

## Molecular epidemiology of hepatitis E virus in humans, pigs and wild boars in Sweden

F. WIDÉN<sup>1\*</sup>, L. SUNDQVIST<sup>2</sup>, A. MATYI-TOTH<sup>1</sup>, G. METREVELI<sup>1</sup>, S. BELÁK<sup>1</sup>,  
G. HALLGREN<sup>3</sup> AND H. NORDER<sup>2</sup>

<sup>1</sup> Joint R&D Division, Department of Virology, Immunobiology and Parasitology, Statens Veterinärmedicinska Anstalt & Department of Biomedicine and Veterinary Public Health, The Swedish University of Agricultural Sciences and the OIE Collaborating Centre for the Application of Biotechnology-based Methods for the Diagnosis of Infectious Diseases in Veterinary Medicine, Uppsala, Sweden

<sup>2</sup> Virological Department, Swedish Institute for Infectious Disease Control, Solna, Sweden

<sup>3</sup> Department of Disease Control and Epidemiology, Statens Veterinärmedicinska Anstalt, Uppsala, Sweden

(Accepted 18 May 2010; first published online 14 June 2010)

### SUMMARY

Hepatitis E infections in humans are usually acquired in endemic countries in Asia or Africa. In Sweden 17 cases infected in Europe, between 1993 and 2009, were identified. All had clinical hepatitis E with unknown source of infection. Hepatitis E virus (HEV) was identified in faecal samples from 63 piglets in 12 pig farms in Sweden. HEV was also identified in blood from 13 out of 159 investigated Swedish wild boars from nine counties. Partial HEV genomes from humans, pigs and wild boars were sequenced and compared by phylogeny. The results showed close relatedness between HEV strains from piglets from the same farm and from wild boars from the same county. HEV strains from humans showed relatedness with strains from pigs and wild boars from the same county. This study showed that HEV strains form geographical clusters in the phylogenetic tree. The methods used in this study may thus be used for tracing the origin of an infecting strain. Furthermore, this study indicated that there are endemic sources of human HEV infections in Sweden.

**Key words:** Hepatitis E, virology, zoonoses.

### INTRODUCTION

Hepatitis E virus (HEV) is a small non-enveloped RNA virus classified in the genus *Hepevirus*, the only member of the Hepeviridae family [1, 2]. There are two species of HEV, mammalian and avian HEV. Antibodies against mammalian HEV have been identified in numerous species worldwide including

humans, pigs, wild boars, rodents, dogs, cats, cows, sheep, and goats [1].

HEV is often responsible for waterborne outbreaks of viral hepatitis in humans, and for a large proportion of cases with sporadic acute hepatitis in endemic regions, such as large parts of Asia, Africa and Latin America [1, 2]. In epidemics in these regions disease morbidity mainly affects young adults, a high mortality of up to 25% is seen in infected pregnant females during the third trimester [3]. Human HEV is mainly transmitted through the faecal/oral route through contaminated water or food, although vertical and bloodborne transmissions have also been

\* Author for correspondence: Dr F. Widén, Statens Veterinärmedicinska Anstalt, Department of Virology, Immunobiology and Parasitology, S-751 89 Uppsala, Sweden.  
(Email: frederik.widen@sva.se)

reported [4–9]. Human-to-human transmission is rare. In developed countries most hepatitis E cases are associated with travel from endemic regions. However, in the last few years an increasing number of sporadic cases have been reported in patients with no known epidemiological risk factor [10–16].

The HEV RNA is about 7.2 kb long and contains three open reading frames, ORF1–ORF3, flanked by short untranslated regions. ORF1 encodes for viral non-structural proteins containing several conserved domains, functioning as the putative methyltransferase, protease, helicase, and RNA-dependent RNA polymerase. ORF2 encodes for the viral capsid protein and ORF3 for a small phosphoprotein with uncertain function [17–19].

Based on divergences of complete genomes, four phylogenetically distinct genotypes of mammalian HEV have been identified, HEV genotypes 1–4. Genotype 1 has so far only been isolated from humans, although one pig in Cambodia has been shown as infected with this genotype [20]. This genotype is the main cause of sporadic and epidemic hepatitis E in Asia, Africa and Latin America. Genotype 2 has so far only been isolated from humans infected in Mexico and sub-Saharan Africa [21–24]. Genotype 3 has a worldwide distribution and has been isolated from humans, swine, wild boars, mongooses and rabbits both in endemic and non-endemic countries, while genotype 4 strains have been isolated from humans, wild boars, pigs and deer in Asia, particularly in Japan, China, Vietnam, Taiwan, and India. It is evident that there is a domestic source of genotype 3 infections, but the infection route is generally not known although spread from animals to humans by ingestion of undercooked liver and deer meat has been documented [6, 8, 9]. Genotype 3 infection leads to hepatitis mainly in persons aged >45 years [10]. In Sweden several cases of non-travel-related HEV genotype 3 infections in humans have occurred and previous studies demonstrated that genotype 3 is also present in Swedish pigs [16, 25, 26]. However, little is known regarding the prevalence of HEV in the Swedish pig population and the prevalence of HEV in the Swedish wild boar population is not known at all. The genetic relationship between strains from humans, pigs and wild boars in Sweden is also not known. In order to gain more information on the prevalence of zoonotic transmission of genotype 3 infections, samples from Swedish domestic pigs and wild boars were collected. Partial ORF1 and ORF2 of the HEV genomes were sequenced, analysed and

compared with the corresponding regions in strains from humans.

