

Outbreaks of non-O157 Shiga toxin-producing *Escherichia coli* infection: USA

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SUMMARY

Non-O157 Shiga toxin-producing *Escherichia coli* (STEC) infections are increasingly detected, but sources are not well established. We summarize outbreaks to 2010 in the USA. Single-aetiology outbreaks were defined as ≥ 2 epidemiologically linked culture-confirmed non-O157 STEC infections; multiple-aetiology outbreaks also had laboratory evidence of ≥ 2 infections caused by another enteric pathogen. Twenty-six states reported 46 outbreaks with 1727 illnesses and 144 hospitalizations. Of 38 single-aetiology outbreaks, 66% were caused by STEC O111 ($n=14$) or O26 ($n=11$), and 84% were transmitted through food ($n=17$) or person-to-person spread ($n=15$); food vehicles included dairy products, produce, and meats; childcare centres were the most common setting for person-to-person spread. Of single-aetiology outbreaks, a greater percentage of persons infected by Shiga toxin 2-positive strains had haemolytic uraemic syndrome compared with persons infected by Shiga toxin 1-only positive strains (7% vs. 0·8%). Compared with single-aetiology outbreaks, multiple-aetiology outbreaks were more frequently transmitted through water or animal contact.

Key words: Diarrhoea, outbreaks, Shiga-like toxin-producing *E. coli*.

INTRODUCTION

Shiga toxin-producing *Escherichia coli* (STEC) infections cause illness ranging from mild diarrhoea to haemorrhagic colitis and life-threatening haemolytic uraemic syndrome (HUS) [1]. In the USA, the serogroup most frequently isolated, most strongly associated with HUS, and responsible for the most outbreaks is O157 [2, 3]. Over 50 non-O157 STEC serogroups also cause illness [4]. Increasing use of Shiga toxin assays by clinical laboratories in recent years has resulted in increased detection of these

infections [5, 6]. In 2010, for the first time, non-O157 STEC infections detected through active sentinel surveillance outnumbered O157 STEC infections [7].

To assess modes of transmission, we describe epidemiological characteristics of all outbreaks of non-O157 STEC infection reported in the USA to 2010. In addition, we assess how outbreak characteristics vary by the presence of STEC virulence factors.

METHODS

Outbreak reports and definitions

An outbreak was defined as epidemiologically linked illnesses (clustered in time or space) with ≥ 2 persons having culture-confirmed non-O157 STEC infection of

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the same serogroup. Single-aetiology outbreaks were defined as isolation of only one serogroup. Multiple-aetiology outbreaks were defined as isolation of only one serogroup of non-O157 STEC from ≥ 2 ill persons and laboratory evidence of another enteric pathogen in ≥ 2 ill persons. Another enteric pathogen was defined as a pathogen other than non-O157 STEC, or another serogroup of non-O157 STEC, or the same non-O157 STEC serogroup that differed by ≥ 4 bands by pulsed-field gel electrophoresis (PFGE) [8].

Since the 1970s, state and local health departments have voluntarily reported waterborne and foodborne disease outbreaks to designated surveillance systems at the Centers for Disease Control and Prevention (CDC) [9, 10]. Before 2009, states informally reported outbreaks of enteric illness transmitted through contact with animals or ill persons and outbreaks with unknown transmission mode; in 2009, states began reporting these outbreaks through the National Outbreak Reporting System (NORS).

We used four methods to identify outbreaks: (1) queries of the Waterborne Disease Outbreak Surveillance System for outbreaks during 1971–2010, the Foodborne Disease Outbreak Surveillance System (FDOSS) for outbreaks during 1973–2010, and NORS for outbreaks transmitted by contact with animals or persons or by unknown mode during 2009–2010, (2) review of informally reported outbreaks that occurred before 2009, (3) direct contact with all state health departments to ascertain unreported outbreaks, and (4) searches of Medline and health department websites. When discrepancies were found between these sources, we used information from published reports or, if available, updated information from state health departments.

Information collected included setting of exposure, date of first illness, state(s) of exposure, pathogens detected, number of illnesses (laboratory-confirmed and clinically compatible cases), mode of transmission, food vehicle (if applicable), demographic characteristics, and frequency of various symptoms, hospitalization, reported HUS, and death.

We defined outbreak onset as onset date of the first illness. For foodborne outbreaks in which exposures were not limited to a single event or location, setting was classified as community. Most outbreaks were classified into one of four transmission modes: foodborne, waterborne, person-to-person, and animal contact. Outbreaks in childcare centres with no reported mode of transmission were classified as person-to-person. An outbreak was classified as foodborne if a

food vehicle was reported, or if there was exposure to a common meal or food establishment. Because outbreak surveillance does not collect laboratory values necessary to confirm HUS, we defined HUS as illnesses reported as HUS by state health departments; it is possible that some of the reported cases did not meet widely accepted HUS case definitions [11].

We defined reported food vehicles as foods implicated through either microbiological or epidemiological evidence and reported to CDC or published in Medline listed reports. If a food contained a single contaminated ingredient or all ingredients belonged to one of 17 defined commodities, the food was classified as that one commodity [12]. Outbreaks that could not be assigned to one commodity, or for which the report contained insufficient information for commodity assignment, were not attributed to any commodity. We defined outbreaks with exposure to the implicated vehicle in more than one state as multi-state.

