

Proceedings of the Nutrition Society (2017), **76**, 378–391 © The Authors 2016 First published online 2 November 2016

Nutrition Society Summer Meeting 2016 held at University College Dublin on 11-14 July 2016

# Conference on 'New technology in nutrition research and practice' Postgraduate Symposium

# Investigating cholesterol metabolism and ageing using a systems biology approach

A. E. Morgan<sup>1</sup>, K. M. Mooney<sup>2</sup>, S. J. Wilkinson<sup>1</sup>, N. A. Pickles<sup>3</sup> and M. T. Mc Auley<sup>1</sup>\*

<sup>1</sup>Department of Chemical Engineering, University of Chester, Thornton Science Park, Chester CH2 4NU, UK

<sup>2</sup>Faculty of Health and Social Care, Edge Hill University, Ormskirk, Lancashire L39 4QP, UK

<sup>3</sup>Department of Biological Sciences, University of Chester, Parkgate Road, Chester CH1 4BJ, UK

CVD accounted for 27 % of all deaths in the UK in 2014, and was responsible for 1.7 million hospital admissions in 2013/2014. This condition becomes increasingly prevalent with age, affecting 34·1 and 29·8 % of males and females over 75 years of age respectively in 2011. The dysregulation of cholesterol metabolism with age, often observed as a rise in LDL-cholesterol, has been associated with the pathogenesis of CVD. To compound this problem, it is estimated by 2050, 22 % of the world's population will be over 60 years of age, in culmination with a growing resistance and intolerance to pre-existing cholesterol regulating drugs such as statins. Therefore, it is apparent research into additional therapies for hypercholesterolaemia and CVD prevention is a growing necessity. However, it is also imperative to recognise this complex biological system cannot be studied using a reductionist approach; rather its biological uniqueness necessitates a more integrated methodology, such as that offered by systems biology. In this review, we firstly discuss cholesterol metabolism and how it is affected by diet and the ageing process. Next, we describe therapeutic strategies for hypercholesterolaemia, and finally how the systems biology paradigm can be utilised to investigate how ageing interacts with complex systems such as cholesterol metabolism. We conclude by emphasising the need for nutritionists to work in parallel with the systems biology community, to develop novel approaches to studying cholesterol metabolism and its interaction with ageing.

Cholesterol: Ageing: Systems biology: Computational modelling

Life expectancy has increased dramatically (Fig. 1). In the UK, males and females born in 1982, had a life expectancy of 71·1 and 77·0 years, respectively, while the projected values for 2082 are 89·7 and 92·6 years<sup>(1)</sup>. Thus, we are witnessing a staggering demographic shift in favour of older people (Fig. 2). For instance, it has been estimated the percentage of individuals in the UK over 60 years of age will double to 22 % by 2050, when compared to 2000<sup>(2)</sup>. Remaining disease free is a significant challenge faced by older people, as the prevalence of many conditions increases with age (Fig. 3). Of the diseases associated with advancing age, CVD is the

leading cause of morbidity in individuals over 60 years of age<sup>(3)</sup>. The dysregulation of cholesterol metabolism is intimately associated with the pathogenesis of CVD<sup>(4)</sup>, and age-related alterations in the metabolism of cholesterol are implicated in the disturbance of this system<sup>(5)</sup>. These include, a decrease in LDL-cholesterol clearance; a potential increase in cholesterol absorption; a decrease in bile acid synthesis; a decrease in bacterial bile acid modification<sup>(6–8)</sup>. It is likely that these alterations play a role in the accumulation of LDL-cholesterol and disease pathogenesis. The accumulation of plasma cholesterol can also be moderated by diet, while

Abbreviations: ABC, ATP-binding cassette; BSH, bile salt hydrolase; CETP, cholesteryl ester transfer protein; CYP7A1, cholesterol 7α-hydroxylase; HMG, 3-hydroxy-3-methylglutaryl; LDLr, LDL receptors; NPC1L1, Niemann–Pick C1-like 1 protein; TC, total cholesterol. \*Corresponding author: M. T. Mc Auley, email m.mcauley@chester.ac.uk





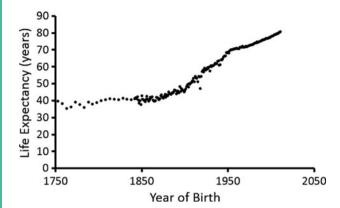


Fig. 1. UK life expectancy by year of birth. Data from Clio-Infra $^{(132)}$ .

pharmaceutical and pre- and probiotic administration have largely been associated with reduced LDL-cholesterol levels and CVD risk<sup>(9-13)</sup>.

