



Conference on ‘Nutrient–nutrient interaction’ Symposium 1: Competition and bioavailability of dietary components

Iron deficiency anaemia: experiences and challenges

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Iron deficiency remains the largest nutritional deficiency worldwide and the main cause of anaemia. Severe iron deficiency leads to anaemia known as iron deficiency anaemia (IDA), which affects a total of 1.24 billion people, the majority of whom are children and women from resource-poor countries. In sub-Saharan Africa, iron deficiency is frequently exacerbated by concomitant parasitic and bacterial infections and contributes to over 120 000 maternal deaths a year, while it irreparably limits the cognitive development of children and leads to poor outcomes in pregnancy.

Currently available iron compounds are cheap and readily available, but constitute a non-physiological approach to providing iron that leads to significant side effects. Consequently, iron deficiency and IDA remain without an effective treatment, particularly in populations with high burden of infectious diseases. So far, despite considerable investment in the past 25 years in nutrition interventions with iron supplementation and fortification, we have been unable to significantly decrease the burden of this disease in resource-poor countries.

If we are to eliminate this condition in the future, it is imperative to look beyond the strategies used until now and we should make an effort to combine community engagement and social science approaches to optimise supplementation and fortification programmes.

Iron deficiency: Iron deficiency anaemia: Capacity building: Iron supplementation

Iron deficiency anaemia: the scale of the problem

Iron deficiency remains the most prevalent nutritional deficiency worldwide, affecting an estimated 4–6 billion people. Iron deficiency is also the main cause of anaemia in children and women in both high or low infection burden settings^(1,2). Two billion people, about 30 % of the world’s population, and 43 % of children between the age 6–59 months, are anaemic and the prevalence of anaemia is five times higher in low- and middle-income countries (LMIC) than in high-income countries^(3,4). As such, iron deficiency anaemia (IDA) is the largest nutritional deficiency disorder in the world and one of the five leading causes of global disease burden⁽³⁾. At any

given moment, more individuals suffer from IDA than any other health problem with a staggering 1.24 billion affected individuals worldwide⁽³⁾ (see Fig. 1).

Iron deficiency is associated with multiple pathologies, including anaemia and defective organ function and formation⁽⁵⁾. Iron deficiency in the absence of anaemia is somewhat subtler in its manifestations than other micronutrient deficiencies, despite being a major contributor to ill health, premature death and lost earnings in developing countries. Even mild iron deficiency appears to impair intellectual development in young children and is lowering national intelligence quotients^(6–10), while overt IDA is associated with an increased risk of serious morbidity, poor motor and mental development in

Abbreviations: IDA, iron deficiency anaemia; LMIC, low- and middle-income countries.

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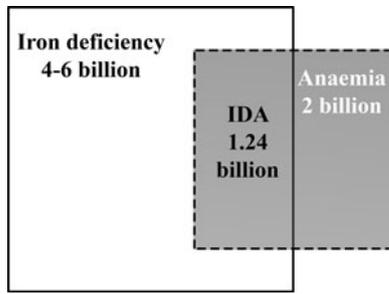


Fig. 1. Prevalence of iron deficiency, iron deficiency anaemia (IDA) and anaemia worldwide. Adapted from⁽³⁾.

children, reduced work capacity in adults, poor pregnancy outcomes and impaired immunity: all of which make it one of the most expensive diseases in the world according to the WHO, causing an estimated 4.05 % of lost gross domestic product globally⁽¹¹⁾. For WHO, reducing the prevalence of IDA is one of its six priorities, and it is estimated that appropriate treatment of IDA not only could restore individual's health, but also raise national productivity levels by as much as 20 %⁽¹¹⁻¹⁴⁾.

Nevertheless, in the past 25 years, not much has changed in LMIC, where IDA maintains its top position as the leading cause of years lived with disability⁽³⁾, and is responsible for over 50 000 deaths annually⁽¹⁵⁾, with iron deficiency contributing to 120 000 maternal deaths per year⁽¹⁶⁾. This is perhaps not surprising, since eliminating IDA in these countries is particularly challenging due to the double burden of IDA and infectious diseases, and malaria, HIV/AIDS, tuberculosis, hookworm and other intestinal parasitic or bacterial infections, all contribute to the anaemia burden.

Iron deficiency in the UK

Recently, Public Health England and the Food Standards Agency has published the combined results from years 7 and 8 (2014/2015–2015/2016) of the National Diet and Nutrition Survey⁽¹⁷⁾, providing an update on results from years 5 and 6 published in 2016. Data on dietary iron intake as well as on biochemical body iron status indicated an increased risk of iron deficiency in girls aged 11–18 years and women aged 19–64 years. When the dietary iron intake was compared with government recommendations, the National Diet and Nutrition Survey reported that 54 % of girls and 27 % of women had iron intakes below the lower reference nutrient intake, which is considered inadequate for most individuals, as it represents the level of intake that is likely to be sufficient to meet the needs of only 2.5 % of the population. Interestingly, the proportion of children aged 1.5–3 years with dietary iron intake below the lower reference nutrient intake was greater than in the previous report (10 % compared with 6 % in the past)^(17,18).

