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Profiling of foxtail millet (*Setaria italica* L.) germplasm using *in vitro* drought screening and SSR markers

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Abstract

Foxtail millet (Setaria spp.) is an ancient cereal crop, having a short cropping cycle. Drought tolerance was assessed in this crop at an early growth stage and the extent of genetic diversity was measured between the foxtail millet genotypes, applying DNA markers. Tolerance of 18 foxtail millet genotypes was studied in vitro under four levels of polyethylene glycol (0, 10, 20 and 30% PEG-6000). PEG-6000 decreased final germination percentage and led to a reduction in shoot and root length with different stress levels. The genotypes ISe 869, ISe 1851 and vellow spike show superiority in stress tolerance for germination and the growth of root and shoot traits. They also clustered together in the biplot diagram and dendrogram of the genotypes based on the morphological traits. Marker polymorphism index (PI) was 80.36% and a total of 132 polymorphic alleles (4.00 alleles/locus) were obtained from 33 polymorphic primers. Polymorphic information content (0.54–0.83) was highly informative with an average value of 0.67. A dendrogram distributed the genotypes into five distinct clusters based on simple-sequence repeat (SSR) data, independent of their geographical distribution. A relationship was established between the SSR markers and the genotypes ability to tolerate drought stress. The SSR markers used could contribute to conducting DNA profiling of foxtail millet, and facilitating their use in future breeding programmes for drought tolerance in this crop. Based on water-stress experiment, three most tolerant genotypes: ISe 869, ISe 1851 and yellow spike are recommended to be cultivated under drought conditions around the world.

Introduction

Foxtail millet (*Setaria italica* (L.) Beauv.) (2n = 18) is an important minor millet in the world, grown under different environmental conditions as a drought-tolerant crop (Reddy *et al.*, 2020). It is an annual and self-pollinating crop of the *Setaria* genus (Rodiansah and Puspita, 2020), and suggested to have been domesticated nearly 8700 years ago (Zohary *et al.*, 2012) from wild green foxtail (*Setaria viridis*), that is native to temperate areas of Asia and Europe (Hu *et al.*, 2018). The crop has a short-life cultivation of around 3 months (Martin *et al.*, 2021), which provides an opportunity for growing this crop in drought-prone and heat-stressed areas. Foxtail millet grain contains high protein of 12.3–12.5% (Ji *et al.*, 2019), 5.5% oil, 60.9% carbohydrates (Dhumketi *et al.*, 2017) and an ash content of 3.0% (Ch *et al.*, 2020), and has many health and medicinal benefits (Ghimire *et al.*, 2018).

Currently, different parts of the world, including Iraq, are facing water scarcity. Up to 40% of the world's population will live under severely water-stressed condition by 2035 (Guppy et al., 2017). One of the solutions for food and nutritional security in such regions is the introduction of new staple and minor crops into agricultural production. Millet is a strong candidate crop in the context of climate-resilient agriculture (Muthamilarasan et al., 2019). It offers better adaptation and reasonable yield under uncertain conditions compared to major crops, having well-developed adventitious root systems (Ceasar et al., 2020). Development of crop varieties with lower water requirement and higher drought tolerance is an important component of water-use efficiency, to satisfy food demand of the steadily increasing world population (Fahad et al., 2017; Ierna, 2021). Drought stress limits the availability of water that is needed for germination and subsequently compromises the establishment of seedlings (Pavli et al., 2020; Kintl et al., 2021). The ability of plants to persist under drought conditions depends on their capacity to initiate internal signal transduction at different stress levels (Gull et al., 2019; Yang et al., 2021). Short-term drought stress has been reported to affect plant growth, leading to declining growth and crop yield (Niu et al., 2018; Kapoor et al., 2020). Selection of drought-tolerant cultivars through in vitro screening is an important approach to improving foxtail millet. PEG-6000 as a high molecular weight that has been reported to induce uniform water stress for drought stress screening, with no physiological damage to the plant (Shivakrishna et al., 2018).

Studying the genetic relationships of the available genotypes is highly relevant to preserving the germplasm of foxtail millet and ensuring sustainable production (Trivedi et al., 2018). Rapid identification of heterozygosity in segregated populations (Ahmad et al., 2016) and true hybrids at an early growth stage (Ahmad, 2014) can be achieved simply with simple-sequence repeats (SSRs). Determining the level of polymorphism and genetic diversity among foxtail millet genotypes is important to preserve the genetic base and conserve the species (Kim et al., 2012). Although the origin and domestication process of foxtail millet (S. italica subsp. italica (L.) P. Beauv.) have been studied by some groups, characterization is still ambiguous and detection of a broader genetic base of the crop is essential. The objectives of this study were to evaluate drought stress tolerance in foxtail millet at an early growth stage and to detect the extent of genetic diversity between the genotypes using SSR marker systems.

Materials and methods

The study was conducted in the central laboratory of the College of Agricultural Engineering Sciences, University of Sulaimani. The genetic materials comprised of 18 millet genotypes (16 foxtail millet with two green foxtail accessions) were selected based on their origin, response to a wide range of drought stress (according to previous studies) and their cultivation in different arid areas of Iraq. They were obtained from different national and international research centres, as described in online Supplementary Table S1.

In vitro experiment (osmotic stress)

Foxtail millet genotypes were analysed for drought stress tolerance at germination and early growth stage. Seeds from the 18 genotypes were surface sterilized with 70% ethanol for 5 min, and then rinsed with distilled water. They were allowed to germinate in 9 cm diameter Petri dishes, using two layers of Whatman no. 1 filter paper. A factorial experiment was conducted in a completely randomized design, for genotypes and osmotic solutions. Different osmotic solutions of 0, 10, 20 and 30% PEG-6000 (Bheemesh et al., 2018; Shivhare and Lata, 2019) were prepared and run for all the genotypes with three replicates. Twenty uniform seeds were allowed to germinate in each Petri dish. An imbibition period of 24 h with distilled water was allowed by placing the Petri dishes in an incubator at 25°C before treating with different solutions. Every 2 d, 2.5 ml of distilled water was supplemented to prevent drying out of the Petri dishes. The Petri dishes were placed in an incubator (M 7040 R Elektro-mag) at $25 \pm 2^{\circ}$ C for 12 d. Germination per cent, shoot length (cm), root length (cm), root/shoot ratio and seed vigour index (SVI) were taken after 12 d of incubation according to the International Seed Testing Association (ISTA, 1993). Seeds with a 0.2 cm length of radicle were considered as germinated. SVI was estimated according to the formula: (root length + shoot length) × germination percentage (Uddin et al., 2021).

Data obtained from the stress experiment were subjected to an analysis of variance (ANOVA), using XLSTAT 2016 software and the comparisons of trait's means for both factors and their interactions were made using Least significant Difference test (LSD) at the 5% level of probability. Principal component analysis (PCA) and principal coordinate analysis (PCoA) were conducted for the genotypes using rank correlation matrix of two-way data from the studied criteria. Cluster analysis was also performed based on squared Euclidean distance to classify the genotypes relatedness.

DNA extraction

Fresh leaf samples of 14 d-old seedlings were used for DNA extraction. Total Genomic DNA was isolated from 3–4 g of bulked leaves for each genotype using the protocol of Dellaporta *et al.* (1983). The quality and concentration of the DNA were measured on 1% (w/v) agarose gel in 1× TBE buffer, using known concentrations of DNA lambda, and the equivalence reaction concentration of 20 ng/µl were produced for the genotypes' DNA by adding sterile distilled water.