Traditionally, when nutritionists have investigated complex metabolic pathways, such as cholesterol metabolism, they have utilised conventional wet laboratory techniques. However, studying cholesterol metabolism and its interaction with both diet and ageing using conventional approaches is challenging, due to the integrated nature of this system, and the time scales involved in studying the effects of the ageing process. Traditional in vivo or in vitro techniques can also be limited when testing a hypothesis, as such approaches can be resource-intensive, expensive, impractical and potentially unethical<sup>(14)</sup>. Thus, utilisation of the systems biology approach is becoming an increasingly important tool in nutrition-based research, as systems biology overcomes a number of the challenges outlined earlier, but more importantly, facilitates the integration of data generated from a diverse range of sources<sup>(14)</sup>, leading to an improved understanding of how cellular dynamics influence the behaviour of tissues and ultimately the health of whole-organ systems (15). Thus, the systems biology approach seeks to understand complex biological systems by studying them in a more holistic manner, in contrast to the reductionist approach regularly adopted in human nutrition. At the core of the systems biology approach is computational modelling. Computational modelling is an abstract process that is used to represent the dynamics of a biological system in a precise manner using mathematics. The steps involved in building a computational model are outlined in Fig. 4. Computational models are now used to model a diverse range of complex nutrient centred pathways including cholesterol metabolism for a number of reasons. Firstly, computational models are capable of providing quantitative data on the interaction of molecular components<sup>(16)</sup>. Secondly, nutrient-based interactions are inherently complex and often non-linear in nature<sup>(17–19)</sup>, and can involve complex feedback and feedforward loops<sup>(20–22)</sup>. Thus, it is challenging and even unfeasible to reason about these by human intuition alone. Computational modelling offers an alternative means of handling this complexity; thus, utilisation of computational modelling alongside the

experimental work provides a means of representing the dynamics of complex biological systems. Models can be used to simulate intrinsic perturbations, such as those associated with ageing and extrinsic perturbations, such as diet. Output from the model provides an overview of how these changes impact the dynamics of the system, and the implications this has for healthspan. In this review, we present an overview of cholesterol metabolism and discuss how ageing impacts its regulatory mechanisms. We also discuss how diet influences cholesterol metabolism, and how the dysregulation of this system influences heath. Next, we discuss therapeutic strategies for the treatment of hypercholesterolaemia, namely dietary, pharmacological and probiotic intervention. Finally, we describe how we are using the systems biology framework to investigate cholesterol metabolism and the impact ageing has on it. Specifically, there is a focus on how we have used computational modelling and how we are exploring this approach with simulated digestive tracks.

## Overview of cholesterol metabolism

Cholesterol plays a vital role in the body as a component of cell membranes, and precursor to steroid hormones and bile acids. Whole-body cholesterol metabolism is encapsulated by cholesterol ingestion, absorption, excretion and synthesis. These factors interact in a coordinated fashion to regulate whole-body cholesterol balance, with subtle changes to individual components influencing the behaviour of the others, so that cholesterol balance is maintained. In the next sections, we will outline in detail the complexities of cholesterol metabolism and how ageing interacts with it, thus emphasising the need for a systems biology approach when investigating it.

# Cholesterol ingestion and absorption

In the UK, the average daily intake of cholesterol is 304 and 213 mg for males and females, respectively<sup>(23)</sup>, 10–15 % of which is in the esterified form<sup>(24)</sup>. In the small intestine, esterified cholesterol is hydrolysed to form free cholesterol, which is more readily incorporated into bile acid micelles, which facilitate the absorption of cholesterol via Niemann-Pick C1-like 1 protein (NPC1L1)<sup>(25,26)</sup>. Additionally, phytosterols can also be absorbed via NPC1L1<sup>(27)</sup>. Intestinal absorption of cholesterol and phytosterols can be limited by heterodimer ATP-binding cassette (ABC) G5/G8, which effluxes these sterols back to the intestinal lumen<sup>(28)</sup>. Acyl CoA: cholesterol acyltransferase 2 esterifies internalised cholesterol, which is then incorporated into a nascent chylomicron via microsomal TAG transfer protein<sup>(29,30)</sup>. The nascent chylomicron then exits the enterocyte by exocytosis into the lymphatic system before entering the blood stream<sup>(31)</sup>. The nascent chylomicron is converted to a mature chylomicron upon acquisition of apo C-II and E from HDL. Apo C-II activates lipoprotein lipase on the capillary endothelium of adipose or muscle tissue,



