



Reassessing growth and mortality estimates for the Ordovician trilobite *Triarthrus eatoni*

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Abstract.—A length–frequency sample (n = 295) from a fossil population of the Ordovician trilobite *Triarthrus eatoni* Hall, 1838, assembled and analyzed by J. L. Cisne in 1973 is here reexamined using methods of length–frequency analysis commonly used in fishery science and marine biology. Theoretical considerations and the empirical data at hand suggest that the growth of *T. eatoni* was not "linear," but asymptotic, as is the growth of most Recent marine invertebrates. The parameters of the von Bertalanffy growth function ($L_{\infty} = 41 \text{ mm}, K = 0.29 \text{ yr}^{-1}$) suggest that *T. eatoni*, which apparently lived in a challenging environment, grew somewhat more slowly than the extant marine isopod *Ceratoserolis trilobitoides* (Eights, 1833), used here as Recent analogue to *T. eatoni*. This trilobite probably lived up to 10 years, rather than the suggested 4 years, and its mortality rate was 15%–20% per year rather than 30%–40% per year. These represent the first estimates of trilobite absolute growth characteristics using methods known to accurately model growth in extant water-breathing ectotherms. These provide a baseline for trilobite growth that can be used to make inferences about growth in other species. The approach used here may also be applied to other trilobites for which suitable length–frequency data exist.

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Introduction

Growth and size represent two of an animal's most important traits (Haldane 1926). Notably, growth provides the framework for an animal's life history (Alonzo and Kindsvater 2008). The study of growth in extinct organisms, particularly in early animals, can help to answer important questions in evolutionary developmental biology, including how developmental processes evolve (e.g., Hughes et al. 2017). However, growth in such organisms cannot be studied directly in the field or aquaria. As a result, paleontological studies of "growth" are often based on relative age data (e.g., in trilobites; Fusco et al. 2012; Hughes et al. 2017). Here, we distinguish between the absolute and relative age an animal can have. In some taxa, absolute age can be readily estimated precisely; for example, by counting daily rings in the otoliths of teleost larvae (Jones 1986) or the statoliths of squid larvae (Pauly 1998b).

When absolute ages cannot be estimated, for example, when analyzing length–frequency (L/F) data, ages are relative, that is, added to an assumed age at some small arbitrary length.

Outside paleontology, "growth" is understood to link an animal's size (length or weight) with time elapsed during its ontogeny (absolute age). Thus, an increase in the maximum size of the species in a clade of water-breathing ectotherms (WBEs) over evolutionary time is not "growth" in this restricted sense, and neither is the identification of successive anatomical stages such as molt instars. On the other hand, assuming that the successive peaks in a length distribution represent annual cohorts (or year classes) is an inference about growth. As a result, any instance in the fossil record where growth can be inferred based on absolute age is of high importance, as it allows direct comparison with models of growth in extant taxa. Such comparative analyses are key to the emerging discipline of paleo-evo-

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devo (Minelli and Fusco 2008). Even a small number of instances where such analysis can be conducted is useful, as the relationship between absolute and relative growth in these examples can then be used to make inferences about other fossil taxa.

Trilobites provide some of our best insights into growth in early animals and are one of the most diverse groups in the fossil record (Paterson 2020). As noted earlier, studies of trilobite growth have generally been based on measures of relative growth and, in particular, patterns of change in relation to the molt cycle. These usually focus on differences in size and shape between specimens (e.g., Hong et al. 2014; Holmes et al. 2020) or in relation to changing morphologies between successive molt instars (e.g., Fusco et al. 2014, 2016; Dai et al. 2021; Holmes et al. 2021a; Hughes et al. 2021). Even in life, trilobites, like other arthropods, lacked accretive hard parts that allow their age and growth to be determined directly, as in some other WBEs like fish and mollusks (Pannella and MacClintock 1968; Jones 1986; Jones and Quitmyer 1996; Pauly 1998b). One potential example of absolute growth in trilobites is the results of Cisne (1973), who presented L/F data on the Late Ordovician trilobite Triarthrus eatoni Hall, 1838 with pronounced size-distribution peaks that he interpreted as annual cohorts. Here, we apply methods developed within fisheries science and marine biology (for studying the growth of fish and invertebrates lacking annual or other skeletal marking) to reanalyze the data presented by Cisne (1973), in order to provide estimates of absolute growth and mortality parameters. For comparison, a similar analysis was also performed on a modern analogue, the marine isopod Ceratoserolis trilobitoides (Eights, 1833).

Application of Growth Models to Trilobites.— The absolute ages of *T. eatoni* specimens estimated by Cisne (1973) assumed a linear model of growth in length with time. However, we know from extant WBEs (and other metazoans) that growth is generally asymptotic, with the growth rate declining with time. Indirect support for this in trilobites comes from the presence of encrusting epibionts on large holaspides of the phacopid *Flexicalymene*, which has been used to suggest increasing time between instances of molting (representing relatively constant size increases due to Dyar's rule; see Fusco et al. 2012), and thus a decline in absolute growth (Brandt 1996). In extant WBEs (including arthropods), this decline is well described by an asymptotic growth curve such as the von Bertalanffy growth function (VBGF), and this is the model adopted here. Given the nature of the fossils at hand, the approach used here must rely on the analysis of L/F data. Moreover, the growth model or equation used to fit the available L/F data analysis must be selected a priori, as the available L/F data do not allow for contrasting the performance of different growth models (see "On the Potential Suitability of Different Mathematical Models to Describe the Growth of Trilobites" in the Supplementary Material for a discussion of alternative models; see https://doi.org/10. 5281/zenodo.6640357).

The growth model chosen here builds on the foundation of Pütter (1920) and von Bertalanffy (1938), who viewed growth rate (dw/dt) as the net result of two processes with opposite signs, that is,

$$\frac{dw}{dt} = Hw^d - kw \tag{1}$$

where the two terms on the right are anabolism and catabolism, respectively, and where d < 1.

