reduce healthcare-associated infections in 2 Asia-Pacific countries.<sup>6,7</sup> These are essential prerequisites that hospitals should have in place to achieve successful MDRO IPC. In this study, we confirm that both hospital-level factors (ie, excellent safety culture and leadership support) and individual-level factors (ie, knowledge regarding implementation of intensified IC programs and involvement in collaborative networks) are important predictors for successful containment of MDRAB in hospitals with IDPs in Thailand.

We are aware of several limitations in this study. First, the small sample size limited our capacity to analyze other important factors potentially associated with successful containment of MDROs (eg, predictors for implementation of individual intervention and level of compliance needed to achieve successful containment of each intervention). Second, survey data were subject to recall bias associated with interventions that each IDP implemented. This bias was likely low given that all information was derived from hospital IC databases, including levels of compliance with each IPC intervention. Nonetheless, our study highlights some important modifiable gaps in MDRO containment. Further education regarding implementation of intensified IC interventions, along with sustainable IDP networks, is needed to contain the increase in MDRAB prevalence in this middle-income country.

#### ACKNOWLEDGMENTS

*Financial support.* No financial support was provided relevant to this article. *Potential conflicts of interest.* A.A. was supported by the National Research University Project of the Thailand Office of Higher Education Commission. L.M.M. conducted this work pro bono and independent of her employment at Luitpold Pharmaceuticals, Inc. (LPI).

## Anucha Apisarnthanarak, MD;<sup>1</sup> Aubonphan Buppajarntham, MD;<sup>1</sup> Linda M. Mundy, MD, PhD<sup>2</sup>

Affiliations: 1. Division of Infectious Diseases, Faculty of Medicine, Thammasat University, Pathumthani, Thailand; 2. Luitpold Pharmaceuticals, Inc., Norristown, Pennsylvania, United States.

Address correspondence to Anucha Apisarnthanarak, MD, Division of Infectious Diseases, Faculty of Medicine, Thammasat University, Pathumthani, Thailand, 10120 (anapisarn@yahoo.com).

Infect. Control Hosp. Epidemiol. 2016;37(4):489–491

© 2016 by The Society for Healthcare Epidemiology of America. All rights reserved. 0899-823X/2016/3704-0023. DOI: 10.1017/ice.2015.343

### REFERENCES

- Dejsirilert S, Tiengrim S, Sawanpanyalert P, Aswapokee N, Malathum K. Antimicrobial resistance of *Acinetobacter baumannii*: six years of National Antimicrobial Resistance Surveillance Thailand (NARST) surveillance. *J Med Assoc Thai* 2009;91(Suppl4):S34–S45.
- Apisarnthanarak A, Buppunharun W, Tiengrim S, Sawanpanyalert P, Aswapokee N. An overview of antimicrobial susceptibility patterns for gram-negative bacteria from the National

Antimicrobial Resistance Surveillance Thailand (NARST) program from 2000 to 2005. *J Med Assoc Thai* 2009;92(Suppl4): S91–S94.

- Apisarnthanarak A, Khawcharoenporn T, Mundy LM. Practices to prevent multidrug-resistant *Acinetobacter baumannii* and methicillin-resistant *Staphylococcus aureus* in Thailand: a national survey. *Am J Infect Control* 2013;41:416–421.
- Siegel JD, Rhinehart E, Jackson M, Chiarello L, Healthcare Infection Control Practices Advisory Committee. Management of multidrug-resistant organisms in healthcare settings, 2006. Am J Infect Control 2007;35(10Suppl2):S165–S193.
- Buppajarntham A, Apisarnthanarak A, Khawcharoenporn T, Rutjanawech S, Singh N. National survey of Thai Infectious Diseases physicians on treatment of carbapenem-resistant *Acinetobacter baumannii* ventilator-associated pneumonia: the role of infection control awareness? *Infect Control Hosp Epidemiol* 2016;37:61–69.
- Apisarnthanarak A, Greene MT, Kennedy EH, Khawcharoenporn T, Krein S, Saint S. National survey of practices to prevent healthcareassociated infections in Thailand: the role of safety culture and collaboratives. *Infect Control Hosp Epidemiol* 2013;33:711–717.
- Sakamoto F, Sakihama T, Saint S, Greene MT, Ratz D, Tokuda Y. Health care-associated infection prevention in Japan: the role of safety culture. *Am J Infect Control* 2014;42:888–893.

# Wisdom of Microbial Pathogens: A Novel Approach to Develop Antimicrobials Against Methicillin-Resistant *Staphylococcus aureus*

To the Editor-Staphylococcus aureus is the one of the most commonly isolated human bacterial pathogens causing skin and soft-tissue infections, endovascular infections, osteomyelitis, pneumonia, endocarditis, septic arthritis, and sepsis. Methicillinresistant S. aureus (MRSA) isolates have developed resistance to all available penicillins and other β-lactam antimicrobial drugs.<sup>1</sup> A few drugs, such as vancomycin (glycopeptide), daptomycin (lipopeptide), and linezolid (oxazolidinone), have been approved for the treatment of serious infections caused by MRSA. However, different MRSA strains have already been emerging with resistance to these last-resort antimicrobial drugs.<sup>2–4</sup> These resistance trends for newer drugs emphasize the ongoing need for new and more potent antimicrobial drugs. Successful pathogenic bacteria may have to outcompete other coinfecting bacteria to stay in their eukaryotic host, such as humans. This interplay between pathogenic bacteria may lead to development of new antimicrobials.