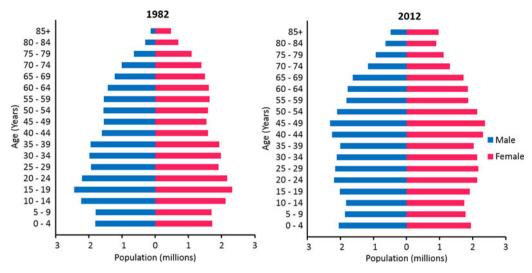


Fig. 2. (Colour online) UK population by age and sex in 1982 and 2012. Data from United Nations Statistics Division<sup>(133)</sup>.

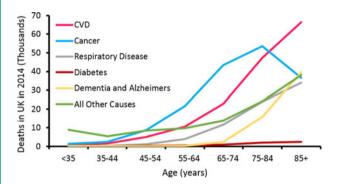


Fig. 3. (Colour online) UK causes of death by age. Data from British Heart Foundation  $^{(13.4)}$ .

which in turn catalyses the hydrolysis of TAG<sup>(32,33)</sup>. Apo C-II is then returned to HDL, and hepatic LDL receptors (LDLr) and LDLr-related protein recognise apo B-48 and E, initiating the absorption of the chylomicron remnants<sup>(34)</sup>.

#### Cholesterol synthesis

Cholesterol is synthesised endogenously in all nucleated cells in the body from acetyl CoA<sup>(35)</sup>. Renfurm *et al.*<sup>(36)</sup> observed endogenous cholesterol was synthesised at a rate of 9.8 (SD 6.2) mg/kg per d in healthy adults with a mean age of 32 years and mean weight of 64 kg. This equates to 627.2 mg/d synthesised cholesterol, a similar value to the 710 mg/d observed by Jones and Schoeller<sup>(37)</sup>. Interestingly, cholesterol consumption can influence the synthesis of endogenous cholesterol; an increase from 173 to 781 mg/d dietary cholesterol has been observed to decrease the rate of sterol synthesis by 34 %<sup>(38)</sup>.

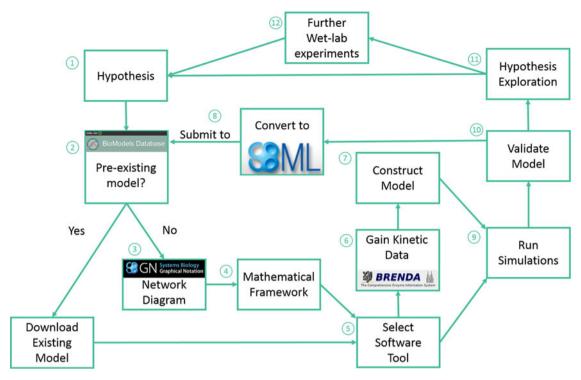
Cholesterol synthesis commences when acetoacetyl CoA thiolase catalyses the interconversion of acetyl CoA and acetoacetyl CoA. One molecule acetyl CoA and one molecule acetoacetyl CoA undergo a condensation