Isolate characterization

We reviewed CDC records for isolates from outbreaks and identified additional isolates by providing lists to states of isolates characterized at CDC to see if any were from patients in an outbreak. For outbreaks for which CDC had no isolate, we contacted state health department laboratories to collect information on strains characterized there.

CDC's National *E. coli* Reference Laboratory serotyped non-O157 STEC isolates and tested for virulence genes *stx*₁ and *stx*₂ (encode Shiga toxins 1 and 2), *eae* (encodes intimin), and the putative virulence factor, *ehxA* (encodes enterohaemolysin), as described previously [13]. Different methods may have been used at state public health laboratories, but results were accepted irrespective of methodology.

Statistical analysis

We compared mode of transmission of single-aetiology and multiple-aetiology outbreaks and characteristics of single-aetiology outbreaks by mode of transmission and by isolate virulence factors using the χ^2 or Fisher's exact (if an expected cell size was <5) tests for categorical variables and the Kruskal–Wallis test for continuous variables.

RESULTS

We identified 46 outbreaks involving non-O157 STEC infection that caused 1727 illnesses (477



Fig. 1. Outbreaks of non-O157 STEC infection, by state to 2010.

laboratory-confirmed), 144 hospitalizations, and one death in 26 states (Fig. 1). The first outbreak was reported in 1990 and the number reported increased after 1998. Most (59%) occurred during 2007–2010 (Fig. 2).

Single-aetiology outbreaks

Of 46 outbreaks, 38 (83%) were of single-aetiology, which accounted for 886 (51%) illnesses. Of patients with recorded information, 112 (13%) of 859 were hospitalized, 289 (36%) of 836 reported bloody stool and 39 (4%) of 875 had HUS. One death occurred in a STEC O111 outbreak. Serogroups O111 (14 outbreaks) and O26 ($n=11$) accounted for 66% of outbreaks, followed by O45 ($n=4$), O103 ($n=2$), O121 ($n=2$), O145 ($n=2$), O104 ($n=1$), O165 ($n=1$), and O undetermined ($n=1$) (Table 1). Most outbreaks (58%) began during June–September. The median outbreak size was 7.5 illnesses (range 2–344). Outbreaks occurred throughout the country (Fig. 1). There were three multistate outbreaks, all in 2010.

Foodborne

Seventeen (45%) outbreaks were foodborne. The median size was 18 illnesses (range 2–344), which

was significantly larger than single-aetiology outbreaks with other modes of transmission (five illnesses, range 2–45, $P=0.02$). Foodborne outbreaks accounted for 36 (92%) of all HUS cases.

A food vehicle was reported for 12 outbreaks and 10 could be classified into commodities: dairy ($n=3$), leafy vegetables ($n=2$), game meat ($n=2$), beef ($n=1$), pork ($n=1$), and fruits or nuts ($n=1$). In one of the remaining two outbreaks the vehicle was unclassifiable (water-based punch); in the other, multiple food vehicles including lettuce-containing salads, corn, dinner rolls, and ice, were implicated. Two multistate outbreaks led to food recalls: STEC O145 in Romaine lettuce and STEC O26 in ground beef.

Two of five outbreaks with no identified vehicle, including the largest outbreak of non-O157 STEC infections reported in the USA, reported ill food workers as a possible contributing factor [14]. Last, in the remaining three outbreaks, ground beef was suspected by states as a possible vehicle on the basis of common food exposures at a catered meal, reported undercooking of ground beef at a barbecue, or tracebacks of ground beef to the same source; however, the states did not feel evidence was sufficient to report ground beef as a vehicle.

