at least one of the search terms.

contained 24. Fifteen papers were repeated across multiple species at this stage of screening. Finally, 40 articles passed the second round of full-text eligibility screening and were used in the final review (Fig. 1). The final collection of 40 papers did not have repeating entries.

Study characteristics and categorization

The final selection of 40 primary research articles that remained post-screening and eligibility testing were categorized into two overarching research objectives, with a third category that linked both objectives, across four different species (Table 2). Fourteen papers (35.0%) pertained to maternal factors influencing non-nutritive bioactive components in milk, 13 papers (32.5%) related to non-nutritive components of milk impacting offspring development, and 13 papers (32.5%) linked maternal factors altering non-nutritive bioactive milk components and, in turn, alterations in offspring outcomes. Of the 40 papers, 25 papers (62.5%) were in humans, 8 papers (20.0%) were in rodents, 2 papers (5.0%) in bovine and 5 papers (12.5%) were in other mammals.

Analysis and synthesis of results

Of the 40 remaining papers, 13 papers (32.5%) characterized non-nutritive bioactive components in milk in relation to maternal

factors impacting developmental outcomes of offspring (Table 2 – Category 3; Table 3). Within this category, six papers (46.1%) were in human, five papers (38.5%) were in rodent, and two papers (15.4%) were in other mammals, specifically rhesus macaques (*Macaca mulatta*). As reported in Table 3, 11 studies (84.6%) investigated maternal overweight or obesity status, which was associated with altering non-nutritive bioactive milk components and impacting offspring outcomes. Maternal gestational stress, metabolic syndrome (gestational diabetes mellitus), social rank, and parity were investigated by only four studies (30.1%). Two of these studies were included in multiple categories since they investigated more than one maternal factor (e.g., obesity, gestational stress and metabolic syndrome). In addition, weight, growth, and development were the most common offspring outcomes that were explored across all 13 papers with respect to maternal factors and non-nutritive milk composition. Five papers (38.4%) measured offspring body fat/adiposity, four papers (30.8%) pertained to offspring metabolism, and two papers (15.4%) examined offspring behavior. Finally, two papers (15.4%) assessed other offspring outcomes, such as microbiome, intestinal, and adipose tissue inflammation.

Supplementary Table 1 contains a complete list of all 40 papers identified in the final selection of primary literature included in this scoping review, along with a short description of each paper's main findings/focus.

Discussion

The overall objective of this scoping review was to explore the current literature pertaining to maternal factors, including nutritional status, psychosocial stress, and environmental factors that may influence non-nutritive bioactive composition in milk, and thereby impact developmental trajectories in offspring (e.g., physiological, neurological, metabolic, and behavioral outcomes). Numerous studies have investigated distinct aspects of this topic and the associations between individual maternal factors, lactation biology, and/or offspring outcomes. However, potential causal relationships and associations between maternal factors, changes in non-nutritive milk composition, and early life programming effects in offspring remain poorly understood.

Maternal factors influencing milk composition and offspring developmental trajectories

Maternal factors, including nutritional status, metabolic disorders, and/or psychosocial stress appear to play important roles in shaping non-nutritive milk composition (explored across 14 papers; Table 2), which in turn may influence the development and early life programming of offspring (explored across 13 papers; Table 2). Maternal nutritional status was the most investigated maternal factor reported to influence lactation biology and offspring outcomes. In particular, there is evidence that maternal obesity status (BMI > 30 kg/m²) alters miRNAs,⁶⁵ immune components,⁶⁶ pro-inflammatory fatty acid,⁵⁰ and carotenoid levels⁵⁰ in milk. These bioactive compounds are critical for offspring growth³⁹, visual and neural development,⁵⁰ intestinal microbiota diversity and metabolic health.⁴² Moreover, human and laboratory rodent studies have demonstrated that the initiation of lactation was delayed in mothers that were on a high fat diet,^{67,68} including disruption of mammary gland morphology and a notable decline of intact alveolar units that are essential for lactogenesis.⁶⁷ Several rodent studies also reported that mothers exposed to a high fat diet during pre-gestation, gestation, and lactation, exhibited increased nursing

behaviors, where time spent in “arched-back”⁶⁹ and “blanket”⁷⁰ nursing increased during the dark phase of the circadian cycle. “Arched-back” nursing is a type of maternal care behavior, where the mother arches her back while nursing pups to allow more access to the teats, and “blanket” nursing is where the dam uses her body to lay over the pups while nursing.^{71,72} We previously postulated that the percent increase in time spent nursing may be a compensatory lactation-specific behavior to combat impaired milk production, prolactin insensitivity⁷³ or delayed milk production due to increased mammary gland inflammation.^{67,70} Several human studies have shown that maternal obesity alters hormonal and immune components in milk, and that being obese and/or overweight during the pre-gestational period is associated with increased bodyweight and alterations in the metabolic profile of offspring during early childhood.^{39,74,75} For example, higher maternal post-pregnancy BMI was associated with increased levels of milk adiponectin, a hormone that affects development by regulating lipid metabolism, reducing pro-inflammatory cytokines and improving insulin sensitivity in offspring.^{76–79} Other studies reported that adiponectin levels in maternal milk from healthy lactating women were found to be positively correlated with adiponectin in infant serum⁸⁰ and inversely associated with growth and weight gain during early infancy.^{79,81} Another study reported that infants who were partially breastfed (defined as the combined use of maternal milk and formula) by mothers with obesity had elevated levels of insulin and adiponectin and lower levels of IGF-I (insulin growth factor 1) in the blood at 9 months of age compared to infants partially breastfed by normal weight mothers.⁷⁴ Thus, the higher concentration of blood adiponectin reported in offspring who were born to as well as partially breastfed by mothers with obesity may indicate a positive, lactation-specific metabolic adaptation to prenatal maternal obesity exposure.