Polymerase chain reaction (PCR) amplification

Thirty-seven SSR primer sets, previously described (Lin et al., 2011; Venkata Suresh et al., 2013; Zhang et al., 2014), were chosen from all the nine chromosomes of the millet genome (online Supplementary Table S2). Annealing temperature was tested for each primer at a range of temperatures (from 50 to 65°C), using a pooled DNA template from the genotypes used in the study. Primer amplifications were performed by using a PCR machine (MultiGene OptiMax Thermal Cycler, Labnet Company) in a total reaction volume of 20 µl. PCR mix was provided from CinnaGen Company (Iran) which contained 10× assay buffer, 2.5 mM MgCl₂, 400 µM dNTP (Fermantas) and 1 U of Taq DNA polymerase. Each amplification contained 10 µl of the reaction mix, 2-4 µl of genomic DNA (20-50 ng in total), 2 µl (20 ng/µl) of each forward and reverse primers and the final volume was made up to 20 µl with distilled water (ddH₂O). PCR programme was set up for one initial denaturation cycle at 94°C for 4 min, followed by 40 cycles of 60 s at 94°C and then 60 s at the appropriate annealing temperature (between 50 and 61°C), followed by 2 min of extension at 72°C. The reaction was terminated with a final extension step of 72°C for 10 min. A 10 µl aliquot of the PCR product was mixed with 5 µl of electrophoresis 6× loading buffer. The samples were loaded onto the agarose gel (2% concentration in $1 \times$ TBE buffer). The agarose gel used here was with a low electroendoosmotic, for satisfactory scoring resolve. An amount of 3-5 µl of ethidium bromide (0.5 µg/ml) was added to the gel before pouring into the tray. The gels of PCR products were run on 80 V for 120 min. The fragments were separated and visualized by a UV-light using electrophoresis (ENDUROTM GDS Touch, Labnet, model no. GDST1302). PCRs were repeated for the primers with smeary or unclear bands on the agarose gel.

SSR data analysis

Fragments amplified with the SSR primers were scored manually; alleles were scored as present (1) or absent (0). Fragment bands of less than 100 bp length on the gel, out of expected size (which could be primer dimer) were excluded from the scoring. The polymorphism percentage was calculated for all polymorphic SSR markers according to the method of Blair *et al.* (1999):

Polymorphism% = total number of bandsnumber of monomorphic bands/ total number of bands × 100 The informativeness of the markers was quantified through the estimation of polymorphic information content (PIC) (Botstein *et al.*, 1980) which also depends on the scored binary data as follows:

PIC =
$$1 - \sum_{i=1}^{n} P_i^2 - \left(\sum_{i=1}^{n} P_i^2\right)^2 + \sum_{i=1}^{n} P_i^4$$

where p_i and p_j = frequency of the marker allele *i* and *j*, respectively, in the genotypes tested, and n = number of different alleles at a certain locus.

Marker index (MI) was estimated to indicate the overall utility of the maker system (Chesnokov and Artemyeva, 2015), as follows:

$$MI = PIC \times n_p (n_p/n)$$

where $n_{\rm p}$ is the number of polymorphic loci, and n is the total loci number.

The amplification products at each SSR locus were converted to binary matrix data and they were utilized to estimate the genetic distance between the genotypes in the study based on Jaccard's coefficient, followed by the unweighted pair group method with the arithmetic mean algorithm (UPGMA). PCA and PCoA were analysed and clusters algorithm on the genetic distances were expressed as a dendrogram with the help of XLSTAT 2016 software.

Results

In vitro drought experiment

Different concentrations of PEG-6000 were used to evaluate the stress-tolerance response of foxtail millet genotypes. Genotype differences under water-stress conditions were determined based on seed germination and early growth parameters. The ANOVA shows highly significant differences ($P \le 0.01$) for the traits studied in the foxtail genotypes at different osmotic potentials induced by different concentrations of PEG-6000 (online Supplementary Table S3). ISe 869 and ISe 1851 had the most germination rate

Table 1. Effect of the genotypes and different drought stress conditions on germination and seedling characters for 18 foxtail millet genotypes

Experiment factor	Germination	Root length (R)	Shoot length (S)	R/S	SVI
Genotype					
ISO313	71.67 ^c	3.85 ^c	3.47 ^c	1.07 ^{gh}	604.46 ^d
Mk1075	71.50 ^c	3.59 ^f	3.49 ^c	1.02 ^{hi}	574.84 ^e
Yellow spike	76.42 ^b	3.91 ^c	3.78 ^b	0.99 ⁱ	663.05 ^c
Acc.67	62.58 ⁱ	3.53 ^{fg}	3.23 ^{de}	1.10 ^{defg}	531.86 ^h
SIA326	65.75 ^h	3.15 ⁱ	2.89 ^{ij}	1.07 ^{fgh}	450.92 ⁿ
GS260	65.83 ^h	3.14 ⁱ	2.95 ^{hi}	1.06 ^{gh}	484.48 ¹
ISe 869	77.75 ^a	4.32 ^a	3.91 ^a	1.11 ^{cdefg}	709.88 ^a
ISe 1851	78.11 ^a	4.15 ^b	3.74 ^b	1.11 ^{cdefg}	690.22 ^b
SiA 1244	60.75 ^j	3.50 ^g	2.93 ^{ij}	1.18 ^{ab}	484.18 ^l
SiA 2644	57.25 ^k	3.33 ^h	2.88 ^{ij}	1.18 ^{ab}	461.63 ^m
BD-897	71.58 ^c	3.93 ^c	3.51 ^c	1.13 ^{bcdef}	609.71 ^d
BD-881	70.17 ^d	3.74 ^d	3.27 ^d	1.15 ^{bcd}	564.00 ^f
Bastan	67.67 ^{fg}	3.53 ^{fg}	3.10 ^{fg}	1.18 ^{ab}	522.99 ^{hi}
P-kf11	67.75 ^{fg}	3.49 ^g	3.02 ^{gh}	1.17 ^{bc}	511.52 ^{jk}
P-kf13	67.58 ^g	3.52 ^{fg}	2.86 ^j	1.23 ^a	501.46 ^k
PI223677	68.50 ^{ef}	3.68 ^{de}	3.18 ^{ef}	1.16 ^{bcd}	548.68 ^g
PI230134	69.16 ^e	3.60 ^{ef}	3.16 ^{ef}	1.14 ^{bcde}	545.90 ^g
Local	65.67 ^h	3.56 ^{fg}	3.27 ^d	1.09 ^{efg}	512.78 ^{ij}
LSD	0.852	0.087	0.084	0.059	10.568
PEG-6000 concentration					
0%	91.99 ^a	5.52 ^a	5.07 ^a	1.17 ^a	975.08 ^a
10%	85.63 ^b	4.65 ^b	4.00 ^b	1.14 ^a	743.46 ^b
20%	72.13 ^c	3.15 ^c	2.80 ^c	1.09 ^b	434.54 ^c
30%	24.85 ^d	1.24 ^d	1.17 ^d	1.07 ^b	63.05 ^d
LSD	0.402	0.041	0.040	0.028	4.982

Note: Mean values in columns with different letters are significantly different according to LSD test at the 5% level of probability.

compared to other genotypes for the different osmotic stress levels (Table 1). ISe 869 also surpassed the others in root length, shoot length and SIV. Both green foxtail genotypes (PI223677 and PI230134) had a reasonable performance for the traits studied. The lowest ratios were recorded by SiA 2644 for all the traits studied except R/S ratio. Maximum reduction was recorded at the highest level of water stress for all the traits.

Percentage of germination

The ability to tolerate the chemical dehydration of PEG was investigated for 18 foxtail millet genotypes during germination and early growth stages. The genotypes had varied responses to germination at different osmotic potentials as induced by PEG-6000 (online Supplementary Fig. S1). Reduction in germination per cent was recorded for all the genotypes at different negative osmotic solutions compared to the control. The solution of 10% PEG had less effect on germination reduction. The higher concentration of PEG (30%) caused a maximum and significant reduction in germination up to 90.5% in SiA 2644 (standard deviation of 0.68) compared to the control.

Root length (cm)

Drought stress imposed a significant reduction on root length in all the genotypes at early growth stages. Root length of the genotype SiA 2644 was reduced to the minimum level by the increased PEG concentration in the irrigation solutions (online Supplementary Fig. S2), while both ISe 869 and ISe 1851 were found to be the most resistant genotypes for the PEG concentrations given the least reduction in root length at the highest concentration of PEG (30%), followed by local genotype.

