

## Ethnic difference in the relationship between acute inflammation and serum ferritin in US adult males

Y. PAN\* AND R. T. JACKSON

*Department of Nutrition and Food Science, University of Maryland, College Park, MD, USA*

*(Accepted 12 February 2007; first published online 22 March 2007)*

### SUMMARY

This study examined the ethnic difference in the association between increased serum ferritin (SF) ( $> 300 \mu\text{g/l}$ ) and acute inflammation (AI) (C-reactive protein  $\geq 1.0 \text{ mg/dl}$ ) between black and white males aged  $\geq 20$  years. Using data from the third National Health and Nutrition Examination Survey (NHANES III), we determined the risk for having elevated SF in black males ( $n = 164$ ) and white males ( $n = 325$ ) with AI present as well as black males ( $n = 1731$ ) and white males ( $n = 2877$ ) with AI absent. Black subjects with AI present were 1.71 times (95% CI 1.18–2.49), and 1.87 times (95% CI 1.46–2.40) more likely to have increased SF than AI absent blacks and AI present whites, respectively. Furthermore, with AI present, every increment of C-reactive protein, white blood cell count, serum albumin, lymphocyte count and platelet count was associated with higher odds of having elevations in SF in blacks than whites. Regardless of AI status, blacks were more likely to have elevations in SF than whites, and the prevalence of elevated SF was significantly higher in blacks than whites. This finding suggested that black males may respond to inflammation with a more aggressive rise in SF compared to white males. Future research is needed to investigate the underlying mechanisms.

### INTRODUCTION

Ferritin is a ubiquitous and highly conserved iron-binding protein. In humans, iron in excess of need is stored primarily as ferritin, principally in the macrophage system of liver, spleen and bone marrow. Increasingly, perturbations in cellular iron and ferritin are emerging as an important element in the pathophysiology of disease. The changes in ferritin are important not only in the classic diseases of iron acquisition, transport, and storage, such as primary haemochromatosis, but also in diseases characterized by inflammation, infection, injury, and repair [1]. Among these are some of the most common diseases that affect human beings: Parkinson's and Alzheimer's disease, cardiac and neuronal ischaemia-reperfusion

injury, atherosclerosis, pulmonary inflammatory states, rheumatoid arthritis, various pre-malignant conditions, and cancers [2–8].

Not only as a member of a group of iron regulatory proteins that maintain cellular and organismal iron homeostasis, ferritin is also a member of the protein family that orchestrates the cellular defence against stress and inflammation. Described as a 'two-faced' protein, ferritin behaves as a positive acute phase reactant, whose serum levels rise rapidly in response to trauma, burn or inflammation [9]. The crucial role that ferritin plays in the cellular and organismal response to inflammation is intimately linked to its primary function in 'iron sequestration', which occurs in animals by sequestering iron in a non-toxic form that minimizes its ability to catalyse the formation of reactive oxygen species (ROS). Therefore, iron status largely affects the generation of oxygen as well as ferryl and nitrogen radicals. In addition, the

\* Author for correspondence: Y. Pan, Ph.D., M.D., 2000 St. James Place, Suite 534, Houston, TX 77056, USA.  
(Email: yapan@na.ko.com)

toxicity of iron at the cellular level is attributable in large part to its capacity to participate in the generation of such reactive species, which can directly damage DNA, lipids, and proteins, leading to profound cellular toxicity. Ferritin, by ‘capturing’ and ‘buffering’ the intracellular labile iron pool [10, 11], thereby plays a key role in maintaining iron homeostasis and reducing the damage from pathogenic invasion.

Health disparities in different ethnic populations, especially between blacks and whites, has become a topic of intense research for the past few decades, yet they are still poorly understood. Of particular interest is the aetiology of ethnic differences in disease susceptibility and response [12]. Inflammation, the underlying pathophysiology of several disorders, is therefore suspected to be one of the factors that may contribute to the ethnic differences in disease progression and outcome. Immune system hyper-responsiveness and inflammation have been associated with a number of disorders that more commonly plague African Americans. Examples include preterm birth [13, 14], atherosclerosis [15–17], autoimmune diseases [18], and transplant rejection [19]. As a key molecule in cellular defence against stress and inflammation, ferritin limits the extent, character, and location of the pro-oxidant stress that typifies inflammatory diseases, cancer and conditions of altered oxygenation [4, 7, 20]. Therefore, an understanding of how ferritin responds to inflammation may lead to insights into the aetiology of the difference in host inflammatory response and clinical outcomes.

In this study, we used data from the third National Health and Nutrition Examination Survey (NHANES III) to examine the ethnic difference in the relationship between acute inflammation (AI) and serum ferritin (SF). Our study thus had three purposes: (1) to examine whether the likelihood of having increased SF differs between subjects with and without AI; (2) to determine whether blacks are more likely to have elevated SF in regard to AI than whites, and (3) to investigate whether there are any differences in the magnitude of having elevated SF between blacks and whites with AI present.

## METHODS

### Data source

NHANES III was a national representative survey conducted by the National Center for Health

Statistics between 1988 and 1994, to assess the health and nutritional status of the US population. The data were collected via standardized questionnaires, physical examinations and laboratory investigations. NHANES III included a sample of 33 994 persons aged  $\geq 2$  months from 89 randomly selected locations throughout the United States. Multistage, stratified sampling design was applied to select the participants constituting a representative sample of the civilian, non-institutionalized national population. Additional details regarding the study design and sample selection were reported previously [21].

### Study sample

A total number of 5007 NHANES III male subjects, aged  $\geq 20$  years, with measured SF and C-reactive protein (CRP) was included in this study. Among them, 3112 were non-Hispanic white (NHW) and 1895 were non-Hispanic black (NHB). Due to the impact each has on ferritin, subjects who were haemophiliacs or who had recently undergone chemotherapy were excluded from the analysis. Subjects who had been treated for anaemia (including diet, iron pills, iron injections, transfusions as treatment) within the past 3 months or who had blood donation within 1 month prior to the survey were also excluded from the analysis. We only chose male subjects to eliminate the additional, potentially confounding effects associated with female subjects, such as menstruation, oral contraceptive use, pregnancy, parity, menopause and hormone replacement therapy.