Maternal psychosocial stress is another prominent factor that may influence cortisol levels in milk,⁸² which in turn has been associated with offspring development and behavioral trajectories. A study in rhesus macaques reported a positive correlation between increased maternal milk cortisol levels and changes in male offspring temperament, including increased activity, boldness, confidence and curiosity.⁸³ Higher milk cortisol was also associated with increased social interactions in female macaques at 4–8 months of age.⁸⁴ However, another study in rhesus macaques found that cortisol levels in milk were positively correlated with nervous temperament, and negatively correlated with confident temperament in offspring.⁸⁵ A study in humans reported a protective effect of increased milk cortisol exposure during early childhood, where increased milk cortisol levels were associated with decreased childhood obesity.⁸⁶ However, there is other evidence that higher milk cortisol levels are associated with increased fear reactivity in female infants at 3 months⁸⁷ and 8 months of age.⁸⁸ Despite mixed findings on the beneficial/deleterious effects of increased milk cortisol transfer across mother-offspring dyads, these studies illustrate the manner in which maternal psychosocial stress may alter milk composition and offspring behavior.

Additional studies in humans and other mammals reported that maternal immune status, group rank, and parity also influence not only cortisol, but adiponectin, growth factors and secretory immunoglobulin levels in milk.^{83,89,90} Gestational age and mode of delivery are two other maternal factors associated with altered milk composition.⁹¹ Specifically, Carney et al.⁹¹ reported that the expression profiles of milk miRNAs in lipid and skim fractions differed in preterm delivery compared to term birth. They found that two miRNAs showed increased transcript abundance, and six

Table 2. Overarching research topics and categorization of the articles included in the final review (*N* = 40)

Research objective	Total papers, <i>n</i> (%)	Human, <i>n</i> (%)	Rodent, <i>n</i> (%)	Bovine, <i>n</i> (%)	Other, <i>n</i> (%)	Citations
1. Maternal factors influencing non-nutritive bioactive components of milk	14 (35.0)	12 (85.7)	2 (14.3)	–	–	(O'Rourke <i>et al.</i> , 2018), (Carney <i>et al.</i> , 2017), (Kugananthan <i>et al.</i> , 2017), (Munch <i>et al.</i> , 2013), (Thibeau <i>et al.</i> , 2016), (Xi <i>et al.</i> , 2016), (D. Chen <i>et al.</i> , 1998), (Fujimori <i>et al.</i> , 2016), (Groer <i>et al.</i> , 2004), (Mirza <i>et al.</i> , 2019), (Browne <i>et al.</i> , 2019), (Aparicio <i>et al.</i> , 2020), (Pomar <i>et al.</i> , 2019), (Hernandez <i>et al.</i> , 2012)
2. Non-nutritive bioactive components of milk influencing offspring development	13 (32.5)	7 (53.8)	1 (7.7)	2 (15.4)	3 (23.1)	(Grey <i>et al.</i> 2013), (Munblit <i>et al.</i> , 2017), (Berger <i>et al.</i> , 2020), (van Rossem <i>et al.</i> , 2019), (Nolvi <i>et al.</i> , 2018), (Hahn-Holbrook <i>et al.</i> , 2016), (Willumsen <i>et al.</i> , 2003), (Melo <i>et al.</i> , 2009), (Lee <i>et al.</i> , 2018), (Li <i>et al.</i> , 2019), (Ma <i>et al.</i> , 2017), (Sullivan <i>et al.</i> , 2011), (Dettmer <i>et al.</i> , 2018)
3. Maternal factors altering non-nutritive bioactive components of milk that in turn influence offspring outcomes	13 (32.5)	6 (46.1)	5 (38.5)	–	2 (15.4)	(Lemas <i>et al.</i> , 2016), (Panagos <i>et al.</i> , 2016), (Young <i>et al.</i> , 2017), (Ellsworth <i>et al.</i> , 2020), (Zamanillo <i>et al.</i> , 2019), (Yu <i>et al.</i> , 2018), (Oben <i>et al.</i> , 2010), (Bautista <i>et al.</i> , 2016), (Monks <i>et al.</i> , 2018), (Y. Chen <i>et al.</i> , 2017), (Purcell <i>et al.</i> , 2011), (Hinde <i>et al.</i> , 2015), (Bernstein & Hinde, 2016)
Total papers <i>n</i> (%)	40 (100.0)	25 (62.5)	8 (20.0)	2 (5.0)	5 (12.5)	

miRNAs showed decreased transcript abundance in both fractions of preterm milk, when compared to term milk. In addition, six miRNAs in maternal milk were correlated (three positively and three negatively) with mode of delivery.⁹¹ None of the expression profiles of the milk miRNAs examined in the study were associated with maternal age, race, or ethnicity.⁹¹

Limitations in the current literature

We identified several common limitations in studies of human lactation to date. First, limited number of studies included large heterogeneous populations representing a wide range of ethnicities, socioeconomic status, and pre-gestational and gestational BMIs. Although it may be challenging to include multi-ethnic populations due to limitations in funding as well as difficulties in the recruitment of mothers belonging to underrepresented groups, the lack of diversity in these datasets limits the generalizability of the interpretations relating to lactation biology and offspring development. Interestingly, the few studies that included diverse populations reported differences in milk composition among ethnic groups as a function of regional maternal diets and breastfeeding practices.^{92,93} Second, a large proportion of studies lack detailed descriptions of the methods used for milk collection and analysis, including the time of milk collection, use of breast pumps, the interval from last feeding to the time of sample collection, storage and processing steps, and lactational age. Consideration of these factors is critically important for studies that use human milk, especially human mature milk since there are notable differences in bioactive composition as mature milk is produced over a longer period of time, ranging from 4 to 6 weeks post-partum until the end of the lactation period.² In addition, milk collected at the beginning of a feed (foremilk) contains lower fat and higher cellular content compared to the milk collected at the end of the feed (hindmilk).¹⁹ Efforts to standardize the methods, protocols, techniques for milk sample collection and processing by following the technical

guidelines set forth by the ISEV (referred to as the Minimal Information for Studies of Extracellular Vesicles, MISEV) could serve to limit these potential confounds.⁶⁴