## MATERIALS

### Human serum samples

A total of 102 human serum samples were used for typing HEV strains. The samples, collected between 1993 and 2009, were reactive for anti-HEV IgM and/or IgG at the Swedish Institute for Infectious Disease Control. All samples were reactive for anti-HEV IgG and 57 of those were also reactive for anti-HEV IgM. The samples were from Swedish patients with clinical signs of hepatitis not caused by hepatitis A, B, C or D virus. Most patients had a recent travel history. The sera were tested for HEV IgG and IgM by a commercial ELISA using two recombinant HEV antigens corresponding to a structural region of HEV (Diagnostic Biotechnology, Singapore). Between 1993 and 2006, all samples were also tested for HEV IgG by the then commercially available ELISA kits from Abbott Laboratories (USA). During 2009 the samples were also tested for anti-HEV reactivity by another commercially available ELISA, Recowell (Mikrogen GmbH, Germany). All reactive sera were tested for HEV RNA by polymerase chain reaction (PCR) and the amplified fragments were sequenced.

### Pig samples

Faecal samples ( $n=240$ ) were collected from 22 pig farms selected at random in south Sweden. The samples were from piglets aged between 2 and 4 months. Fresh faecal samples were collected from ten pigs per farm. For each farm, each pig sampled was kept in a box separate from the others. Faecal samples were stored at  $-20^{\circ}\text{C}$  until processed. All pig samples were tested for HEV RNA by real-time PCR as described previously [25].

### Wild boar samples

A total of 159 wild boar blood samples were selected from samples sent in by hunters. The wild boars were from nine different counties in southern Sweden. Sixty-four (40%) were piglets and 95 (60%) were yearlings. The age of the sampled wild boars was estimated by the hunter and classified as either piglet or yearling. Samples exhibiting excessive haemolysis

or foul smell were not tested. The sera were stored at  $-20^{\circ}\text{C}$  until processed. All samples were tested for HEV RNA by real-time PCR as described previously [25].

## METHODS

### Homogenization of faecal samples

Pig faecal samples were diluted about 1:10 in TE buffer (Tris EDTA; pH 7.6), homogenized in grinding tubes as described previously [25] and stored at  $-80^{\circ}\text{C}$ .

### RNA extraction

HEV RNA was extracted from 200  $\mu\text{l}$  human sera as previously described [16]. HEV RNA was extracted from 140- $\mu\text{l}$  faecal sample supernatant or wild boar sera with the QIAamp viral RNA mini kit (Qiagen, Germany) and stored at  $-80^{\circ}\text{C}$ .

### cDNA synthesis

Five microlitres of RNA were used for cDNA synthesis in a 20- $\mu\text{l}$  mix that included 100 U Superscript II (Invitrogen, USA), which was performed according to the manufacturer's instructions.

### Screening for HEV in pig and wild boar samples

#### *TaqMan*<sup>®</sup> assay for faecal samples

Assays were run in 12  $\mu\text{l}$  reaction mix per tube as described previously [25], with 2  $\mu\text{l}$  cDNA added as template.

#### *TaqMan* assay for serum samples from wild boars

RNA from wild boar serum samples were analysed by a combined reverse transcription (RT) and PCR program using the Qiagen OneStep RT-PCR kit (Germany). From each RNA sample, 2.5  $\mu\text{l}$  was analysed. Assays were run in 12  $\mu\text{l}$  reaction mix per tube with the following cycling conditions:  $50^{\circ}\text{C}$  for 30 min,  $95^{\circ}\text{C}$  for 2 min followed by two-step cycling 45 times at  $94^{\circ}\text{C}$  for 15 s and  $56^{\circ}\text{C}$  for 60 s. Fluorescence was monitored during the annealing step of each cycle.

#### PCR amplification in ORF1

A nested PCR was carried out in a 50  $\mu\text{l}$  reaction with 5  $\mu\text{l}$  cDNA,  $10\times$  *Taq* buffer, 2 mM  $\text{MgCl}_2$  (Applied Biosystems, Roche Molecular Systems, USA), 4 U

*Taq* polymerase, 0.2 mM dNTP (Thermo Scientific, Abgene<sup>®</sup>, UK) or as a combined RT-PCR protocol with 5  $\mu\text{l}$  of the extracted RNA (OneStep master mix, OneStep RT-PCR enzyme mix; Qiagen). The primers ISP-4232 and EAP-4576 [27] (0.06  $\mu\text{l}$  of 0.2 mM of each) were used. RT was performed at  $50^{\circ}\text{C}$  for 30 min. The PCR reaction was performed for 40 cycles, each cycle consisting of denaturation at  $94^{\circ}\text{C}$  for 20 s, annealing at  $60^{\circ}\text{C}$  for 30 s and extension at  $72^{\circ}\text{C}$  for 60 s. Cycling conditions for the second round were identical to the first. Five microlitres of the first-round product was added as template for the second round and 2.5 mM  $\text{MgCl}_2$  and 0.06  $\mu\text{l}$  of 0.2 mM of each primer ISP-4232 and IAP-4561 [27] were used.

#### PCR amplification in ORF2

The PCR in ORF2 was performed, as described previously [28], in a 50  $\mu\text{l}$  reaction with 10  $\mu\text{l}$  cDNA as template. Primers HE110 and HE041 were used for the first round, and primers HE110 and HE3159 for the second round.

### Sequencing

The amplified products were purified using the EZNA Cycle Pure kit (Omega Bio-Tek, USA) or the Qiagen PCR purification kit (Qiagen). The sequencing reaction was made with BigDye Terminator Cycle Sequencing Ready reaction kit version 3.1 (Applied Biosystems, USA) with the primers used in the PCR as sequencing primers. The ABI Prism 3100 genetic analyser (Applied Biosystems) was used for electrophoresis and data collection.