### Variables

Elevated SF was defined as a ferritin concentration  $> 300 \mu\text{g/l}$  in men [22]. Presence of AI was defined as a CRP  $\geq 1.0 \text{ mg/dl}$  [23]. Disease status including cancer (various types), congestive heart failure, diabetes mellitus, heart attack, hypertension, and stroke was determined by the responses to questions of whether a doctor has ever told the respondent that he had such conditions. Undiagnosed diabetes was identified by a fasting plasma glucose concentration  $> 126 \text{ mg/dl}$  [24]. Hypertension was defined as a systolic blood pressure  $\geq 140 \text{ mmHg}$  or/and diastolic blood pressure  $\geq 90 \text{ mmHg}$  [25], or if a doctor has told the participant that he had hypertension.

Several inflammation markers including plasma fibrinogen, serum albumin, white blood cell count

(WBC), lymphocyte number, monocyte number and platelet count were examined. Other laboratory variables including haemoglobin (Hb), serum iron, transferrin saturation (TS), total iron binding capacity (TIBC), total triglycerides (TG), total cholesterol, high density lipoprotein (HDL) and low density lipoprotein (LDL) cholesterol were also included in our study. Detailed description of laboratory measurements of these variables has been published elsewhere [26].

Ethnicity, age, education, poverty income ratio (PIR), body mass index (BMI, kg/m<sup>2</sup>), and cigarette smoking may be associated with both AI and the levels of SF. These variables were, therefore, included in the logistic regression models as potential confounders. Age was modelled as a continuous variable, and was further divided into seven groups: 20–29, 30–39, 40–49, 50–59, 60–69, 70–79, and ≥80 years, in order to present the age-stratified distribution of elevated SF between black and white subjects with AI present. Education was measured as the highest year or grade completed by the respondent, and was further dichotomized as higher than or less than high-school education. PIR was measured based on total household income adjusted for household size, and was used as an index of relative socioeconomic status in the NHANES III survey. Three PIR strata: <1.3, 1.3–3.5, and >3.5 were used to represent low, moderate and high income levels, respectively according to the analytical and reporting guidelines of NHANES III [27]. BMI was further divided into four categories: underweight (BMI <18.5), normal (BMI ≥18.5 to <25), overweight (BMI ≥25 to <30), and obese (BMI ≥30) [28]. Cigarette smoking was classified as never smoked (if they had smoked <100 cigarettes in their lifetime), former smoker (≥100 lifetime cigarettes, not currently smoking), and current smoker (≥100 lifetime cigarettes, currently smoking).

### Statistical analysis

The aims of our analyses were, first, to examine whether the likelihood of having increased SF differ between subjects with and without AI, within each ethnic group. This was determined by using multivariate logistic regression models to compare the odds ratios of having increased SF between subjects with and without AI present, stratified by ethnicity, after adjusting for the potential confounding effects.

The second aim of our study was to investigate whether blacks are more likely to have elevated SF than whites in regard to AI. This was determined by comparing the odds of having elevated SF between blacks and whites, stratified by AI status, after adjusting for the potential confounders.

The third aim of our analyses was to examine whether the magnitude of the elevated SF is different between blacks and whites with AI present. This was determined by comparing the risks of having increased SF per unit change in several inflammation markers including CRP, fibrinogen, WBC, albumin and platelet count between black and white subjects with AI present, after adjusting for the potential confounding effects. An interaction term constructed as cross-product of ethnicity and selected inflammation markers (as continuous variables) was included in the logistic regression models to allow calculation of  $\beta$  coefficients.

In addition to the confounders described earlier, cardiovascular diseases/diabetes mellitus/hypertension status, and cardiovascular disease risks including TG, total cholesterol, HDL cholesterol, LDL cholesterol, waist circumference, systolic and diastolic blood pressure were also adjusted for in logistic regression models because of their known relationship to ferritin and CRP [29]. However, LDL cholesterol and waist circumference were excluded from logistic models because of their unacceptable percentage of missing values (LDL cholesterol 59.65%, waist circumference 15.54%). Multicollinearity among independent variables for each logistic model was tested by checking the variance inflation factor (VIF). Any variable with a VIF that exceeded 4 was excluded from the model (no variable was detected with VIF greater than 4).

Data were analysed by using SAS 9.1 (SAS Institute Inc., Cary, NC, USA) and SUDAAN 9.0 (Research Triangle Institute, Research Triangle Park, NC). Descriptive statistics were computed for all variables, including means for continuous data, frequencies for categorical variables, and standard error of the mean (S.E.M.). SUDAAN's *t* test and SUDAAN's  $\chi^2$  test were used to compare the means of all continuous variables and frequencies of all categorical variables between NHWs and NHBs stratified by status of AI, respectively. Logistic regression analysis was applied for our likelihood and magnitude tests. The statistical significance level was set at  $P < 0.05$ . Sampling weights were applied to all analyses to account for the complex design effect and for non-response.

## RESULTS

### Subjects characteristics

The overall prevalence of AI (CRP  $\geq 1.0$  mg/dl) was 7.97% (399/5007), including 235 NHWs and 164 NHBs (Table 1). The mean SF was significantly higher in NHBs than NHWs (212.2  $\mu\text{g/l}$  vs. 175.9  $\mu\text{g/l}$ ,  $P < 0.01$ ). In individuals with AI present, NHBs were younger, poorer, less likely to be a former smoker but more likely to smoke currently, and had a higher percentage of subjects with less than high school education compared to NHWs. Overall, there was no significant difference in reported disease prevalence including cancer, congestive heart failure, diabetes mellitus, heart attack, hypertension and stroke between NHW and NHB subjects with AI. There were no significant differences in the values of CRP, serum albumin, platelet count, plasma fibrinogen and lymphocyte between NHBs and NHWs. However, NHWs with AI present were found to have significantly higher values of WBC and mononuclear counts compared to their black counterparts. NHBs had significant lower values of Hb, but their SF concentrations were significantly higher compared to NHWs. Significant differences in HDL and LDL cholesterol were also found between these two groups (Table 1).