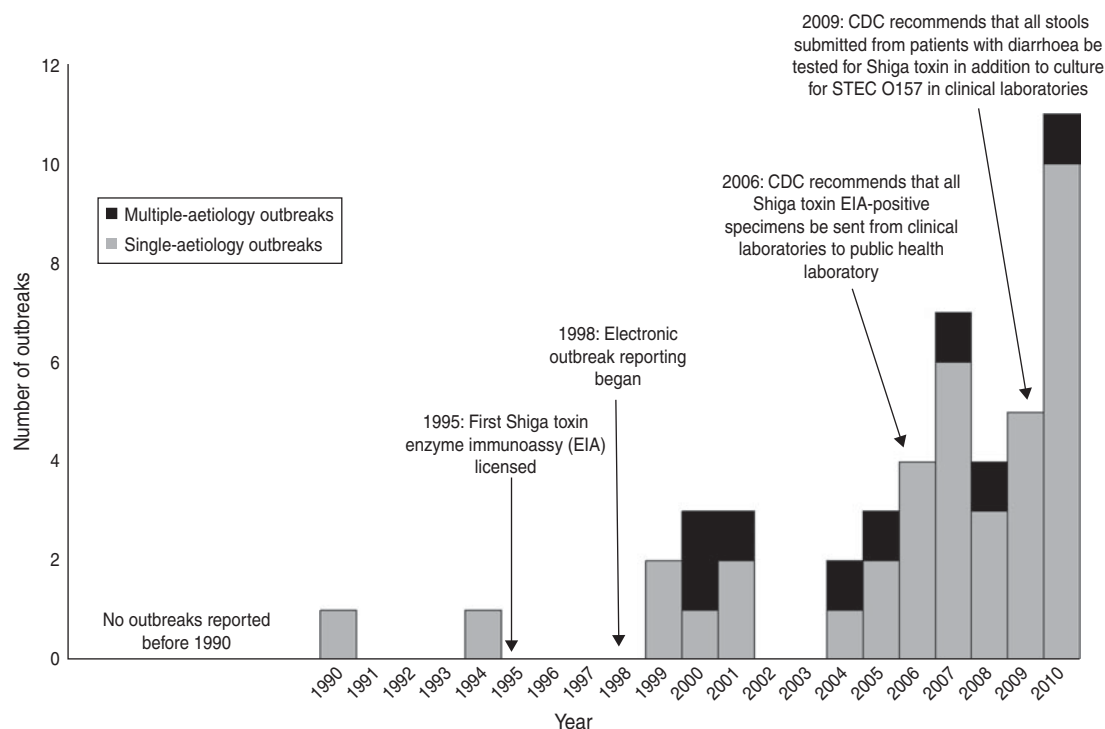


Fig. 2. Number of outbreaks of non-O157 STEC infection, by year to 2010.

Person-to-person

Illnesses in 15 (39%) outbreaks were transmitted from one person to another. The median size was four illnesses (range 2–45). Thirteen (87%) occurred in child-care centres. One outbreak occurred in a home setting and another in an elementary school. Two outbreaks (caused by serotypes O111:non-motile and O111:H8) resulted in two HUS cases.

Other transmission modes

Four outbreaks were attributed to water ($n=1$), animal contact ($n=1$), or an unknown mode ($n=2$). The remaining two outbreaks were propagated through contact with both animals and ill persons. In a STEC O45 outbreak, the index patient visited a goat farm and then attended daycare facilities where other children became ill; other persons who only visited the goats also became ill. In a STEC O111 outbreak in a correctional facility, a subset of ill inmates worked directly with cattle at a dairy and probably spread illness to others.

Multiple-aetiology outbreaks

Eight (17%) of the 46 outbreaks were multiple-aetiology, accounting for 841 (49%) illnesses. Of patients with recorded information, 32 (4%) of 797

were hospitalized and 150 (19%) of 788 reported bloody stool; none developed HUS or died. Two outbreaks were associated with multiple non-O157 STEC strains: one with >1 serogroup and one with a single serogroup displaying two PFGE patterns. The most commonly isolated serogroup was O111 (4 outbreaks), followed by O26 ($n=2$), with one outbreak each for O69, O84, O121, O141, O145, and O undetermined. The proportion of patients with culture-confirmed non-O157 STEC infections was generally small relative to those with other laboratory-confirmed infections (Table 2). Other pathogens detected in six outbreaks were *Cryptosporidium* ($n=3$), STEC O157:H7 ($n=3$), *Campylobacter* ($n=3$), *Shigella* ($n=1$), *Salmonella* serotype Typhimurium ($n=1$), and norovirus ($n=1$). The most frequent pathogen pairings were STEC O111 and *Cryptosporidium* ($n=3$), and STEC O111 and *Campylobacter* ($n=3$). Most (75%) multiple-aetiology outbreaks occurred during June–September.

Three multiple-aetiology outbreaks were associated with food; the vehicle in one was unpasteurized apple cider. In the two remaining outbreaks ill food workers were reported as a possible contributing factor: one additionally implicated two dairy products.

The other modes of transmission in multiple-aetiology outbreaks were animal contact ($n=2$),