As interpreted by Pauly (2019, 2021), anabolism refers to the synthesis of body tissues. This process requires an amino acid pool to provide building blocks for proteins and ATP to provide the "energy" required for their synthesis. Here, ingested food is not "energy"; rather, food is oxidized (i.e., "burned") to generate ATP, which may be considered to be "energy" (Cox and Nelson 2008). Thus, the process of anabolism requires oxygen, which must enter the body through some permeable surface (e.g., in trilobites, the exopod functioned as a gill; Hou et al. 2021). Therefore, the parameter d in equation (1) is equivalent to the exponent of a relationship linking respiratory surface area (*S*) to body weight of the form $S = a \cdot W^d$. Because *S* is a surface, it cannot keep up with a growing volume (or weight), and hence *d* is generally 0.6–0.7 in small fishes (Pauly 2019, 2021) and 2/3 in small crustaceans (von Bertalanffy and Krywienczyk 1953). Thus, assuming that d was 2/3 in trilobites is a parsimonious hypothesis. In contrast to anabolism, catabolism is directly proportional to body weight, because it consists of the spontaneous denaturation of protein and other molecules contributing to that weight (Brandts 1967; Goldberg and St. John 1976; Hawkins 1991). Spontaneous denaturation, that is, loss of quaternary structure by molecules, occurs throughout the body and requires neither "energy" nor oxygen to be supplied. It is therefore weight proportional, even if denaturation proceeds at different rates in different molecular types.

The integration of equation (1) when assuming d = 2/3 results in the standard VBGF, which implies that asymptotic size is reached when $Hw^d = kw$. The curvilinear VBGFs for length and weight growth resulting from this integration have found wide application in fisheries and marine biology because they describe very well the growth of WBEs, that is, fish and marine invertebrates (see www.fishbase. org and www.sealifebase.org). On the other hand, "linear" growth of WBEs in length and/or weight appears to have never been demonstrated in Recent organisms (see, e.g., Wang et al. 2017). Thus, there is no reason to assume that linear growth would have occurred in ancient WBEs such as trilobites. This provides a rationale for reexamining the inferences on growth and related attributes of T. eatoni, notably mortality and longevity estimated by Cisne (1973), based on his assumption of "linear" growth, with some emphasis on comparison with the growth of crustaceans in general and the benthic marine isopod C. trilobitoides in particular.

Materials and Methods

Triarthrus eatoni Sample of Cisne (1973).—The data analyzed here are those of Cisne (1973), based on a sample of the olenid trilobite *T. eatoni* (Fig. 1) from the base of "Beecher's Trilobite bed," a ca. 4 cm mudstone turbidite bed present at a locality within the Frankfort Shale, near Rome, New York, U.S.A. (Farrell et al. 2009, 2011). Specimens range in size from protaspides (the smallest postembryonic



FIGURE 1. A ventral view of the olenid trilobite *Triarthrus eatoni* from the Ordovician of New York State (U.S.A.) showing exceptional preservation of appendages. Scale bar, 5 mm. (Natural History Museum, London; photo by J.D.H.)

stage, <1 mm long) to large holaspides (adults, up to ca. 40 mm). Pervasive soft-part preservation of non-biomineralized structures such as appendages (e.g., Whittington and Almond 1987; Hou et al. 2021) allowed Cisne (1973) to identify 467 specimens that were alive at the time of burial (from a total of 574), of which 295 were complete enough following preparation to allow sagittal length to be measured. As such, the sample represents one of the most unbiased estimates of a census population known from the trilobite fossil record.

In his analysis, Cisne (1973: fig. 2, table 1) recognized several peaks in the population size (length) distribution, which he interpreted as representing annual cohorts. He reported the two largest groupings (his groups II and III) as having mean lengths of ca. 10.2 mm and ca. 19.1 mm. These clearly do not represent molt

instars, as this would imply an unrealistic permolt growth rate (~2) between stages (see Fusco et al. 2012). The third peak is much lower than the second, while the fourth and fifth peaks continue a more modest decline. While there appears to be substantial size variation within the cohorts, this is expected, given that individuals grow and molt at different rates (e.g., see the comments by R. G. Hartnoll in Sheldon [1988]).

Annual cohorts have not yet been reliably identified in other trilobites, most likely because instances in the fossil record that might accurately preserve census samples of living populations are extremely rare. These invariably rely on a representative sample being captured by a single event (e.g., a sediment gravity flow or turbidite). Even then, there is no way of knowing whether the sample is truly unbiased. For example, it may have been easier for larger specimens to escape the turbidite, and they may be underrepresented in the sample. Likewise, members of different size classes may live in different environments, making them more or less likely to be captured in such an event. However, it should be noted that although such a sample will be biased in terms of reflecting the population structure as a whole, this does not necessarily preclude the identification of certain population characteristics, such as recruitment pulses (if present). Even after such a deposit has been identified, strict sampling procedures are required to maintain the integrity of the sample. These are often not met, particularly with historical collections (Whitaker and Kimmig 2020). In this case, Cisne (1973) noted that a number of specimens (~100) were given away to other collections and that these were probably mostly of medium size. However, he considered any bias introduced from this to be minimal (as do we).