Analysis of blood samples of 704 adults and 329 children in the same UK survey provided evidence of IDA

(as indicated by low Hb levels) and low iron stores (plasma ferritin) in all age/sex groups in the population, with a higher prevalence in females. Based on the WHO criteria for the definition of iron deficiency and anaemia, the prevalence of IDA in the UK was 9 % for girls aged 11–18 years (v. 5 % in the previous report), while low iron stores were evident in 24 % of adolescent girls, 5 % of adult women and 1 % of older women⁽¹⁷⁾. It is important to note that pregnant or breast-feeding women who may have different requirements were among the population groups that were excluded from the survey.

These data support that iron deficiency, and IDA, remain impactful nutritional deficiency disorders in the UK, despite the wide-ranging availability of fortified foods and iron supplements.

Strategies to prevent and treat iron deficiency

The most commonly used strategies to control iron deficiency are supplementation (including multi-micronutrient powders), food fortification, dietary diversification and control of parasitic and other infections. Supplementation or fortification programmes are often the selected short-term approaches because they are cost-effective and relatively easy to implement.

The current WHO recommendation for the prevention of IDA is for daily iron supplementation for 3 consecutive months in a year to all pre-menopausal women, adolescent girls and young children in countries with over 40 % anaemia prevalence (i.e. the majority of countries in sub-Saharan Africa and South East Asia) and intermittent supplementation in settings with lower prevalence^(19,20).

Iron compounds are widely available but typically generate a non-physiological bolus of bio-accessible and reactive ionic iron that can cause significant adverse effects, either in the colon (i.e. unabsorbed iron fraction) or in circulation (i.e. absorbed iron fraction)⁽²¹⁻²⁴⁾. Meta-analysis of trials involving nearly 10 000 young children in developing countries have consistently shown that conventional soluble oral iron supplementation used to treat IDA is associated with increased infection including bloody diarrhoea^(22,25-27) and detrimental changes to the gut microbiome and gut inflammation^(27,28), further increasing the burden from enteric infection and environmental enteropathy (i.e. persistent gut damage and inflammation that leads to malabsorption), which is a major cause of growth failure in children in resource-poor environments^(29,30). Wide-scale home fortification programmes hold promise for reaching at-risk populations with the use of multi-micronutrient powders⁽³¹⁾. Nonetheless, progress has been painstakingly slow, in spite of the dimension of the problem, and strategies to control iron deficiency have failed to decrease the global burden of this deficiency. This is particularly the case for young children in sub-Saharan Africa, where iron supplementation has consistently shown limited efficacy and a potential for increasing infection risk^(21,26,32).

Our biggest challenge: iron bioavailability in high-infection settings

Infection

Many LMIC have high prevalence of malnutrition and infection, both of which contribute to a vicious cycle⁽³³⁾. First, malnutrition weakens barrier and immune functions, allowing pathogens easier access and overall impairing the host's ability to fight pathogens⁽³⁴⁾. Secondly, damage to the intestinal mucosa lining caused by enteric infection and inflammation will decrease nutrient status due to impaired absorption and diarrhoea; therefore predisposing to further infection and worsening nutritional status, and perpetuating a cycle of chronic infection and malnutrition⁽³⁴⁾. This cycle of infection, malnutrition and persistent inflammation will lead to malabsorption and in the long term to growth failure and mortality^(34–36).

The state of chronic infection leads to high levels of hepcidin and anaemia. Elevated hepcidin can decrease iron absorption, irrespective of the iron source, and also leads to low circulating serum iron and transferrin saturation (hypoferraemia), through the mobilisation of circulating iron to the iron stores in macrophages and the liver^(37–40).

Conversely, iron supplementation regimens can cause the release of a high bolus of iron in the blood, which may transiently overwhelm transferrin's iron binding capacity and generate redox-reactive non-transferrin-bound iron, which could be available to extra-cellular pathogenic organisms causing infection^(41–45). Indeed, a committee chaperoned by the WHO concluded that these non-physiological high doses of highly absorbable iron may bypass the naturally evolved systems that safely chaperone iron and be the cause of the excess non-transferrin-bound iron, therefore causing the afore-mentioned harmful effects⁽⁴⁶⁾. This conclusion, supported by a recent study, has shown that 3 h after consumption of a standard iron supplement dose, human serum greatly supports enhanced rates of replication of pathogenic bacteria⁽⁴⁷⁾.

Nutritional immunity

Iron is an essential nutrient to virtually all human pathogens, and the most virulent and invasive strains are those with multiple iron acquisition and utilisation mechanisms^(48–51). Furthermore, iron can also regulate evolutionary transitions between commensal and pathogenic states in microbes⁽⁵²⁾. Nutritional immunity is an innate defence mechanism characterised by the sequestration of iron and other essential trace elements from the circulation to reduce iron availability and limit pathogen growth and virulence^(34,53,54).