For subjects without AI, significant differences in age, education, income level and smoking status were found between NHWs and NHBs. Generally, NHBs were younger, less educated, more likely to be non-smokers and current smokers, and had less income compared to their NHW counterparts. Disease status was not significantly different between the two groups, except for the significantly higher prevalence of heart attack in NHWs. The overall BMI and its categories were not found to be significantly different between NHBs and NHWs, but NHWs were observed to have significantly higher waist circumference compared to NHBs. All the infection markers were found to be significantly different between these two groups except for plasma fibrinogen. Haematological markers were also significantly different between NHWs and NHBs. Compared to NHWs, NHBs had significantly lower Hb but higher SF concentration. NHWs showed significantly higher levels of TG, total cholesterol and significantly lower levels of HDL cholesterol compared to NHBs. However, systolic blood pressure was significantly higher in NHBs (Table 1).

### Distribution of elevated SF

The proportion of subjects with AI present who had elevated SF concentrations ( $> 300 \mu\text{g/l}$ ) stratified by age and ethnicity is shown in the Figure. NHBs generally showed a higher proportion of individuals with elevations in SF in regard to AI than their white counterparts at each age group except for age 20–29 years. The greatest ethnic difference in the percentage of individuals with elevated SF concentrations was seen in the 40–49 years age group.

The prevalence of elevated SF was significantly higher in NHBs (35.23%) with AI present than NHWs (21.78%). In the absence of AI, the prevalence of elevated SF was also significantly higher in NHBs (18.71%) than NHWs (13.14%). Generally, the proportion of subjects with elevated SF was lower in subjects without AI than in subjects with AI. Regardless of AI status, the proportion of individuals with normal ferritin concentrations was significantly higher in NHWs compared to NHBs (Table 2).

### Multivariate analysis

Compared with NHB males without AI, NHBs with AI present were 1.60 times more likely (95% CI, 1.11–2.35) to have elevations in SF after adjustment for age, PIR, education, BMI, smoking status, and disease status. When the relationship was further adjusted for cardiovascular disease risks including TG, total cholesterol, HDL cholesterol, systolic and diastolic blood pressure, NHBs with AI present were associated with a 1.71-fold greater risk (95% CI 1.18–2.49) of having elevated SF than NHBs without AI. In contrast, there was no significant association between elevated SF and AI found in NHWs (Table 3).

Irrespective of AI status, NHBs were more likely to have elevations in SF than NHWs (Table 4). With AI present, NHBs were 2.15 times more likely (95% CI 1.23–3.75) to have elevations in SF compared to their white counterparts after adjusting for age, ethnicity, PIR, education, BMI, cigarette smoking and disease status. The relationship remained significant after further adjustment for cardiovascular risks, and the odds ratio of having elevated SF in NHBs was 2.24 (95% CI 1.33–3.76). In the absence of AI, NHBs had a 1.81-fold greater chance (95% CI 1.43–2.30) of having elevated SF than did NHWs. After additional adjustment for cardiovascular risks, the odds ratio of having increased SF in NHBs was 1.87 (95% CI 1.46–2.40) (Table 4).

Table 1. Characteristics of 5007 male NHANES III participants aged  $\geq 20$  years by status of acute inflammation (AI) and ethnicity