Limitations of this scoping review

While we have attempted to provide a comprehensive analysis of the current literature on the biological associations between maternal factors, lactation biology and developmental outcomes in offspring, this scoping review is not without limitations. Despite compiling 40 papers from four different databases, we may have missed relevant studies in our search. Only peer-reviewed, primary literature published in English was included in our search criteria. A few published studies required specialty permission and journal subscriptions that limited our ability to access the full-print version to apply the exclusion and inclusion criteria. Moreover, the anthropometric controls, such as maternal obesity, used as fixed variables in epidemiological studies varied widely, and many studies investigated only one or two maternal factors at distinct timeframes. For instance, while some studies examined maternal pre-gestational obesity,^{39,42,50} others investigated maternal gestational or post-gestational obesity.^{11,74} This imposed limitations on our ability to draw clear conclusions from such findings concerning the roles of specific factors.

Conclusion

There is strong evidence that maternal factors, including nutritional status, psychosocial stress, and environmental factors, influence the non-nutritive bioactive components in milk, which are associated with developmental trajectories in offspring. These outcomes include early life growth, weight, height, language, and emotional development. Studies using non-human models have provided important information concerning the mechanistic role of non-nutritive bioactive components of milk on offspring development. Understanding the association between milk composition

Table 3. Non-nutritive bioactive components in milk linking maternal factors with offspring outcomes

Maternal factors	Milk components	Species	Offspring outcomes					Citations	
			Growth ^b / Development ^c	Metabolism	Body fat/ adiposity	Microbiome	Inflammation Intestine Adipose tissue Behavior ^d		
Obesity ^a	Hormones (Insulin, leptin, adiponectin)	Human	1, 2, 3, 4		1	1	1	(Lemas <i>et al.</i> , 2016) ¹ , (Ellsworth <i>et al.</i> , 2020) ² , (Zamanillo <i>et al.</i> , 2019) ³ , (Yu <i>et al.</i> , 2018) ⁴	
		Rodent	5, 6, 7	5, 6, 8	5, 6, 7		6	7	(Bautista <i>et al.</i> , 2016) ⁵ , (Monks <i>et al.</i> , 2018) ⁶ , (Purcell <i>et al.</i> , 2011) ⁷ , (Oben <i>et al.</i> , 2010) ⁸
	microRNAs	Human	3						(Zamanillo <i>et al.</i> , 2019) ³
		Rodent	9	9					(Y. Chen <i>et al.</i> , 2017) ⁹
	Carotenoid	Human	10		10				(Panagos <i>et al.</i> , 2016) ¹⁰
	Inflammatory cytokines and markers	Human	11						(Young <i>et al.</i> , 2017) ¹¹
Gestational stress	Hormones (Insulin, leptin, adiponectin)	Rodent	7		7			7	(Purcell <i>et al.</i> , 2011) ⁷
Metabolic syndrome (Gestational diabetes mellitus)	Hormones (adiponectin, ghrelin, insulin)	Human	4						(Yu <i>et al.</i> , 2018) ⁴
Social rank and parity	Hormone (cortisol, adiponectin), epidermal growth factor and transforming growth factor	Rhesus macaques	12, 13					13	(Bernstein & Hinde, 2016) ¹² , (Hinde <i>et al.</i> , 2014) ¹³

¹⁻¹³Numerical values in "Offspring outcomes" correspond to the references listed in the "Citations" column.

^aCharacterization of obesity in human studies: overweight: BMI > 25 kg/m², and obese: BMI > 30 kg/m².

^bGrowth: Z-scores for weight-for-age, length-for-age, head circumference-for-age, weight-for-length, body muscle index.

^cDevelopment: brain, immunity.

^dBehavior: nervous and confident temperament, milk ingestion.

and offspring developmental outcomes is essential in the development of strategies to enhance maternal nutrition and mitigate risk factors, leading to the improved health of both the mother and offspring. Milk compositional analysis may also inform enhancement of the quality of infant formula, given that exclusive breastfeeding is not feasible for all mothers due to health and/or lactational complications. Future studies with an increased ethnic and socioeconomic diversity of participants, a more extensive collection of participant phenotypes, and more standardized procedures for human milk collection, processing, storage, and analysis will enhance the generalizability and reproducibility of knowledge in this emerging field of research.

Supplementary materials. For supplementary material for this article, please visit <https://doi.org/10.1017/S2040174422000149>

Acknowledgements. We would like to thank Ms. Sara Guay, Liaison Librarian at University of Toronto – Scarborough for assisting with the literature search. The authors are entirely responsible for the scientific content of the paper.

Financial support. This work was supported by the Natural Sciences and Engineering Council of Canada (NSERC) (P.O.M., grant number RGPIN-2019-493091), (L.J., Undergraduate Student Research Award), and (S.W., Post-doctoral Research Fellowship).

Conflicts of interest. The authors declare that they have no competing or financial interests.

Ethical standards. None.