There is a constant competition for iron between the human host and invasive pathogens⁽³⁹⁾. Anaemia is protective against malaria and iron supplementation removes this protective effect^(55,56). Iron supplementation may also increase the risk of respiratory infections and other systemic infections, due to iron's ability to increase virulence and multiplication of pathogens^(47,57,58). Our

body has evolved very effective ways to intentionally reduce circulating iron when challenged by infection in an attempt to 'starve' the pathogen. Central to nutritional immunity are high-affinity iron transport proteins, such as transferrin and lactoferrin, which allow us to 'starve' pathogens by maintaining free iron atoms scarce at concentrations of 10^{-18} M^(54,59,60). As aforementioned, a key aspect of our response to infection is the more recently discovered hormone hepcidin, the master regulator of body iron homeostasis, which allows our body to reduce circulating iron by suppressing intestinal iron absorption and decreasing its release from body iron stores. Any iron supplementation regimen that aims to be effective in a high-infection burden setting needs to acknowledge the close interaction between iron and infection and this key aspect of our natural immunity cannot be overlooked.

Hepcidin/inflammation

Our body has no means of excreting excess iron and the control of body iron levels occurs by regulation of iron absorption. Absorption of iron from the gastrointestinal tract is tightly regulated by the systemic need for iron through the action of the hormone hepcidin^(39,61,62). Hepcidin responds to changes in body iron stores, tissue hypoxia and demand for iron, and it alters absorption accordingly. Hepcidin, as the master regulator of iron homeostasis, is the main inhibitor of iron export from cells (including enterocytes, hepatocytes and macrophages) into blood circulation⁽⁵⁾. When hepcidin is elevated, it binds to ferroportin (the only cellular iron exporter known) and it causes its internalisation and degradation, so that cellular iron is locked inside cells and cannot (1) be absorbed into the bloodstream or (2) be released from body iron stores^(63,64).

In IDA, hepcidin levels tend to be low to allow iron absorption, but in the presence of inflammation or in iron-replete individuals, hepcidin levels are usually high^(38,65,66). Anaemia of inflammation, also called anaemia of chronic disease, is characterised by high hepcidin, high iron stores and low circulation iron⁽⁶⁷⁾.

In anaemia of inflammation, when hepcidin levels are high, if non-physiological bolus doses of highly absorbable iron supplements are initiated, this iron will be poorly absorbed, while unabsorbed iron will just be excreted in the stools⁽⁶⁶⁾. Iron absorption can be as high as 50% when hepcidin is switched off and is virtually zero when hepcidin is high⁽⁶⁶⁾.

Microbiome

Low 'free' iron availability in the colon is a major modulator of our gut microbiome particularly in early life or disease. Both iron deficiency and iron supplementation have an impact on the gut microbiome^(68,69). The establishment of an adult-like microbiome requires bio-accessible iron to maintain diversity^(69,70), but in certain population groups, iron supplementation could cause or contribute to microbiome dysbiosis (see Fig. 2).

Excess unabsorbed dietary iron through fortification or supplementation can modify the gut microbiota

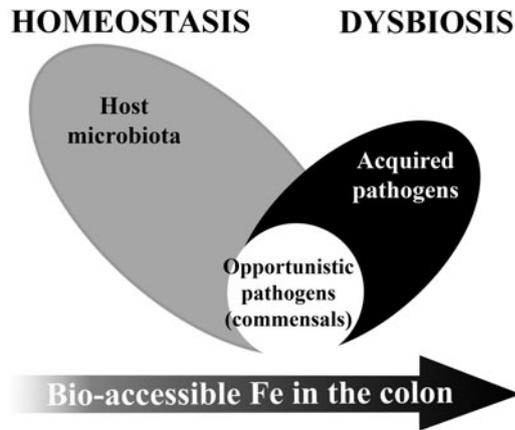


Fig. 2. Model of microbiome dysbiosis caused by an excess of bio-accessible iron (Fe) in the colon.

equilibrium and favour the growth of pathogenic strains over barrier strains. Iron is essential for the virulence and colonisation of most enteropathogenic strains^(71–74). However, gut bacteria species considered most beneficial to the host, i.e. barrier function bacteria such as members of the *Lactobacillaceae* and *Bifidobacteriaceae* families, have either no or a very low requirement for iron^(75,76). Indeed, studies in children have shown the detrimental effects of currently used soluble forms of iron on the gut microbiome, suggesting that iron supplements decrease the abundance of beneficial bacteria groups such as *Lactobacillus* and *Bifidobacterium* and increase the relative abundance of potential enteropathogenic bacteria belonging to the *Enterobacteriaceae* family, and also increase intestinal inflammation^(27,28,76). This gut microbiome dysbiosis combined with mucosal inflammation can lead to reduced resistance to infection, an increase in the risk of diarrhoea and an increase in gut permeability leading to endotoxaemia^(22,27,76–79). In this way, current iron supplementation could be contributing to the vicious cycle of infection, diarrhoea and eventually causing further anaemia. Use of prebiotics together with iron supplements may be able to alleviate part of the gut-related adverse effects of the iron supplements, but further studies are needed⁽⁷⁶⁾.