### Phylogenetic analysis

The sequences obtained were analysed in the programs DNASTAR SeqMan and Sequencing Analysis. The sequences were aligned with the corresponding region of 554 sequences obtained from GeneBank. Phylogenetic analysis was carried out with the PHYLIP package version 3.65 [29]. Evolutionary distances were calculated using the F84 algorithm in the DNADIST program with a transition/transversion ratio of 4.29. Phylogenetic trees were constructed using the unweighted pair-group method using arithmetic averages (UPGMA) and the neighbour-joining method in the NEIGHBOR program of the PHYLIP package. The trees were visualized using the program Tree View, version 1.6.6. Bootstrap analysis of 1000

Table 1. Country of infection of humans with IgG or IgM reactivity reactive to IgM and IgG and to IgG alone against hepatitis E virus (HEV) in relation to HEV genotype obtained by sequencing HEV strains in the serum samples

Country of infection	No. of samples	No. positive for HEV RNA	Samples sequenced/ IgM pos.	Samples sequenced/ IgG pos.	Genotype	
					1	3
Europe	12	7	5/5	2/7	1	6
Sweden	1	0	—	0/1	—	—
Bulgaria	1	0	—	0/1	—	—
Spain	1	0	—	0/1	—	—
Italy	1	1	1/1	—	0	1
Mallorca/Austria	1	1	1/1	—	0	1
Serbia/Montenegro						
Subtotal	17	9	7/7 (100%)	2/10 (20%)	1	8
Asia	1	0	—	0/1	—	—
Afghanistan	4	1	1/3	0/1	1	0
Bangladesh	14	10	7/11	1/3	8	0
India	31	25	20/23	4/8	24	0
Nepal	4	4	4/4	—	4	0
Pakistan	8	8	6/6	2/2	8	0
Thailand	1	0	—	0/1	—	—
Vietnam	1	0	—	0/1	—	—
Mongolia	1	1	1/1	—	0	1
Subtotal	65	49	39/48 (81%)	7/17 (41%)	45	1
Middle East						
Syria	1	0	—	0/1	—	—
Turkey	1	0	—	0/1	—	—
Subtotal	2	0	0	0/2	0	0
South America						
Dominican Republic	1	0	—	0/1	0	0
Not reported	17	1	0/3	1/14	1	0
Total	102	58	45/57 (84%)	10/45 (22%)	47	9

replicas was performed with the programs SEQBOOT and CONSENSE in the PHYLIP package.

## RESULTS

HEV RNA could be amplified in 58/102 human anti-HEV-positive samples (Table 1). Most of the samples with detectable HEV RNA were from patients with anti-HEV IgM (48/58), while 10 (22%) were from patients with detectable anti-HEV IgG only without anti-HEV IgM. Sixty-seven (64%) of the 102 patients had been infected while in Asia or the Middle East. However, 17 had been infected in Europe (Table 1). The HEV strains could be sequenced in 56 samples, 46 of which were from patients infected outside Europe. All these patients were infected with genotype 1 (Table 1). One genotype

3-infected patient was a newly arrived immigrant from Mongolia. It was not known if he was infected en route to Sweden. The HEV strains in sera from the 17 patients infected in Europe were all but one of genotype 3 (Table 1). The patient infected with genotype 1 in Sweden, was infected while taking care of his brother, who became infected during a visit in Pakistan [16]. The patients infected with genotype 3 were all aged >46 years, and the majority (7/9), were male (Table 2).

HEV RNA could be amplified by real-time PCR in 71 (29.6%) of 240 faecal samples collected from pigs aged 2–4 months from 22 pig farms (Fig. 1). HEV RNA was detected in samples from piglets in 16 (72.7%) of the farms. Thirty of the HEV RNA samples were sequenced in ORF1 and 26 samples were sequenced in ORF2. All were genotype 3 strains.

Table 2. Strains of HEV genotype 3, sex, age, and country of infection of patients positive for the virus

Strain	Sex	Age (yr)	Country of infection
26-02	M	60	Sweden
560-05	M	67	Sweden
1459-06	F	62	Spain/Austria
1559-07	F	65	Serbia/Montenegro
668-08	M	49	Sweden
366-09	M	67	Sweden
640-09	M	46	Sweden
748-09	M	68	Sweden
995-09	M	Not known	Mongolia immigrant

The age of the Mongolian immigrant was not known.

Thirteen (8.2%) of 159 samples from wild boars had detectable HEV RNA by real-time PCR. The infected wild boars were from five of the nine investigated counties (Fig. 1). Eight (14.8%) of the wild boar piglets, and five (4.8%) of the older wild boars were infected. ORF1 was partially sequenced in seven and ORF2 in six wild boar samples, all were of genotype 3. In five of these samples both ORF1 and ORF2 were partially sequenced.

Phylogenetic comparison of the sequences demonstrated that the genotype 3 strains subdivided into two groups, designated 3-I and 3-II both in ORF1 and ORF2 as previously shown for ORF2 (Figs 2 (a), 3, 4; Supplementary Table 1 (available online) [16]). Both groups contained strains from pigs and humans intermixed in the tree. Based on analysis of the sequences, the 3-I group contained strains of subtypes 3a, b, c, h, and j as well as three additional groups of strains with undefined subtypes (Fig. 2). The subtype 3a strains were from the USA, South Korea, and Japan. Those of subtype 3b were from Japan and China, the 3h strain was from Mongolia, and the 3j strain from Canada (Fig. 2). Ten Swedish strains were found in this group. One strain was from a woman infected in Serbia/Montenegro. The HEV strain infecting her was similar to strains from South Korea. One separate branch was formed by strains from a Swedish man and a wild boar. One additional branch was formed by two strains from German wild boars and a strain from a Swedish man. A third branch was formed by six strains from Swedish piglets at farm 11 (Fig. 2, b). These subdivisions of the strains were confirmed in ORF2 (Supplementary Fig. 1, available online).