Table 1. *Single-aetiology outbreak of non-O157 STEC, 1990–2010*

Year	Month	State (reference)	Serotype	<i>stx</i> ₁	<i>stx</i> ₂	<i>eae</i>	<i>ehxA</i>	Mode of transmission	Vehicle or animal	Setting	Total ill	HUS reported (no. cases)
1990	Apr.	Ohio [30]	O111:NM	+	+	+	+	Unknown		Home	5	Yes (1)
1994	Feb.	Montana [31]	O104:H21	–	+	–	+	Food	Pasteurized milk	Community	18	No
1999	June	Texas [32]	O111:H8	+	+	+	+	Food	Multiple foods ^a	Camp	55	Yes (2)
1999	Dec.	Minnesota	O145:NM	–	+	+	+	P-to-P		Childcare	2	No
2000	July	Washington	O103:H2	+	–	+	+	Food	Water-based punch	Event ^b	18	Yes (2)
2001	Aug.	Minnesota	O26:NM	+	+	+	+	Water	Recreational lake water	Lake	4	No
2001	Sep.	South Dakota	O111:NM	+	+	+	+	P-to-P		Childcare	3	No
2004	Mar.	Maryland ^c	OUnd:H8	+	–	+	+	P-to-P		Childcare	4	No
2005	Aug.	New York [33]	O45:NM	+	–	+	+	Food	Unknown; ill food worker(s)	Correctional facility	52	No
2005	Oct.	Nevada	O26:H11	+	–	+	+	P-to-P		Childcare	12	No
2006	Apr.	North Carolina	O45:H2	+	–	+	+	Animal & P-to-P	Goats	Farm and childcare	11	No
2006	June	Oregon	O165:NM	–	+	+	+	Unknown		Correctional facility	3	No
2006	July	Utah	O121:H19	–	+	+	+	Food	Lettuce-based salad	Event	42	Yes (3)
2006	July	Massachusetts	O26:H11	+	–	+	+	Food	Blueberries, or strawberries, or both	Farm	5	No
2007	Mar.	Maine ^d	O111:NM	U	U	U	U	P-to-P		Childcare	8	No
2007	Apr.	North Dakota	O111:NM	+	+	+	+	P-to-P		School	6	No
2007	June	Iowa	O26:H11	+	–	+	+	P-to-P		Childcare	3	No
2007	June	Colorado	O111:NM	+	+	+	+	P-to-P		Home	3	No
2007	July	North Dakota	O111:NM	+	+	+	+	Food	Unknown ^e	Event	23	No
2007	Oct.	New Hampshire	O45	U	U	U	U	Animal	No specific animal implicated	Fair	5	No
2008	July	Nebraska	O111	U	U	U	U	Food	Barbecued pork	Event	34	No
2008	Aug.	Oklahoma [14]	O111:NM	+	+	+	+	Food	Unknown; ill food worker(s)	Restaurant	344	Yes (25)
2008	Sep.	Minnesota	O111:NM	+	–	+	+	P-to-P		Childcare	3	No
2009	May	Wisconsin ^f	O26:H11	+	–	+	+	Food	Unknown	Event	8	No
2009	July	South Dakota	O111:H8	+	+	+	+	P-to-P		Childcare	13	Yes (1)
2009	July	Wyoming ^g	O111:NM	+	+	+	+	Food	Unknown	Community	2	No
2009	Aug.	California	O26:H11	+	+	U	U	P-to-P		Childcare	3	No
2009	Sep.	Washington ^h	O121:H19	–	+	+	+	Food	Unpasteurized milk			
2010	Feb.	Washington	O26:H11	+	–	+	+	Food	Unpasteurized milk	Community	6	No
2010	Apr.	Multiple	O145:NM	–	+	+	+	Food	Romaine lettuce	Community	45	Yes (4)
2010	Apr.	Colorado [34]	O111:NM	+	–	+	+	Animal & P-to-P	Cattle	Correctional facility	24	No
2010	May	Colorado [35]	O26:H11	+	–	+	+	P-to-P		Childcare	45	No
2010	June	Multiple	O26:NM	+	–	+	+	Food	Ground beef	Community	3	No

Table 1 (cont.)

Year	Month	State (reference)	Serotype	<i>stx</i> ₁	<i>stx</i> ₂	<i>eae</i>	<i>ehxA</i>	Mode of transmission	Vehicle or animal	Setting	Total ill	HUS reported (no. cases)
2010	Aug.	Colorado	O26:H11	+	-	+	+	P-to-P		Childcare	21	No
2010	Sep.	Tennessee	O111:NM	+	+	+	+	P-to-P		Childcare	3	Yes (1)
2010	Sep.	Multiple	O45:H2	+	-	+	+	Food	Smoked game meat	Community	7	No
2010	Oct.	Oregon	O26:H11	+	-	+	+	P-to-P		Childcare	11	No
2010	Nov.	Minnesota ¹ [36]	O103:H2	+	-	+	+	Food	Venison	School	29	No

STEC, Shiga toxin-producing *Escherichia coli*; HUS, haemolytic uraemic syndrome; NM, non-motile; OUnd, O undetermined; P-to-P, person-to-person; U, unknown.

^a Cob corn, food from salad bar, dinner roll, and ice.

^b Events were banquets, wedding receptions, catered events, customer appreciation days, and family outings.

^c Rotavirus was isolated from one patient.

^d One patient tested positive for *Cryptosporidium*.

^e The reporting state suspected ground beef as a vehicle on the basis of common food exposures at a catered meal.

^f The reporting state suspected ground beef as a vehicle on the basis of reported consumption of undercooked ground beef.

^g The reporting state suspected ground beef as a vehicle on the basis of tracebacks of consumed ground beef to the same source.

^h *E. coli* O157:H7 was isolated from one patient.

ⁱ Additional patients had *E. coli* O145:NM isolated; however, neither isolate had Shiga toxin genes. One patient had norovirus identified.

water ($n=2$), and person-to-person ($n=1$). Both outbreaks transmitted through animal contact or water occurred in camp settings.

Compared with single-aetiology outbreaks, multiple-aetiology outbreaks had more illnesses transmitted through water or animal contact, and more illnesses were seen in children aged 5–19 years (Table 3).

Virulence factor characterization

Shiga toxin information was available for 35/38 single-aetiology outbreak strains and for 11 strains from 7/8 multiple-aetiology outbreaks (Tables 1 and 2). All but one strain in each group (single-aetiology and multiple-aetiology outbreaks), for which information was available, was *eae* positive and all were *ehxA* positive.

Outbreaks with *stx*₂-positive strains (with or without *stx*₁) were significantly more likely to have HUS cases reported than outbreaks with *stx*₁-only positive strains (32% vs. 4% of outbreaks, $P=0.02$). In an analysis of single-aetiology outbreaks, 7% of patients in outbreaks with a *stx*₂-positive strain (with or without *stx*₁) developed HUS compared with 0.8% of patients in outbreaks with a *stx*₁-only positive strain ($P<0.001$).

DISCUSSION

Non-O157 STEC strains are increasingly recognized as causes of enteric disease outbreaks in the USA. Of the 38 outbreaks we identified, none occurred before 1990 and over half occurred during 2007–2010. Most outbreaks were transmitted through contaminated food or from one person to another. Nearly one-fifth of all outbreaks involved more than one pathogen, and these multiple-aetiology outbreaks were epidemiologically different from single-aetiology outbreaks. Over half of all outbreaks involved STEC serogroups O111 or O26.