References

- Andreas NJ, Kampmann B, Mehring Le-Doare K. Human breast milk: a review on its composition and bioactivity. *Early Hum Dev.* 2015; 91(11), 629–635. DOI [10.1016/j.earlhumdev.2015.08.013](https://doi.org/10.1016/j.earlhumdev.2015.08.013).
- Ballard O, Morrow AL. Human milk composition. Nutrients and bioactive factors. *Pediatr Clin North Am.* 2013; 60(1), 49–74. DOI [10.1016/j.pcl.2012.10.002](https://doi.org/10.1016/j.pcl.2012.10.002).
- Beghetti I, Biagi E, Martini S, Brigidi P, Corvaglia L, Aceti A. Human milk's hidden gift: implications of the milk microbiome for preterm infants' health. *Nutrients.* 2019; 11(12), 2944. DOI [10.3390/nu11122944](https://doi.org/10.3390/nu11122944).
- Kramer MS, Kakuma R. The optimal duration of exclusive breastfeeding: a systematic review. *Adv Exp Med Biol.* 2004; 554, 63–77. DOI [10.1007/978-1-4757-4242-8_7](https://doi.org/10.1007/978-1-4757-4242-8_7)
- Allen LH. B vitamins in breast milk: relative importance of maternal status and intake, and effects on infant status and function. *Adv Nutr.* 2012; 3(3), 362–369. DOI [10.3945/an.111.001172](https://doi.org/10.3945/an.111.001172).
- Gates A, Marin T, De Leo G, Stansfield BK. Review of preterm human-milk nutrient composition. *Nutr Clin Pract.* 2020; 36, 1163–1172. DOI [10.1002/npc.10570](https://doi.org/10.1002/npc.10570).
- Keikha M, Bahreynian M, Saleki M, Kelishadi R. Macro- and micronutrients of human milk composition: are they related to maternal diet? A comprehensive systematic review. *Breastfeed Med.* 2017; 12(9), 517–527. DOI [10.1089/bfm.2017.0048](https://doi.org/10.1089/bfm.2017.0048).
- Keikha M, Shayan-Moghadam R, Bahreynian M, Kelishadi R. Nutritional supplements and mother's milk composition: a systematic review of interventional studies. *Int Breastfeed J.* 2021; 16(1), 1–30. DOI [10.1186/s13006-020-00354-0](https://doi.org/10.1186/s13006-020-00354-0).
- Skibiell AL, Hood WR. Milk matters: offspring survival in Columbian ground squirrels is affected by nutrient composition of mother's milk. *Front Ecol Evol.* 2015; 3, 111. DOI [10.3389/fevo.2015.00111](https://doi.org/10.3389/fevo.2015.00111).
- Stannard HJ, Miller RD, Old JM. Marsupial and monotreme milk—a review of its nutrient and immune properties. *PeerJ.* 2020; 8(5), e9335. DOI [10.7717/peerj.9335](https://doi.org/10.7717/peerj.9335).
- Zamanillo R, Sánchez J, Serra F, Palou A. Breast milk supply of microRNA associated with leptin and adiponectin is affected by maternal overweight/obesity and influences infancy BMI. *Nutrients.* 2019; 11(11), 2589. DOI [10.3390/nu11112589](https://doi.org/10.3390/nu11112589).
- Munblit D, Treneva M, Peroni DG, et al. Immune components in human milk are associated with early infant immunological health outcomes: a prospective three-country analysis. *Nutrients.* 2017; 9(6), 532. DOI [10.3390/nu9060532](https://doi.org/10.3390/nu9060532).
- Alsaweed M, Hartmann PE, Geddes DT, Kakulas F. Micronas in breastmilk and the lactating breast: potential immunoprotectors and developmental regulators for the infant and the mother. *Int J Environ Res Public Health.* 2015; 12(11), 13981–14020. DOI [10.3390/ijerph121113981](https://doi.org/10.3390/ijerph121113981).
- Le Doare K, Holder B, Bassett A, Pannaraj PS. Mother's milk: a purposeful contribution to the development of the infant microbiota and immunity. *Front Immunol.* 2018; 9, 361. DOI [10.3389/fimmu.2018.00361](https://doi.org/10.3389/fimmu.2018.00361).
- Moossavi S, Sepehri S, Robertson B, et al. Composition and variation of the human milk microbiota are influenced by maternal and early-life factors. *Cell Host Microbe.* 2019; 25(2), 324–335. DOI [10.1016/j.chom.2019.01.011](https://doi.org/10.1016/j.chom.2019.01.011).