Most of the Swedish strains were found in group 3-II. This group contained strains of subtypes 3e, 3f,

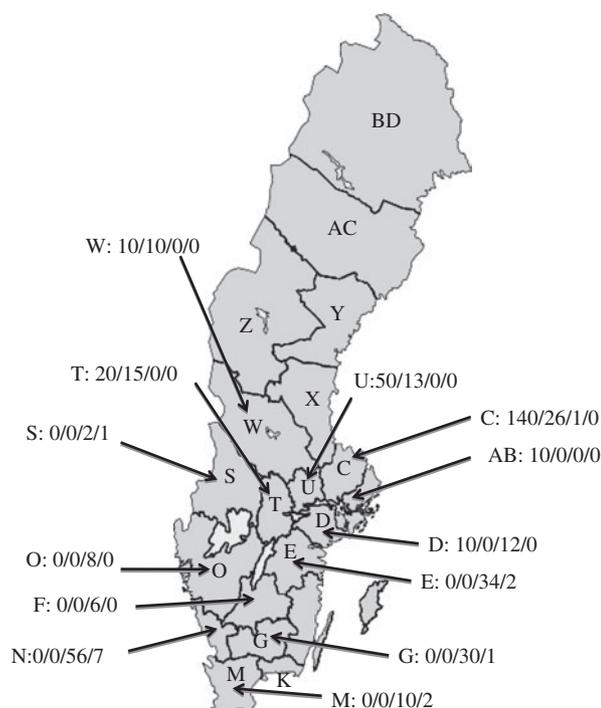
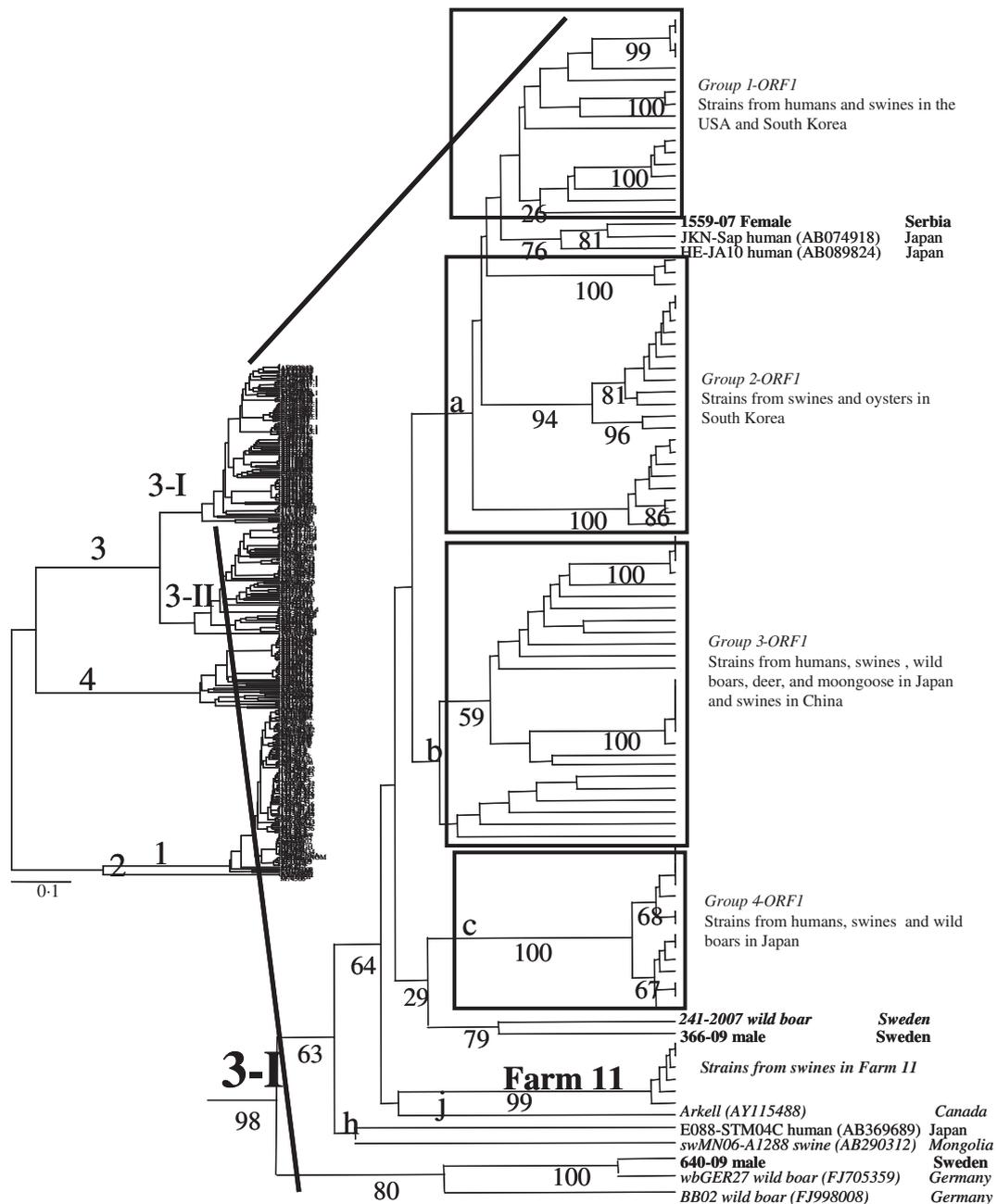


Fig. 1. Map of Sweden showing the counties labelled with the official county letter. Counties where samples were collected are indicated with arrows. The arrows are labelled with the official county letter followed by number of pigs sampled/number of PCR-positive pigs/number of wild boar sampled /number of PCR-positive wild boar in the county indicated. AB, Stockholm county; C, Uppsala county; D, Sörmland; E, Östergötland; F, Jönköping county; G, Kronoberg county; H, Kalmar county; K, Blekinge county; M, Skåne region; N, Halland county; O, Västra Götaland region; S, Värmland county; T, Örebro county; U, Västmanland county; W, Dalarna county; X, Gävleborg county; Y, Västernorrland; Z, Jämtland county; AC, Västerbotten county; BD, Norrbotten county; I, Gotland municipality.