The increasing detection of non-O157 STEC outbreaks coincides with a marked rise in reporting of sporadic non-O157 STEC infections, which is related to the increasing use of enzyme immunoassays or polymerase chain reaction by clinical laboratories to detect Shiga toxin (or the genes that encode them) in the stool of patients with diarrhoea [5, 6]. These tests, first licensed in 1995, are currently the only widely available, practical means of detecting non-O157 STEC in clinical specimens.

Table 2. Multiple-aetiology outbreaks in which non-O157 STEC were isolated, USA, 1990–2010

Year	Month	State (reference)	Serotype(s)	<i>stx</i> ₁	<i>stx</i> ₂	<i>eae</i>	<i>ehxA</i>	Other pathogen(s) ^a	Total ill	Mode of transmission	Vehicle or animal	Setting
2000	June	Minnesota [37]	O111:H8 (1) O111:NM (1)	+ +	– –	+ +	+ +	<i>Cryptosporidium</i> (9), <i>Campylobacter</i> (1), <i>Salmonella</i> serotype Typhimurium (1)	61	Animal contact	Calves	Camp
2000	Aug.	Utah	O111:NM (3)	+	+	+	+	<i>E. coli</i> O157:H7 (6), <i>Campylobacter</i> (51), <i>Shigella</i> (1)	126	Water	Irrigation water	Camp
2001	June	Minnesota [37]	O111:NM (2) O69:H11 (2) ^b OUnd:HU (1)	+ + U	– – U	+ + U	+ + U	<i>E. coli</i> O157:H7 (4), <i>Cryptosporidium</i> (8)	31	Animal contact	Calves/cattle	Camp
2004	Sep.	New York	O111:NM (25)	+	–	+	+	<i>Cryptosporidium</i> (42)	213	Food	Unpasteurized apple cider	Orchard
2005	May	Oregon [9]	O145:NM (2)	U	U	U	U	<i>Campylobacter</i> (6), <i>E. coli</i> O157:H7 (12)	60	Water	Drinking water	Camp
2007	July	Colorado	O121:H19 (35) O84:H2 (1) O26:H11(4)	– + +	+ – –	+ + +	+ + +		135	Food	Margarine, pasteurized American cheese, ill foodworker(s)	Correctional facility
2008	Mar.	Maryland	O141:H49 (2)	–	+	–	+	Norovirus (9)	191	Food	Unknown, ill foodworker(s)	Catered event
2010	June	Washington	O26:NM (5) O26:H11 (8) ^c	+ +	+ –	+ +	+ +		24	P-to-P		Childcare

STEC, Shiga toxin-producing *Escherichia coli*; NM, non-motile; Und, undetermined; U, unknown; P-to-P, person-to-person.

^a Number isolated from patients, not mutually exclusive.

^b These specimens were originally reported as O rough:H11 in the reference and it was concluded that they were O51:H11 for which the O51 antigen could not be identified. These specimens were retested at CDC in 2011 and were determined to be O69:H11.

^c PFGE patterns differed by four bands.

Table 3. Transmission, demographic, and clinical characteristics of patients in single- and multiple-aetiology outbreaks^a

	Single-aetiology outbreak illnesses (<i>n</i> = 886)		Multiple-aetiology outbreak illnesses (<i>n</i> = 841)		<i>P</i> value
	No.	(%)	No.	(%)	
Mode of transmission^b	<i>(n</i> = 886)		<i>(n</i> = 841)		
Person-to-person	140	(16)	24	(3)	
Food	694	(78)	539	(64)	
Water	4	(0.5)	186	(22)	<0.001
Animal	5	(0.6)	92	(11)	
Unknown	8	(1)	0	(0)	
Mixed ^c	35	(4)	0	(0)	
Demographic characteristics					
Female ^d	472/842	(56)	378/837	(45)	<0.001
Age distribution^b	<i>(n</i> = 781)		<i>(n</i> = 752)		
<1 year	13	(2)	1	(0.2)	
1–4 years	104	(13)	29	(4)	
5–19 years	146	(19)	276	(37)	<0.001
20–49 years	297	(38)	361	(48)	
≥ 50 years	221	(28)	85	(11)	

^a All *P* values based on the χ^2 test unless otherwise noted.

^b Fisher's exact test.

^c Mixed modes of transmission were defined as outbreaks with ≥ 1 mode of transmission. Two outbreaks met this criterion. Both involved contact with animals and person-to-person contact.

^d Gender differences ceased to be significant when four outbreaks of ≥ 50 illnesses in which $\geq 85\%$ of patients are of one gender were excluded: a STEC O111 outbreak at a cheerleading camp (55 illnesses, 96% female), a STEC O45 outbreak at a correctional facility (52 illnesses, 100% male), an outbreak involving STEC O121:H19, O26:H11, and O84:H2 infections at a correctional facility (135 illnesses, 85% male), and an outbreak involving STEC O111, *Campylobacter*, *Shigella*, and STEC O157:H7 at a football camp (126 illnesses, 94% male). Removing these outbreaks increases the percentage of female patients to 57% and 61% in single-aetiology and multiple-aetiology outbreaks, respectively (*P* = 0.14).