There are a number of published examples that have been interpreted as trilobite "census samples." Paterson et al. (2007: fig. 5A) described a sample of the early Cambrian *Balcoracania dailyi* from the Warragee Member of the Billy Creek Formation that likely represents a largely unbiased sample of a living population. It is noteworthy that this sample has a similar right-skewed distribution to that of the *T. eatoni* sample analyzed here. However, the scarcity of larger specimens in the *B. dailyi* sample makes any identification of recruitment cohorts problematic. Brezinksi (1986) interpreted a sample of the Ordovician trilobite Ampyxina bellatula as a census population, the distribution of which shows a roughly normal distribution with eight to nine subordinate peaks consistent with his interpretation of molt instars (based on size differences between peaks). However, this sample only contains holaspid (adult) specimens, and thus represents only a part of ontogeny. The peak of the overall distribution itself may represent a recruitment cohort; however, based on the limited size interval, it is difficult to determine whether this is the case. Incidentally, Brezinski (1986) apparently made several errors in his analysis, including the interpretation of molt instars as recruitment pulses and confusing relative and absolute growth (see Sheldon 1988). This highlights the importance of the issues discussed in the present contribution.

Despite the absence of other examples in the fossil record, our assumption that the peaks in the *T. eatoni* size distribution represent recruitment cohorts is justified, for the following reasons: (1) We have fit Gaussian finite mixture models to a simulated dataset based on the size distribution of Cisne (1973), and the best-

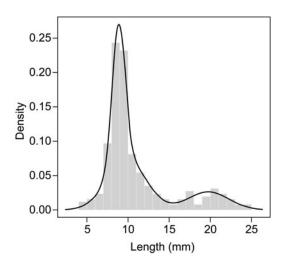


FIGURE 2. Density estimate (line) of the Gaussian finite mixture model fit to the simulated size (length) distribution of the trilobite *Triarthrus eatoni* (based on data from Cisne [1973]). Results suggest that the two peaks are drawn from separate distributions.

supported models suggest that groups II and III are drawn from separate distributions (Fig. 2; see "Testing for Multiple Distributions"). (2) Specimens can be clearly identified as being alive when buried (or very recently deceased) based on both articulation state and soft-part preservation. This is essentially a "gold standard" for recognizing a census sample and not present in the examples discussed earlier. (3) The collection biases are known, and it is unlikely that these would affect the ability to discern recruitment cohorts. (4) The observed size distances between annual peaks are consistent with modern crustaceans, for example, mean lengths of 29.3, 44.4, 55.8 and 62.9 mm were reported for four annual cohorts of the deepwater shrimp Aristeus antennatus, implying percentage increases of 151%, 126%, and 112% (Ragonese and Bianchini 1996), compared with 10.2, 19.1, 32.0, and 38.2 mm (187%, 167%, and 119%) for the larger groups in T. eatoni (note the decreasing growth in percentage length in both cases). Likewise, two annual cohorts with peaks of ca. 12 and 40 mm (333%) were observed for juveniles of the crab Ranina ranina (Kirkwood et al. 2005). The size distribution of R. ranina is also very similar to that of T. eatoni, showing a very large younger cohort and a much smaller older one (Kirkwood et al. 2005: fig. 3). Annual peaks identified in the marine isopod Ceratoserolis trilobitoides (represented by the molt classes III and V of Luxmoore [1981]) varied from ca. 15 to 23 mm (153%). These examples suggest that T. eatoni size-distribution peaks are well within the range of annual growth for extant crustaceans. (5) If groups II and III are assumed to be annual cohorts (representing average size increase of 187%), it can be estimated that T. eatoni probably molted (on average) approximately five to seven times during this period. Per-molt growth rates of trilobites were largely constant across ontogeny (consistent with Dyar's rule; e.g., Fusco et al. 2012), and trilobites living in environments similar to that of T. eatoni show size increase of ca. 110%–115% per molt (Fusco et al 2012; Hou et al. 2017; Holmes et al. 2021b; Hopkins 2021). This is consistent with extant arthropods, for example, decapod crustaceans (e.g., Corgos et al. 2007). Here, therefore, we proceed with the explicit assumption that Cisne (1973) was correct in identifying his groups (specifically II and III) as annual cohorts.

Testing for Multiple Distributions.—Cisne (1973: table 1) only provided summary statistics for his L/F *T. eatoni* data. While this and the size distribution (Cisne 1973: fig. 2) provide the input data required for the growth models employed here, the lengths of individual specimens were not provided. However, to test for multiple distributions, we can simulate these data based on information from the published size distribution. We randomly generated the same number of length measurements within each 1 mm size bin of Cisne (1973: fig. 2) for groups II and III to create a dataset for testing whether the observed data are likely to be drawn from one or more distributions.

Using the function Mclust() from the mclust package (Scrucca et al. 2016), we fit Gaussian finite mixture models to the simulated dataset to test its suspected multidistribution nature. This function tests models with different numbers of components and selects the most likely model based on the Bayesian information criterion. Note that these models assume that the data are drawn from one or more normal distributions. Models were fit to both logged and unlogged data (group II appears to be right skewed, and logging normalizes this somewhat). In both cases, models with three components were the most likely. The models suggest that group II is comprised of two distributions with similar mean values, while group III is a separate distribution. In the simulation we ran, the means of the two group II distributions were 8.78 and 9.88 mm, while the mean of the group III distribution was 19.80 mm (i.e., very similar to the mean estimates of Cisne [1973]). It is likely that the somewhat nonnormal group II distribution is causing the models to suggest that this is composed of two normal distributions rather than one. However, the important point is that group III is clearly identified as a distribution separate from group II. Density estimates were then produced for each data point based on the three-component Gaussian finite mixture model (using the function *densityMclust(*) and plotted for visual confirmation of how the model fits the data; Fig. 2). The R code for this analysis is brief and appended to the Supplementary Material rather than as a separate R script file (see https://doi.org/10.5281/ zenodo.6640357).

Growth Models.—For length, the VBGF has the form:

$$L_t = L_{\infty} \cdot (1 - e^{-K \cdot (t - t_0)})$$
(2)

where L_t is the mean length at age t of the WBEs; L is their asymptotic length, that is, the mean length attained by the surviving WBEs after an infinitely long time; K (here: yr⁻¹) expresses how rapidly L_{∞} is approached; and t_0 is a parameter that corrects for the fact that the VBGF generally fails to describe the growth of the larval stages of WBEs (Pauly 1998a). When t_0 is assumed to be zero, age becomes "relative age" (t').