and 3g (Fig. 3). The 3f strains originated from Europe, Thailand, and Mongolia, while the 3e strains originated from Germany and Japan. The 3g strain was from a pig from Kyrgyzstan. Moreover, in the 3-II group, the strains from the Swedish piglets formed separate branches according to origin. The strains from farms A, 1, 3, 4, 12, and 46, were of subtype 3f while the strains from farms 10 and BT were of subtype 3e (Fig. 3). One strain from farm A was similar to seven strains from farm 1, and clustered with those on the same branch. Another three strains from farm 1 were divergent and formed a separate branch (Fig. 3). Six strains from Swedish wild boars belonged to subtype 3f. All strains from Halland county were grouped as a separate branch. One of the

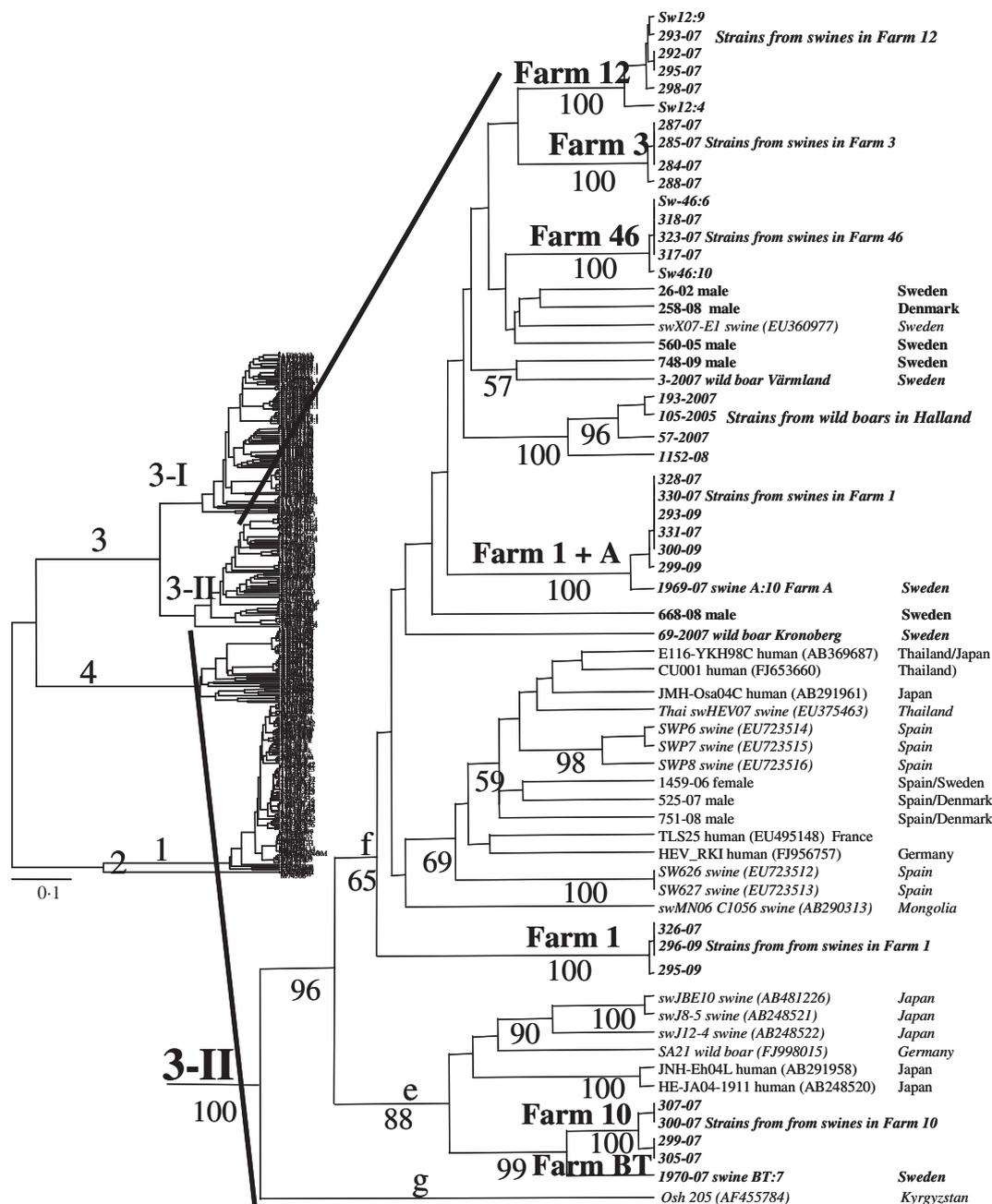


**Fig. 2.** UPGMA dendrogram based on 346 nucleotides of ORF1 in 309 HEV strains. The genotypes and subdivision of genotype 3 into two groups are shown on the branches. The branch of 3-I strains is shown. The subtypes are indicated on the branches. Origin of strains from Swedish pigs is indicated on the branch, by the designation of the pig farm. Strains sequenced and described in this study are shown in bold. Strains from animals are indicated in italic. Bootstrap values of 1000 replications are indicated below the branches. Accession numbers of sequences obtained from GenBank are given at the nodes or for sequences within groups 1–4 in Supplementary Table 1 (available online).

other strains was distantly similar to a strain from a man infected in Sweden (Fig. 3). The humans infected in Spain all were infected by strains similar to strains from Spanish pigs. The similarities and sub-clustering of the strains were confirmed on analysis of ORF2 (Fig. 4).