Characteristics	Presence of AI				Absence of AI			
	Non-Hispanic white ( <i>n</i> = 235)		Non-Hispanic black ( <i>n</i> = 164)		Non-Hispanic white ( <i>n</i> = 2877)		Non-Hispanic black ( <i>n</i> = 1731)	
	<i>n</i>	Mean (S.E.M.)	<i>n</i>	Mean (S.E.M.)	<i>n</i>	Mean (S.E.M.)	<i>n</i>	Mean (S.E.M.)
Age (yr)	235	56.0 (0.3)	164	50.7 (1.2)	2877	44.5 (0.5)	1731	40.2 (0.5)**
Education (yr)	235	12.00 (0.29)	164	12.18 (1.19)	2877	13.11 (0.11)	1731	12.35 (0.26)**
$\leq 12$ yr (%)	114	33.76 (4.22)	99	50.54 (3.52)*	865	20.37 (1.22)	647	31.76 (1.66)**
$> 12$ yr (%)	121	66.24 (4.22)	65	49.46 (3.52)*	2012	79.63 (1.22)	1084	68.24 (1.66)**
Poverty income ratio	217	2.94 (0.14)	149	1.95 (0.15)*	2877	3.52 (0.08)	1731	2.28 (0.08)**
$< 1.3$ (%)	69	26.91 (3.95)	85	45.79 (5.51)*	527	14.52 (1.00)	704	37.79 (1.91)**
$1.3-3.5$ (%)	106	46.06 (5.14)	61	41.23 (5.21)	1309	42.78 (1.41)	750	43.39 (1.58)**
$\geq 3.5$ (%)	60	27.02 (3.53)	18	12.98 (3.37)*	1041	42.70 (1.65)	277	18.82 (1.30)**
Prevalence (%)								
Cancer (various types)	0	0	0	0	0	0	0	0
Congestive heart failure	22	5.69 (2.13)	14	9.66 (3.37)	128	2.07 (0.25)	55	2.19 (0.38)
Diabetes mellitus	4	1.21 (0.64)	8	4.87 (1.97)	41	0.94 (0.28)	38	1.60 (0.36)
Heart attack	42	12.71 (2.83)	14	8.99 (2.73)	267	4.74 (0.49)	60	2.68 (0.38)**
Hypertension	82	28.30 (4.55)	65	33.98 (3.39)	819	19.23 (1.18)	473	23.30 (1.18)**
Stroke	26	6.36 (0.94)	14	6.12 (1.50)	116	1.85 (0.23)	39	1.48 (0.29)**
Body mass index (kg/m <sup>2</sup> )	235	27.50 (0.50)	164	28.45 (0.62)	2877	26.69 (0.13)	1728	26.42 (0.13)
$< 18.5$ (%)	8	2.02 (0.82)	6	3.34 (1.43)	40	1.02 (0.24)	29	1.52 (0.29)
$\geq 18.5$ to $< 25$ (%)	82	33.08 (5.13)	53	28.08 (3.68)	1087	38.28 (1.20)	718	41.83 (1.33)
$\geq 25$ to $< 30$ (%)	96	40.75 (4.72)	54	37.17 (5.30)	1215	40.96 (1.14)	630	36.61 (1.08)
$\geq 30$ (%)	49	24.15 (3.13)	51	31.42 (3.84)	535	19.73 (0.90)	351	20.04 (1.02)
Waist circumference (cm)	190	100.77 (0.99)	147	99.31 (1.55)	2706	96.17 (0.27)	1657	91.47 (0.38)**
Smoking status (%)								
Non-smoker	57	20.67 (2.81)	43	25.99 (2.53)	933	34.87 (1.16)	660	40.59 (1.63)**
Former smoker	112	37.24 (3.96)	37	17.89 (2.73)*	1233	34.41 (1.23)	399	20.88 (1.16)**
Current smoker	66	42.10 (4.54)	84	56.12 (3.75)*	711	30.72 (1.28)	671	38.53 (1.27)**
CRP (mg/dl)	235	2.27 (0.15)	164	2.19 (0.12)	2877	0.27 (0.01)	1731	0.29 (0.01)**
Serum albumin (g/dl)	232	3.97 (0.05)	163	3.85 (0.03)	2854	4.32 (0.02)	1712	4.20 (0.02)**
WBC ( $10^3$ cells/mm <sup>3</sup> )	235	9.17 (0.18)	163	7.45 (0.18)*	2828	7.23 (0.05)	1714	6.37 (0.06)**
Platelet count ( $10^3$ cells/mm <sup>3</sup> )	235	281.69 (4.9)	163	282.06 (8.47)	2826	254.42 (2.28)	1714	260.89 (2.51)**
Plasma fibrinogen (mg/dl)	216	668.01 (11.45)	131	614.76 (11.23)	2095	516.12 (56.64)	868	540.17 (52.69)
Lymphocyte ( $10^3$ cells/mm <sup>3</sup> )	235	2.18 (0.07)	163	2.37 (0.09)	2827	2.21 (0.02)	1714	2.32 (0.02)**
Mononuclear ( $10^3$ cells/mm <sup>3</sup> )	233	0.56 (0.01)	160	0.47 (0.02)*	2794	0.45 (0.01)	1669	0.41 (0.01)**
Haemoglobin (g/dl)	235	14.63 (0.14)	163	13.83 (0.11)*	2828	15.17 (0.04)	1714	14.52 (0.04)**
Serum iron ( $\mu$ g/l)	234	62.07 (3.04)	164	61.17 (2.70)	2875	100.57 (0.93)	1729	93.60 (1.06)
Serum ferritin ( $\mu$ g/l)	235	212.19 (11.26)	164	274.64 (14.90)*	2877	174.10 (4.07)	1731	207.29 (6.14)**
TS (%)	234	19.63 (0.96)	164	19.81 (0.89)	2870	29.45 (0.32)	1725	28.07 (0.30)**
TIBC	234	329.37 (4.56)	164	315.16 (5.08)	2872	347.34 (2.46)	1726	338.62 (2.21)**
Triglycerides (mg/dl)	233	162.32 (9.83)	164	141.87 (6.08)	2867	159.05 (3.69)	1727	120.85 (2.52)**
Total cholesterol (mg/dl)	235	198.56 (3.68)	164	206.52 (5.11)	2876	203.25 (1.09)	1727	197.77 (1.21)**
HDL cholesterol (mg/dl)	232	41.99 (0.93)	164	47.63 (1.58)*	2858	45.05 (0.46)	1720	52.79 (0.53)**
LDL cholesterol (mg/dl)	89	122.63 (3.43)	72	144.19 (7.04)*	1202	131.35 (1.13)	718	125.66 (1.85)**
Plasma glucose (mg/dl)	207	104.60 (2.31)	159	115.75 (5.44)	2792	99.95 (0.64)	1722	98.94 (0.80)
Systolic blood pressure (mmHg)	206	130.24 (1.76)	158	133.08 (1.17)	2796	124.54 (0.52)	1725	126.41 (0.49)**
Diastolic blood pressure (mmHg)	206	76.64 (0.71)	158	78.33 (1.04)	2796	76.63 (0.28)	1725	77.98 (0.43)**

CRP, C-reactive protein; WBC, white blood cell; TS, transferrin saturation; TIBC, total iron binding capacity; HDL, high density lipoprotein; LDL, low density lipoprotein.

\* The means of all continuous variables were compared between AI present non-Hispanic white and non-Hispanic black male subjects using SUDAAN's *t* test. The frequencies of all categorical variables were compared between these two groups by using SUDAAN's  $\chi^2$  test. Statistical significance was set at  $P < 0.05$ .

\*\* The means of all continuous variables were compared between AI absent non-Hispanic white and non-Hispanic black male subjects using SUDAAN's *t* test. The frequencies of all categorical variables were compared between these two groups by using SUDAAN's  $\chi^2$  test. Statistical significance was set at  $P < 0.05$ .

Table 2. Association between serum ferritin levels† and acute inflammation (AI) status, by ethnicity

Ethnicity	Presence of acute inflammation			
	Yes		No	
	<i>n</i>	% (S.E.M.)	<i>n</i>	% (S.E.M.)
Non-Hispanic white ( <i>n</i> = 3112)				
Elevated ferritin	54	21.78 (2.67)	417	13.14 (0.91)
Normal ferritin	172	75.40 (3.00)	2364	84.02 (1.01)
Decreased ferritin	9	2.82 (0.88)	96	2.84 (0.35)
Non-Hispanic black ( <i>n</i> = 1985)				
Elevated ferritin	56	35.23 (3.60)*	330	18.71 (1.15)**
Normal ferritin	105	63.20 (3.68)*	1359	79.45 (1.13)**
Decreased ferritin	3	1.58 (1.26)	42	1.85 (0.36)

† A serum ferritin concentration <25 µg/l, 25–300 µg/l, or >300 µg/l was used to define decreased, normal and elevated ferritin level, respectively. SUDAAN's  $\chi^2$  test was used to compare the number of individuals with decreased, normal, and elevated ferritin level between blacks and whites, respectively, stratified by AI status.