Future outlook: science and beyond

Micronutrient nutrition is a key aspect of the international development agenda and is integral to the UN sustainable development goals. Recognising the importance of micronutrient nutrition is also paramount in defining strategies to address the double burden of undernutrition and overnutrition and the double burden of malnutrition and infection, both widely prevalent across most countries in the world. The WHO recently recognised that a considerable investment in building public health nutrition capacity is required, particularly in LMIC, alongside a persistent effort in obtaining scientific evidence for the effectiveness of existing and new nutrition programmes and interventions⁽⁸⁰⁾. Additionally,

obtaining scientific evidence through clinical trials and nutrition intervention studies in ‘virgin’ rural settings in LMIC also carries the ethical responsibility to add social value, with community engagement and capacity building, which goes beyond the scientific discovery. Traditionally, research communication has been reserved for scientific publications and news pieces in research websites. However, this strategy creates a science isolation that is not effective in today’s world. Therefore, we must train researchers not only in their ability to conduct nutrition science research but also in their ability to communicate science to the wider community and general public. Equally we must equip public health professionals, nurses and clinicians in LMIC in their ability to engage with the local communities as much as in their knowledge of how to manage the prevention and treatment of various forms of malnutrition. A practical example of how we can combine scientific discovery, capacity building and community engagement is outlined later with the IHAT-GUT iron supplementation clinical trial in The Gambia.

Case study: IHAT-GUT trial in The Gambia

Since iron delivered naturally through foods may be a safer alternative⁽⁵⁾, there is a growing interest in developing novel nano iron compounds or delivery systems to use in fortification and supplementation^(81–84). One of the strategies proposed is to use a mimetic of the iron core of ferritin, namely iron hydroxide adipate tartrate (IHAT), to alleviate the adverse effects associated with reactive ionic iron⁽⁸⁵⁾. The hypothesis is that supplementation with IHAT will cause a slow release of iron into the blood, a lower rise of transferrin saturation, resembling that of natural food iron^(85–87), and may constitute a safer option for supplementation, particularly in population settings with high-infection burden. Pre-clinical data indicate that IHAT does not accumulate in the intestinal mucosa and can promote a beneficial microbiota after dysbiosis^(85,87). IHAT and standard-of-care ferrous sulphate, are currently being tested in a randomised placebo-controlled double-blind clinical trial taking place at the MRC Unit The Gambia at the London School of Hygiene & Tropical Medicine. The IHAT-GUT trial (NCT02941081) is enrolling about 700 young children living in some of the poorest and more rural communities in The Gambia, where risk of infection and diarrhoea are very high.

As add-on projects to the IHAT-GUT trial, we have conducted nutrition training workshops and we are also conducting a social science project, where we document the ‘ins’ and ‘outs’ of setting up a nutrition intervention clinical trial, aimed at translating novel chemistry into public benefit, in a rural setting in The Gambia (see Fig. 3). We believe this social science aspect of research training is as important to global health and nutrition science researchers as the training in nutrition science itself. The social science project is gathering the personal views from clinical and field study staff, all of whom were based in The Gambia before the trial commenced, and will, hopefully, demonstrate that it is

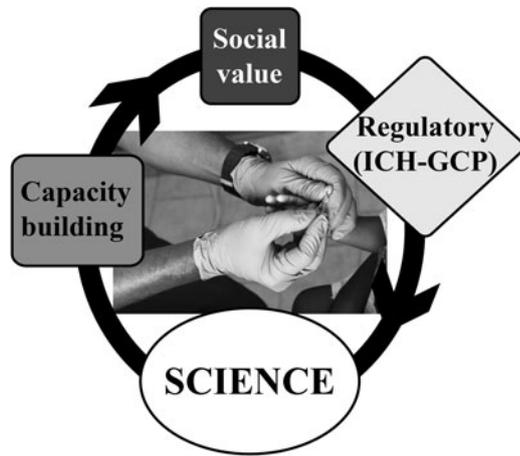


Fig. 3. Systems model integrating social value and capacity building in an iron supplementation trial conducted to ICH-GCP standards in The Gambia.

possible to translate international science to rural Africa and that it is possible to conduct nutrition clinical trials in these rural settings to ICH-GCP international standards. Likewise, it will illustrate the many challenges faced by the researchers, for example, how to implement clinical studies in new settings, how to attract motivated and skilled staff, etc. This project will help staff and communities to embrace these studies and to acknowledge the added value of being part of nutrition research. We envisage that such information will inspire the next generation of researchers in global health and international nutrition, from academia or industry, to translate more international science to rural resource-poor settings.