Typically, the parameters of the VBGF are estimated by nonlinear fitting of length-at-age data pairs. However, we cannot use this technique here because—contrary to Cisne's (1973) assumption—only two of the size groups he identified (II and III in our Fig. 3A) could be reliably assumed to be successive annual groups. However, two alternative methods of estimating the parameters of the VBGF are available. The first is approximate and involves identifying only two size groups (L_1 , L_2) separated by a known or assumed time interval ($t_2 - t_1$).

Given such data, the parameter K of the VBGF can be estimated from:

$$K \approx [(L_2 - L_1)/(t_2 - t_1)]/L_{\infty}$$
 (3)

where L_{∞} is an estimate of the asymptotic length (Gulland and Holt 1959), which can be approximated by the largest specimen in an L/F sample where the sampling procedure is not biased against larger individuals.

The second method for estimating the parameters of the VBGF is electronic length frequency analysis (ELEFAN; Pauly and David 1981). ELEFAN is a quasi-Bayesian method in which the L/F data at hand are re-expressed in the form of positive "points" corresponding to peaks and negative points corresponding to the troughs separating the peaks. Then, hundreds of VBGF parameter sets are tested, and the set is retained that, by

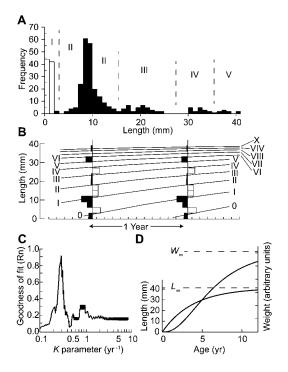


FIGURE 3. Aspects of the growth of the olenid trilobite *Triarthrus eatoni;* A, Length–frequency (L/F) sample assembled by Cisne (1973), with his assumed annual groups (I to V); B, electronic length frequency analysis (ELE-FAN) analysis of the same L/F data, after regrouping them in 3 mm bins and restructuring via a smoothing procedure to identify peaks (black, positive histograms) and troughs (white, negative histograms), which allowed a von Bertalanffy growth function (VBGF) parameter set to be determined that produced a growth curve that hit most peaks and avoided most troughs; C, goodness of fit of a VBGF with $L_{\infty} = 41$ mm with over a wide range of *K* values, identifying K = 0.29 yr⁻¹ as the best estimate; D, the length and likely weight growth curves for *T. eatoni*.

accumulating "points," best connects the peaks while avoiding the troughs (Pauly and David 1981; Gayanilo et al. 2005).

Once L_{∞} and *K* estimates are available, a rate of mortality (*M*) can be estimated from a length-converted catch curve (Pauly et al. 1995), which has the form:

$$ln(N/\Delta t) = a - Mt' \tag{4}$$

where *N* is the number of individuals in a given length class, Δt is the time required for individuals to grow through that length class (as can be determined from the VBGF), *a* is a scaling parameter, *t'* is the relative age as defined earlier, and *M* is the instantaneous rate of mortality (Pauly 1998a). Longevity (t_{max}) can be estimated from *K* if it is assumed that the longest-lived WBEs grow to 95% of their asymptotic size; in such a case, $t_{max} \approx 3/K$ (Taylor 1958). Another interesting property of the VBGF is that, when combined with an instantaneous rate of mortality, this rate (i.e., *M*) is also the rate of biological production (*P*; Winberg 1971) in a population (*B*), that is, $P = M \cdot B$ (Allen 1971), which is a required parameter for constructing trophic food web models (Pauly and Christensen 2002).

For growth comparison, we selected crustaceans as analogues to trilobites, and the benthic marine isopod *C. trilobitoides* as an analogue to *T. eatoni*, as justified by its comparable size and shape (hence its species name). Then we used L/F samples from Luxmoore (1981: fig. 4) and the ELEFAN method to estimate growth parameters for *C. trilobitoides*, using the width of the third pereonal segment as the length input. We also created an auximetric plot for the growth parameters of 462 populations in 95 crustacean species in SeaLifeBase (www.sealifebase), whose asymptotic lengths ranged from 1 to 50 cm.

Note that in *C. trilobitoides*, as in other crustaceans, the growth curve for a population is an average resulting from different individuals, each molting and increasing its size at different times. This will tend to produce a smooth growth curve that can be fit without considering instars. There is no evidence that this was different in trilobites, hence the application of a "smooth" growth model to *T. eatoni*.

Results

The most prominent peak in Figure 3A, ranging from 4.2 to 14.4 mm, has a mean length of 10.2 mm, while the second prominent peak, ranging from 14.9 to 24.8 mm, has a mean length of 19.1 mm (Cisne 1973: table 1). Hence, if we follow Cisne (1973) in the assumption that these two peaks are separated by 1 year and use 41 mm as input for L_{∞} (1 mm longer than the maximum reported length), a first estimate of the parameter *K* (yr⁻¹) for *T. eatoni* can be obtained from:

$$19.1 - 10.2/41 \approx 0.22 \tag{5}$$

This estimate served as prior to the ELEFAN analysis, which yielded, for $L_{\infty} = 41 \text{ mm}$, an estimate of $K = 0.29 \text{ yr}^{-1}$ (Fig. 3B). Given $t_{\rm max} \approx 3/K$, this allowed estimating longevity as $3/0.29 \approx 10$ years, while constructing a length-converted catch curve provided an estimate of $M = 0.48 \text{ yr}^{-1}$, with a 95% confidence interval of 0.38-0.58 yr⁻¹ (Fig. 4B). This corresponds to 15%-20% of a population dying every year, rather than 30%–40%, as suggested by Cisne (1973; see Fig. 4A). Three data points in Figure 4B, shown as open circles and ranging from 24 to 33 mm, were not used to estimate M, because their low ordinate values resulted from "medium-sized specimens" having been given away (Cisne 1973). Indeed, their ordinate values could be made to meet the regression