reaction by 3-hydroxy-3-methylglutaryl (HMG) CoA synthase to form a molecule of HMG CoA. HMG CoA reductase, with the addition of two NADPH molecules, and then catalyse the conversion of HMG CoA to mevalonate. As the rate-limiting enzyme of cholesterol biosynthesis, HMG CoA reductase is the therapeutic target of statins, for the treatment of hypercholesterolaemia, and the prevention of atherosclerosis (39). Phosphorylation of mevalonate by the enzyme mevalonate kinase forms mevalonate-5P, which undergoes further phosphorylation to form mevalonate-5PP via the enzyme phosphomevalonate kinase. Decarboxylation and dehydration by mevalonate-5PP decarboxylase create isopentenyl-PP and thus its isoform dimethylallyl-PP via isopentenyl diphosphate delta isomerase. Farnesyl diphosphate synthase initiates the condensation of dimethylallyl-PP with one molecule of isopentenyl-PP and NADPH to create geranyl-PP. Further condensation and the addition of another molecule of isopentenyl-PP and NADPH create farnesyl-PP. Condensation of two farnesyl-PP molecules by squalene synthase and NADPH forms squalene, which is then converted to squalene epoxide by squalene epoxidase, NADPH, and O2, before undergoing cyclisation by oxidosqualene cyclase to form lanosterol<sup>(40)</sup>. A series of reactions, including the branching of 7-dehydrodesmosterol to either desmosterol or 7-dehydrocholesterol, both of which can then be converted to the end product cholesterol via the enzymes 24-dehydrocholesterol reductase and 7-dehydrocholesterol reductase concludes the de novo synthesis of cholesterol(41,42).

#### Lipoprotein dynamics and reverse cholesterol transport

VLDL-cholesterol is formed from the hepatic pool of cholesterol to transport endogenously synthesised TAG to the tissues<sup>(43)</sup>. Partial hydrolysis of VLDL by lipoprotein lipase forms intermediate density lipoprotein, with subsequent hydrolysis of intermediate-density lipoprotein





**Fig. 4.** (Colour online) Modelling overview. (1) Identify the system to model and hypothesis formation. (2) Identify pre-existing models; using the BioModels Database, a repository for peer reviewed models. (3) If no model of the system of interest exists: produce a network diagram. If a model does exist: download model and move to step 5, then step 7. (4) Establish mathematical framework. (5) Identify a suitable modelling tool; there are several available including: COPASI, which we utilised in our updated model of cholesterol metabolism<sup>(123)</sup>, CellDesigner, Mathematica and MATLAB. (6) Obtain initial concentrations of species, rate laws and kinetic data to construct the model. The online resources BRENDA and SABIO-RK provide a substantial volume of kinetic data. (7) Run simulations. (8) Validate the model. (9) Explore the hypotheses, and determine if the model accurately represents the biological system, and can be used to make predictions, or if the model needs refining. (10) Conduct further wet laboratory experiments based upon model output. (11) Code the model in the exchange format, Systems Biology Markup Language and deposit in the BioModels Database. Adapted from Mc Auley and Mooney<sup>(135)</sup>.

by hepatic lipase forming LDL, which acts to deliver cholesterol to the peripheral tissue<sup>(43)</sup>. VLDL-cholesterol, intermediate-density lipoprotein-cholesterol and LDLcholesterol can be removed from the circulation by hepatic LDLr, while LDL-cholesterol can also be absorbed independently (44,45). Reverse cholesterol transport transfers cholesterol from the tissues to the liver via HDL, reducing the risk of cholesterol accumulation and atherosclerosis (46). Cholesterol can be effluxed from the tissues by the receptors ABC-A1, and scavenger receptor class B member 1, or via receptor independent passive diffusion to nascent HDL<sup>(47–49)</sup>. The incorporated cholesterol is then esterified by lecithin-cholesterol acyltransferase<sup>(50)</sup>. Cholesterol enters the liver either directly, via the receptor scavenger receptor class B member 1, or via the enzyme cholesteryl ester transfer protein (CETP). CETP mediates the 1:1 exchange of cholesterol from HDL with TAG from VLDL and LDL<sup>(51)</sup>. Once in the liver, cholesterol can be removed from the body.

#### Cholesterol excretion

Cholesterol can be removed from the body by two mechanisms: directly via the hepatic ABCG5/G8

receptor and effluxed to the gall bladder or alternatively, cholesterol can be converted to bile acids for faecal excretion<sup>(52,53)</sup>. Approximately 98 % of bile acids are conjugated to either taurine or glycine, as conjugation increases polarity, which reduces passive transport from the intestinal lumen into enterocytes, and allows the movement of bile acids to be tightly regulated, and under receptor control; in addition to improving solubility<sup>(54)</sup>. Removal of the amino acid from conjugated bile acids, by bacterial bile salt hydrolase (BSH), decreases reabsorption efficiency, thus unconjugated bile acids make up 98 % of the 5 % of bile acids that are excreted daily (55,56). This modification is of significant interest, as the production of more readily excreted bile acids, may lead to the increased conversion of cholesterol to bile acids to replace those lost, in turn lowering serum cholesterol<sup>(57)</sup>.