Conclusion

Anaemia is multifactorial as it is caused by both poor diet and high levels of infection and inflammation. Nowhere more than in sub-Saharan Africa do these causes coexist, and eliminating iron deficiency and anaemia in African children and women still remains one of the main nutritional challenges of the twenty-first century. Iron supplementation in malaria-endemic areas or in regions with high-infection burden needs to be implemented with caution, should target those who are anaemic or at a high risk of deficiency, and be accompanied by strategies to prevent and treat malaria, hookworm, schistosomiasis and other nutrient deficiencies, such as vitamin B₁₂, folate and vitamin A^(32,46).

In spite of many successes, it is clear that more needs to be done to eliminate iron deficiency in LMIC. Today, public health interventions that can correct or prevent iron and other micronutrient deficiencies merit the highest priority for national and international organisations. Routine monitoring for compliance, research into the best delivery vehicles, ensuring the continuous availability of supplements or fortified foods at the community level, engaging local community and community leaders, and capacity building are the key factors for sustaining achievements and increasing the success of such programmes. Only

then will we fully value the World Bank comment: ‘the control of vitamin and mineral deficiencies is one of the most extraordinary development-based scientific advances of recent years. Probably no other technology available today offers as large an opportunity to improve lives and accelerate development at such low cost in such short time.’

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Conflicts of Interest

D. I. A. P. is one of the inventors of the IHAT iron supplementation technology for which she could receive future awards to inventors through the MRC Awards to Inventor scheme. Notwithstanding, the authors declare no conflict of interest.

Authorship

All authors contributed to the writing of this manuscript. The opinions expressed in this article are the authors’ own and do not reflect necessarily the view of their affiliated institutions.

References

1. Wirth JP, Woodruff BA, Engle-Stone R *et al.* (2017) Predictors of anemia in women of reproductive age: Biomarkers Reflecting Inflammation and Nutritional Determinants of Anemia (BRINDA) project. *Am J Clin Nutr* **106**, 416S–427S.
2. Engle-Stone R, Aaron GJ, Huang J *et al.* (2017) Predictors of anemia in preschool children: Biomarkers Reflecting Inflammation and Nutritional Determinants of Anemia (BRINDA) project. *Am J Clin Nutr* **106**, 402S–415S.
3. GBD 2016 Disease and Injury Incidence and Prevalence Collaborators (2017) Global, regional, and national incidence, prevalence, and years lived with disability for 328 diseases and injuries for 195 countries, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet* **390**, 1211–1259.
4. Miller JL (2013) Iron deficiency anemia: a common and curable disease. *Cold Spring Harb Perspect Med* **3**, a011866.