**DISCUSSION**

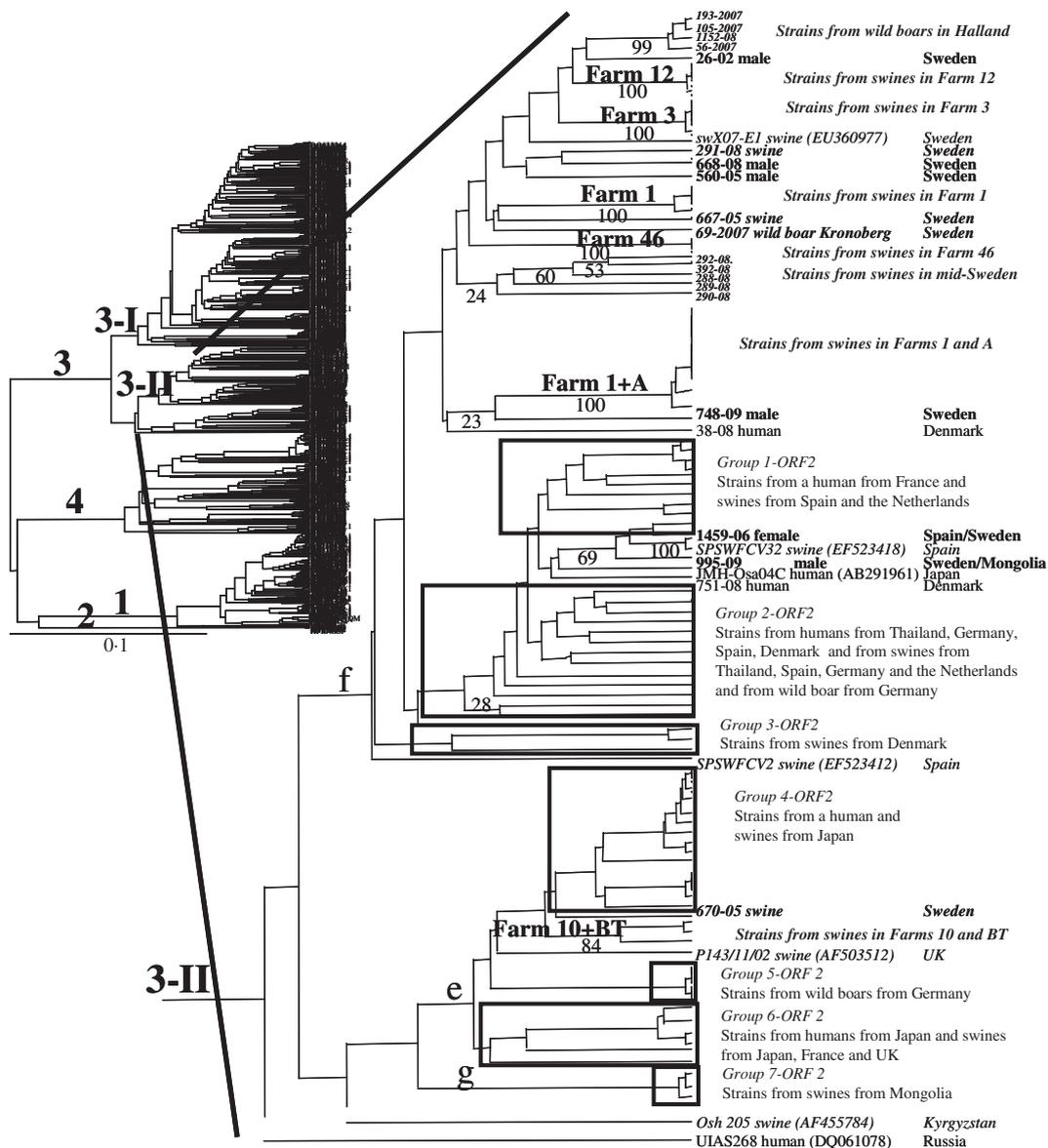
This study confirmed the prevalence of hepatitis E in humans, domestic pigs and wild boars in Sweden. Most identified human cases were travel related, mainly from Asia, and were infected with the HEV



**Fig. 3.** UPGMA dendrogram based on 346 nucleotides of ORF1 in 309 HEV strains. The genotypes and subdivision of genotype 3 into two groups are shown on the branches. The branch formed by 3-II strains is shown. Origin of strains from Swedish pigs is indicated on the branch, by the designation of the pig farm. Strains sequenced and described in this study are shown in bold. Strains from animals are indicated in italic. Bootstrap values of 1000 replications are indicated below the branches. Accession numbers of sequences obtained from GenBank are given at the nodes or for sequences within groups 1–4 in Supplementary Table 1 (available online).

genotype 1. However, those infected in Sweden or in other European countries were all infected with genotype 3. This was the only genotype found in Swedish domestic pigs and wild boars. Based on the geographical distribution of infected animals and human cases, this study demonstrates a probable endemic circulation of HEV genotype 3 in Sweden.

Phylogenetic analysis of the sequences showed that the previously described subdivision of genotype 3 strains into two groups, 3-I and 3-II, when comparing ORF2 sequences [17], was confirmed when more samples were analysed both in ORF1 and ORF2. Most of the Swedish strains were found in group 3-II and belonged to subtype 3f, although piglets from two



**Fig. 4.** UPGMA dendrogram based on 276 nucleotides of ORF2 in 640 HEV strains. The genotypes and subdivision of genotype 3 into two groups are shown on the branches. The details of the branch formed by 3-II strains are shown. The subtypes are indicated on the branches. Origin of strains from Swedish pigs is indicated on the branch, by the designation of the pig farm. Strains sequenced and described in this study are shown in bold. Strains from animals are indicated in italic. Bootstrap values of 1000 replications are indicated below the branches. Accession numbers of sequences obtained from GenBank are given at the nodes or for sequences within groups 1-3 in Supplementary Table 1 (available online). The details of branch 3-I strains are not shown here but are available in Supplementary Fig. 1 (available online).

farms were infected by 3e strains. Subtype 3f is the most common subtype found in pigs in Europe, e.g. France, The Netherlands, and Spain [30–34] although subtype 3e has also been found in European pigs [30, 34]. This study showed that HEV prevalence in the Swedish pig population is high. The high prevalence in Swedish piglets aged 2–4 months, which is in accord with a study from France [30], indicates that this is the age when HEV is spreading most

rapidly. In the French study piglets aged 2 months had been moved to a farm, where they became infected by the same strains [30]. This French farm had two strains circulating – subtypes 3e and 3f. In our study each Swedish farm also had one, or sometimes two specific HEV strains, which could be distinguished from all other strains by phylogenetic analysis. The high prevalence of HEV in the majority of the Swedish farms may be due to a mixing of young

piglets from different litters leading to an efficient spread of HEV.