## Cholesterol and healthspan

#### Intrinsic ageing

The ageing process has been associated with an increase in both total cholesterol (TC) and LDL-cholesterol. For





instance, Ericsson et al. reported an increase in TC from 4.8 mm/l in the young (aged 20–39 years), to 5.14 mm/l in the middle aged (aged 40-59 years), and to 5.44 mm/l in old aged (aged 60-80 years) healthy Scandinavian volunteers (58). Furthermore, LDL-cholesterol increased with age, from 3.37 in the young, to 3.76 in the middle aged and to 4.05 mm/l in the old aged. Additionally, VLDLcholesterol has been observed to either remain steady or increase with age, while HDL-cholesterol appears to be unaffected by the ageing process<sup>(58,59)</sup>. Abbott *et al.* also found sex influences the lipoprotein profile. For example, females exhibited higher levels of LDL-cholesterol, especially in those using oestrogen hormones, and increased HDL-cholesterol, whereas VLDL-cholesterol was greater in males<sup>(59)</sup>. The age-associated dysregulation of cholesterol metabolism, and accumulation of LDL-cholesterol, has been associated with alterations to several key mechanisms, including cholesterol absorption, LDL-cholesterol clearance, bile acid synthesis and subsequent intestinal bacterial modification (Fig. 5)<sup>(5)</sup>.

Intestinal cholesterol absorption varies greatly among individuals, with estimates ranging from 20.0 to 80.1 % (59,60). Evidence from murine models suggests cholesterol absorption increases significantly with age<sup>(7)</sup>. Rodent studies have demonstrated this is mediated by a significant increase in NPC1L1 in both the duodenum and jejunum, while ABCG5/G8 expression is suppressed<sup>(61)</sup>. It has been estimated this increase in cholesterol absorption and concurrent reduction in efflux from enterocytes, confers a 19-40 % increase in cholesterol absorption with age<sup>(61)</sup>. Moreover, it was observed that high levels of oestrogen up-regulated NPC1L1 and ABCG5/G8 mRNA expression. Oestrogen and ageing have been reported to enhance cholesterol absorption via the estrogen receptorα pathway<sup>(61)</sup>. It is important to note that these findings have not yet been observed in human individuals<sup>(60)</sup>.

Bile acid metabolism is also affected by the ageing process, most significantly, there is a reduction in bile acid synthesis. Wang<sup>(7)</sup> found ageing resulted in a significantly reduced biliary bile acid output, from 192 to 211 to 124– 157 µm/h per kg in mice. Additionally, Wang<sup>(7)</sup> demonstrated intrinsic ageing resulted in a reduction in bile acid synthesis, with a 33·3-57·1 and 41·7-56·3 % decrease in cholesterol 7α-hydroxylase (CYP7A1) activity in male and female mice, respectively, dependent on dietary and genetic factors. Similarly in human subjects, an inverse correlation between age and CYP7A1 expression has been described<sup>(62)</sup>. For instance, cholesterol  $7\alpha$ -hydroxylation rates were reduced by 50 % for individuals over 65 years, compared with individuals below 65 years of age in one Italian cohort<sup>(63)</sup>. Bertolotti et al.<sup>(63)</sup> estimated by linear regression analysis a 60 mg/d (about 150 μм/ d) decline every 10 years, in cholesterol undergoing cholesterol 7α-hydroxylation, while Einarsson et al. (64) estimated an 80 mg/d (200  $\mu$ M/d) reduction over the same time period. Bertolotti *et al.* (62) propose the age-related reduction in CYP7A1 expression could be related to the concomitant decline in hepatic nuclear factor four and co-activator CYP7A1 promoter-binding factor/liver receptor homologue-1, mediated by the decline in growth hormone and insulin-like growth factor.