5. Prentice AM, Mendoza YA, Pereira D *et al.* (2017) Dietary strategies for improving iron status: balancing safety and efficacy. *Nutr Rev* **75**, 49–60.
6. Grantham-McGregor S & Ani C (2001) A review of studies on the effect of iron deficiency on cognitive development in children. *J Nutr* **131**, 649S–666S; discussion 666S–668S.
7. Pollitt E, Saco-Pollitt C, Leibel RL *et al.* (1986) Iron deficiency and behavioral development in infants and preschool children. *Am J Clin Nutr* **43**, 555–565.
8. McCann JC & Ames BN (2007) An overview of evidence for a causal relation between iron deficiency during development and deficits in cognitive or behavioral function. *Am J Clin Nutr* **85**, 931–945.
9. Lozoff B, Jimenez E & Smith JB (2006) Double burden of iron deficiency in infancy and low socioeconomic status: a longitudinal analysis of cognitive test scores to age 19 years. *Arch Pediatr Adolesc Med* **160**, 1108–1113.
10. Halterman JS, Kaczorowski JM, Aligne CA *et al.* (2001) Iron deficiency and cognitive achievement among school-aged children and adolescents in the United States. *Pediatrics* **107**, 1381–1386.
11. Horton S & Ross J (2007) The economics of iron deficiency (vol 28, pg 51, 2003). *Food Policy* **32**, 141–143.
12. McGuire S (2015) World Health Organization. Comprehensive implementation plan on maternal, infant, and young child nutrition. Geneva, Switzerland, 2014. *Adv Nutr* **6**, 134–135.
13. Haas JD & Brownlie TT (2001) Iron deficiency and reduced work capacity: a critical review of the research to determine a causal relationship. *J Nutr* **131**, 676S–688S; discussion 688S–690S.
14. WHO (2014) *Global Nutrition Targets 2025: Anaemia Policy Brief (WHO/NMH/NHD/14.4)*. Geneva: World Health Organization.
15. Wang H, Naghavi M, Allen C *et al.* (2017) Global, regional, and national life expectancy, all-cause mortality, and cause-specific mortality for 249 causes of death, 1980–2015: a systematic analysis for the Global Burden of Disease Study 2015. *Lancet* **388**, 1459–1544.
16. WHO (2009) *Global Health Risks. Mortality and Burden of Disease Attributable to Selected Major Risks*. Geneva: World Health Organization.
17. Public Health England and Food Standards Agency (2018) *National Diet and Nutrition Survey. Results from Years 7–8 (combined) of the Rolling Programme (2014/15 to 2015/16)*. London: Public Health England.
18. Geissler C & Singh M (2011) Iron, meat and health. *Nutrients* **3**, 283–316.
19. WHO (2016) *Guideline: Daily Iron Supplementation in Infants and Children*. Geneva, Switzerland: World Health Organization.
20. WHO (2016) *Guideline: Daily Iron Supplementation in Adult Women and Adolescent Girls*. Geneva, Switzerland: World Health Organization.
21. Sazawal S, Black RE, Ramsan M *et al.* (2006) Effects of routine prophylactic supplementation with iron and folic acid on admission to hospital and mortality in preschool children in a high malaria transmission setting: community-based, randomised, placebo-controlled trial. *Lancet* **367**, 133–143.
22. Soofi S, Cousens S, Iqbal SP *et al.* (2013) Effect of provision of daily zinc and iron with several micronutrients on growth and morbidity among young children in Pakistan: a cluster-randomised trial. *Lancet* **382**, 29–40.
23. Prentice AM, Verhoef H & Cerami C (2013) Iron fortification and malaria risk in children. *JAMA* **310**, 914–915.
24. Prentice AM (2008) Iron metabolism, malaria, and other infections: what is all the fuss about? *J Nutr* **138**, 2537–2541.
25. Mayo-Wilson E, Imdad A, Junior J *et al.* (2014) Preventive zinc supplementation for children, and the effect of additional iron: a systematic review and meta-analysis. *BMJ Open* **4**, e004647.
26. Zlotkin S, Newton S, Aimone AM *et al.* (2013) Effect of iron fortification on malaria incidence in infants and young children in Ghana: a randomized trial. *JAMA* **310**, 938–947.
27. Jaeggi T, Kortman GA, Moretti D *et al.* (2015) Iron fortification adversely affects the gut microbiome, increases pathogen abundance and induces intestinal inflammation in Kenyan infants. *Gut* **64**, 731–742.
28. Zimmermann MB, Chassard C, Rohner F *et al.* (2010) The effects of iron fortification on the gut microbiota in African children: a randomized controlled trial in Cote d'Ivoire. *Am J Clin Nutr* **92**, 1406–1415.
29. Naylor C, Lu M, Haque R *et al.* (2015) Environmental enteropathy, oral vaccine failure and growth faltering in infants in Bangladesh. *EBioMedicine* **2**, 1759–1766.
30. Lin A, Arnold BF, Afreen S *et al.* (2013) Household environmental conditions are associated with enteropathy and impaired growth in rural Bangladesh. *Am J Trop Med Hyg* **89**, 130–137.
31. De-Regil LM, Suchdev PS, Vist GE *et al.* (2013) Home fortification of foods with multiple micronutrient powders for health and nutrition in children under two years of age (review). *Evid Based Child Health Cochrane Rev J* **8**, 112–201.
32. Mwangi MN, Phiri KS, Abkari A *et al.* (2017) Iron for Africa-Report of an expert workshop. *Nutrients* **9**, E576.
33. Muller O & Krawinkel M (2005) Malnutrition and health in developing countries. *CMAJ* **173**, 279–286.
34. Calder PC & Jackson AA (2000) Undernutrition, infection and immune function. *Nutr Res Rev* **13**, 3–29.
35. Deen JL, Walraven GE & von Seidlein L (2002) Increased risk for malaria in chronically malnourished children under 5 years of age in rural Gambia. *J Trop Pediatr* **48**, 78–83.
36. Moore SE, Cole TJ, Poskitt EM *et al.* (1997) Season of birth predicts mortality in rural Gambia. *Nature* **388**, 434.
37. Atkinson SH, Armitage AE, Khandwala S *et al.* (2014) Combinatorial effects of malaria season, iron deficiency, and inflammation determine plasma hepcidin concentration in African children. *Blood* **123**, 3221–3229.
38. Jaeggi T, Moretti D, Kvalsvig J *et al.* (2013) Iron status and systemic inflammation, but not gut inflammation, strongly predict gender-specific concentrations of serum hepcidin in Infants in Rural Kenya. *PLoS ONE* **8**, e57513.
39. Drakesmith H & Prentice AM (2012) Heparin and the iron-infection axis. *Science* **338**, 768–772.
40. Nemeth E & Ganz T (2014) Anemia of inflammation. *Hematol Oncol Clin North Am* **28**, 671–681, vi.
41. Brittenham GM, Andersson M, Egli I *et al.* (2014) Circulating non-transferrin-bound iron after oral administration of supplemental and fortification doses of iron to healthy women: a randomized study. *Am J Clin Nutr* **100**, 813–820.
42. Hutchinson C, Al-Ashgar W, Liu DY *et al.* (2004) Oral ferrous sulphate leads to a marked increase in pro-oxidant nontransferrin-bound iron. *Eur J Clin Invest* **34**, 782–784.
43. Schumann K, Solomons NW, Orozco M *et al.* (2013) Differences in circulating non-transferrin-bound iron after oral administration of ferrous sulfate, sodium iron