Shoot length (cm)

A similar pattern for shoot length was realized for the genotypes under different PEG concentrations. An osmotic solution of 30% PEG-6000 had the highest effect on the reduction in shoot length for the genotypes. SiA 2644 had the highest reduction in shoot length at 30% PEG-6000, while ISe 869 and ISe 1851 were highly resistant to negative water potentials. The last two genotypes recorded the highest shoot length along the concentrations of 10, 20 and 30% PEG-6000 compared to other genotypes, followed by yellow spike (online Supplementary Fig. S3).

Root/shoot ratio (R/S)

Variable effects of R/S ratio were recorded for the genotypes at different PEG concentrations. Most of the genotypes had an increased ratio at the most negative osmotic solutions (online Supplementary Fig. S4). Less fluctuation in this ratio for different PEG concentration was observed for some genotypes such as ISe 869 and local, showing non-significant differences in R/S ratios at different negative solutions compared to the control.

Seed vigour index

It was observed that SVI was reduced by increasing the PEG concentration. The genotypes SiA 2644 and SiA 1244 had lower values for 30% PEG, while ISe 869 and ISe 1851 showed the minimum decline in SVI value for the same solution (30% PEG-6000), see online Supplementary Fig. S5. Yellow spike was similar in ISe 1851 in terms of its resistance to the reduction in SVI value with the increased concentration of PEG-6000.

PCA for morphological data

The relationships among different parameters were displayed in a graphical biplot of PCA. The first and second components accounted for 95.91% of the total variation (Fig. 1). The genotypes ISe 869 and ISe 1851 were shown to be highly associated, having high values for germination, root and shoot length and SVI, while they both recording lower R/S ratio based on the biplot diagram.

Agglomerative hierarchical clustering using morphological data

All the genotypes were included in pairwise comparisons, the average dissimilarity values were calculated based on data of germination and seedling growth stage. Dissimilarity values for the proximity matrix ranged from 0.36 between Bastan and P-kf11, to 6.75 between ISe 869 and SiA 2644 (data not shown).

Cluster analysis based on the morphological data

A dendrogram was created for the foxtail millet genotypes based on the data of germination and early growth characteristics. Clustering was verified using Euclidean distance of Ward's method for dissimilarity matrix. The genotypes were divided into three main groups (Fig. 2). The first group contains the genotypes SiA 2644, SiA 1244, GS 260 and SiA 326. The second group consists of eight genotypes, both green foxtail millets with the other six foxtail millet genotypes. While the third group includes the genotypes yellow spike, Mk1075, ISO313 from Australia that clustered in a subgroup with the genotype BD-897 from Bangladesh, ISe 869 and ISe 1851 from India were in the second subgroup of the third clade.

SSR analysis

Of the 37 primers used in this study, 33 amplified genomic DNA from the genotypes and revealed the existence of genetic variation between them. A total of 33 polymorphic primers showed the presence of 132 polymorphic alleles (out of 166 loci) at the rate of 4.00 alleles per primer. PIC had the average value of 0.67 for the primers used. The highest value was recorded for primer SICAAS1065 with value 0.83, while the lowest PIC is referred to primer SICAAS6023 with the value of 0.37 (Table 2). The same patterns of the highest MI value and the least value were recorded for primer SICAAS1065 and SICAAS6023, respectively.

PCoA for SSR data

The relation between the foxtail genotypes was determined using PCoA based on the SSR genotypic data (Fig. 3). The biplot clearly shows the distribution of the genotypes based on SSR data. ISe 869 and ISe 1851 were found very close to each other with yellow spike. Both green foxtail millet genotypes (PI230134 and PI223677) were positioned close to each other in a group. The genotypes SiA 2644 and SiA 1244 were placed close to the local genotype based on the SSR data.

Agglomerative hierarchical clustering using SSR markers

Genetic distance was estimated for the 18 foxtail millet genotypes using the amplification products of SSR markers to produce a dissimilarity matrix (data not shown). The results showed the highest similarity was between the two green foxtail millet, PI223677 and



Fig. 1. Scatter plot of PCA for the 18 foxtail millet genotypes. The biplot demonstrates the relationship between the genotypes based on the morphological traits.

PI230134 genotypes, those sourced from Iran, giving the lowest value of genetic distance (5.32). While the lowest percentage of genetic similarity (corresponds to the largest genetic distances 19.17) was obtained between ISe 1851 (an Indian origin) and local genotypes.

Cluster analysis based on SSR marker data

Genetic relationships between the 18 genotypes of foxtail millet from different geographical regions identified five main clusters (each with two to five genotypes) in the UPGMA scheme (Fig. 4). The first main cluster includes the genotypes Bastan, MK1075, Acc.67, BD-881 and GS 260. Two green foxtail genotypes (PI223677 and PI230134) were clustered in the second group. P-KF11 and P-KF13 were clustered in the subgroup of the third cluster, while the other subgroup represents the genotypes BD-897, SiA326 and ISO313. The fourth cluster includes the local foxtail genotypes with the other two Indian genotypes (SiA 2644, SiA 1244). Other two Indian genotypes (ISe 1851, ISe 869) were grouped with the Australian foxtail millet yellow spike in the fifth cluster.

Discussion

In vitro experimental analysis

PEG technique was used successfully for current foxtail millet genotypes to identify potentially more stress-tolerant genotypes.

Effects of genotypes, different water-stress conditions and their interaction, induced by PEG-6000 indicated highly significant differences in the response of these genotypes in vitro. This outcome might be a good indication for rapid identification of tolerant foxtail millet genotypes to drought stress. The measurement of seed germination and early seedling growth characteristics have been shown to be useful evaluation for selecting stress tolerance in plants (Islam et al., 2018; Cai et al., 2020). Different foxtail millet genotypes and simulated water-deficit conditions had distinct effects on the decrease in both germination per cent and seedling growth (Yu et al., 2020). The current genotypes are found to have a wide response to the characteristics measured at early growth stages. Both ISe 869 and ISe 1815 genotypes showed better performance than all the others for germination and early growth traits at the different stress levels of 10, 20 and 30% PEG, while SiA 2644 had poorer response. High PEG concentrations affected the traits with a highly significant reduction and enabled the discrimination putative of tolerant genotypes among the foxtail millet genotypes. The increased negative water potential at 30% PEG-6000 caused the greatest reduction in germination; however, it had not reached the upper limit of the physiological activity to stop germination totally. A higher dosage of 40% of PEG-6000 caused germination of pearl millet reported to be stopped (Sani and Boureima, 2014). However, the reduction in germination is due to the development of osmotically enforced dormancy and enzymatic activity reduction under water-deficit conditions (Muscolo et al., 2014; Pérez-Hernández et al., 2018), the genotypes ISe 869, ISe 1851 and yellow spike were indicated as





Fig. 2. Phylogenetic relationship between the 18 foxtail millet genotypes based on morphological data at germination and early growth stage. The genotypes were divided into three main groups. Clustering could be demonstrated by Euclidean distance of Ward's method for dissimilarity matrix at 8 (vertical line).

resistant to the osmotic stresses, while the most sensitive genotype was SiA 2644 at the treatment levels of 10, 20 and 30% PEG.

Root length is among the major determinant indicators in screening genotypes for dryland conditions. Root length of the current genotypes was also affected by the increased PEG concentration in the irrigation solutions. Like germination, root length was reduced to a minimum value for SiA 2644 at 30% PEG, while the least reduction in root length at the high PEG concentration (30%) was ISe 869 and ISe 1851. Local genotype was also identified as a good resistant type, producing a reasonable root length at a maximum concentration of 30% PEG. An association between root length and drought tolerance at early growth stress in millet was reported previously (Lata, 2015; Passot, 2016). Seedlings with longer roots are more adapted to drought would be more able to cope with water-deficit conditions better than those with shorter roots (Ghatak *et al.*, 2021).