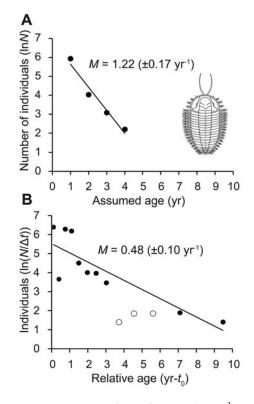


FIGURE 4. Estimation of mortality rates (M, yr^{-1}) in *Triarthrus eatoni* through catch curves. A, The catch curve proposed by Cisne (1973), assuming that the four size groups in Fig. 3A are successive annual reproductive "pulses"; B, a length-converted catch curve based on the length–frequency (L/F) data in Fig. 3A but using the VBGF in Fig. 3B,D to convert length to relative ages. The three open circles were omitted from the computation because they belong to the "medium" size, many of which had been given away to other collections (see text).

line by adding a total of 21 specimens to the three depleted size classes.

The ELEFAN growth parameters estimated for the isopod *Ceratoserolis trilobitoides* were $L_{\infty} = 54$ mm and K = 0.48 yr⁻¹, with a relatively high goodness of fit (Fig. 5B). This implies, if $t_{max} \approx 3/K$, that *C. trilobitoides* may reach an age of 6 years. These parameters imply much faster growth than in *Triarthrus eatoni*. This difference is also visible in Figure 6, which presents an auximetric plot (Pauly 1998a, 2019), enabling the comparison of the single estimate of trilobite growth parameters with the growth parameters of crustaceans in general and *C. trilobitoides* in particular.

Discussion

The asymptotic growth curve presented above for *Triarthrus eatoni* and its derived statistics are, as far as could be ascertained, the first

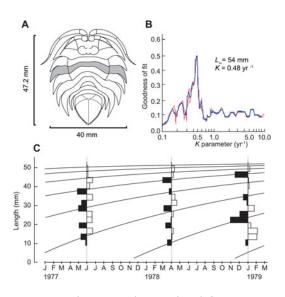


FIGURE 5. Analysis using electronic length frequency analysis (ELEFAN) of length–frequency (L/F) data of the isopod *Ceratoserolis trilobitoides*. A, Showing (in gray) the third pereonal segment, whose width is here used as measure of length, modified by E. Chu from fig. 2 of Luxmoore (1981); B, goodness of fit of a von Bertalanffy growth function (VBGF) with $L_{\infty} = 54$ mm with over a wide range of *K* values, identifying K = 0.48 yr⁻¹ as the best estimate; C, the three L/F samples in fig. 4 of Luxmoore (1981), initially in 0.5 mm classes, regrouped in 2.5 mm classes and with superposed best-fitting growth curve. Note that no a priori assumption was made about the time separating the peaks (black histograms), whose height is similar despite wide differences between recruitment pulses.

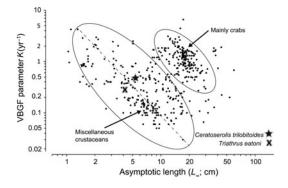


FIGURE 6. Auximetric plot with each of the small back dots documenting one of 462 pairs of L_{∞} and *K* values (i.e., a growth curve) in 98 species of crustaceans in SeaLifeBase (www.sealifebase.org). "Shrimps" (mainly of the family Penaeidae) were excluded, as many of their L_{∞} and *K* data pairs originated from farmed populations. Note that the position of the L_{∞} and *K* data pair for *Triarthrus eatoni* is below the large ellipsoid's main axis (dotted line), suggesting a slower growth than for its analogue, the isopod *Ceratoserolis trilobitoides*, but still well within the crustacean range. VBGF, von Bertalanffy growth function.

application of these concepts to trilobites, derived from what Cisne (1973) called "a tolerable sample of a natural population." Indeed, this sample appears to suffer from biases roughly similar to those sampled by trawling from a Recent population of WBEs (Pauly 1998a). In such samples, the biases generally affect the estimation of mortality, notably because large individuals, which occur in deeper waters, are not well represented. However, these biases tend to have a minimum impact on the estimation of growth parameters, especially when ELEFAN is used, which is nonparametric, and whose identification of the "peaks" and "troughs" representing recruitment pulses is largely independent of their absolute frequency.

Figure 6 suggests that the growth of *T. eatoni* is slow (i.e., to the lower left of the dotted line representing the main axis of the large ellipsoid) when compared with that of the isopod *Ceratoserolis trilobitoides*, but not exceptionally so when compared with that of Recent crustaceans in general. However, it is appropriate to ask whether the parameters estimated here are internally consistent and consistent with knowledge about olenid trilobites. One aspect of these results' internal consistency is the ratio M/K = 1.7, expressing the relationship of

mortality to somatic growth. Values of M/K between 1.5 and 2.0 are common for WBEs, notably in fishes (Pauly 1980; see also www. fishbase.org) and invertebrates, including crustaceans (Pauly et al. 1984; see also www. sealifebase.org). However, consistency with external evidence is still required.