- Golan-Gerstl R, Elbaum Shiff Y, Moshayoff V, Schecter D, Leshkowitz D, Reif S. Characterization and biological function of milk-derived miRNAs. *Mol Nutr Food Res.* 2017; 61(10), 1700009. DOI [10.1002/mnfr.201700009](https://doi.org/10.1002/mnfr.201700009).
- de la Torre Gomez C, Goreham RV, Bech Serra JJ, Nann T, Kussmann M. Exosomes—A review of biophysics, biology and biochemistry of exosomes with a focus on human breast milk. *Front Genet.* 2018; 9, 92. DOI [10.3389/fgene.2018.00092](https://doi.org/10.3389/fgene.2018.00092).
- Zempleni J, Aguilar-Lozano A, Sadri M, et al. Biological activities of extracellular vesicles and their cargos from bovine and human milk in humans and implications for infants. *J Nutr.* 2017; 147(1), 3–10. DOI [10.3945/jn.116.238949](https://doi.org/10.3945/jn.116.238949).
- McGuire MK, Seppo A, Goga A, et al. Best practices for human milk collection for COVID-19 research. *Breastfeed Med.* 2021; 16(1), 29–38. DOI [10.1089/bfm.2020.0296](https://doi.org/10.1089/bfm.2020.0296).
- Bernstein RM, Hinde K. Bioactive factors in milk across lactation: maternal effects and influence on infant growth in rhesus macaques (*Macaca mulatta*). *Am J Primatol.* 2016; 78(8), 838–850. DOI [10.1002/ajp.22544](https://doi.org/10.1002/ajp.22544).
- Puppel K, Gołębiewski M, Grodkowski G, et al. Composition and factors affecting quality of bovine colostrum: a review. *Animals.* 2019; 9(12), 1070. DOI [10.3390/ani9121070](https://doi.org/10.3390/ani9121070).
- Boss M, Gardner H, Hartmann P. Normal human lactation: closing the gap. *F1000Research.* 2018; 7, F1000. DOI [10.12688/F1000RESEARCH.14452.1](https://doi.org/10.12688/F1000RESEARCH.14452.1).
- Christian P, Smith ER, Lee SE, Vargas AJ, Bremer AA, Raiten DJ. The need to study human milk as a biological system. *Am J Clin Nutr.* 2021; 113(5), 1063–1072. DOI [10.1093/AJCN/NQAB075](https://doi.org/10.1093/AJCN/NQAB075).
- Langley-Evans SC. Developmental programming of health and disease. *Proc Nutr Soc.* 2006; 65(1), 97–105. DOI [10.1079/pns.2005478](https://doi.org/10.1079/pns.2005478).
- Alfaradhi M, Ozanne S. Developmental programming in response to maternal overnutrition. *Front Genet.* 2011; 2, 27. DOI [10.3389/FGENE.2011.00027](https://doi.org/10.3389/FGENE.2011.00027).
- Gluckman PD, Hanson MA, Pinal C. The developmental origins of adult disease. *Matern Child Nutr.* 2005; 1, 130–141. DOI [10.1111/j.1740-8709.2005.00020.x](https://doi.org/10.1111/j.1740-8709.2005.00020.x).
- Aizer A, Stroud L, Buka S. Maternal stress and child outcomes: evidence from siblings. *J Hum Resour.* 2016; 51(3), 523–555. DOI [10.3386/W18422](https://doi.org/10.3386/W18422).
- Oftedal OT. The evolution of milk secretion and its ancient origins. *Animal.* 2012; 6(3), 355–368. DOI [10.1017/S1751731111001935](https://doi.org/10.1017/S1751731111001935).
- Robertson RC, Manges AR, Finlay BB, Prendergast AJ. The human microbiome and child growth – first 1000 days and beyond. *Trends Microbiol.* 2019; 27(2), 131–147. DOI [10.1016/j.tim.2018.09.008](https://doi.org/10.1016/j.tim.2018.09.008).
- Forbes JD, Azad MB, Vehling L, et al. Association of exposure to formula in the hospital and subsequent infant feeding practices with gut microbiota and risk of overweight in the first year of life. *JAMA Pediatr.* 2018; 172(7), e181161. DOI [10.1001/jamapediatrics.2018.1161](https://doi.org/10.1001/jamapediatrics.2018.1161).
- Azad MB, Konya T, Maughan H, et al. Gut microbiota of healthy Canadian infants: profiles by mode of delivery and infant diet at 4 months. *Can Med Assoc J.* 2013; 185(5), 385–394. DOI [10.1503/cmaj.121189](https://doi.org/10.1503/cmaj.121189).
- Badillo-Suárez PA, Rodríguez-Cruz M, Nieves-Morales X. Impact of metabolic hormones secreted in human breast milk on nutritional programming in childhood obesity. *J Mammary Gland Biol Neoplasia.* 2017; 22(3), 171–191. DOI [10.1007/s10911-017-9382-y](https://doi.org/10.1007/s10911-017-9382-y).