Although non-O157 STEC are estimated to cause nearly twice as many infections as STEC O157 in the USA [15] the number of detected outbreaks caused by non-O157 STEC is substantially less. This is probably due in part to more severe illness in persons with STEC O157 than in the broad group of non-O157 STEC infections [6]. Moreover, clinical laboratories can more readily identify STEC O157. Unlike for STEC O157, few states were routinely subtyping non-O157 STEC isolates by PFGE until the past 5 years; submitting PFGE patterns of subtyped isolates to PulseNet facilitates detection of outbreaks. As more clinical laboratories test for Shiga toxin and more non-O157 STEC infections are reported, we can expect greater detection of non-O157 STEC outbreaks.

The number of outbreaks we describe does not include all those reported by state health departments. Our case definition (at least two epidemiologically linked culture-confirmed cases) excluded seven outbreaks [16]. In one, non-O157 STEC were isolated from two persons but the serogroups differed. In the

other six, only one case was culture-confirmed (four with STEC O121, two with O111). Three included reports of HUS, including one published as an O121 outbreak [17].

The proportion of single-aetiology non-O157 STEC outbreaks attributed to food (43%) and, especially, person-to-person transmission (35%) differs from that found for US STEC O157 outbreaks during 1982–2002 (52% foodborne, 14% person-to-person) [2]. These differences may relate to past challenges in detecting non-O157 STEC infections and, consequently, limited detection of outbreaks via laboratory-based surveillance. A greater portion of non-O157 STEC outbreaks may have been detected because patients or physicians contacted health departments after noticing a localized cluster of illness. Our findings suggest this may be especially true for outbreaks in childcare centres, the setting for nearly 90% of outbreaks due to person-to-person transmission, and camps, the setting of half of multiple-aetiology outbreaks. Person-to-person trans-

mission also accounts for a considerable number of the non-O157 STEC outbreaks reported from other countries [18, 19]. Likewise, the historical effect of limited laboratory surveillance of non-O157 STEC would probably result in disproportionately under-detecting foodborne outbreaks. Without sensitive surveillance, contaminated commercially distributed products that cause widely dispersed infections may go unnoticed. All three multistate outbreaks of non-O157 STEC infection were foodborne and detected through the combination of appropriate testing in clinical laboratories and PulseNet surveillance.

Of the single-aetiology foodborne disease outbreaks, vehicles included those implicated in STEC O157 outbreaks in the USA, including leafy vegetables, dairy, game meat, and ground beef [2]. Of reported non-O157 STEC foodborne outbreaks in other countries, implicated foods have also included beef and dairy products [20–23]. We are unaware of any reports from outside the USA that implicate leafy vegetables as a non-O157 STEC outbreak vehicle. However, contaminated raw sprouts caused a severe STEC O104:H4 outbreak in Europe [24]. The first reported non-O157 STEC outbreak in the USA associated with raw sprouts occurred in 2012 [25].

Environmental exposures play an important role in multiple-aetiology outbreaks. Exposures to cattle or contaminated water (for drinking, swimming, or irrigation) accounted for a greater proportion of illnesses in multiple-aetiology than in single-aetiology outbreaks. Half of all multiple-aetiology outbreaks occurred at camps, where children may have contact with animals and untreated water. This may explain the greater frequency of illnesses in children aged 5–19 years in multiple-aetiology outbreaks compared with single-aetiology outbreaks. Furthermore, the other pathogens most frequently identified in these outbreaks (*Cryptosporidium*, *E. coli* O157, *Campylobacter*) are carried by ruminant animals and have caused waterborne outbreaks. Similarly, animal contact may have caused multiple-aetiology outbreaks in Belgium and Australia [20, 26].

Outbreaks of infection with STEC strains that contain *stx*₂ or *eae* genes carry a greater risk of bloody diarrhoea or HUS [27] probably making them easier to detect. Enterohaemolysin (encoded by *ehxA*) is a putative virulence factor. A greater percentage of the 34 single-aetiology outbreak strains we identified carried *stx*₂ (50%), *eae* (97%), or *ehxA* (100%) than commonly seen in non-O157 STEC isolated from ill persons. In a USA convenience sample of >900

non-O157 STEC isolates from ill persons, 39% carried *stx*₂, 84% carried *eae* and 86% carried *ehxA* [13].

The clinical characteristics of infections caused by STEC of different serogroups may affect the number of outbreaks observed. Two serogroups were responsible for two-thirds of the 38 single-aetiology outbreaks, O111 (38%) and O26 (29%). The high frequency of STEC O26 outbreaks could simply reflect the high frequency of STEC O26 infections in the USA. From 2000 to 2010, 1708 non-O157 STEC infections with a known serogroup were reported to FoodNet; O26 accounted for 26%, followed by O103 (22%) and O111 (19%) [6]. Alternatively, O111 and O26 may predominate because of low infectious doses, facilitating person-to-person transmission [28]. Nearly 90% of person-to-person outbreaks were caused by these two serogroups. The high frequency of STEC O111 outbreaks may relate to the greater propensity of this serogroup to produce Stx₂ and cause severe illness compared with STEC O26 and STEC O103, thereby facilitating outbreak detection [3, 13]. Serogroup O111 caused 83% of all outbreaks with HUS cases.