Olenid trilobites like T. eatoni are characteristic of low-oxygen environments (Fortey 2000), and Beecher's Trilobite Bed and intervals with similar exceptional preservation in the Frankfort Shale are interpreted as being within the lower part of the dysaerobic zone (Farrell et al. 2011). In fact, a similar environment is interpreted for many deposits that preserve a high proportion of articulated trilobites, many of which have similar olenid-type morphologies (e.g., Hughes et al. 1999, 2014; Gaines and Droser 2003; Paterson et al. 2016). Trilobites living in such conditions are known to exhibit low per-molt growth rates relative to those from less stressful environments (Fusco et al. 2012; Hou et al. 2017; Holmes et al. 2021a,b; Hopkins 2021). Extant marine arthropods exposed to hypoxic conditions also exhibit low per-molt growth rates, as well as lower rates of molting in general (i.e., fewer molts per unit time and lower absolute growth), compared with growth under more oxygenated conditions (e.g., Das and Stickle 1993; Burnett and Stickle 2001). This is consistent with growth experiments on Recent WBEs, whose (absolute) growth is impaired by reduction in ambient oxygen, as illustrated by, for example, Kolding et al. (2008) for bony fish and Wu and Or (2005) for amphipod crustaceans. Thus, based on the finding that T. eatoni exhibited low rates of growth relative to C. trilobitoides but within the bounds for extant crustaceans provides strong evidence for the validity of the approach used here (including the assumptions of annual growth cohorts and model choice). The estimated maximum age for T. eatoni of 10 years is also consistent with what is known for modern WBE arthropods, although this can be highly variable. For example, C. trilobitoides is estimated to have lived for up to 6 years (while growing faster and dying younger than T. eatoni), as does the shrimp Aristeus antennatus (discussed earlier; e.g., Ragonese and Bianchini 1996). In contrast, the American horseshoe crab (*Limulus polyphe-mus*), also often considered a modern analogue of trilobites (e.g., Bicknell et al. 2018, 2021), can live for up to 20 years (e.g., Carmichael et al. 2003), and European lobsters (*Homarus gammarus*) for more than 50 years (Sheehy et al. 1999). It should be noted that estimates of maximum age in such extant WBE arthropods must often be estimated in a similar manner as was done here for *T. eatoni* and *C. trilobitoides*.

These consistencies are encouraging and suggest that the methods proposed here could be profitably applied to other species of trilobites fossilized under circumstances allowing for a least two size groups, representing annual reproductive "pulses," to be identified. Even in isolation, the results presented here represent the first reasonable estimates of absolute ages based on methods known to accurately characterize growth in modern WBEs. This provides a baseline for trilobite growth and allows inferences to be made about absolute age in other taxa.

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Data Availability Statement

Data available from the Zenodo Digital Repository: https://doi.org/10.5281/zenodo. 6640357.

Literature Cited

- Allen, K. R. 1971. Relation between production and biomass. Journal of the Fisheries Research Board of Canada 28:1573–1581.
- Alonzo, S. H., and H. K. Kindsvater. 2008. Life-history patterns. Pp. 2175–2180 in S. E. Jørgensen and B. D. Fath, eds. Encyclopedia of ecology. Academic Press, Oxford.
- Bertalanffy, L. von. 1938. A quantitative theory of organic growth (inquiries on growth laws. II). Human Biology 10:181–213.
- Bertalanffy, L. von, and J. Krywienczyk. 1953. The surface rule in crustaceans. American Naturalist 87:107–110.
- Bicknell, R. D. C., J. A. Ledogar, S. Wroe, B. C. Gutzler, W. H. Watson III, and J. R. Paterson. 2018. Computational biomechanical analyses demonstrate similar shell-crushing abilities in modern and ancient arthropods. Proceedings of the Royal Society of London B 285:20181935.
- Bicknell, R. D. C., J. D. Holmes, G. D. Edgecombe, S. R. Losso, J. Ortega-Hernández, S. Wroe, and J. R. Paterson. 2021. Biomechanical analyses of Cambrian euarthropod limbs reveal their

effectiveness in mastication and durophagy. Proceedings of the Royal Society of London B 288:20202075.

- Brandt, D. S. 1996. Epizoans on *Flexicalymene* (Trilobita) and implications for trilobite paleoecology. Journal of Paleontology 70:442–449.
- Brandts, T. F. 1967. Heat effects on protein and enzymes. Pp. 25–72 in A. H. Rose, ed. Thermobiology. Academic Press, New York.
- Brezinski, D. K. 1986. An opportunistic Upper Ordovician trilobite assemblage from Missouri. Lethaia 19:315–325.
- Burnett, L. E., and W. B. Stickle. 2001. Physiological responses to hypoxia. Pp. 101–114 in N. N. Rabalais and R. E. Turner, eds. Coastal hypoxia: consequences for living resources and ecosystems. American Geophysical Union, Washington, D.C.
- Carmichael, R. H., D. Rutecki, and I. Valiela. 2003. Abundance and population structure of the Atlantic horseshoe crab *Limulus polyphemus* in Pleasant Bay, Cape Cod. Marine Ecology Progress Series 246:225–239.
- Cisne, J. L. 1973. Life history of an Ordovican trilobite *Triarthrus* eatoni. Ecology 54:135–142.
- Corgos, A., M. P. Sampedro, E. González-Gurriarán, and J. Freire. 2007. Growth at moult, intermoult period, and moulting seasonality of the spider crab *Maja brachydactyla*: combining information from mark-recapture and experimental studies. Journal of Crustacean Biology 27:255–262.
- Cox, M. M., and D. L. Nelson. 2008. Lehninger principles of biochemistry. Freeman, New York.
- Dai, T., N. C. Hughes, X. Zhang, and G. Fusco. 2021. Absolute axial growth and trunk segmentation in the early Cambrian trilobite Oryctocarella duyunensis. Paleobiology 47:517–532.
- Das, T., and W. B. Stickle. 1993. Sensitivity of crabs Callinectes sapidus and C. similis and the gastropod Stramonita haemastoma to hypoxia and anoxia. Marine Ecology Progress Series 98:263–274.
- Eights, J. 1833. Description of a new Crustaceous animal found on the shores of the South Shetland Islands. Transactions of the Albany Institute 2:53–57.
- Farrell, Ú. C., M. J. Martin, J. W. Hagadorn, T. Whiteley, and D. E. G. Briggs. 2009. Beyond Beecher's Trilobite Bed: widespread pyritization of soft tissues in the Late Ordovician Taconic foreland basin. Geology 37:907–910.
- Farrell, Ú. C., D. E. G. Briggs, and R. R. Gaines. 2011. Paleoecology of the olenid trilobite *Triarthrus*: new evidence from Beecher's Trilobite Bed and other sites of pyritization. Palaios 26:730–742.
- Fortey, R. 2000. Olenid trilobites: the oldest known chemoautotrophic symbionts? Proceedings of the National Academy of Sciences USA 97:6574–6578.
- Fusco, G., T. Garland, G. Hunt, and N. C. Hughes. 2012. Developmental trait evolution in trilobites. Evolution 66:314–329.
- Fusco, G., P. S. Hong, and N. C. Hughes. 2014. Positional specification in the segmental growth pattern of an early arthropod. Proceedings of the Royal Society of London B 281:20133037.
- Fusco, G., P. S. Hong, and N. C. Hughes. 2016. Axial growth gradients across the postprotaspid ontogeny of the Silurian trilobite *Aulacopleura koninckii*. Paleobiology 42:426–438.
- Gaines, R. R., and M. L. Droser. 2003. Paleoecology of the familiar trilobite *Elrathia kingii*: an early exaerobic zone inhabitant. Geology 31:941–944.
- Gayanilo, F. C., P. Sparre, and D. Pauly. 2005. FAO-ICLARM stock assessment tools II (FiSAT II). Revised version user's guide. FAO Computerized Information Series (Fisheries) 8. FAO, Rome.
- Goldberg, A. L., and A. C. St John. 1976. Intracellular protein degradation in mammalian and bacterial cells: Part 2. Annual Review of Biochemistry 45:747–803.
- Gulland, J. A., and S. J. Holt. 1959. Estimation of growth parameters for data at unequal time intervals. ICES Journal of Marine Science 25:47–49.
- Haldane, J. B. S. 1926. On being the right size. Harper's Magazine 152:424–427.