33. Mirza AH, Kaur S, Nielsen LB, et al. Breast milk-derived extracellular vesicles enriched in exosomes from mothers with type 1 diabetes contain aberrant levels of micRNAs. *Front Immunol.* 2019; 10, 2543. DOI [10.3389/fimmu.2019.02543](https://doi.org/10.3389/fimmu.2019.02543).
34. Chen Y, Wang J, Yang S, et al. Effect of high-fat diet on secreted milk transcriptome in midlactation mice. *Physiol Genomics.* 2017; 49(12), 747–762. DOI [10.1152/physiolgenomics.00080.2017](https://doi.org/10.1152/physiolgenomics.00080.2017).
35. Fodor A, Zelena D. The effect of maternal stress activation on the offspring during lactation in light of vasopressin. *Sci World J.* 2014; 2014(5), 1–15. DOI [10.1155/2014/265394](https://doi.org/10.1155/2014/265394).
36. Monks J, Orlicky DJ, Stefanski AL, et al. Maternal obesity during lactation may protect offspring from high fat diet-induced metabolic dysfunction. *Nutr Diabetes.* 2018; 8(1), 55. DOI [10.1038/s41387-018-0027-z](https://doi.org/10.1038/s41387-018-0027-z).
37. Sasaki A, de Vega W, St-Cyr S, Pan P, McGowan P. Perinatal high fat diet alters glucocorticoid signaling and anxiety behavior in adulthood. *Neuroscience.* 2013; 240(Suppl. 1), 1–12. DOI [10.1016/j.neuroscience.2013.02.044](https://doi.org/10.1016/j.neuroscience.2013.02.044).
38. Winther G, Elfving B, Müller HK, Lund S, Wegener G. Maternal high-fat diet programs offspring emotional behavior in adulthood. *Neuroscience.* 2018; 388(4), 87–101. DOI [10.1016/j.neuroscience.2018.07.014](https://doi.org/10.1016/j.neuroscience.2018.07.014).
39. Ellsworth L, Perng W, Harman E, Das A, Pennathur S, Gregg B. Impact of maternal overweight and obesity on milk composition and infant growth. *Matern Child Nutr.* 2020; 16(3), 12979. DOI [10.1111/mcn.12979](https://doi.org/10.1111/mcn.12979).
40. Forno E, Young OM, Kumar R, Simhan H, Celedón JC. Maternal obesity in pregnancy, gestational weight gain, and risk of childhood asthma. *Pediatrics.* 2014; 134(2), e535–e546. DOI [10.1542/peds.2014-0439](https://doi.org/10.1542/peds.2014-0439).
41. Chan D, Goruk S, Becker A, et al. Adiponectin, leptin and insulin in breast milk: associations with maternal characteristics and infant body composition in the first year of life. *Int J Obes Adv online Publ.* 2017; 42(1), 36–43. DOI [10.1038/ijo.2017.189](https://doi.org/10.1038/ijo.2017.189).
42. Lemas DJ, Young BE, Baker PR, et al. Alterations in human milk leptin and insulin are associated with early changes in the infant intestinal microbiome. *Am J Clin Nutr.* 2016; 103(5), 1291–1300. DOI [10.3945/ajcn.115.126375](https://doi.org/10.3945/ajcn.115.126375).
43. Davis EP, Sandman CA. The timing of prenatal exposure to maternal cortisol and psychosocial stress is associated with human infant cognitive development. *Child Dev.* 2010; 81(1), 131–148. DOI [10.1111/j.1467-8624.2009.01385.x](https://doi.org/10.1111/j.1467-8624.2009.01385.x).
44. Browne PD, Aparicio M, Alba C, et al. Human milk microbiome and maternal postnatal psychosocial distress. *Front Microbiol.* 2019; 10, 2333. DOI [10.3389/fmicb.2019.02333](https://doi.org/10.3389/fmicb.2019.02333).
45. Pannaraj PS, Li F, Cerini C, et al. Association between breast milk bacterial communities and establishment and development of the infant gut microbiome. *JAMA Pediatr.* 2017; 171(7), 647–654. DOI [10.1001/jamapediatrics.2017.0378](https://doi.org/10.1001/jamapediatrics.2017.0378).
46. Carlson AL, Xia K, Azcarate-Peril MA, et al. Infant gut microbiome associated with cognitive development. *Biol Psychiatry.* 2018; 83(2), 148–159. DOI [10.1016/j.biopsych.2017.06.021](https://doi.org/10.1016/j.biopsych.2017.06.021).
47. Milani C, Duranti S, Bottacini F, et al. The first microbial colonizers of the human gut: composition, activities, and health implications of the infant gut microbiota. *Microbiol Mol Biol Rev.* 2017; 81(4), 26050. DOI [10.1128/mmb.00036-17](https://doi.org/10.1128/mmb.00036-17).
48. Turrioni F, Milani C, Duranti S, et al. The infant gut microbiome as a microbial organ influencing host well-being. *Ital J Pediatr.* 2020; 46(1), 1–13. DOI [10.1186/s13052-020-0781-0](https://doi.org/10.1186/s13052-020-0781-0).
49. Bautista C, Montaña S, Ramirez V, et al. Changes in milk composition in obese rats consuming a high-fat diet. *Br J Nutr.* 2016; 115(3), 538–546. DOI [10.1017/S0007114515004547](https://doi.org/10.1017/S0007114515004547).
50. Panagos PG, Vishwanathan R, Penfield-Cyr A, et al. Breastmilk from obese mothers has pro-inflammatory properties and decreased neuroprotective factors. *J Perinatol.* 2016; 36(4), 284–290. DOI [10.1038/jp.2015.199](https://doi.org/10.1038/jp.2015.199).
51. Bravi F, Wiens F, Decarli A, Dal Pont A, Agostoni C, Ferraroni M. Impact of maternal nutrition on breast-milk composition: a systematic review. *Am J Clin Nutr.* 2016; 104(3), 646–662. DOI [10.3945/ajcn.115.120881](https://doi.org/10.3945/ajcn.115.120881).
52. Innis SM. Impact of maternal diet on human milk composition and neurological development of infants. *Am J Clin Nutr.* 2014; 99(3), 734S–741S. DOI [10.3945/AJCN.113.072595](https://doi.org/10.3945/AJCN.113.072595).