\* Significant difference between AI present non-Hispanic white and non-Hispanic black male subjects,  $P < 0.05$ .

\*\* Significant difference between AI absent non-Hispanic white and non-Hispanic black male subjects,  $P < 0.05$ .

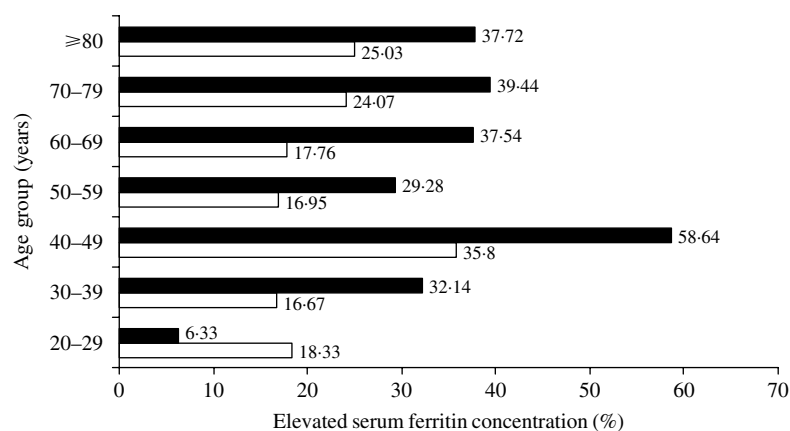


Fig. The proportion of subjects with acute inflammation present who had elevated serum ferritin concentration (> 300 µg/l), stratified by age and ethnicity. ■, Non-Hispanic black; □, non-Hispanic white.

With AI present, every increment of WBC, serum albumin, lymphocyte count and platelet count was associated significantly higher risks of having elevated SF in NHBs than NHWs in both reduced and full logistic models (Table 5). With additional adjustment for cardiovascular disease risks in the full logistic regression model, every unit change in CRP was associated with significantly higher odds ratios of having elevations in SF in NHBs than NHWs. However, no significant difference in the odds ratios of having

elevated SF with unit change in monocyte count was found between NHBs and NHWs (Table 5).

## DISCUSSION

In the present study, the prevalence of elevated SF was significantly higher in NHBs than NHWs, regardless of AI status. Overall, the mean SF concentration of NHBs was significantly higher than that of

Table 3. Association between acute inflammation (AI) and elevated serum ferritin (SF) concentration, stratified by ethnicity†

Ethnicity	Adjusted odds ratio			
	Model no. 1‡		Model no. 2§	
	<i>n</i>	OR (95% CI)	<i>n</i>	OR (95% CI)
Non-Hispanic white	2910	1.43 (0.87–2.34)	2787	1.38 (0.89–2.12)
Non-Hispanic black	1729	1.60 (1.09–2.35)	1709	1.71 (1.18–2.49)

OR, Odds ratio; CI, confidence interval.

† Odds ratios of having elevated SF concentrations were compared between AI present (CRP  $\geq$  1.0 mg/dl) subjects and AI absent subjects (reference group) within each ethnicity. Elevated SF concentration defined as  $> 300 \mu\text{g/l}$  for male.

‡ Model no. 1 adjusted for age, poverty income ratio, education, body mass index, smoking status, chronic diseases including cancer, congestive heart failure, heart attack, hypertension, diabetes and stroke.

§ Model no. 2 further adjusted for cardiovascular risk factors including triglycerides, total cholesterol, high density lipoprotein cholesterol, systolic and diastolic blood pressure in addition to the effects adjusted in model no. 1.

Table 4. Comparisons of non-Hispanic whites and non-Hispanic blacks in regard to the risk of having elevated serum ferritin (SF), stratified by acute inflammation (AI) status†

Logistic regression models	Presence of AI			
	Yes		No	
	<i>n</i>	aOR (95%CI)	<i>n</i>	aOR (95%CI)
Model no. 1‡	366	2.15 (1.23–3.75)	4273	1.81 (1.43–2.30)
Model no. 2§	333	2.24 (1.33–3.76)	4163	1.87 (1.46–2.40)

aOR, Adjusted odds ratio.

† Odds ratios of having elevated SF in regard to AI were compared between non-Hispanic white (reference group) and non-Hispanic black male subjects, stratified by AI status. Elevated SF defined as  $> 300 \mu\text{g/l}$  for male.

‡ Model no. 1 adjusted for age, ethnicity, poverty income ratio, education, body mass index, smoking status, chronic diseases including cancer, congestive heart failure, heart attack, hypertension, diabetes and stroke.

§ Model no. 2 further adjusted for cardiovascular risk factors including triglycerides, total cholesterol, high density lipoprotein cholesterol, systolic and diastolic blood pressure in addition to the effects adjusted in model no. 1.

NHWs. This finding is consistent with the results from previous studies in which blacks were found to have higher SF concentrations than their white counterparts [30–33]. Although most probably the higher SF in blacks results from a complex interplay of individual attributes, minority, socioeconomic stress and environmental characteristics, the possible influence of different response to inflammation on the ethnic differences in ferritin distribution has caught increasing research interest.