- EDTA, or iron polymaltose in women with marginal iron stores. *Food Nutr Bull* **34**, 185–193.
44. Brissot P, Ropert M, Le Lan C *et al.* (2012) Non-transferrin bound iron: a key role in iron overload and iron toxicity. *Biochim Biophys Acta* **1820**, 403–410.
 45. van der A DI, Marx JJ, Grobbee DE *et al.* (2006) Non-transferrin-bound iron and risk of coronary heart disease in postmenopausal women. *Circulation* **113**, 1942–1949.
 46. WHO (2007) Conclusions and recommendations of the WHO Consultation on prevention and control of iron deficiency in infants and young children in malaria-endemic areas. *Food Nutr Bull* **28**, S621–S627.
 47. Cross JH, Bradbury RS, Fulford AJ *et al.* (2015) Oral iron acutely elevates bacterial growth in human serum. *Sci Rep* **5**, 16670.
 48. Subashchandrabose S & Mobley HLT (2015) Virulence and fitness determinants of uropathogenic *Escherichia coli*. *Microbiol Spectr* **3**, UTI-0015-2012.
 49. Okeke IN, Scaletsky ICA, Soars EH *et al.* (2004) Molecular epidemiology of the iron utilization genes of enteroaggregative *Escherichia coli*. *J Clin Microbiol* **42**, 36–44.
 50. Connolly JPR, Finlay BB & Roe AJ (2015) From ingestion to colonization: the influence of the host environment on regulation of the LEE encoded type III secretion system in enterohaemorrhagic *Escherichia coli*. *Front Microbiol* **6**, 568.
 51. Parkhill J, Wren BW, Thomson NR *et al.* (2001) Genome sequence of *Yersinia pestis*, the causative agent of plague. *Nature* **413**, 523–527.
 52. Barber MF & Elde NC (2015) Buried treasure: evolutionary perspectives on microbial iron piracy. *Trends Genet* **31**, 627–636.
 53. Barber MF & Elde NC (2014) Escape from bacterial iron piracy through rapid evolution of transferrin. *Science* **346**, 1362.
 54. Bullen JJ, Rogers HJ, Spalding PB *et al.* (2006) Natural resistance, iron and infection: a challenge for clinical medicine. *J Med Microbiol* **55**, 251–258.
 55. Goheen MM, Wegmuller R, Bah A *et al.* (2016) Anemia offers stronger protection than sickle cell trait against the erythrocytic stage of falciparum malaria and this protection is reversed by iron supplementation. *EBioMedicine* **14**, 123–130.
 56. Clark MA, Goheen MM, Fulford A *et al.* (2014) Host iron status and iron supplementation mediate susceptibility to erythrocytic stage *Plasmodium falciparum*. *Nat Commun* **5**, 4446.
 57. Armitage AE, Stacey AR, Giannoulatou E *et al.* (2014) Distinct patterns of hepcidin and iron regulation during HIV-1, HBV, and HCV infections. *Proc Natl Acad Sci USA* **111**, 12187–12192.
 58. McDermid JM, Hennig BJ, van der Sande M *et al.* (2013) Host iron redistribution as a risk factor for incident tuberculosis in HIV infection: an 11-year retrospective cohort study. *BMC Infect Dis* **13**, 48.
 59. Bilitewski U, Blodgett JAV, Duhme-Klair AK *et al.* (2017) Chemical and biological aspects of nutritional immunity-perspectives for new anti-infectives that target iron uptake systems. *Angew Chem Int Ed Engl* **56**, 14360–14382.
 60. Weinberg ED (1975) Nutritional immunity. Host's attempt to withhold iron from microbial invaders. *JAMA* **231**, 39–41.
 61. Ganz T (2003) Hepcidin, a key regulator of iron metabolism and mediator of anemia of inflammation. *Blood* **102**, 783–788.
 62. Nemeth E, Tuttle MS, Powelson J *et al.* (2004) Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science* **306**, 2090–2093.
 63. Ganz T & Nemeth E (2012) Hepcidin and iron homeostasis. *Biochim Biophys Acta* **1823**, 1434–1443.
 64. Deschemin JC & Vaulont S (2013) Role of hepcidin in the setting of hypoferrremia during acute inflammation. *PLoS ONE* **8**, e61050.
 65. Bregman DB, Morris D, Koch TA *et al.* (2013) Hepcidin levels predict nonresponsiveness to oral iron therapy in patients with iron deficiency anemia. *Am J Hematol* **88**, 97–101.
 66. Prentice AM, Doherty CP, Abrams SA *et al.* (2012) Hepcidin is the major predictor of erythrocyte iron incorporation in anemic African children. *Blood* **119**, 1922–1928.
 67. Pasricha SR, Atkinson SH, Armitage AE *et al.* (2014) Expression of the iron hormone hepcidin distinguishes different types of anemia in African children. *Sci Transl Med* **6**, 235re233.
 68. Kortman GA, Dutilh BE, Maathuis AJ *et al.* (2015) Microbial metabolism shifts towards an adverse profile with supplementary iron in the TIM-2 *In vitro* model of the human colon. *Front Microbiol* **6**, 1481.
 69. Pereira DIA, Aslam MF, Frazer DM *et al.* (2015) Dietary iron depletion at weaning imprints low microbiome diversity and this is not recovered with oral nano Fe(III). *Microbiologyopen* **4**, 12–27.
 70. Timmerman HM, Rutten N, Boekhorst J *et al.* (2017) Intestinal colonisation patterns in breastfed and formula-fed infants during the first 12 weeks of life reveal sequential microbiota signatures. *Sci Rep* **7**, 8327.
 71. Kortman GA, Mulder ML, Richters TJ *et al.* (2015) Low dietary iron intake restrains the intestinal inflammatory response and pathology of enteric infection by food-borne bacterial pathogens. *Eur J Immunol* **45**, 2553–2567.
 72. Kortman GA, Roelofs RW, Swinkels DW *et al.* (2014) Iron-induced virulence of *Salmonella enterica* serovar typhimurium at the intestinal epithelial interface can be suppressed by carvacrol. *Antimicrob Agents Chemother* **58**, 1664–1670.
 73. Kortman GA, Boleij A, Swinkels DW *et al.* (2012) Iron availability increases the pathogenic potential of *Salmonella typhimurium* and other enteric pathogens at the intestinal epithelial interface. *PLoS ONE* **7**, e29968.
 74. Jadhav S, Hussain A, Devi S *et al.* (2011) Virulence characteristics and genetic affinities of multiple drug resistant uropathogenic *Escherichia coli* from a semi urban locality in India. *PLoS ONE* **6**, e18063.
 75. Weinberg ED (1997) The lactobacillus anomaly: total iron abstinence. *Perspect Biol Med* **40**, 578–583.
 76. Paganini D, Uyoga MA, Kortman GAM *et al.* (2017) Prebiotic galacto-oligosaccharides mitigate the adverse effects of iron fortification on the gut microbiome: a randomised controlled study in Kenyan infants. *Gut* **66**, 1956–1967.
 77. Mullen A, Gosset L, Larke N *et al.* (2012) The effects of micronutrient-fortified complementary/replacement food on intestinal permeability and systemic markers of inflammation among maternally HIV-exposed and unexposed Zambian infants. *Br J Nutr* **107**, 893–902.
 78. Brunser O, Espinoza J, Araya M *et al.* (1993) Chronic iron intake and diarrhoeal disease in infants. A field study in a less-developed country. *Eur J Clin Nutr* **47**, 317–326.
 79. Cani PD, Bibiloni R, Knauf C *et al.* (2008) Changes in gut microbiota control metabolic endotoxemia-induced