Once bile acids reach the small intestine, modification by digestive microflora occurs influencing enterohepatic circulation. Many digestive bacteria produce the enzyme BSH, which deconjugates bile acids, decreasing reabsorption efficiency, and enhancing excretion. For example, Tanaka et al. (65) reported 59 and 98 % of Lactobacillus and Bifidobacteria strains, isolated from faeces are BSH positive. With age there are several changes to the gut microflora, including a decline in the number and species diversity of Lactobacillus and Bifidobacterium (8,66). Therefore, it is possible that the age-related decline of these bacterial species reduces bile acid deconjugation, and in turn reduces the conversion of cholesterol to bile acid. This may play a role in the accumulation of cholesterol with age (5).

It has also been reported, the clearance rate of LDLcholesterol is affected by the ageing process. The apo B-100 containing lipoproteins, LDL-cholesterol and VLDL-cholesterol are removed from the blood via hepatic LDLr, for elimination, either by direct efflux or conversion to bile acids. Millar et al. (6) determined the mean LDL apo B-100 residence time was 2.42 d for younger male adults (mean age 31 (sp 6) years) and 3.46 d for older male adults (mean age 61 (sp 10) years). With age, a decline in LDLr activity and/or numbers is thought to be responsible for the reduction in LDL clearance rate and increase in residence time. The reduced conversion of cholesterol by CYP7A1 may play a role in reduction of LDLr. Additionally, proprotein convertase subtilisin/kexin type 9, a proprotein convertase responsible for the degradation of LDLr has been correlated with age<sup>(67)</sup>. Interestingly, proprotein convertase subtilisin/kexin type 9 has also been correlated with BMI, TC, LDL-cholesterol and TAG<sup>(68)</sup>. Furthermore. Millar et al. (6) observed, that although VLDL apo B-100 residence time was not affected by age, the production rate of VLDL apo B-100 was correlated with age. The age-related increase in body fat and elevated plasma free fatty acids were attributed to this increase in VLDL apo B-100 production rate.

## Cholesterol metabolism and diet

Herron et al. (69) examined the effect of about 640 mg/d cholesterol feeding on men aged 18-57 years, and determined that 37.5 % of subjects behaved as hyper-responders, with an increase of >0.06 mm/l in TC, while 62.5 % behaved as hypo-responders, with an increase in TC of <0.05 mm/l. Hyper-responders exhibited a significant 23.0, 7.8 and 18.0 % increase in LDL-cholesterol and HDL-cholesterol and LDL:HDL ratio, respectively, whereas changes to LDL-cholesterol, HDL-cholesterol, TG and LDL:HDL ratio were NS for hypo-responders. Interestingly, hyper-responders exhibited elevated lecithin-cholesterol acyltransferase and CETP activity, suggesting the up-regulation of reverse cholesterol transport as a compensatory mechanism, to reduce the risk of atherosclerosis. Quintao et al. (70) propose tissue pools of cholesterol may rapidly expand in response to cholesterol feeding, even in the absence of aberrations to plasma cholesterol levels. Typically, there are two





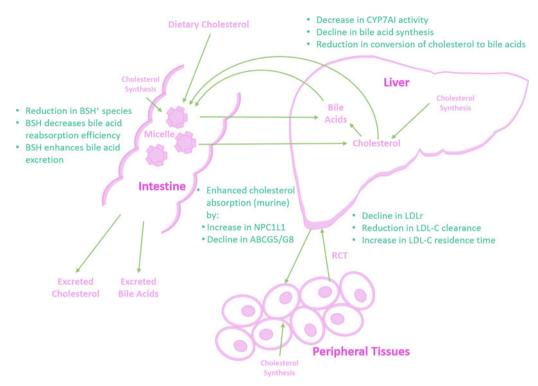


Fig. 5. (Colour online) Overview of age-related changes to cholesterol metabolism.

