

Application of Analytical Electron Tomography to the Study of Pathogenic Protozoa

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Understanding mechanisms involved in osmoregulation control in protozoan parasites has been a challenge for many research groups. Over the past years, a number of key players in cell signaling in trypanosomatid parasites have been identified. Among these, cyclic AMP (cAMP) has been shown to play a key role in osmoregulation, through a mechanism that involves a cAMP-dependent pathway that leads to the efflux of osmolytes across the parasite surface, and water elimination through a contractile vacuole complex (CVC). In *Trypanosoma cruzi*, the CVC is formed by a central vacuole surrounded by a collection of interconnected vesicles and tubules that undergo dynamic changes upon osmotic stress. A unique characteristic of this system is the presence of acidic calcium-rich organelles named acidocalcisomes, whose structural organization, chemical properties and physiological activity may vary upon events of osmotic stress. The structure of the osmoregulatory system of Trypanosomes has been described in a few Works [1]. One of the main restrictions for the electron microscopy characterization of this system is that it is highly sensitivity to chemical fixation. This becomes specially critical in trypanosomatids since these cells present a unique osmoregulatory system that seem to congregate a contractile vacuole complex that operates with acidocalcisomes, also been shown to be highly sensitive to chemical fixation [2-4]. To minimize structural changes and ion extraction during sample preparation, cells were submitted to quick freezing and freeze substitution or freeze drying prior to electron microscopy analysis. Sections of freeze-substituted cells or freeze-dried whole cells were submitted to FIB-SEM, serial electron tomography and STEM tomography. For analytical tomography, cells were submitted to plunge freezing and freeze drying. Results showed the detailed organization of the contractile vacuole complex of *Trypanosoma cruzi* containing a central bladder and a series of tubules that form a spongione. In addition, a close contact with ion containing organelles named acidocalcisomes was observed. Analytical electron tomography showed an heterogeneous 3D distribution of ions such as phosphate, calcium, magnesium and zinc within the organellar matrix. Electron tomography has tremendously contributed to the understanding of the structural organization of the osmoregulatory system in *Trypanosoma cruzi* and the use of analytical electron tomography to define the 3D distribution of diffusible ions within the organelles of the parasite demonstrate a clear contribution of this technique to this field of life sciences.

[1] Girard-Dias *et al*, Histochemistry and Cell Biology **138** (2012),821-831.

[2] Miranda *et al*, Histochemmistry and Cell Biology **121** (2004), 407-418.

[3] Miranda *et al*, Protist **155** (2004), 395-405.

[4] Ramos *et al.*, PLoS One 6 (2011), e27276.

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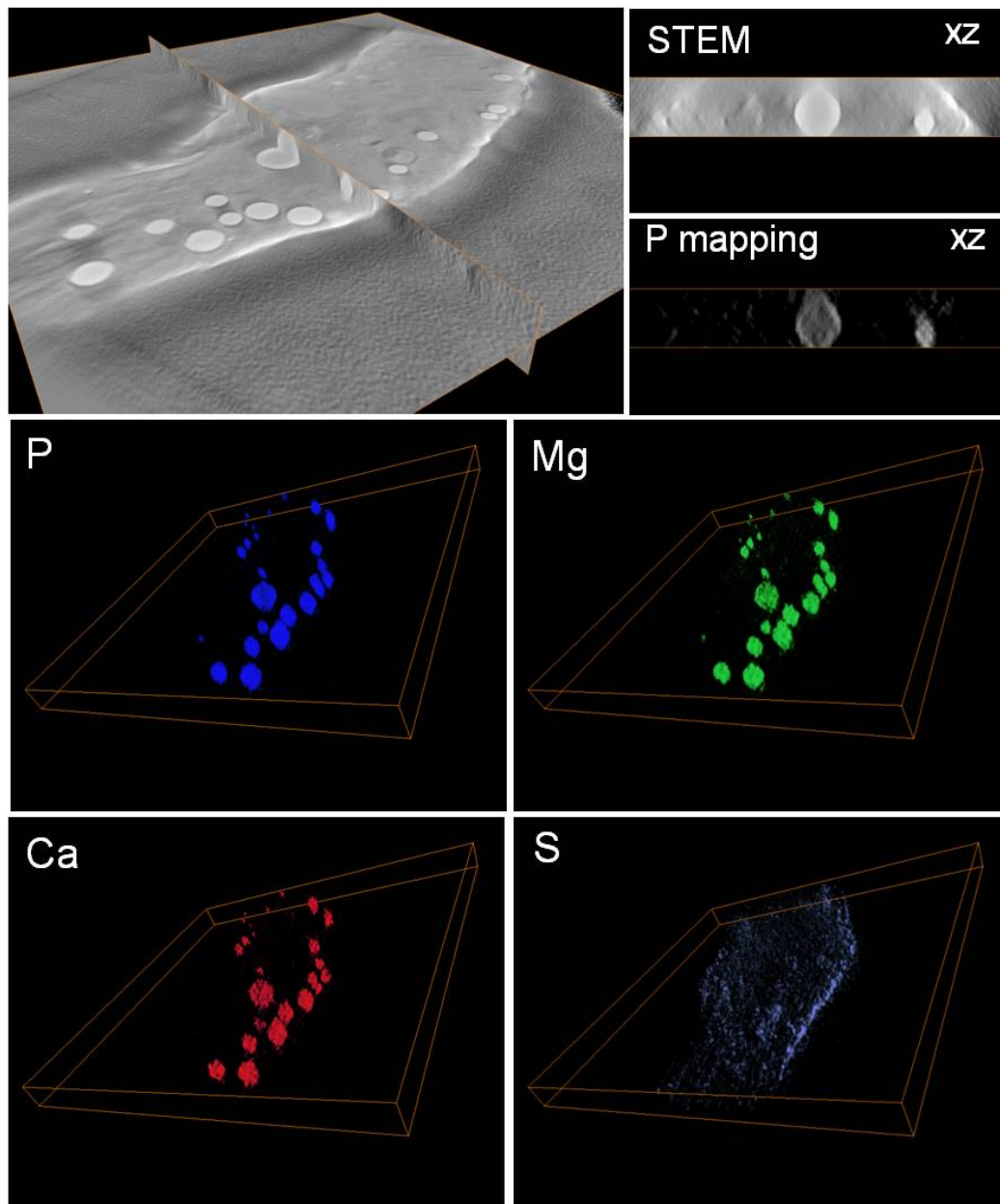


Figure 1. Analytical electron tomography of plunge frozen and freeze substituted *Trypanosoma cruzi* cells, showing the 3D distribution of phosphorus, magnesium, calcium and sulfur within the parasite body.