Pioloform F Instead of Formvar as a Support Film for TEM

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Pioloform F (Wacker Chemie Co., Munich, Germany) is an excellent plastic for use as a support film for electron microscopy. The advantages of Pioloform F are high mechanical and thermal stability, and low material bulk (Stockem, 1970). Because of these properties, thinner films than Formvar or collodion may be used and remain stable under the electron beam without subsequent stabilization by carbon coating. Thin support films of gray interference color are prepared by dissolving 0.5 grams of Pioloform F powder in 100 milliliters of anhydrous chloroform (Stockem, 1970), using the dip method described by Reimer (1967). The Pioloform solution can be stored indefinitely in a brown bottle containing molecular sieve; this shortens preparation time and involves less handling of the components which are carcinogenic. Just before use, pipet the necessary amount of solution from the bottle without disturbing the molecular sieve. Our laboratory routinely uses Pioloform to support single slot grids, unstable ultrathin sections, and as a substrate for negative staining.

Pioloform films are similar to Formvar films in that the film surface is hydrophobic and carries a net positive charge. In negative stain techniques, the hydrophobic property of both films causes uneven spreading of the specimen and the stain. Common methods for rendering hydrophobic support films hydrophilic such as glow discharge, coating with cationic dyes, and the use of surfactants are effective for Pioloform films (for an excellent review of these methods, see Hayat and Miller, 1990).

A common pitfall is when films such as Pioloform are used in immunocytochemical procedures involving colloidal gold probes. Non-specific background labeling of the film often results from the electrostatic attraction of the

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negatively charged regions of the gold particle to the positively charged surface" of the film. Using fish gelatin as a protein stabilizer for the immunogold complex. (Birrell, 1987) or incorporation of gelatin (Bloom #60-100, 1% w/v) in the dilution and rinse buffers (Behnke, 1986) can reduce background problems. Other effec- 🕫 tive remedies are to use surfactants such as Tween-20 (0.05-0.1% v/v) in the dilution and rinse buffers or by increasing the salt concentration of the rinse buffer after the immunogold incubation step (i.e., use 1.0 M Tris-buffered saline instead of 0.1 M). Typically, we use a combination of several methods to prevent background when using Pioloform-coated grids with immunogold staining. For § example, the dilution and rinse buffers usually contain 0.1 M Tris - or phosphatebuffered saline (at the appropriate pH for the antibodies), 0.1% cold-water fish a skin gelatin, 0.05% Tween-20, and for the last two rinses after incubating in im- 2 munogold and before post-fixing in glutaraldehyde, rinse in 1.0 M Tris-HCI, pH 8.2. As for any immunolabeling procedure, different specimen types and gold probes will require modifying or combining these methods to achieve optimal a results.

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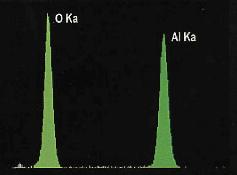
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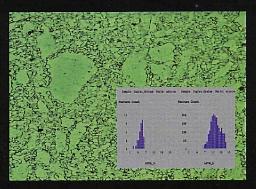
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