The prevalence of HEV in the wild boars in our study is lower than in domestic pigs. However, the age distribution and sample type were not the same in the two groups and it is anticipated that the viral load is lower in sera than in liver, bile or faeces. The lower prevalence in the wild boars may also reflect a slower spread of the virus due to a lower population density than in a pig farm. Most of the HEV strains from wild boars belonged to subtype 3f but one was found in group 3-I and may represent a new subtype. The strains clustered mainly according to origin with all strains from Halland county forming one branch in the phylogenetic tree. This geographical clustering was also found in German wild boars, although the strains were more diverse with representatives from several subtypes [35, 36]. Furthermore, in the German study a difference between urban and rural regions could be seen. This could not be studied in Sweden since there are no urban wild boar populations in Sweden. The difference in prevalence of HEV between the two studies may also be attributed to difference in sample material.

Most HEV strains from humans infected in Europe were of subtype 3f. Those infected in Spain had strains related to strains from Spanish pigs, while most of the strains from those infected in Sweden were related to strains from Swedish domestic pigs or wild boars. One Swedish man was infected with a strain related to a German wild boar. The similarity between strains from humans and pigs or deer from the same country has previously been demonstrated [16, 37–41]. However, the analysis of ORF1 and ORF2 sequences in this study shows that the country or even county of origin of the infecting HEV strain can be determined. Further studies are needed to investigate if there is a permanent spread of specific strains in each pig farm and if there are specific strains spread in the wild boars in the different Swedish counties.

Since HEV infections are frequent in Swedish domestic pigs and wild boars the risk of human infection from them is possible. This can occur for example, when wild boars are handled after being shot by hunters or if undercooked pig and wild boar products are consumed, as has been shown in Japan [8]. This study demonstrates that phylogeny can be a useful tool for tracing the source of non-travel-related HEV infections and in determining the geographical origin of HEV genotype 3 strains infecting humans.

The usefulness of this tool for tracing human infections is dependent on regular screenings of pig and the wild boar population. It is reasonable to assume that if more information regarding HEV strains circulating in regional pig and wild boar populations was available, the precision and reliability of this model for tracing infections would increase significantly.

## CONCLUSION

This study demonstrates that HEV is common in Swedish pigs and wild boars. It also demonstrates that phylogeny based on short sequences from ORF1 and ORF2 is a useful tool for tracing the origin of human HEV infections. By using this tool it was concluded that there is a strong similarity between HEV in humans on the one hand and in pigs and wild boars on the other. This study indicates that in Sweden human HEV infection may be zoonotic. It is evident that pigs and wild boars may be the reservoirs.

## NOTE

Supplementary material accompanies this paper on the Journal's website (<http://journals.cambridge.org/hyg>).

## ACKNOWLEDGEMENTS

Thanks are due to Maria Lindberg and Marie Sjölund at Swedish Animal Health for collecting samples from pigs, and Per Wallgren for establishing the contact with Swedish Animal Health. Thanks are also due to the Swedish hunters that collected and sent in wild boar blood samples and to Maj Hjort for preparing the samples. This study was supported by Pathogen Combat, contract no. FOOD-CT-2005-007081, European Union Network of Excellence 'Network for the prevention and control of Zoonoses (Med-Vet-Net)', contract no. 506122. Support was also received from the 'Ivar and Elsa Sandberg Foundation', Sweden. Part of this work was also supported by the European Commission DG Research Quality of Life Program, 6th Framework (EVENT, SP22-CT-2004-502571).

## DECLARATION OF INTEREST

None.