ISe 869 and ISe 1851 genotypes were realized to be highly persistent for shoot length in the negative water potential induced with PEG-6000. They consistently gave the highest length for control and different PEG concentrations compared to the others. The greatest reduction in root length was within the susceptible genotypes such as SiA 2644 and SiA 1244 at 30% PEG. Influence of high PEG concentrations on the reduction of root and shoot lengths in foxtail millet genotypes was also reported by Bheemesh *et al.* (2018); however, shoot length was more inhibited than root length by increasing the concentration of PEG, because root length has the strongest dehydration tolerance (Yu *et al.*, 2020). Drought stress also led to a reduction in the length of both root and shoot for all the genotypes, which is consistent with the observations made by Shivhare and Lata (2019) and Baloch *et al.* (2012). Decrease in seedling growth under drought stress is reported to be the result of the reduced cell division and cell elongation (Hasanuzzaman *et al.*, 2019; Abro *et al.*, 2020).

Non-significant differences in the R/S ratio were observed for both genotypes ISe 869 and local across the different negative solution strengths. The least fluctuation in R/S ratio indicates that physiological activities of the root system in such genotypes are less sensitive to low relative water content of the plant (Sani and Boureima, 2014). This case was not indicated for other genotypes, as their R/S ratios were decreased by increasing the PEG contents. The most effected genotype for the R/S ratio is identified for SiA 1244 which is also the most sensitive to the other studied traits.

SVI was reduced for all the genotypes with different rates for the different PEG solutions. ISe 869 and ISe 1851 were shown to have a lower reduction in SVI than others with the increased PEG concentration of 30% PEG, and they could be regarded as more drought tolerance genotypes (Baloch *et al.*, 2012). The maximum decline in SVI values for the genotypes SiA 2644 and SiA 1244 indicating their sensitivity to drought stress conditions. High seedling vigour at early seedling growth is important and it is associated with rapid and high emergence rate, and quick establishment of crop stand (Abati *et al.*, 2018).

Selection of genotypes based on the seedling characteristics could be marked as a selection criterion for identification of the right genotypes under water-stress conditions. Characterization of genotypes based on different seedling traits under the induced

Table 2. Motif repeat, number of alleles	, PICs and MI of the SSF	primers used in the genetic	divergence analy	sis of the 18 foxtail millet ge	enotypes
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				Allele number		nber			
No.	Locus	Repeat motif	Fragment size range (bp)	No. of Amplified band	Total allele	Polymorphic allele	Polymorphism (%)	PIC	МІ
1	SICAAS7094	(ATACAT)7	88-470	58	6	5	83	0.70	0.58
2	SICAAS-KC8034	(AAT)16	80-250	78	6	4	67	0.76	0.51
3	SICAAS9027	(TC)23	80-210	53	5	5	100	0.67	0.67
4	SieSSR81	(CCT)5	180-1500	51	5	4	80	0.78	0.62
5	SieSSR237	(GCT)5	80-1200	71	7	6	86	0.58	0.50
6	SieSSR274	(CGC)5	80-310	68	5	3	60	0.59	0.35
7	SICAAS1008	(GA)23	145-200	43	4	3	75	0.68	0.51
8	SICAAS4004	(GA)19	80-320	35	4	4	100	0.67	0.67
9	SICAAS3090	(ATATAC)13	100-500	68	5	4	80	0.74	0.59
10	SICAAS2004	(CT)19	88-400	65	6	5	83	0.76	0.63
11	SICAAS5051	(AT)12	80-300	57	5	4	80	0.54	0.43
12	SICAAS6049	(CA)17	170-180	45	4	3	75	0.69	0.52
13	SITM04	(TG)13	160-600	69	6	4	67	0.59	0.40
14	SITM06	(CA)10	80-195	54	4	3	75	0.57	0.43
15	SITM11	(ATC)6	115-200	32	3	3	100	0.58	0.58
16	SITTM22	(ATG)7	80-900	43	5	4	80	0.74	0.59
17	SITM18	(CAT)8	90-200	79	7	5	71	0.70	0.50
18	SICAAS1006	(AT)24	120-500	40	4	4	100	0.59	0.59
19	SICAAS1065	(CTA)8	160-550	23	3	3	100	0.83	0.83
20	SICAAS2072	(ACT)10	200-850	78	7	6	86	0.59	0.51
21	SICAAS2084	(TAT)12	150-400	48	4	3	75	0.70	0.53
22	SICAAS4022	(AG)9	120-600	34	4	4	100	0.70	0.70
23	SICAAS5089	(ATA)19	180-350	57	5	4	80	0.77	0.62
24	SICAAS5072	(CTT)19	200-600	63	6	5	83	0.70	0.58
25	SICAAS6054	(AC)12	160-480	38	4	4	100	0.70	0.70
26	SICAAS6023	(TC)12	130-250	57	5	4	80	0.37	0.30
27	SICAAS6024	(AT)9	130-550	55	4	2	50	0.76	0.38
28	SICAAS7040	(CT)10	150-650	86	7	5	71	0.76	0.54
29	SICAAS9004	(GA)8	120-480	68	6	5	83	0.70	0.58
30	SICAAS9075	(AG)24	170-460	63	5	4	80	0.70	0.56
31	SICAAS9030	(GA)24	150-400	57	6	4	67	0.59	0.40
32	SieSSR346	(GCG)5	200-660	45	4	3	75	0.58	0.44
33	SieSSR275	(AT)6	160-640	73	5	3	60	0.74	0.44
Average	9				5.03	4.00	80.36	0.67	0.54

drought stress caused by PEG-6000 was also reported previously in foxtail millet (Niu *et al.*, 2018), finger millet (Mundada *et al.*, 2020) and other cereals such as wheat (Ahmad *et al.*, 2017; Abro *et al.*, 2020), rice (Purbajanti *et al.*, 2019), maize (Magar *et al.*, 2019) and tomato (Ali *et al.*, 2018).

The first two principal components (PC), PC1 and PC2 covered nearly all phenotypic variations (95.91%), across the eighteen genotypes, for germination and early growth characteristics. This result might indicate a high contribution of these two components in determining the relationship between the foxtail genotypes (Fig. 1). The cosine of the angle between the studied traits except R/S ratio indicating their positive association, also indicate their negative relation to the R/S ratio. Such correlations will also be helpful in predicting the traits through others associated traits that are more easily measured (Harrison and LaForgia, 2019). The genotypes have a different degree of relationship based on the





Fig. 3. Scatter plot of PCoA for the 18 millet genotypes. The biplot demonstrates the relationship between the genotypes based on the SSR data.

morphological traits. ISe 869 and ISe 1851 were realized to be highly associated, having high values for germination, root and shoot lengths and SVI, while they both recorded lowered R/S ratio based on the biplot diagram. These two genotypes also performed very well and highly associated under field and greenhouse conditions for an investigation of phenotypic responses and phosphate content in India (Ceasar *et al.*, 2020). The designated angles here reflect the importance of the largest contributor to the total variation and association between the genotypes at each axis of differentiation (Abdi and Williams, 2010).

Reasonable variability was indicated based on the wide range of dissimilarity values (0.36-6.75) between the genotypes using the morphological matrix data, applying Euclidean distance dissimilarity of Ward's method of agglomerative hierarchical clustering analysis. Determination of the genetic distance will be highly helpful in conducting hybridization programme to involve highly distinct genotypes, while for the genotypes with low genetic distance selection programmes could be followed to emphasize additive alleles for desirable traits (Govindaraj et al., 2015). To further understanding the relationship between the studied genotypes, UPGMA clustering analysis was able to classify the foxtail millet genotypes into three groups based on the morphological data. According to the studied characteristics most of the genotypes located in the first group are identified to be the most sensitive to the induced drought stress especially SiA 2644 and SiA 1244. Grouping the green foxtail millet genotypes with other foxtail millets in the second group indicates their origin similarity or ancestral relationship, and both could be regarded as the same species (Diao and Jia, 2017). While the third cluster includes the genotypes with the most drought-resistant based on the traits studied. ISe 869 and ISe 1851 were superior for germination and other seedling traits under induced drought stress. Yellow spike, Mk1075, ISO313 and BD-897 are also clustered together within a subgroup from the third cluster, showing moderate tolerance to stress conditions. It has been identified that based on screening the genotypes under drought-induced stress that foxtail millets were separated into these groups independent of their origins. Biplot results of the current study are in agreement with cluster analysis of the morphological data in identifying the same tolerant genotypes for the induced water stress. Cluster analysis has been utilized previously to describe the variation and grouping foxtail millet genotypes based on agronomic traits (Brunda et al., 2015; Yadav et al., 2018; Kumar et al., 2019).