The percentage of illnesses reported as being complicated by HUS in single-aetiology outbreaks was relatively high (4%). Only about 1% of non-O157 STEC infections in the USA lead to physician-diagnosed HUS compared with 11% of STEC O157 infections [6]. This higher than expected percentage of reported HUS in non-O157 STEC outbreaks suggests that outbreaks with more severe illnesses were more likely to be detected. Alternatively, reporting officials may not have applied appropriate criteria in reporting HUS cases. We were unable to verify that all reported HUS cases met widely accepted laboratory-defined HUS definitions because HUS-defining laboratory values including serum creatinine, haemoglobin (or haematocrit), and platelet count are not collected through outbreak surveillance. However, many of the reported cases of HUS (25/39, 64%) in our analysis were from a single outbreak of STEC O111 infections that did apply laboratory criteria to define HUS [14].

Reports to the outbreak surveillance system include both laboratory-confirmed and other epidemiologically linked illnesses [16]. In this analysis, the percentage of reported infections that were not laboratory confirmed was relatively large (72%). Thus, for every laboratory-confirmed illness reported (28% of all illnesses) in the outbreaks described there were 3·6 non-confirmed illness reported. It is possible that some

illnesses were misclassified as being part of an outbreak. However, it is unlikely that inclusion of some non-confirmed cases would have resulted in considerable overestimation of outbreak size. Most people with diarrhoeal illness do not seek medical care and even fewer submit a stool specimen. It has been estimated that for every confirmed non-O157 STEC infection there may be as many as 107 undetected cases in the population [15].

Because of increasing use of assays that detect Shiga toxins in clinical laboratories, non-O157 STEC strains have increasingly been detected as causes of outbreaks in the USA during the past two decades. These infections are acquired through several routes of transmission. Stools of patients with community-acquired diarrhoea for which an aetiology is sought should be tested for Shiga toxin and all positive samples should be sent to public health laboratories for STEC isolation and characterization [29]. Such an approach can identify all pathogenic non-O157 STEC including important novel strains such as STEC O104:H4 and can improve our understanding of these important pathogens.

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DECLARATION OF INTEREST

None.

REFERENCES

1. Tarr PI, Gordon CA, Chandler WL. Shiga-toxin-producing *Escherichia coli* and haemolytic uraemic syndrome. *Lancet* 2005; **365**: 1073–1086.
2. Rangel JM, *et al.* Epidemiology of *Escherichia coli* O157:H7 outbreaks, United States, 1982–2002. *Emerging Infectious Diseases* 2005; **11**: 603–609.
3. Mody RK, *et al.* Infections in pediatric postdiarrheal hemolytic uremic syndrome: factors associated with identifying Shiga toxin-producing *Escherichia coli*. *Archives of Pediatrics and Adolescent Medicine* 2012; **166**: 902–909.
4. Strockbine N, *et al.* Overview of detection and subtyping methods. In: Kaper J, O'Brien A, eds. *Escherichia coli O157:H7 and Other Shiga Toxin-Producing E. coli Strains*. Washington, DC: ASM Press, 1998, pp. 331–347.
5. Hoefler D, *et al.* Laboratory practices for the identification of Shiga toxin-producing *Escherichia coli* in the United States, FoodNet Sites, 2007. *Foodborne Pathogens and Disease* 2011; **8**: 555–560.
6. Gould LH, *et al.* Increased recognition of non-O157 Shiga toxin-producing *Escherichia coli* infections in the United States During 2000–2010: epidemiologic features and comparison with *E. coli* O157 infections. *Foodborne Pathogens and Disease* 2013; **10**: 453–460.
7. Centers for Disease Control and Prevention. Vital Signs: incidence and trends of infection with pathogens transmitted commonly through food—Foodborne Diseases Active Surveillance Network, 10 U.S. Sites, 1996–2010. *Morbidity and Mortality Weekly Report* 2011; **60**: 749–755.
8. Tenover FC, *et al.* Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *Journal of Clinical Microbiology* 1995; **33**: 2233–2239.
9. Yoder J, *et al.* Surveillance for waterborne disease and outbreaks associated with drinking water and water not intended for drinking—United States, 2005–2006. *Morbidity and Mortality Weekly Report. Surveillance Summaries* 2008; **57**: 39–62.
10. Bean NH, *et al.* Surveillance for foodborne-disease outbreaks—United States, 1988–1992. *Morbidity and Mortality Weekly Report. Surveillance Summaries* 1996; **45**: 1–66.
11. Tarr PI, Karpman D. *Escherichia coli* O104:H4 and hemolytic uremic syndrome: the analysis begins [Editorial commentary]. *Clinical Infectious Diseases* 2012; **55**: 760–763.
12. Painter JA, *et al.* Recipes for foodborne outbreaks: a scheme for categorizing and grouping implicated foods. *Foodborne Pathogens and Disease* 2009; **6**: 1259–1264.
13. Brooks JT, *et al.* Non-O157 Shiga toxin-producing *Escherichia coli* infections in the United States, 1983–2002. *Journal of Infectious Diseases* 2005; **192**: 1422–1429.
14. Bradley KK, *et al.* Epidemiology of a large restaurant-associated outbreak of Shiga toxin-producing *Escherichia coli* O111:NM. *Epidemiology and Infection* 2012; **140**: 1644–1654.
15. Scallan E, *et al.* Foodborne illness acquired in the United States—major pathogens. *Emerging Infectious Diseases* 2011; **17**: 7–15.
16. NORS—National Outbreak Reporting System. Guidance document ([http://www.cdc.gov/outbreaknet/pdf/NORS_Guidance_5213_06232009\(compliant\).pdf](http://www.cdc.gov/outbreaknet/pdf/NORS_Guidance_5213_06232009(compliant).pdf)). Accessed 3 April 2013.
17. McCarthy TA, *et al.* Hemolytic-uremic syndrome and *Escherichia coli* O121 at a lake in Connecticut, 1999. *Pediatrics* 2001; **108**: E59.