In the present study, NHBs consistently had a higher percentage of individuals with elevated SF than that of NHWs after age 30 years, when AI was present. In contrast to NHWs, NHBs were more likely to have elevated SF with respect to AI. Moreover, with AI present, NHB ethnicity was not only associated with greater risks of having increased SF, but also associated with a much greater magnitude of elevations in SF per unit change in inflammation markers. Therefore, the results raised the

Table 5. Comparisons of having elevated serum ferritin per unit change in inflammation markers between non-Hispanic whites and non-Hispanic blacks, with acute inflammation present†

Inflammation marker	Model 1‡		Model 2§	
	<i>n</i>	$\hat{\beta}$ ( <i>P</i> )¶	<i>n</i>	$\hat{\beta}$ ( <i>P</i> )¶
C-reactive protein	366	0.10 (0.3556)	333	0.21 (0.0005)*
White blood cell	365	0.06 (<0.0001)*	332	0.05 (0.0001)*
Serum albumin	364	0.08 (0.0201)*	331	0.09 (0.0031)*
Lymphocyte count	365	0.15 (0.0040)*	332	0.14 (0.0038)*
Monocyte count	361	0.38 (0.0987)	328	0.32 (0.1264)
Platelet count	365	0.002 (0.0002)*	332	0.002 (0.0003)*

† Logistic regression models were used to estimate the magnitude of serum ferritin elevation associated with one unit change in inflammation markers. Interaction term (composite of ethnicity and selected infection markers as continuous variables) was included to allow calculation of  $\beta$  coefficients.

‡ Model no. 1 adjusted for age, ethnicity, poverty income ratio, education, body mass index, smoking status, chronic diseases including cancer, congestive heart failure, heart attack, hypertension, diabetes and stroke.

§ Model no. 2 further adjusted for cardiovascular risk factors including triglycerides, total cholesterol, high density lipoprotein cholesterol, systolic and diastolic blood pressure in addition to the effects mentioned in Model no. 1.

¶ *P* value corresponding to the comparisons between the  $\beta$  of non-Hispanic black and non-Hispanic white (reference group), respectively.

\* Statistical significance set as  $P < 0.05$ .

possibility that there might be different modes of inflammation-mediated ferritin increase between blacks and whites.

Cytokines directly regulate ferritin secretion both transcriptionally and post-transcriptionally. A number of cytokines including interleukin-1-alpha (IL-1 $\alpha$ ), interleukin 6 (IL-6), tumour necrosis factor-alpha (TNF- $\alpha$ ) have been shown to up-regulate ferritin production [1, 9]. Pro-inflammatory cytokines TNF- $\alpha$  and IL-1 $\alpha$  transcriptionally induce the H chain of ferritin. As a result, the selective induction of ferritin H mRNA will lead to the accumulation of a population of H-rich ferritin protein and substantially increase the content of ferritin [1]. In primary cultured human hepatocytes, IL-1 $\alpha$  and IL-6 can induce a transient secretion of ferritin at 24 h followed by a decline to baseline [34]. Results from these studies suggest another possibility that the differential patterns of raised ferritin observed between blacks and whites could be a result of different cytokine-induced ferritin synthesis during inflammation. However, prospective studies are required to test the hypothesis.

In our study, NHBs demonstrated a greater capability and propensity to have increased stored iron, reflected as elevated SF, than NHWs in regard to AI.

One potential explanation for this finding is that the observed disparate patterns of ferritin increase may be a result of the different cytokine genotype distribution between blacks and whites. Several studies have shown that significantly different allelic and genotypic cytokine distribution exists between blacks and whites [35–37]. Most studies reported that blacks have a greater predisposition to inflammation because of the more up-regulated inflammation cytokine genotypes they carry, IL-1 $\alpha$ , IL-6 and TNF- $\alpha$  in particular. Therefore, the higher risks of having raised SF observed in blacks with inflammation present may be the functional consequences of their predisposition to the higher levels of pro-inflammatory cytokines which are known to stimulate ferritin secretion.

We also observed that, with AI present, every unit increment of selected inflammation markers was associated with significantly higher risks of having elevated SF in NHBs than in NHWs. The stronger inflammation–ferritin relationship observed in blacks raised another possibility that the amplified increase in stored iron may predispose blacks to worse clinical outcomes and poorer treatment responses. Indeed, observations over the last few decades have identified ethnic differences in response to therapies in many



diseases, including hypertension [38, 39], diabetes [40], renal transplantation [41], and heart failure [42]. In addition, there are striking ethnic differences in response to interferon therapy for hepatitis C (HCV). Long-term sustained responses to alpha interferon are substantially lower in black patients than white patients with HCV [43]. Interestingly, the fact that iron depletion by phlebotomy in patients with HCV reduces serum aminotransferase levels [44], and in combination with interferon, may have improved antiviral efficacy compared with interferon alone [45] suggests that increased iron stores may be the cause of the reduced response to interferon. Although the presence of increased iron stores is a recognized predictor of poor response to interferon monotherapy [46–48], the association between increased iron stores and treatment response for other diseases has not yet been adequately investigated. Given the fact that ferritin plays a prominent role in the cytokine response and inflammatory processes, perhaps the more aggressive iron accumulation pattern, reflected by the higher likelihood and greater magnitude of elevated SF is a reflection of the greater intensity of inflammation and the stronger host immunoresponsiveness underlying a number of physiopathological processes. Therefore, larger, prospective studies are needed to investigate whether the different patterns of increase in iron stores are associated with different therapy response among various ethnicities.

To date, little is known about the underlying mechanisms of the disease disparities highlighted by the strikingly higher morbidity and mortality rates in blacks. Compared with whites, blacks have ~1.5 times higher death rates from all causes, worse survival after myocardial infarction [49], a twofold-higher incidence of and morbidity from stroke [50, 51], and significantly higher morbidity and mortality rates from diabetes [52], AIDS [53], and cancer [54]. Given the pivotal role ferritin plays in the cellular inflammatory response and pathogenesis of disease, the question of whether there is a difference in responsive ferritin synthesis during inflammation between blacks and whites, and whether the differences in iron store play a role in disease disparities merits further research.