main mechanisms to compensate for an increase in dietary cholesterol; elevated cholesterol excretion and decreased cholesterol synthesis. It has been suggested a reduction in cholesterol intake should be considered unnecessary for individuals who have already reduced SFA intake, and increased PUFA:SFA ratio<sup>(71)</sup>. Edington et al.(71) determined a 2-fold increase or decrease in dietary cholesterol in participants who also reduced dietary fat with an increased PUFA:SFA ratio had no effect on TC or LDL-cholesterol after 8 weeks. This is due to the significant impact SFA had on serum LDL-cholesterol, by influencing several regulatory mechanisms. Firstly, it has been observed there is a reduction in LDLr; resulting in reduced LDL-cholesterol clearance and increased LDL-cholesterol<sup>(72)</sup>. Mustad et al. (72) demonstrated an 8-week reduction in SFA resulted in a 10.5 % increase in LDLr and subsequent 11.8 % decrease in LDL-cholesterol. It has been estimated for every 1 % increase in LDLr, there is a 0.74 % reduction in LDL-cholesterol. Secondly, SFA may influence cholesterol synthesis<sup>(73)</sup>. Glatz and Katan<sup>(73)</sup> determined a low PUFA:SFA ratio diet resulted in increased cholesterol synthesis, compared with a high PUFA:SFA ratio diet (1.86 v. 1.55 mm/d). Additionally, Jones et al. (74) demonstrated maize oil increased absolute cholesterol synthesis from 13.9 mg/kg per d at baseline, to 21.3 mg/kg per d. Thirdly, it has been demonstrated SFA influences the concentration of CETP. For example, Jansen et al. (75) observed that CETP concentrations were significantly elevated by 12 and 11 %, in individuals on a high SFA diet, compared with individuals on the National Cholesterol Education Program StepI diet and MUFA diet, respectively. Additionally, an elevation of SFA from 8.4 to 11 % decreased lecithin-cholesterol acyltransferase activity from 56 to 74 nm/ml per h, which may result in decreased reverse cholesterol transport and influence CVD risk<sup>(76)</sup>.

#### Cholesterol metabolism and disease

The age-related accumulation of both TC and LDLcholesterol has been associated with the pathogenesis of several diseases (77–79). In a meta-analysis of sixty-two studies, a 17.5 % reduction in relative risk for all-cause mortality for every 1 mm/l decrease in TC was reported, while each 1 mm/l decrease in LDL-cholesterol was associated with a 15.6 % relative risk reduction for all-cause mortality<sup>(4)</sup>.

CHD has been associated with significantly elevated TC, LDL-cholesterol, TAG, apo B and Lp(a) and reduced HDL-cholesterol and apo A-I in both American males and females (79). Gould *et al.* (4) reported a 24.5 and 29.5% reduction in relative risk for CHDrelated mortality and CHD event respectively for every 1 mm/l decline in TC. Additionally, each 1 mm/l decline in LDL-cholesterol was associated with a 28.0 and 26.6 % decline in relative risk for CHD-related mortality and CHD event, respectively. Sharrett et al. (79) found that each 1 mm/l increase in LDL-cholesterol was associated with a relative risk for CHD of between 1.36 and 1.44 for males and 1.19 and 1.32 for females, while a 0.4 mm/l increase in HDL-cholesterol was associated with a 0.64-0.72 and 0.64-0.76 relative risk for CHD for males and females, respectively. Interestingly, the





decreased risk for mortality with reduced TC has been observed to decline with age. For example, a meta-analysis conducted by Lewington *et al.*<sup>(80)</sup> concluded a 1 mm/l reduction in TC was associated with an about 50 % reduction in IHD mortality for 40–49 year olds, which decreased to an about 33 % reduction for 50–69 year olds and about 17 % for 70–89 year olds.

Intriguingly, several studies have described an inverse relationship between cholesterol and mortality<sup>(81)</sup>. Lv et al.<sup>(82)</sup> observed a 19 % decrease in relative risk for each 1 mm/l increase in LDL-cholesterol in Chinese ≥80 year olds. Additionally, a 40 % lower risk for mortality was associated with those with abnormally high LDL-cholesterol (≥3·37 mm/l), compared with those with a lower plasma concentration of LDL-cholesterol. Similarly, Takata et al.<sup>(83)</sup> observed a 0·8 % decrease in mortality for each 1 mg/dl (0·026 mm/l) increase in LDL-cholesterol in Japanese ≥80 year olds. In addition, each 1 mg/dl increase in TC was associated with a 0·9 % reduction in mortality.

# Targeting cholesterol metabolism as a therapeutic strategy

There are many strategies utilised in the treatment of hypercholesterolaemia. These can be used singularly or in combination, and include pharmacological intervention, changes to diet and exercise regimens, and dietary supplementation<sup>(9,11,84,85)</sup>.