## REFERENCES

1. Purcell RH, Emerson SU. Hepatitis E: an emerging awareness of an old disease. *Journal of Hepatology* 2008; **48**: 494–503.
2. Meng XJ. Recent advances in hepatitis E virus. *Journal of Viral Hepatitis* 2010; **17**: 153–161.
3. Khuroo MS, *et al.* Incidence and severity of viral hepatitis in pregnancy. *American Journal of Medicine* 1981; **70**: 252–255.
4. Balayan MS, *et al.* Evidence for a virus in non-A, non-B hepatitis transmitted via the fecal-oral route. *Inter-virology* 1983; **20**: 23–31.
5. Tei S, *et al.* Zoonotic transmission of hepatitis E virus from deer to human beings. *Lancet* 2003; **362**: 371–373.
6. Khuroo MS, Kamili S, Jameel S. Vertical transmission of hepatitis E virus. *Lancet* 1995; **345**: 1025–1026.
7. Matsuda H, *et al.* Severe hepatitis E virus infection after ingestion of uncooked liver from a wild boar. *Journal of Infectious Diseases* 2003; **188**: 944.
8. Takahashi K, *et al.* Complete or near-complete nucleotide sequences of hepatitis E virus genome recovered from a wild boar, a deer, and four patients who ate the deer. *Virology* 2004; **330**: 501–505.
9. Gérolami R, Moal V, Colson P. Chronic hepatitis E with cirrhosis in a kidney-transplant recipient. *New England Journal of Medicine* 2008; **358**: 859–860.
10. Dalton HR, *et al.* Hepatitis E: an emerging infection in developed countries. *Lancet Infectious Diseases* 2008; **8**: 698–709.
11. Ijaz S, *et al.* Non-travel-associated hepatitis E in England and Wales: demographic, clinical, and molecular epidemiological characteristics. *Infectious Diseases* 2005; **192**: 1166–1172.
12. Mansuy JM, *et al.* Hepatitis E in the south west of France in individuals who have never visited an endemic area. *Journal of Medical Virology* 2004; **74**: 419–424.
13. Tsang TH, *et al.* Acute hepatitis E infection acquired in California. *Clinical Infectious Diseases* 2000; **30**: 618–619.
14. Widdowson MA, *et al.* Cluster of cases of acute hepatitis associated with hepatitis E virus infection acquired in the Netherlands. *Clinical Infectious Diseases* 2003; **36**: 29–33.
15. Norder H, *et al.* Endemic hepatitis E in two Nordic countries. *Eurosurveillance* 2009; **14**.
16. Lopes Dos Santos DR, *et al.* First report of a human autochthonous hepatitis E virus infection in Brazil. *Journal of Clinical Virology* 2010; **47**: 276–279.
17. Chandra V, *et al.* The hepatitis E virus ORF3 protein modulates epidermal growth factor receptor trafficking, STAT3 translocation, and the acute-phase response. *Journal of Virology* 2008; **82**: 7100–7110.
18. Moin SM, *et al.* The hepatitis E virus ORF3 protein stabilizes HIF-1 $\alpha$  and enhances HIF-1-mediated transcriptional activity through p300/CBP. *Cellular Microbiology* 2009; **11**: 1409–1421.
19. Zafrullah M, *et al.* The ORF3 protein of hepatitis E virus is a phosphoprotein that associates with the cytoskeleton. *Journal of Virology* 1997; **71**: 9045–9053.
20. Caron M, *et al.* Identification of genotype 1 hepatitis E virus in samples from swine in Cambodia. *Journal of Clinical Microbiology* 2006; **44**: 3440–3442.
21. Buisson Y, *et al.* Identification of a novel hepatitis E virus in Nigeria. *Journal of General Virology* 2000; **81**: 903–909.
22. van Cuyck-Gandré H, *et al.* Characterization of hepatitis E virus (HEV) from Algeria and Chad by partial genome sequence. *Journal of Medical Virology* 1997; **53**: 340–347.
23. Huang CC, *et al.* Molecular cloning and sequencing of the Mexico isolate of hepatitis E virus (HEV). *Virology* 1992; **191**: 550–558.
24. Nicand E, *et al.* Genetic heterogeneity of hepatitis E virus in Darfur, Sudan, and neighboring Chad. *Journal of Medical Virology* 2005; **77**: 519–521.
25. Gyarmati P, *et al.* Universal detection of hepatitis E virus by two real-time PCR assays: TaqMan and Primer-Probe energy transfer. *Journal of Virological Methods* 2007; **146**: 226–235.
26. Xia H, *et al.* Molecular characterization and phylogenetic analysis of the complete genome of a hepatitis E virus from European swine. *Virus Genes* 2008; **37**: 39–48.
27. Zhai L, Dai X, Meng J. Hepatitis E virus genotyping based on full-length genome and partial genomic regions. *Virus Research* 2006; **120**: 57–69.
28. Mizuo H, *et al.* Polyphyletic strains of hepatitis E virus are responsible for sporadic cases of acute hepatitis in Japan. *Journal of Clinical Microbiology* 2002; **40**: 3209–3218.
29. Felsenstein, J. PHYLIP (Phylogeny Inference Package) version 3.6. Distributed by the author. 2005. Department of Genome Sciences, University of Washington, Seattle (<http://evolution.genetics.washington.edu/phylip/getme.html>). Accessed 14 December 2009.
30. Kaba M, *et al.* Frequent transmission of hepatitis E virus among piglets in farms in Southern France. *Journal of Medical Virology* 2009; **81**: 1750–1759.
31. Legrand-Abravanel F, *et al.* Hepatitis E virus genotype 3 diversity, France. *Emerging Infectious Diseases* 2009; **15**: 110–114.
32. van der Poel WH, *et al.* Hepatitis E virus sequences in swine related to sequences in humans, The Netherlands. *Emerging Infectious Diseases* 2001; **7**: 970–976.
33. Peralta B, *et al.* Genetic characterization of the complete coding regions of genotype 3 hepatitis E virus isolated from Spanish swine herds. *Virus Research* 2009; **139**: 111–116.
34. Rutjes SA, *et al.* Sources of hepatitis E virus genotype 3 in The Netherlands. *Emerging Infectious Diseases* 2009; **15**: 381–387.
35. Schielke A, *et al.* Detection of hepatitis E virus in wild boars of rural and urban regions in Germany and whole genome characterization of an endemic strain. *Virology Journal* 2009; **6**: 58.
36. Baechlein C, *et al.* Prevalence of Hepatitis E virus-specific antibodies in sera of German domestic pigs

- estimated by using different assays. *Veterinary Microbiology*. Published online: 16 December 2009. doi:10.1016/j.vetmic.2009.12.011.
37. **Meng XJ, et al.** A novel virus in swine is closely related to the human hepatitis E virus. *Proceedings of the National Academy of Sciences USA* 1997; **94**: 9860–9865.
38. **Schlauder GG, et al.** The sequence and phylogenetic analysis of a novel hepatitis E virus isolated from a patient with acute hepatitis reported in the United States. *Journal of General Virology* 1998; **79**: 447–456.
39. **Kaba M, et al.** Detection of hepatitis E virus in wild boar (*Sus scrofa*) livers. *Veterinary Journal*. Published online: 10 September 2009. doi:10.1016/j.tvjl.2009.08.008.
40. **Fogeda M, et al.** Imported and autochthonous hepatitis E virus strains in Spain. *Journal of Medical Virology* 2009; **81**: 1743–1749.
41. **Reuter G, et al.** Characterization and zoonotic potential of endemic hepatitis E virus (HEV) strains in humans and animals in Hungary. *Journal of Clinical Virology* 2009; **44**: 277–281.