Molecular data analysis

SSRs are useful markers for studying genetic diversity, the hypervariable nature of SSRs results in high levels of allelic variation, even among very closely related varieties (Vieira *et al.*, 2016; Ma *et al.*, 2019). In this study, we have demonstrated a successful application of SSR analysis to study genetic distance among the genotypes of foxtail millet from different geographic backgrounds



Dendrogram

Fig. 4. Phylogenetic relationship between the 18 foxtail millet genotypes based on 132 allele data. The genotypes were divided into five main groups. Clustering could be demonstrated at a Jaccard's coefficient, following UPGMA for dissimilarity matrix at 215 (vertical line).

and to be in association with drought stress tolerance. Obtaining relatively high polymorphism of SSR primers average (four alleles/locus, Table 2) indicates the utility of these markers in determining unique genetic profiles of the individual foxtail millet genotypes. The majority of primers used in this study were previously applied on foxtail millet genotypes and tested for polymorphism by various researchers (Lin et al., 2011; Zhang et al., 2014); however, seven primers were taken from genic SSR of expressed sequence tags (NCBIdbEST) (Venkata Suresh et al., 2013) of the foxtail millet marker database (FmMDb; http:// www.nipgr.res.in/foxtail.html), and used successfully in this study for the first time. Five out of the seven genic SSR primers were polymorphic giving three to six alleles per locus. Fewer alleles per locus (2.52) were observed in the foxtail millet crop in a previous study (Trivedi et al., 2018), and a higher mean number of alleles per locus (4.62 alleles) were produced in pearl millet from a total of 74 SSR markers (Kumar et al., 2020). In green foxtail millet 6.1 alleles per locus were detected (Hsieh et al., 2021), while, in foxtail millet a higher number of alleles per primer (10.6 and 17.87 alleles/primer) were obtained by Kim et al. (2012) and Jia et al. (2015), respectively. Similar ranges of alleles per primer for SSR markers were obtained in other diverse studies of foxtail millet (Chander et al., 2017), pearl millet (Adeoti et al., 2017), finger millet (Lee et al., 2017) and barley (Tahir et al., 2021). Polymorphism of the current primers used here was reasonably high (80.36%), demonstrating the effectiveness of such SSR

markers in the assessment of genetic diversity in foxtail millet (Chander *et al.*, 2017).

The discrimination power of the SSR markers was determined based on PICs. All the primers used seem to be reasonably informative because they had PIC and MI values greater than 0.5, indicating highly informative SSR marker for diversity assessment (Kumar *et al.*, 2020). A high MI of the marker indicates the better ability of their combination to detect the differences between larger populations of foxtail millet (Gilbert *et al.*, 1999). In this investigation the number of alleles was found to have a positive correlation ($r^2 = 0.86$, P = 0.000) with PIC values, implying that the alleles number amplified could be used indirectly to assess PIC of the SSR primers in the foxtail millet genotypes. Association of allele number and PIC value was a result of microsatellite marker genotyping (Lebedev *et al.*, 2020).

The extent of genetic variation in among the *Setaria* genotypes selected from diverse geographical locations around the world could be one of the reasons of high polymorphism for these primers. Highly polymorphic and effective SSRs with high alleles ranged from 2 to 16 were obtained using genome-wide microsatellite variant analysis of *S. italica* L. (Yang *et al.*, 2010; Zhang *et al.*, 2014) and number of targeted loci sited by SSR markers (Hayden and Sharp, 2001). Detection rate of the genotypes by SSRs depends on the length of primers. Those with more than 20 bp could able to detect the variation between genotypes more efficiently compared to shorter primers (Wang *et al.*, 201

2015), which is the case for most of the primers used here. In addition, these primers could have better detection for the heterogeneous alleles, especially in the related varieties (Ho *et al.*, 2011).

The biplot diagram for structuring genetic distance indicates a wide distribution of the genotypes on the plate using SSR alleles' data. For ISe 869 and ISe 1851 originated from India are in close relation to yellow spike. These genotypes were identified to have similar potential power to tolerate the drought stress induced by PEG-6000. Response of ISe 1851 to abiotic stress such as salinity was identified previously (Upadhyaya et al., 2015). The two green foxtail millet genotypes (PI230134 and PI223677) positioned close to each other for indicating their similarity. They both are weedy relative to foxtail millet and integrating them in the study would suitably attribute to the genetic analysis of foxtail millet characteristics (Li and Brutnell, 2011). The genotypes SiA 2644 and SiA 1244 were genetically close to local genotype based on the SSR alleles, which is useful for transferring superior genes via hybridization programme to the local, or vice versa. High relatedness of both lines P-kf11 and P-kf13 are also indicated in this diagram. Other genotypes are distributed with variable distance on the plot. Similar patterns of clustering groups for the genotypes were obtained for the morphological data. The relationship between SiA 2644 and SiA 1244, and between both lines P-kf11 and P-kf13 based on the induced stress experiment and the DNA marker data is likely to confirm the value of relying on morphological and molecular data. It is expected that the SSR markers applied on the foxtail genotypes might position close to alleles responsible for stress tolerance in the foxtail millet genome.

Foxtail genotypes sourced from India were distributed in four groups. This clustering format might be supported with prehistorical evidence (Stevens and Fuller, 2017; Martin et al., 2021), as foxtail millet is originated somewhere in Asia, then they were further diversified and spread around the world. Ungrouping the genotypes from the same region in the current study indicating that foxtail millet possesses a wide genetic base (Diao et al., 2014; Sapkota et al., 2016) and diverse germplasm collection from diverging areas of Eurasia with different genetic background might be introduced and cultivated in suitable environments of the same region (Motuzaite-Matuzeviciute et al., 2008). This could support the idea that such clustering did not appear to be a result of recent hybridization events (Hirano et al., 2011). However, some evidence suggesting that the genetic clustering across the foxtail species coincides with the geographical areas more than taxonomy (Lin, 2012), combination of foxtail genotypes from Australia, Russia, India and Iran in the first group indicates their genetic relation independent of their geographical distribution (Ali et al., 2016), as they might have the ancestor relation (Wei et al., 2012). The results obtained here, to cluster foxtail genotypes from diverse geographical regions in one group, are in accordance with what was investigated by other researchers (Lin et al., 2011; Gupta et al., 2012; Kim et al., 2012), to find out that geographical distribution of the foxtail millet is not able to frame their genetic relationship. Wide growing of foxtail millet in semi-arid areas of the Middle East and large areas of India (Krishna, 2013) belonging to eastern complex (race Maxima) would indicate their genetic similarity between the accessions from different countries there. The current diversion of foxtail genotypes will help the breeder in tagging the right foxtail genotypes for development programmes, especially in crossing programmes to guarantee the production of unique offspring from diverse genotypes.

The accessions of green foxtail millet (PI230134 and PI223677) were shown to locate within the same group. Green foxtail millet identified to be the ancestor of foxtail millet (Hsieh et al., 2021). Indeed, repeated genetic introgression has likely resulted from the interspecific hybridization of green foxtail and foxtail millet, due to cross-compatibility of S. italica and S. viridis (Li and Brutnell, 2011), and established close genetic architecture for both species, as foxtail millet maintained 45% of its wild diversity (Wang et al., 2010). This fact will facilitate the introgression of desirable genes from the wild ancestor (green foxtail) into the foxtail cultivars easily, and reduce the risk of genetic drift of elite varieties due to continued inbreeding of foxtail genotypes (Bhandari et al., 2017; Allaby et al., 2019). Krishna (2013) has stated that foxtail millet is grown widely in semi-arid areas in Turkey, Syria, northern Iran, Iraq and large areas of India. The highest values of genetic similarity were found between some accessions of these areas; as they might belong to eastern Asia and European complexes (races Maxima and Moharia). While the Australian genotypes (yellow spike) and two Indian genotypes ISe 869 and ISe 1851 show high deviation from other genotypes, which allows them to be structured in a different cluster. This combination between yellow spike from Australia and two Indian genotypes (ISe 869 and ISe 1851) could be referring to their origin from another complex (race Indica) in spite of their geographical distance (Donnelly et al., 2014; Upadhyaya et al., 2015).