To our knowledge, this is the first study to assess the ethnic difference in the association between AI and increased iron stores, defined as elevated SF between blacks and whites. Despite the fact that ethnic differences in disease burden, progressions, complications, outcomes and treatment response have been

observed between blacks and whites, the explanation for this remains unknown. Of great interest is to discover whether there are ethnic differences in the inflammatory process and host immunoresponsiveness that underlie a variety of clinical pathophysiological conditions. Within this work, we presented evidence that blacks seem to have a more aggressive pattern of having elevated SF in regard to AI compared to whites. And the stronger inflammation–ferritin relationship observed in blacks may reflect their more robust immune response and, therefore, may contribute to the poorer prognosis and treatment resistance. However, more prospective data are required to further investigate the hypothesis.

Despite the significant findings provided, our analysis is not powered to address the underlying mechanisms of the observed ethnic differences in the association between AI and raised ferritin. Due to the cross-sectional nature of the design, causal relationship cannot be inferred between AI and raised SF. Therefore, more specific investigation is needed to further explore the responsible biological rationale. However, our study does raise the possibility that different patterns of iron accumulation may play a role in disease processes and result in different clinical outcomes and drug response. Future studies will address this topic.

In conclusion, the current results indicated that different patterns of iron accumulation in regard to AI exist between blacks and whites. Further elucidation is needed about how ferritin regulation is perturbed in diseases, and whether increased iron stores play a role in different clinical outcomes and drug response. To what extent different levels of iron stores contribute to the disparate ethnic responses to disease and treatments requires further investigation. A more complete knowledge of the aetiology of the differences in host inflammatory response could provide clinicians with the ability to optimize the therapeutic regimens tailored to the individual.

## DECLARATION OF INTEREST

None.

## REFERENCES

1. Torti FM, Torti SV. Regulation of ferritin genes and protein. *Blood* 2002; **99**: 3505–3516.
2. Linert W, Jameson GNL. Redox reactions of neurotransmitters possibly involved in the progression of

- Parkinson's Disease. *Inorganic Biochemistry* 2000; **79**: 319–326.
3. **Kondo T, et al.** Embryonic genes expressed in Alzheimer's disease brains. *Neuroscience Letters* 1996; **209**: 157–160.
  4. **Pang JH, et al.** Increased ferritin gene expression in atherosclerotic lesions. *Journal of Clinical Investigation* 1996; **97**: 2204–2212.
  5. **Ryan TP, et al.** Pulmonary ferritin: differential effects of hyperoxic lung injury on subunit mRNA levels. *Free Radical Biology & Medicine* 1997; **22**: 901–908.
  6. **Biamond P, et al.** Intraarticular ferritin-bound iron in rheumatoid arthritis – a factor that increases oxygen free radical-induced tissue destruction. *Arthritis & Rheumatism* 1986; **29**: 1187–1193.
  7. **Ahmadzadeh N, et al.** Correlation of metal-binding proteins and proteinase inhibitors with immunological parameters in rheumatoid synovial fluids. *Clinical and Experimental Rheumatology* 1990; **8**: 547–551.
  8. **Wu CG, et al.** Rat ferritin-H: CDNA cloning, differential expression and localization during hepatocarcinogenesis. *Carcinogenesis* 1997; **18**: 47–52.
  9. **Torti SV, et al.** The molecular cloning and characterization of murine ferritin heavy chain, a tumor necrosis factor-inducible gene. *Journal of Biological Chemistry* 1988; **263**: 12 638–12 644.
  10. **Picard V, et al.** Overexpression of the ferritin H subunit in cultured erythroid cells changes the intracellular iron distribution. *Blood* 1996; **87**: 2057–2064.
  11. **Picard V, et al.** Role of ferritin in the control of the labile iron pool in murine erythroleukemia cells. *Journal of Biological Chemistry* 1998; **273**: 15 382–15 386.
  12. **Risch N, et al.** Categorization of humans in biomedical research: genes, race and disease. *Genome Biology* 2002; **3**: 1–12.
  13. **Ahern J, et al.** Preterm birth among African-American and white women: a multilevel analysis of socioeconomic characteristics and cigarette smoking. *Journal of Epidemiology and Community Health* 2003; **57**: 606–611.
  14. **Simhan HN, et al.** Interleukin-6 promoter-174 polymorphism and spontaneous preterm birth. *American Journal of Obstetrics and Gynecology* 2003; **189**: 915–918.
  15. **Wong MD, et al.** Contribution of major diseases to disparities in mortality. *New England Journal of Medicine* 2002; **347**: 1585–1592.
  16. **Padovani JC, et al.** Gene polymorphisms in the TNF locus and the risk of myocardial infarction. *Thrombosis Research* 2000; **100**: 263–269.
  17. **Losito A, et al.** Association of interleukin-6 -174G/C promoter polymorphism with hypertension and left ventricular hypertrophy in dialysis patients. *Kidney International* 2003; **64**: 616–622.
  18. **Petri M.** Epidemiology of systemic lupus erythematosus. *Best Practice & Research Clinical Rheumatology* 2002; **16**: 847–858.
  19. **Hoffmann SC, et al.** Ethnicity greatly influences cytokine gene polymorphism distribution. *American Journal of Transplantation* 2002; **2**: 560–567.
  20. **Biamond P, et al.** Intraarticular ferritin-bound iron in rheumatoid arthritis: a factor that increases oxygen free radical induced tissue destruction. *Arthritis & Rheumatism* 1986; **29**: 1187–1193.
  21. **National Center for Health Statistics, Centers for Disease Control.** Plan and operation of the Third National Health and Nutrition Examination Survey, 1988–1994. Series 1: programs and collection procedures. *Vital Health Statistics* 1994; **1**: 1–407.
  22. **Barton JC, et al.** Management of hemochromatosis. Hemochromatosis Management Working Group. *Annals of Internal Medicine* 1998; **129**: 932–939.
  23. **Young B, Gleeson M, Cripps AW.** C-reactive protein: a critical review. *Pathology* 1991; **23**: 118–124.
  24. **Expert Committee on the Diagnosis and Classification of Diabetes Mellitus.** Report of the expert committee on the diagnosis and classification of diabetes mellitus. *Diabetes Care* 2000; **23** (Suppl. 1): S4–19.
  25. **Chobanian AV, Bakris GL, Black HR.** The Seventh Report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure: The JNC 7 Report. *Journal of the American Medical Association* 2003; **289**: 2560–2571.
  26. **Gunter EW, Lewis BG, Koncickowski SM.** Laboratory procedures used for the Third National Health and Nutrition Examination Survey (NHANES III), 1988–1994. Hyattsville: Centers for Disease Control and Prevention, 1996.
  27. **National Center for Health Statistics.** Analytic and Reporting Guidelines: The Third National Health and Nutrition Examination Survey, NHANES III 1988–1994. October 1996.
  28. **World Health Organization.** Obesity: Preventing and Managing the Global Epidemic. Report of a WHO Consultation on Obesity. Geneva: World Health Organization, 1998.
  29. **Mainous AG 3rd, et al.** Association of ferritin and lipids with C-reactive protein. *American Journal of Cardiology* 2004; **93**: 559–562.
  30. **Williams DM.** Racial differences of hemoglobin concentration: measurements of iron, copper, and zinc. *American Journal of Clinical Nutrition* 1981; **34**: 1694–1700.
  31. **Jackson RT, et al.** Comparison of hemoglobin values in black and white male U.S. military personnel. *Journal of Nutrition* 1983; **113**: 165–171.
  32. **Bazzano GS, et al.** Racial differences in iron-related hematological parameters. In: *Iron Fortification Research on Iron Fortification and Iron Supplementation Conducted in the U.S., Touro Research Institute*, New Orleans, LA, 1986, pp. 47–57.
  33. **Perry GS, et al.** Iron nutrition does not account for the hemoglobin differences between blacks and whites. *Journal of Nutrition* 1992; **122**: 1417–1424.
  34. **Muntanerelat J, et al.** Differential effects of cytokines on the inducible expression of CYP1A1, CYP1A2, and CYP3A4 in human hepatocytes in primary culture. *Hepatology* 1995; **22**: 1143–1153.
  35. **Cox ED, et al.** Cytokine polymorphic analyses indicate ethnic differences in the allelic distribution of