18. **Boudailliez B, et al.** Possible person-to-person transmission of *Escherichia coli* O111 – associated hemolytic uremic syndrome. *Pediatric Nephrology* 1997; **11**: 36–39.
19. **Sonoda C, et al.** An enterohemorrhagic *Escherichia coli* O26 outbreak at a nursery school in Miyazaki, Japan. *Japanese Journal of Infectious Diseases* 2008; **61**: 92–93.
20. **De Schrijver K, et al.** Outbreak of verocytotoxin-producing *E. coli* O145 and O26 infections associated with the consumption of ice cream produced at a farm, Belgium, 2007. *Eurosurveillance* 2008; **13**: 1–7.
21. **Allerberger F, et al.** Hemolytic-uremic syndrome associated with enterohemorrhagic *Escherichia coli* O26:H infection and consumption of unpasteurized cow's milk. *Journal of Infectious Diseases* 2003; **7**: 42–45.
22. **Werber D, et al.** A multistate outbreak of Shiga toxin-producing *Escherichia coli* O26:H11 infections in Germany, detected by molecular subtyping surveillance. *Journal of Infectious Diseases* 2002; **186**: 419–422.
23. **Ethelberg S, et al.** Outbreak of non-O157 Shiga toxin-producing *Escherichia coli* infection from consumption of beef sausage. *Clinical Infectious Diseases* 2009; **48**: e78–81.
24. **Buchholz U, et al.** German outbreak of *Escherichia coli* O104:H4 associated with sprouts. *New England Journal of Medicine* 2011; **365**: 1763–1770.
25. **Centers for Disease Control and Prevention.** Multistate outbreak of Shiga toxin-producing *Escherichia coli* O26 infections linked to raw clover sprouts at Jimmy John's restaurants (<http://www.cdc.gov/ecoli/2012/o26-02-12/index.html>). Accessed 8 March 2012.
26. **Hanna JN, et al.** Haemolytic uraemic syndrome associated with a family cluster of enterohaemorrhagic *Escherichia coli*. *Communicable Diseases Intelligence* 2007; **31**: 300–303.
27. **Ethelberg S, et al.** Virulence factors for hemolytic uremic syndrome, Denmark. *Emerging Infectious Diseases* 2004; **10**: 842–847.
28. **Paton AW, et al.** Molecular microbiological investigation of an outbreak of hemolytic-uremic syndrome caused by dry fermented sausage contaminated with Shiga-like toxin-producing *Escherichia coli*. *Journal of Clinical Microbiology* 1996; **34b**: 1622–1627.
29. **Gould LH, et al.** Recommendations for diagnosis of Shiga toxin-producing *Escherichia coli* infections by clinical laboratories. *Morbidity and Mortality Weekly Report. Recommendations and Reports* 2009; **58** (RR-12): 1–14.
30. **Banatvala N, et al.** Shiga-like toxin-producing *Escherichia coli* O111 and associated hemolytic-uremic syndrome: a family outbreak. *Pediatric Infectious Disease Journal* 1996; **15**: 1008–1011.
31. **Centers for Disease Control and Prevention.** Outbreak of acute gastroenteritis attributable to *Escherichia coli* serotype O104:H21 – Helena, Montana, 1994. *Morbidity and Mortality Weekly Report* 1995; **44**: 501–503.
32. **Brooks JT, et al.** Outbreak of Shiga toxin-producing *Escherichia coli* O111:H8 infections among attendees of a high school cheerleading camp. *Clinical Infectious Diseases* 2004; **38**: 190–198.
33. **Centers for Disease Control and Prevention.** Importance of culture confirmation of Shiga toxin-producing *Escherichia coli* infection as illustrated by outbreaks of gastroenteritis – New York and North Carolina, 2005. *Morbidity and Mortality Weekly Report* 2006; **55**: 1042–1045.
34. **Centers for Disease Control and Prevention.** Outbreak of Shiga toxin-producing *Escherichia coli* O111 infections associated with a correctional facility dairy – Colorado, 2010. *Morbidity and Mortality Weekly Report* 2012; **61**: 149–152.
35. **Brown JA, et al.** Outbreak of Shiga toxin-producing *Escherichia coli* serotype O26: H11 infection at a child care center in Colorado. *Pediatric Infectious Disease Journal* 2012; **31**: 379–383.
36. **Rounds JM, et al.** Non-O157 Shiga toxin-producing *Escherichia coli* associated with venison. *Emerging Infectious Diseases* 2012; **18**: 279–282.
37. **Smith KE, et al.** Outbreaks of enteric infections caused by multiple pathogens associated with calves at a farm day camp. *Pediatric Infectious Disease Journal* 2004; **23**: 1098–1104.