- Hall, J. 1838. Description of two species of trilobites, belonging to the genus *Paradoxides*. American Journal of Science and Arts 33:139–142.
- Hawkins, A. J. S. 1991. Protein turnover: a functional appraisal. Functional Ecology 5:222–233.
- Holmes, J. D., J. R. Paterson, and D. C. García-Bellido. 2020. The trilobite *Redlichia* from the lower Cambrian Emu Bay Shale Konservat-Lagerstätte of South Australia: systematics, ontogeny and soft-part anatomy. Journal of Systematic Palaeontology 18:295–334.
- Holmes, J. D., J. R. Paterson, and D. C. García-Bellido. 2021a. Complex axial growth patterns in an early Cambrian trilobite from South Australia. Proceedings of the Royal Society of London B 288:20212131.
- Holmes, J. D., J. R. Paterson, and D. C. García-Bellido. 2021b. The post-embryonic ontogeny of the early Cambrian trilobite *Estaingia bilobata* from South Australia: trunk development and phylogenetic implications. Papers in Palaeontology 7:931–950.
- Hong, P. S., N. C. Hughes, and H. D. Sheets. 2014. Size, shape, and systematics of the Silurian trilobite *Aulacopleura koninckii*. Journal of Paleontology 88:1120–1138.
- Hopkins, M. J. 2021. Ontogeny of the trilobite *Elrathia kingii* (Meek) and comparison of growth rates between *Elrathia kingii* and *Aulacopleura koninckii* (Barrande). Papers in Palaeontology 7:985–1002.
- Hou, J., N. C. Hughes, J. Yang, T. Lan, X. Zhang, and C. Dominguez. 2017. Ontogeny of the articulated yiliangellinine trilobite *Zhang-shania typica* from the lower Cambrian (Series 2, Stage 3) of southern China. Journal of Paleontology 91:86–99.
- Hou, J., N. C. Hughes, and M. J. Hopkins. 2021. The trilobite upper limb branch is a well-developed gill. Science Advances 7: eabe7377.
- Hughes, N. C., R. E. Chapman, and J. M. Adrain. 1999. The stability of thoracic segmentation in trilobites: a case study in developmental and ecological constraints. Evolution and Development 1:24–35.
- Hughes, N. C., J. Kříž, J. H. S. Macquaker, and W. D. Huff. 2014. The depositional environment and taphonomy of the Homerian "Aulacopleura shales" fossil assemblage near Loděnice, Czech Republic (Prague Basin, Perunican microcontinent). Bulletin of Geosciences 89:219–238.
- Hughes, N. C., P. S. Hong, J. Hou, and G. Fusco. 2017. The development of the Silurian trilobite *Aulacopleura koninckii* reconstructed by applying inferred growth and segmentation dynamics: a case study in paleo-evo-devo. Frontiers in Ecology and Evolution 5:37.
- Hughes, N. C., J. M. Adrain, J. D. Holmes, P. S. Hong, M. J. Hopkins, J.-B. Hou, A. Minelli, T.-Y. S. Park, J. R. Paterson, J. Peng, M. Webster, X.-G. Zhang, X.-L. Zhang, and G. Fusco. 2021. Articulated trilobite ontogeny: suggestions for a methodological standard. Journal of Paleontology 95:298–304.
- Jones, C. 1986. Determining age of larval fish with the otolith increment technique. Fishery Bulletin of the United States 84:91–104.
- Jones, D. S., and I. R. Quitmyer. 1996. Marking time with bivalve shells: oxygen isotopes and season of annual increment formation. Palaios 11:340.
- Kirkwood, J. M., I. W. Brown, S. W. Gaddes, and S. Hoyle. 2005. Juvenile length-at-age data reveal that spanner crabs (*Ranina ranina*) grow slowly. Marine Biology 147:331–339.
- Kolding, J., L. Haug, and S. Stefansson. 2008. Effect of ambient oxygen on growth and reproduction in Nile tilapia (*Oreochromis niloticus*). Canadian Journal of Fisheries and Aquatic Sciences 65:1413–1424.
- Luxmoore, R. A. 1981. Moulting and growth in serolid isopods. Journal of Experimental Marine Biology and Ecology 56:63–85.
- Minelli, A., and G. Fusco. 2008. Evolving pathways—key themes in evolutionary developmental biology. Cambridge University Press, Cambridge.