Depending on the above analysis the genotypes used here were found to be divergent, and they could be utilized successfully in a hybridization programme for the genetic improvement of drought resistance in foxtail millet. High genetic diversity in foxtail millet was also found by other researchers (Yadav *et al.*, 2018; Zhang *et al.*, 2019). The direct relationship of this crop to humans from ancient cultures as a food and feed had a big influence on its distribution. Foxtail millet was distributed around the world through domestication and new introduction, to become a worldwide crop (Diao and Jia, 2017). Due to its adaptability, this crop should be involved in cereal developing programme for yield, offering better promising to cope with the abiotic stresses.

Conclusion

The foxtail millet genotypes were screened for their resistance under induced drought stress using PEG-6000 at germination and seedling stage. Variable distances were identified between the genotypes based on the morphological traits. ISe 869 and ISe 1851, followed by yellow spike are highly associated with tolerance to drought stress. The analysis of phenotypic data was likely in accordance with the results from the SSR allele data to cluster the genotypes in different groups. Green foxtail millet genotypes clustered together for both morphological and SSR data set. ISe 1851, ISe 809 and yellow spike were clustered together in a group for both morphological and SSR data analysis. Close position of both DNA markers and alleles of drought tolerance in this crop could answer similar clustering of both data set analyses.

Extra insight into the genetic resources can be achieved through this investigation, to facilitate their use in future genetic improvement programmes. Indeed, based on the *in vitro* experiment, the three most tolerant genotypes ISe 869, ISe 1851 and yellow spike are recommended to be cultivated under drought conditions in Iraq and similar localities subject to drought stress around the world. Further investigations including of more millet germplasm, greater number of SSRs and other DNA markers parallel to field trials would be useful to clarify their genetic relationship, origins and their genetic background for the rapid development of this crop under different rainfed conditions in the world.

Supplementary material. The supplementary material for this article can be found at https://doi.org/10.1017/S1479262123000151

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References

- Abati J, Brzezinski CR, Zucareli C, Foloni JSS and Henning FA (2018) Growth and yield of wheat in response to seed vigor and sowing densities. *Revista Caatinga* **31**, 891–899.
- Abdi H and Williams LJ (2010) Principal component analysis. Wiley Interdisciplinary Reviews: Computational Statistics 2, 433–459.
- Abro A, Memon S, Abro S, Sam E, Ry H, Rind M, Solangi Z, Muhammad T, Ali B and Ahmed W (2020) Evaluation of drought tolerance in wheat (*Triticum aestivum L.*) cultivars at early seedling stage using polyethylene glycol induced osmotic stress. *The Journal of Animal & Plant Sciences* **30**, 950–957.
- Adeoti K, Djedatin G, Ewedje E, Beulé T, Santoni S, Rival A and Jaligot E (2017) Assessment of genetic diversity among cultivated pearl millet (*Pennisetum glaucum*, Poaceae) accessions from Benin, West Africa. *African Journal of Biotechnology* **16**, 782–790.
- Ahmad NS (2014) DNA profiling of F1 hybrid in Bambara groundnut (Vigna subterranea L. Verdc.) using simple sequence repeat marker. Journal of Zankoy Sulaimani-Part A 16, 85–94.
- Ahmad NS, Redjeki ES, Ho WK, Aliyu S, Mayes K, Massawe F, Kilian A and Mayes S (2016) Construction of a genetic linkage map and QTL analysis in Bambara groundnut. *Genome* 59, 459–472.
- Ahmad NS, Kareem SH, Mustafa KM and Ahmad DA (2017) Early screening of some Kurdistan wheat (*Triticum aestivum* L.) cultivars under drought stress. *Journal of Agricultural Science* 9, 88–103.
- Ali A, Choi YM, Do YH, Lee S, Oh S, Park HJ, Cho YH and Lee MC (2016) EST-SSR based genetic diversity and population structure among Korean landraces of foxtail millet (*Setaria italica* L.). *Korean Journal of Plant Resources* 29, 322–330.
- Ali N, Schwarzenberg A, Yvin JC and Hosseini SA (2018) Regulatory role of silicon in mediating differential stress tolerance responses in two contrasting tomato genotypes under osmotic stress. *Frontiers in Plant Science* 9, 1475.
- Allaby RG, Ware RL and Kistler L (2019) A re-evaluation of the domestication bottleneck from archaeogenomic evidence. *Evolutionary Applications* 12, 29–37.
- Baloch MJ, Dunwell J, Khakwani AA, Dennett M, Jatoi WA and Channa SA (2012) Assessment of wheat cultivars for drought tolerance via osmotic stress imposed at early seedling growth stages. *Journal of Agricultural Research* 50, 299–310.
- Bhandari H, Nishant Bhanu A, Srivastava K, Singh M and Shreya HA (2017) Assessment of genetic diversity in crop plants an overview. Advances in Plants & Agriculture Research 7, 279–286.
- Bheemesh SJ, Rao MS, Sekhar MR and Latha P (2018) Effect of PEG-induced drought stress on germination early seedling growth of foxtail millet varieties. *International Journal of Current Microbiology and Applied Sciences* **6**, 2674–2681.
- Blair M, Panaud O and McCouch S (1999) Inter-simple sequence repeat (ISSR) amplification for analysis of microsatellite motif frequency and fingerprinting in rice (*Oryza sativa* L.). *Theoretical and Applied Genetics* 98, 780–792.

- Botstein D, White RL, Skolnick M and Davis RW (1980) Construction of a genetic linkage map in man using restriction fragment length polymorphisms. *American Journal of Human Genetics* **32**, 314.
- Brunda S, Kamatar M, Naveenkumar K and Hundekar R (2015) Genetic diversity in the foxtail millet (*Setaria italica*) germplasm as determined by phenotypical traits. *The Ecoscan* 8, 483–488.
- Cai K, Chen X, Han Z, Wu X, Zhang S, Li Q, Nazir MM, Zhang G and Zeng F (2020) Screening of worldwide barley collection for drought tolerance: the assessment of various physiological measures as the selection criteria. *Frontiers in Plant Science* 11, 1159.
- Ceasar SA, Ramakrishnan M, Vinod K, Roch GV, Upadhyaya HD, Baker A and Ignacimuthu S (2020) Phenotypic responses of foxtail millet (*Setaria italica*) genotypes to phosphate supply under greenhouse and natural field conditions. *PLoS ONE* **15**, e0233896.
- Ch H, Patro T, Anuradha N, Triveni U and Jogarao P (2020) Estimation of nutritive composition of seven small millets. *Journal of Pharmacognosy and Phytochemistry* 9, 1871–1874.
- Chander S, Bhat K, Kumari R, Sen S, Gaikwad A, Gowda M and Dikshit N (2017) Analysis of spatial distribution of genetic diversity and validation of Indian foxtail millet core collection. *Physiology and Molecular Biology of Plants* 23, 663–673.
- Chesnokov YV and Artemyeva A (2015) Evaluation of the measure of polymorphism information of genetic diversity. Agricultural Biology 50, 571–578.
- **Dellaporta SL, Wood J and Hicks BJ** (1983) A plant DNA minipreparation version II. *Plant Molecular Biology Reporter* **1**, 19–21.
- Dhumketi K, Singh A and Rajput L (2017) Suitability of foxtail millet semolina and soy grits for the formulation of instant upma mix. *International Journal of Chemical Studies* 5, 75–79.
- Diao X and Jia G (2017) Origin and domestication of foxtail millet. In Doust A and Diao X (eds), *Genetics and Genomics of Setaria*, Vol. 19. Switzerland: Springer International Publishing, pp. 61–72.
- Diao X, Schnable J, Bennetzen JL and Li J (2014) Initiation of Setaria as a model plant. Frontiers of Agricultural Science and Engineering 1, 16–20.
- Donnelly JL, Adams DC and Dekker J (2014) Weedy adaptation in *Setaria* spp. VI. S. *faberi* seed hull shape as soil germination signal antenna, Iowa State University, Department of Ecology, Evolution, and Organismal Biology (EEOB), p. 21.
- Fahad S, Bajwa AA, Nazir U, Anjum SA, Farooq A, Zohaib A, Sadia S, Nasim W, Adkins S and Saud S (2017) Crop production under drought and heat stress: plant responses and management options. *Frontiers in Plant Science* 8, 1147.
- Ghatak A, Chaturvedi P, Bachmann G, Valledor L, Ramšak Ž, Bazargani MM, Bajaj P, Jegadeesan S, Li W and Sun X (2021) Physiological and proteomic signatures reveal mechanisms of superior drought resilience in pearl millet compared to wheat. *Frontiers in Plant Science* 11, 1–24.
- Ghimire K, Joshi B, Gurung R and Sthapit B (2018) Nepalese foxtail millet [Setaria italica (L.) P. Beauv.] genetic diversity revealed by morphological markers. Genetic Resources and Crop Evolution 65, 1147–1157.
- Gilbert J, Lewis R, Wilkinson M and Caligari P (1999) Developing an appropriate strategy to assess genetic variability in plant germplasm collections. *Theoretical and Applied Genetics* 98, 1125–1131.
- Govindaraj M, Vetriventhan M and Srinivasan M (2015) Importance of genetic diversity assessment in crop plants and its recent advances: an overview of its analytical perspectives. *Genetics Research International* **2015**, 1–14.
- Gull A, Lone AA and Wani NUI (2019) Biotic and abiotic stresses in plants. Division of Genetics and Plant Breeding, SKUAST-Kashmir, India, 1–19.
- Guppy L, Anderson K, Mehta P and Nagabhatla N (2017) Global water crisis: The facts. Hamilton. United Nations University Institute for Water, Environment and Health, Canada.
- Gupta S, Kumari K, Sahu PP, Vidapu S and Prasad M (2012) Sequence-based novel genomic microsatellite markers for robust genotyping purposes in foxtail millet [Setaria italica (L.) P. Beauv.]. Plant Cell Reports 31, 323–337.
- Harrison S and LaForgia M (2019) Seedling traits predict drought-induced mortality linked to diversity loss. Proceedings of the National Academy of Sciences 116, 5576–5581.
- Hasanuzzaman M, Shabala L, Brodribb TJ, Zhou M and Shabala S (2019) Understanding physiological and morphological traits contributing to