53. Samuel TM, Zhou Q, Giuffrida F, Munblit D, Verhasselt V, Thakkar SK. Nutritional and non-nutritional composition of human milk is modulated by maternal, infant, and methodological factors. *Front Nutr.* 2020; 7, e0197713. DOI [10.3389/FNUT.2020.576133](https://doi.org/10.3389/FNUT.2020.576133).
54. Kuganathan S, Gridneva Z, Lai CT, et al. Associations between maternal body composition and appetite hormones and macronutrients in human milk. *Nutrients.* 2017; 9(3), 252. DOI [10.3390/nu9030252](https://doi.org/10.3390/nu9030252).
55. Young BE, Patinkin ZW, Pyle L, et al. Markers of oxidative stress in human milk do not differ by maternal BMI but are related to infant growth trajectories. *Matern Child Health J.* 2017; 21(6), 1367–1376. DOI [10.1007/s10995-016-2243-2](https://doi.org/10.1007/s10995-016-2243-2).
56. Oben JA, Mouralidarane A, Samuelsson AM, et al. Maternal obesity during pregnancy and lactation programs the development of offspring non-alcoholic fatty liver disease in mice. *J Hepatol.* 2010; 52(6), 913–920. DOI [10.1016/j.jhep.2009.12.042](https://doi.org/10.1016/j.jhep.2009.12.042).
57. Pomar CA, Castro H, Picó C, Serra F, Palou A, Sánchez J. Cafeteria diet consumption during lactation in rats, rather than obesity per se, alters miR-222, miR-200a, and miR-26a levels in milk. *Mol Nutr Food Res.* 2019; 63(8), 1800928. DOI [10.1002/mnfr.201800928](https://doi.org/10.1002/mnfr.201800928).
58. Thibeau S, D'Apolito K, Minnick AF, et al. Relationships of maternal stress with milk immune components in African American mothers of healthy term infants. *Breastfeed Med.* 2016; 11(1), 6–14. DOI [10.1089/bfm.2015.0117](https://doi.org/10.1089/bfm.2015.0117).
59. Chen DC, Nommsen-Rivers L, Dewey KG, Lönnerdal B. Stress during labor and delivery and early lactation performance. *Am J Clin Nutr.* 1998; 68(2), 335–344. DOI [10.1093/ajcn/68.2.335](https://doi.org/10.1093/ajcn/68.2.335).
60. Denić M, Sunarić S, Genčić M, et al. Maternal age has more pronounced effect on breast milk retinol and β -carotene content than maternal dietary pattern. *Nutrition.* 2019; 65(suppl 1), 120–125. DOI [10.1016/j.NUT.2019.02.019](https://doi.org/10.1016/j.NUT.2019.02.019).
61. Butts C, Hedderley D, Herath T, et al. Human milk composition and dietary intakes of breastfeeding women of different ethnicity from the Manawatu-Wanganui region of New Zealand. *Nutrients.* 2018; 10(9), 1231. DOI [10.3390/NU10091231](https://doi.org/10.3390/NU10091231).
62. Tricco AC, Lillie E, Zarin W, et al. PRISMA extension for scoping reviews (PRISMA-ScR): checklist and explanation. *Ann Intern Med.* 2018; 169(7), 467–473. DOI [10.7326/M18-0850](https://doi.org/10.7326/M18-0850).
63. Edwards PD, Lavergne SG, McCaw LK, et al. Maternal effects in mammals: broadening our understanding of offspring programming. *Front Neuroendocrinol.* 2021; 62(2), 100924. DOI [10.1016/j.YFRNE.2021.100924](https://doi.org/10.1016/j.YFRNE.2021.100924).
64. Théry C, Witwer KW, Aikawa E, et al. Minimal information for studies of extracellular vesicles 2018 (MISEV2018): a position statement of the International Society for Extracellular Vesicles and update of the MISEV2014 guidelines. *J Extracell Vesicles.* 2018; 7(1). DOI [10.1080/20013078.2018.1535750](https://doi.org/10.1080/20013078.2018.1535750).
65. Xi Y, Jiang X, Li R, Chen M, Song W, Li X. The levels of human milk microRNAs and their association with maternal weight characteristics. *Eur J Clin Nutr.* 2016; 70(4), 445–449. DOI [10.1038/ejcn.2015.168](https://doi.org/10.1038/ejcn.2015.168).
66. Fujimori M, França EL, Morais TC, Fiorin V, de Abreu LC, Honório-França AC. Cytokine and adipokine are biofactors can act in blood and colostrum of obese mothers. *BioFactors.* 2016; 43(2), 243–250. DOI [10.1002/biof.1339](https://doi.org/10.1002/biof.1339).
67. Hernandez LL, Grayson BE, Yadav E, Seeley RJ, Horseman ND. High fat diet alters lactation outcomes: possible involvement of inflammatory and serotonergic pathways. *PLoS One.* 2011; 7(3), e32598. DOI [10.1371/journal.pone.0032598](https://doi.org/10.1371/journal.pone.0032598).
68. Liu J, Smith MG, Dobre MA, Ferguson JE. Maternal obesity and breastfeeding practices among white and black women. *Obesity.* 2010; 18(1), 175–182. DOI [10.1038/oby.2009.182](https://doi.org/10.1038/oby.2009.182).
69. Purcell RH, Sun B, Pass LL, Power ML, Moran TH, Tamashiro KLK. Maternal stress and high-fat diet effect on maternal behavior, milk composition, and pup ingestive behavior. *Physiol Behav.* 2011; 104(3), 474–479. DOI [10.1016/j.physbeh.2011.05.012](https://doi.org/10.1016/j.physbeh.2011.05.012).
70. Abuaiash S, Spinieli RL, McGowan PO. Perinatal high fat diet induces early activation of endocrine stress responsivity and anxiety-like behavior in neonates. *Psychoneuroendocrinology.* 2018; 98, 11–21. DOI [10.1016/j.psyneuen.2018.08.003](https://doi.org/10.1016/j.psyneuen.2018.08.003).
71. Champagne F, Diorio J, Sharma S, Meaney MJ. Naturally occurring variations in maternal behavior in the rat are associated with differences in