- interleukin-2 and interleukin-6. *Transplantation* 2001; **72**: 720–726.
36. **Hoffmann SC, et al.** Ethnicity greatly influences cytokine gene polymorphism distribution. *American Journal of Transplantation* 2002; **2**: 560–567.
  37. **Ness RB, et al.** Differential distribution of allelic variants in cytokine genes among African Americans and White Americans. *American Journal of Epidemiology* 2004; **160**: 1033–1038.
  38. **Seedat YK.** Varying responses to hypotensive agents in different racial groups: black versus white differences. *Journal of Hypertension* 1989; **7**: 515–518.
  39. **Weir MR, et al.** Differing mechanisms of action of angiotensin-converting enzyme inhibition in black and white hypertensive patients. The Trandolapril Multicenter Study Group. *Hypertension* 1995; **26**: 124–130.
  40. **Lillie-Blanton M, et al.** Racial differences in health: not just black and white, but shades of gray. *Annual Review of Public Health* 1996; **17**: 411–448.
  41. **Neylan JF.** Racial differences in renal transplantation after immunosuppression with tacrolimus versus cyclosporine. FK506 Kidney Transplant Study Group. *Transplantation* 1998; **65**: 515–523.
  42. **Carson P, et al.** Racial differences in response to therapy for heart failure: analysis of the vasodilator-heart failure trials. Vasodilator-Heart Failure Trial Study Group. *Journal of Cardiac Failure* 1999; **5**: 178–187.
  43. **Reddy KR, et al.** Racial differences in responses to therapy with interferon in chronic hepatitis C. Consensus Interferon Study Group. *Hepatology* 1999; **30**: 787–793.
  44. **Di Bisceglie AM, et al.** Iron reduction as an adjuvant to interferon therapy in patients with chronic hepatitis C who have previously not responded to interferon: a multicenter, prospective, randomized, controlled trial. *Hepatology* 2000; **32**: 135–138.
  45. **Fontana RJ, et al.** Iron reduction before and during interferon therapy of chronic hepatitis C: results of a multicenter, randomized, controlled trial. *Hepatology* 2000; **31**: 730–736.
  46. **Clemente MG, et al.** Effect of iron overload on the response to recombinant interferon-alfa treatment in transfusion-dependent patients with thalassemia major and chronic hepatitis C. *Journal of Pediatrics* 1994; **125**: 123–128.
  47. **Piperno A, et al.** Iron stores, response to alpha-interferon therapy, and effects of iron depletion in chronic hepatitis C. *Liver* 1996; **16**: 248–254.
  48. **Kageyama F, et al.** Failure to respond to interferon-alpha 2a therapy is associated with increased hepatic iron levels in patients with chronic hepatitis C. *Biological Trace Element Research* 1998; **64**: 185–196.
  49. **Gillum RF, Mussolino ME, Madans JH.** Coronary artery disease incidence and survival in African-American women and men: the NHANES I epidemiology follow-up study. *Annals of Internal Medicine* 1997; **127**: 111–118.
  50. **Sacco RL, et al.** Stroke incidence among white, black, and Hispanic residents of an urban community: the Northern Manhattan Stroke Study. *American Journal of Epidemiology* 1998; **147**: 259–268.
  51. **Broderick J, et al.** The greater Cincinnati/Northern Kentucky Stroke Study: preliminary first-ever and total incidence rates of stroke among blacks. *Stroke* 1998; **29**: 415–421.
  52. **Carter JS, Pugh JA, Monterrosa A.** Non-insulin-dependent diabetes mellitus in minorities in the United States. *Annals of Internal Medicine* 1996; **125**: 221–232.
  53. **Murrain M.** Differential survival in blacks and Mexican Americans with AIDS. *Ethnicity & Health* 1996; **1**: 373–382.
  54. **Parker SL, et al.** Cancer statistics by race and ethnicity. *CA: A Cancer Journal for Clinicians* 1998; **48**: 31–48.