drought tolerance in barley. *Journal of Agronomy and Crop Science* **205**, 129–140.

- Hayden MJ and Sharp P (2001) Targeted development of informative microsatellite (SSR) markers. *Nucleic Acids Research* **29**, e44–e44.
- Hirano R, Naito K, Fukunaga K, Watanabe KN, Ohsawa R and Kawase M (2011) Genetic structure of landraces in foxtail millet (*Setaria italica* (L.) P. Beauv.) revealed with transposon display and interpretation to crop evolution of foxtail millet. *Genome* 54, 498–506.
- Ho CW, Wu TH, Hsu TW, Huang JC, Huang CC and Chiang TY (2011) Development of 12 genic microsatellite loci for a biofuel grass, *Miscanthus sinensis* (Poaceae). American Journal of Botany 98, e201–e203.
- Hsieh WH, Chen YC, Liao HC, Lin YR and Chen CH (2021) High differentiation among populations of green foxtail, *Setaria viridis*, in Taiwan and adjacent islands revealed by microsatellite markers. *Diversity* 13, 159.
- Hu H, Mauro-Herrera M and Doust AN (2018) Domestication and improvement in the model C4 grass, Setaria. Frontiers in Plant Science 9, 719.
- **Ierna A** (2021) Towards sustainable intensification of crop production Yield gaps and water use efficiency in farming systems. *Agronomy* **11**, 710.
- Islam M, Kayesh E, Zaman E, Urmi T and Haque M (2018) Evaluation of rice (*Oryza sativa* L.) genotypes for drought tolerance at germination and early seedling stage. *The Agriculturists* 16, 44–54.
- **ISTA** (1993) International rules for seed testing. Rules 1985. Seed Science and Technology **21**, 1–288.
- Ji J, Liu Y, Ge Z, Zhang Y and Wang X (2019) Oleochemical properties for different fractions of foxtail millet bran. *Journal of Oleo Science* 68, ess19063.
- Jia G, Liu X, Schnable JC, Niu Z, Wang C, Li Y, Wang S, Wang S, Liu J and Guo E (2015) Microsatellite variations of elite *Setaria* varieties released during last six decades in China. *PLoS ONE* 10, e0125688.
- Kapoor D, Bhardwaj S, Landi M, Sharma A, Ramakrishnan M and Sharma A (2020) The impact of drought in plant metabolism: how to exploit tolerance mechanisms to increase crop production. *Applied Sciences* 10, 5692.
- Kim EJ, Sa KJ, Park KC and Lee JK (2012) Study of genetic diversity and relationships among accessions of foxtail millet [Setaria italica (L.) P. Beauv.] in Korea, China, and Pakistan using SSR markers. Genes & Genomics 34, 529–538.
- Kintl A, Huňady I, Vymyslický T, Ondrisková V, Hammerschmiedt T, Brtnický M and Elbl J (2021) Effect of seed coating and PEG-induced drought on the germination capacity of five clover crops. *Plants* 10, 724.
- Krishna KR (2013) Agroecosystems: Soils, Climate, Crops, Nutrient Dynamics and Productivity. Toronto: CRC Press.
- Kumar GP, Prasad AD and Reddy CCM (2019) Hierarchical cluster analysis for yield and nutritional traits in elite foxtail millet genetic resources. *International Journal of Innovative Science and Research Technology* 4, 1253–1257.
- Kumar S, Hash CT, Singh G, Basava RK and Srivastava RK (2020) Identification of polymorphic SSR markers in elite genotypes of pearl millet and diversity analysis. *Ecological Genetics and Genomics* 14, 100051.
- Lata C (2015) Advances in omics for enhancing abiotic stress tolerance in millets. Proceedings of the Indian National Science Academy 81, 397–417.
- Lebedev VG, Subbotina NM, Maluchenko OP, Lebedeva TN, Krutovsky KV and Shestibratov KA (2020) Transferability and polymorphism of SSR markers located in flavonoid pathway genes in Fragaria and Rubus species. *Genes* 11, 11.
- Lee KJ, Yoon MS, Shin MJ, Lee JR, Cho YH, Lee HS, Ma KH and Lee GA (2017) Development of SSR markers and their use in studying genetic diversity and population of finger millet (*Eleusine coracana* L. Gaertn.). *Plant Breeding and Biotechnology* 5, 183–191.
- Li P and Brutnell TP (2011) Setaria viridis and Setaria italica, model genetic systems for the Panicoid grasses. Journal of Experimental Botany 62, 3031–3037.
- Lin HS (2012) Genetic diversity in the foxtail millet (*Setaria italica*) germplasm as determined by agronomic traits and microsatellite markers. *Australian Journal of Crop Science* 6, 342–349.
- Lin HS, Chiang CY, Chang SB and Kuoh CS (2011) Development of simple sequence repeats (SSR) markers in *Setaria italica* (Poaceae) and crossamplification in related species. *International Journal of Molecular Sciences* 12, 7835–7845.