- estrogen-inducible central oxytocin receptors. *Proc Natl Acad Sci*. 2001; 98(22), 12736–12741. DOI [10.1073/PNAS.221224598](https://doi.org/10.1073/PNAS.221224598).
72. Myers M, Brunelli S, Squire J, Shindeldecker R, Hofer M. Maternal behavior of SHR rats and its relationship to offspring blood pressures. *Dev Psychobiol*. 1989; 22(1), 29–53. DOI [10.1002/DEV.420220104](https://doi.org/10.1002/DEV.420220104).
73. Buonfiglio DC, Ramos-Lobo AM, Freitas VM, et al. Obesity impairs lactation performance in mice by inducing prolactin resistance. *Sci Rep*. 2016; 6(1), 225. DOI [10.1038/srep22421](https://doi.org/10.1038/srep22421).
74. Larnkjær A, Ong KK, Carlsen EM, Ejlerskov KT, Mølgaard C, Michaelsen KF. The influence of maternal obesity and breastfeeding on infant appetite and growth-related hormone concentrations: the SKOT cohort studies. *Horm Res Paediatr*. 2018; 90(1), 28–38. DOI [10.1159/000490114](https://doi.org/10.1159/000490114).
75. Lagström H, Rautava S, Ollila H, et al. Associations between human milk oligosaccharides and growth in infancy and early childhood. *Am J Clin Nutr*. 2020; 111(4), 769–778. DOI [10.1093/ajcn/nqaa010](https://doi.org/10.1093/ajcn/nqaa010).
76. Martin L, Woo J, Geraghty S, et al. Adiponectin is present in human milk and is associated with maternal factors. *Am J Clin Nutr*. 2006; 83(5), 1106–1111. DOI [10.1093/AJCN/83.5.1106](https://doi.org/10.1093/AJCN/83.5.1106).
77. Wulster-Radcliffe M, Ajuwon K, Wang J, Christian J, Spurlock M. Adiponectin differentially regulates cytokines in porcine macrophages. *Biochem Biophys Res Commun*. 2004; 316(3), 924–929. DOI [10.1016/J.BBRC.2004.02.130](https://doi.org/10.1016/J.BBRC.2004.02.130).
78. Combs T, Berg A, Obici S, Scherer P, Rossetti L. Endogenous glucose production is inhibited by the adipose-derived protein Acrp30. *J Clin Invest*. 2001; 108(12), 1875–1881. DOI [10.1172/JCI14120](https://doi.org/10.1172/JCI14120).
79. Yu X, Rong SS, Sun X, et al. Associations of breast milk adiponectin, leptin, insulin and ghrelin with maternal characteristics and early infant growth: a longitudinal study. *Br J Nutr*. 2018; 120(12), 1380–1387. DOI [10.1017/S0007114518002933](https://doi.org/10.1017/S0007114518002933).
80. Savino F, Lupica M, Benetti S, Petrucci E, Liguori S, Cordero Di Montezemolo L. Adiponectin in breast milk: relation to serum adiponectin concentration in lactating mothers and their infants. *Acta Paediatr*. 2012; 101(10), 1058–1062. DOI [10.1111/J.1651-2227.2012.02744.X](https://doi.org/10.1111/J.1651-2227.2012.02744.X).
81. van Rossem L, Smit H, Lentjes E, et al. Does breast milk adiponectin affect BMI and cardio-metabolic markers in childhood? *Br J Nutr*. 2019; 121(8), 905–913. DOI [10.1017/S0007114519000266](https://doi.org/10.1017/S0007114519000266).
82. Aparicio M, Browne PD, Hechler C, et al. Human milk cortisol and immune factors over the first three postnatal months: relations to maternal psychosocial distress. *PLoS One*. 2020; 15(5), e0233554. DOI [10.1371/journal.pone.0233554](https://doi.org/10.1371/journal.pone.0233554).
83. Sullivan E, Hinde K, Mendoza S, Capitano J. Cortisol concentrations in the milk of rhesus monkey mothers are associated with confident temperament in sons, but not daughters. *Dev Psychobiol*. 2011; 53(1), 96–104. DOI [10.1038/jid.2014.371](https://doi.org/10.1038/jid.2014.371).
84. Dettmer AM, Murphy AM, Guitarra D, et al. Cortisol in neonatal mother's milk predicts later infant social and cognitive functioning in rhesus monkeys. *Child Dev*. 2018; 89(2), 525–538. DOI [10.1111/cdev.12783](https://doi.org/10.1111/cdev.12783).
85. Hinde K, Skibiell AL, Foster AB, Del Rosso L, Mendoza SP, Capitano JP. Cortisol in mother's milk across lactation reflects maternal life history and predicts infant temperament. *Behav Ecol*. 2015; 26(1), 269–281. DOI [10.1093/beheco/aru186](https://doi.org/10.1093/beheco/aru186).
86. Hahn-Holbrook J, Le TB, Chung A, Davis EP, Glynn LM. Cortisol in human milk predicts child BMI. *Obesity*. 2016; 24(12), 2471–2474. DOI [10.1002/oby.21682](https://doi.org/10.1002/oby.21682).
87. Grey KR, Davis EP, Sandman CA, Glynn LM. Human milk cortisol is associated with infant temperament. *Psychoneuroendocrinology*. 2013; 38(7), 1178–1185. DOI [10.1016/j.psyneuen.2012.11.002](https://doi.org/10.1016/j.psyneuen.2012.11.002).
88. Nolvi S, Uusitupa H-M, Bridgett DJ, et al. Human milk cortisol concentration predicts experimentally induced infant fear reactivity: moderation by infant sex. *Dev Sci*. 2018; 21(4), 12625. DOI [10.1111/desc.12625](https://doi.org/10.1111/desc.12625).
89. Groer M, Davis M, Steele K. Associations between human milk SIgA and maternal immune, infectious, endocrine, and stress variables. *J Hum Lact*. 2004; 20(2), 153–158. DOI [10.1177/0890334404264104](https://doi.org/10.1177/0890334404264104).
90. Bunnay PE, Zink AN, Holm AA, Billington CJ, Kotz CM. Orexin activation counteracts decreases in nonexercise activity thermogenesis (NEAT) caused by high-fat diet. *Physiol Behav*. 2017; 176(12), 139–148. DOI [10.1002/ajp.22544](https://doi.org/10.1002/ajp.22544).
91. Carney MC, Tarasiuk A, Diangelo SL, et al. Metabolism-related microRNAs in maternal breast milk are influenced by premature delivery. *Pediatr Res*. 2017; 82(2), 226–236. DOI [10.1038/pr.2017.54](https://doi.org/10.1038/pr.2017.54).
92. Gay MCL, Koleva PT, Slupsky CM, et al. Worldwide variation in human milk metabolome: indicators of breast physiology and maternal lifestyle? *Nutrients*. 2018; 10(9), 1151. DOI [10.3390/nu10091151](https://doi.org/10.3390/nu10091151).
93. Su LL, Thamarai Chelvi SK, Lim SL, et al. The influence of maternal ethnic group and diet on breast milk fatty acid composition. *Ann Acad Med Singapore*. 2010; 39(9), 675–679.