In the present study, 39 *Pseudomonas aeruginosa* isolates were screened against MRSA. *P. aeruginosa* were isolated from various patients admitted in different Indian health centers. Four different strains of MRSA were used for susceptibility assays. *S. aureus* ATCC 25923 was used as a control strain.

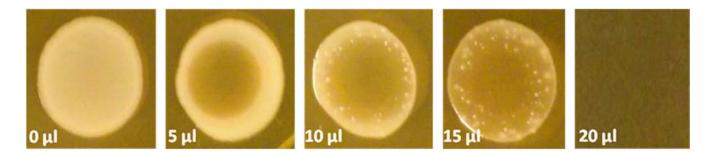


FIGURE 1. Antimicrobial activity of Pseudomonas aeruginosa P-149 supernatant (5-20 µL) against methicillin-resistant Staphylococcus aureus.

S. aureus cultures, grown overnight in Mueller-Hinton broth, were diluted to 0.5 OD<sub>600</sub> (optical density at wavelength of 600 nm) and uniformly spread on Mueller-Hinton agar plate by using a sterilized cotton swab. Similarly, P. aeruginosa cultures were grown in Mueller-Hinton broth and centrifuged (10,000 g, 10 minutes) to collect the supernatant that was then filter sterilized. An aliquot (25 µL) was added to the S. aureus lawn followed by incubation at 37°C. The level of inhibition based on the presence of halo formation around the supernatant spot was defined as follows: "no inhibition" indicated as a halo less than 8 mm; "weak inhibition," 8-15 mm; "strong inhibition," 16-25 mm; "very strong inhibition," greater than 25 mm. Of the 39 Pseudomonas isolates, 28 (72%) failed to inhibit the growth of S. aureus whereas 7 (18%) and 3 (8%) showed weak and strong inhibition, respectively. Only 1 isolate (3%) was able to show very strong inhibition of S. aureus (>30 mm halo). The isolate P-149 that showed maximum inhibition was selected for further analysis. Supernatant aliquots of varying volumes from P-149 (0–20 µL) were spotted on S. aureus lawns. After 24 h of incubation, an inhibition halo was visible around the spot of supernatant (5 µL) with profound growth of S. aureus on the edges compared with the control (Figure 1). However, with further increase in the supernatant volume (10–15 µL), only discrete colonies were visible on the edges of the inhibition zone. Growth of S. aureus was completely inhibited with 20 µL aliquot (Figure 1). Even after 72 h of incubation, growth of S. aureus remained inhibited. The same trend of inhibition was observed with other S. aureus strains including the ATCC control strain. The study clearly suggests that P-149 supernatant could be a potential source for the development of an antimicrobial drug against S. aureus. It should be further noted that crude supernatant was used without any purification step. Extraction of the active antimicrobial drug with organic solvents (acetone, hexane, and dichloromethane; 1:1) was unsuccessful. Treatment with nucleases did not alter the antimicrobial activity either. Certain pathogenic bacteria have been reported to secrete proteins to outgrow or outcompete other pathogens in the human body.<sup>5</sup> However, even after treatment with proteinase K, the P-149 supernatant still retained its activity. Cytotoxicity assays revealed that the P-149 supernatant did not affect the growth of Saccharomyces cerevisiae.

Evidence demonstrates the increasing ineffectiveness of methicillin and newer agents, such as vancomycin and linezolid. The decrease in drug efficacy for *S. aureus* represents a looming threat to patient health with the fear of returning to the morbidity and mortality levels present before antibiotics were developed. Strategies adopted by pathogenic bacteria to keep their enemies at bay may provide an insight into the production of smart weapons.

### ACKNOWLEDGMENTS

I thank E. Subudhi, Siksha 'O' Anusandhan University, Bhubaneswar, Odisha, India, and Dinesh Goyal, Shiv Astha Clinic, Haryana, India, for kindly providing the samples.

*Financial support.* Science and Engineering Research Board, Department of Science & Technology, New Delhi, India.

Potential conflicts of interest. The author reports no conflicts of interest relevant to this article.

### Mohit Kumar, PhD

Affiliations: Biotechnology and Bioinformatics, NIIT University, Neemrana, Rajasthan, India.

Address correspondence to Mohit Kumar, PhD, Biotechnology and Bioinformatics, NIIT University, Neemrana, Rajasthan-301705, India (kumarmohit @yahoo.com).

Infect. Control Hosp. Epidemiol. 2016;37(4):491–492

© 2016 by The Society for Healthcare Epidemiology of America. All rights reserved. 0899-823X/2016/3704-0024. DOI: 10.1017/ice.2015.330

### REFERENCES

- David MZ, Daum RS. Community-associated methicillin-resistant *Staphylococcus aureus*: epidemiology and clinical consequences of an emerging epidemic. *Clin Microbiol Rev* 2010;23:616–687.
- 2. Weigel LM, Clewell DB, Gill SR, et al. Genetic analysis of a high-level vancomycin-resistant isolate of *Staphylococcus aureus*. *Science* 2003;302:1569–1571.
- Mangili A, Bica I, Snydman DR, et al. Daptomycin-resistant methicillin-resistant *Staphylococcus aureus* bacteremia. *Clin Infect Dis* 2005;40:1058–1060.
- 4. Tsiodras S, Gold HS, Sakoulas G, et al. Linezolid resistance in a clinical isolate of *Staphylococcus aureus*. *Lancet* 2001;358:207–208.
- Fialho AM, Stevens FJ, Das Gupta TK, et al. Beyond host-pathogen interactions: microbial defense strategy in the host environment. *Curr Opin Biotechnol* 2007;18:279–286.