- inflammation in high-fat diet-induced obesity and diabetes in mice. *Diabetes* **57**, 1470–1481.
80. Delisle H, Shrimpton R, Blaney S *et al.* (2017) Capacity-building for a strong public health nutrition workforce in low-resource countries. *Bull World Health Organ* **95**, 385–388.
 81. Powell JJ, Bruggraber SFA, Faria N *et al.* (2014) A nano-disperse ferritin-core mimetic that efficiently corrects anemia without luminal iron redox activity. *Nanomed Nanotechnol Biol Med* **10**, 1529–1538.
 82. von Moos LM, Schneider M, Hilty FM *et al.* (2017) Iron phosphate nanoparticles for food fortification: biological effects in rats and human cell lines. *Nanotoxicology* **11**, 496–506.
 83. Hilty FM, Arnold M, Hilbe M *et al.* (2010) Iron from nanocompounds containing iron and zinc is highly bioavailable in rats without tissue accumulation. *Nat Nanotechnol* **5**, 374–380.
 84. Pisani A, Riccio E, Sabbatini M *et al.* (2015) Effect of oral liposomal iron versus intravenous iron for treatment of iron deficiency anaemia in CKD patients: a randomized trial. *Nephrol Dial Transplant* **30**, 645–652.
 85. Pereira DIA, Bruggraber SFA, Faria N *et al.* (2014) Nanoparticulate iron(III) oxo-hydroxide delivers safe iron that is well absorbed and utilised in humans. *Nanomed Nanotechnol Biol Med* **10**, 1877–1886.
 86. Pereira DI, Mergler BI, Faria N *et al.* (2013) Caco-2 cell acquisition of dietary iron(III) invokes a nanoparticulate endocytic pathway. *PLoS ONE* **8**, e81250.
 87. Aslam MF, Frazer DM, Faria N *et al.* (2014) Ferroportin mediates the intestinal absorption of iron from a nanoparticulate ferritin core mimetic in mice. *FASEB J* **28**, 3671–3678.