- Pannella, G., and C. MacClintock. 1968. Biological and environmental rhythms reflected in molluscan shell growth. Journal of Paleontology 2:64–80.
- Paterson, J. R. 2020. The trouble with trilobites: classification, phylogeny and the cryptogenesis problem. Geological Magazine 157:35–46.
- Paterson, J. R., J. B. Jago, G. A. Brock, and J. G. Gehling. 2007. Taphonomy and palaeoecology of the emuellid trilobite *Balcoracania dailyi* (early Cambrian, South Australia). Palaeogeography, Palaeoclimatology, Palaeoecology 249:302–321.
- Paterson, J. R., D. C. García-Bellido, J. B. Jago, J. G. Gehling, M. S. Y. Lee, and G. D. Edgecombe. 2016. The Emu Bay Shale Konservat-Lagerstätte: a view of Cambrian life from East Gondwana. Journal of the Geological Society 173:1–11.
- Pauly, D. 1980. On the interrelationships between natural mortality, growth parameters, and mean environmental temperature in 175 fish stocks. ICES Journal of Marine Science 39:175–192.
- Pauly, D. 1998a. Beyond our original horizons: the tropicalization of Beverton and Holt. Reviews in Fish Biology and Fisheries 8:307–334.
- Pauly, D. 1998b. Why squids, though not fish, may be better understood by pretending they are. *In* A. I. L. Payne, M. R. Lipinski, M. R. Clarke, and M. A. C. Roeleveld, eds. Cephalopod biodiversity, ecology and evolution. South African Journal of Marine Science Series 20:47–58.
- Pauly, D. 2019. Gasping fish and panting squids: oxygen, temperature and the growth of water breathing animals. Excellence in Ecology Series. International Ecology Institute, Oldendorf/ Luhe, Germany.
- Pauly, D. 2021. The gill-oxygen limitation theory (GOLT) and its critics. Science Advances 7:eabc6050.
- Pauly, D., and V. Christensen. 2002. Ecosystem models. Pp. 211–227 in P. J. B. Hart and J. D. Reynolds, eds. Handbook of fish biology and fisheries. Wiley, Chichester, U.K.
- Pauly, D., and N. David. 1981. ELEFAN I, a BASIC program for the objective extraction of growth parameters from length-frequency data. Berichte der Deutschen wissenschaftlichen Kommission für Meeresforschung 28:205–211.
- Pauly, D., J. Ingles, and R. Neal. 1984. Application to shrimp stocks of objective methods for the estimation of growth, mortality and recruitment-related parameters from length-frequency data

(ELEFAN I and II). Pp. 220–234 *in* J. A. Gulland and B. I. Rothschild, eds. Penaeid shrimps—their biology and management. Fishing News Books, Farnham, U.K.

- Pauly, D., J. Moreau, and N. Abad. 1995. Comparison of age-structured and length-converted catch curves of brown trout *Salmo trutta* in two French rivers. Fisheries Research 22:197–204.
- Pütter, A. 1920. Studien über physiologische Ähnlichkeit VI. Wachstumsähnlichkeiten. Pflügers Archive 180:298–340.
- Ragonese, S., and M. L. Bianchini. 1996. Growth, mortality and yield-per-recruit of the deep-water shrimp *Aristeus antennatus* (Crustacea-Aristeidae) of the Strait of Sicily (Mediterranean Sea). Fisheries Research 26:125–137.
- Scrucca, L., M. Fop, T. B. Murphy, and A. E. Raftery. 2016. mclust 5: clustering, classification and density estimation using Gaussian finite mixture models. R Journal 8:289–317.
- Sheehy, M. R. J., R. C. A. Bannister, J. F. Wickins, and P. M. J. Shelton. 1999. New perspectives on the growth and longevity of the European lobster (*Homarus gammarus*). Canadian Journal of Fisheries and Aquatic Sciences 56:1904–1915.
- Sheldon, P. R. 1988. Trilobite size-frequency distributions, recognition of instars, and phyletic size changes. Lethaia 21:293–306.
- Taylor, C. C. 1958. Cod growth and temperature. ICES Journal of Marine Science 23:366–370.
- Wang, K., B. P. V. Hunt, C. Liang, D. Pauly, and E. A. Pakhomov. 2017. Reassessment of the life cycle of the pteropod *Limacina helicina* from a high resolution interannual time series in the temperate North Pacific. ICES Journal of Marine Science 74:1906–1920.
- Whitaker, A., and J. Kimmig. 2020. Anthropologically introduced biases in natural history collections, with a case study on the invertebrate paleontology collections from the middle Cambrian Spence Shale Lagerstätte. Palaeontologia Electronica 23:a58.
- Whittington, H. B., and J. E. Almond. 1987. Appendages and habits of the Upper Ordovician trilobite *Triarthrus eatoni*. Philosophical Transactions of the Royal Society of London B 317:1–46.
- Winberg, G. G. 1971. Methods for the estimation of production of aquatic animals. Academic Press, New York.
- Wu, R. S. S., and Y. Y. Or. 2005. Bioenergetics, growth and reproduction of amphipods are affected by moderately low oxygen regimes. Marine Ecology Progress Series 297:215–223.