- Ma SJ, Sa KJ, Hong TK and Lee JK (2019) Genetic diversity and population structure analysis in Perilla crop and their weedy types from northern and southern areas of China based on simple sequence repeat (SSRs). *Genes & Genomics* 41, 267–281.
- Magar MM, Parajuli A, Sah BP, Shrestha J, Sakh BM, Koirala KB and Dhital SP (2019) Effect of PEG induced drought stress on germination and seedling traits of maize (*Zea mays* L.) lines. *Türk Tarım ve Doğa Bilimleri Dergisi* 6, 196–205.
- Martin L, Messager E, Bedianashvili G, Rusishvili N, Lebedeva E, Longford C, Hovsepyan R, Bitadze L, Chkadua M and Vanishvili N (2021) The place of millet in food globalization during Late Prehistory as evidenced by new bioarchaeological data from the Caucasus. *Scientific Reports* 11, 1–11.
- **Motuzaite-Matuzeviciute G, Colledge S and Jones MK** (2008) Millets across Eurasia: chronology and context of early records of the genera *Panicum* and *Setaria* from archaeological sites in the Old World.
- Mundada P, Nikam T, Kumar SA, Umdale S and Ahire M (2020) Morpho-physiological and biochemical responses of finger millet (*Eleusine coracana* (L.) Gaertn.) genotypes to PEG-induced osmotic stress. *Biocatalysis and Agricultural Biotechnology* 23, 101488.
- Muscolo A, Sidari M, Anastasi U, Santonoceto C and Maggio A (2014) Effect of PEG-induced drought stress on seed germination of four lentil genotypes. *Journal of Plant Interactions* **9**, 354–363.
- Muthamilarasan M, Singh NK and Prasad M (2019) Multi-omics approaches for strategic improvement of stress tolerance in underutilized crop species: a climate change perspective. In Kumar D (ed.), Advances in Genetics, Vol. 103. New Delhi: Elsevier, pp. 1–38.
- Niu X, Song L, Xiao Y and Ge W (2018) Drought-tolerant plant growthpromoting rhizobacteria associated with foxtail millet in a semi-arid agroecosystem and their potential in alleviating drought stress. *Frontiers in Microbiology* **8**, 2580.
- Passot S (2016) Exploring pearl millet root system and its outcome for drought tolerance, Université Montpellier.
- Pavli OI, Foti C, Skoufogianni G, Karastergiou G, Panagou A and Khah EM (2020) PEG-induced drought stress during germination effects on soybean germplasm. Agricultural Research & Technology 23, 70-80.
- Pérez-Hernández Y, Navarro-Boulandier M, Rojas-Sánchez L, Fuentes-Alfonso L and Sosa-del Castillo M (2018) Effect of hydric stress on the germination of seeds from Sorghum bicolor (L.) Moench cv. UDG-110. Pastos y Forrajes 41, 243–252.
- Purbajanti E, Kusmiyati F, Fuskhah E, Rosyida R, Adinurani P and Vincēviča-Gaile Z (2019) Selection for drought-resistant rice (Oryza sativa L.) using polyethylene glycol. IOP Conference Series: Earth and Environmental Science 293, 012014.
- Reddy BV, Srinivasulu K, Reddy CCM, Sekhar AC and Shanthi P (2020) Assessment of genetic diversity in foxtail millet using clustering and principal component analysis. *Plant Cell Biotechnology and Molecular Biology* 21, 74–85.
- Rodiansah A and Puspita MI (2020) In vitro polyploidy induction of foxtail millet (Setaria italica (L) Beauv) cv. buru hotong using colchicine treatment. IOP Conference Series: Earth and Environmental Science 484, 012031.
- Sani DO and Boureima MM (2014) Effect of polyethylene glycol (PEG) 6000 on germination and seedling growth of pearl millet [*Pennisetum glaucum* (L.) R. Br.] and LD50 for *in vitro* screening for drought tolerance. *African Journal of Biotechnology* **13**, 3742–3747.
- Sapkota M, Pandey MP, Thapa DB, Yadav MK, Ghimire S and Timalsina D (2016) Diversity assessment of foxtail millet (*Setaria Italica* L. Beauv) accessions collected from different locations of Nepal. *International Journal of Applied Sciences and Biotechnology* **4**, 483–488.
- Shivakrishna P, Reddy KA and Rao DM (2018) Effect of PEG-6000 imposed drought stress on RNA content, relative water content (RWC), and chlorophyll content in peanut leaves and roots. *Saudi Journal of Biological Sciences* 25, 285–289.
- Shivhare R and Lata C (2019) Assessment of pearl millet genotypes for drought stress tolerance at early and late seedling stages. *Acta Physiologiae Plantarum* **41**, 1–10.

- Stevens CJ and Fuller DQ (2017) The spread of agriculture in Eastern Asia: archaeological bases for hypothetical farmer/language dispersals. *Language Dynamics and Change* 7, 152–186.
- Tahir NAR, Ahmad N, Mustafa K and Kareem D (2021) Diversity maintenance of some barley (*Hordeum* spp.) genetic resources using SSR-based marker. *Journal of Animal and Plant Sciences* **31**, 221–234.
- Trivedi A, Arya L, Verma STR, Hemantaranjan A, Verma M, Sharma V and Saha D (2018) Molecular profiling of foxtail millet (*Setaria italica* (L.) P. Beauv) from Central Himalayan Region for genetic variability and nutritional quality. *The Journal of Agricultural Science* 156, 333–341.
- Uddin S, Ullah S and Nafees M (2021) Effect of seed priming on growth and performance of *Vigna radiata* L. under induced drought stress. *Journal of Agriculture and Food Research* **4**, 100140.
- Upadhyaya HD, Vetriventhan M, Deshpande SP, Sivasubramani S, Wallace JG, Buckler ES, Hash CT and Ramu P (2015) Population genetics and structure of a global foxtail millet germplasm collection. *The Plant Genome* 8, 1–13.
- Venkata Suresh B, Muthamilarasan M, Misra G and Prasad M (2013) FmMDb: a versatile database of foxtail millet markers for millets and bioenergy grasses research. *PLoS ONE* 8, 71418.
- Vieira MLC, Santini L, Diniz AL and Munhoz CDF (2016) Microsatellite markers: what they mean and why they are so useful. *Genetics and Molecular Biology* 39, 312–328.
- Wang C, Chen J, Zhi H, Yang L, Li W, Wang Y, Li H, Zhao B, Chen M and Diao X (2010) Population genetics of foxtail millet and its wild ancestor. *BMC Genetics* 11, 1–13.
- Wang Y, Yang C, Jin Q, Zhou D, Wang S, Yu Y and Yang L (2015) Genome-wide distribution comparative and composition analysis of the SSRs in Poaceae. *BMC Genetics* 16, 18.

- Wei L, Hui Z, Wang YF, Li H and Diaw XM (2012) Assessment of genetic relationship of foxtail millet with its wild ancestor and close relatives by ISSR markers. *Journal of Integrative Agriculture* 11, 556–566.
- Yadav RK, Adhikari AR, Gautam S, Ghimire KH and Dhakal R (2018) Diversity sourcing of foxtail millet through diversity assessment and on-farm evaluation. *Cogent Food & Agriculture* **4**, 1482607.
- Yang X, Yan J, Shah T, Warburton ML, Li Q, Li L, Gao Y, Chai Y, Fu Z and Zhou Y (2010) Genetic analysis and characterization of a new maize association mapping panel for quantitative trait loci dissection. *Theoretical and Applied Genetics* **121**, 417–431.
- Yang X, Lu M, Wang Y, Wang Y, Liu Z and Chen S (2021) Response mechanism of plants to drought stress. *Horticulturae* 7, 50.
- Yu A, Zhao J, Wang Z, Cheng K, Zhang P, Tian G, Liu X, Guo E, Du Y and Wang Y (2020) Transcriptome and metabolite analysis reveal the drought tolerance of foxtail millet significantly correlated with phenylpropanoidsrelated pathways during germination process under PEG stress. *BMC Plant Biology* 20, 1–17.
- Zhang S, Tang C, Zhao Q, Li J, Yang L, Qie L, Fan X, Li L, Zhang N and Zhao M (2014) Development of highly polymorphic simple sequence repeat markers using genome-wide microsatellite variant analysis in foxtail millet [Setaria italica (L.) P. Beauv.]. BMC Genomics 15, 78.
- Zhang L, Liu P, Zhang A, Zheng G, Xing G and Jia J (2019) Analysis of genetic diversity in foxtail millet (*Setaria italica*) using ISSR markers. *Agrociencia (Montecillo)* 53, 133–144.
- Zohary D, Hopf M and Weiss E (2012) Domestication of Plants in the Old World: The Origin and Spread of Domesticated Plants in Southwest Asia, Europe, and the Mediterranean Basin. Oxford: Oxford University Press.