# Pharmacological intervention

Lipid-lowering drugs are often utilised in the treatment of CVD (Fig. 6). Statins are often recommended for those diagnosed with CVD, or those with a high risk of developing the disease, due to their tolerability and efficacy<sup>(86)</sup>. Statins act by competitively inhibiting HMG CoA reductase, the rate-limiting enzyme of cholesterol synthesis in order to lower serum cholesterol, thus reducing CVD risk<sup>(87)</sup>. Simivastatin reduced LDLcholesterol by 33 % after 12 weeks, in patients with a history of myocardial infarction, while atorvastatin exhibited a 49 % reduction (88). A meta-analysis of four trials demonstrated standard-dose statin therapy reduced LDL-cholesterol by 22 %, to a mean of 2.59 mm/l (101 mg/dl), while high-dose statin therapy reduced LDLcholesterol by 42 %, to a mean of 1.92 mm/l (75 mg/ dl)<sup>(89)</sup>. The standard-dose therapy was associated with a coronary death or myocardial event rate of 9.4 %, whereas high-dose therapy was associated with an 8.0 % event rate. Significantly, overall statin therapy resulted in a 16 % odds reduction in coronary death or myocardial infarction<sup>(89)</sup>. Ridker et al.<sup>(90)</sup> observed that patients whose statin therapy reduced LDL-cholesterol to <70 mg/dl (<1.81 mm/dl), had a reduced rate of recurrent myocardial infarction or death from coronary events (2.7 events per 100 person-years), than individuals whose statin therapy did not reach this goal (4.0 events per 100 person-years). However, it has also been demonstrated non-statin therapy (diet, bile acid sequesterants

and ileal bypass surgery) reduced CHD risk equally to that of statins, with each 1% reduction in LDL-cholesterol corresponding to a 1% decrease in CHD risk<sup>(10)</sup>. Statins can also be used in combination with drugs such as Ezetimbibe, which targets NPC1L1 to lower cholesterol absorption. Cannon *et al.*<sup>(84)</sup> observed that combination therapy lowered LDL-cholesterol by a further 24% than statin therapy alone. Combination therapy also significantly reduced myocardial infarction and ischaemic stroke risk.

#### Dietary intervention

A reduced intake of SFA and an increase in MUFA and PUFA may be useful in maintaining a healthy lipid profile, and reducing CHD risk<sup>(11)</sup>. An 8-week reduction in SFA was observed to reduce TC and LDL-cholesterol by 9·3 and 11·8 %, respectively, in healthy males and females aged between 20 and 65 years<sup>(72)</sup>. Additionally, Berry *et al.*<sup>(91)</sup> reported a 12-week MUFA or PUFA diet reduced TC by about 10 and 16 %, respectively, while reducing LDL-cholesterol by 14 and 21 %. A meta-analysis of 13 614 participants participating in diets where SFA was replaced with PUFA, found that on average there was a 0·76 mm/l (29 mg/dl) reduction in TC, and that each 1 mm/l reduction in TC was associated with 24 % reduced risk for a CHD event. Moreover, each additional year of diet was related to a further 9·2 % risk reduction<sup>(11)</sup>.

Additionally, the use of phytosterols is recommended as a therapeutic strategy for the treatment of hypercholesterolaemia. It is estimated a 2 g/d dose of phytosterols lowers LDL-cholesterol by  $10\,\%^{(92)}$ . A meta-analysis of 124 studies concluded 0.6–3.3 g/d phytosterols reduced LDL-cholesterol by 6–12 %, with stanols and sterols showing similar LDL-cholesterol lowering efficacies (85). The effect of phytosterols diminishes above about 2–3 g/d (92), consequently 2 g/d is generally accepted as the daily amount required to help treat hypercholesterolaemia (93).

Furthermore, meta-analysis of sixty-seven trials revealed that soluble fibre could lower TC and LDLcholesterol by 0.045 and 0.057 mm/l per g soluble fibre, respectively<sup>(94)</sup>. There are a variety of mechanisms that dietary fibre could influence; for example, bile acid micelle formulation, fat excretion, intestinal motility, SCFA production and absorption of macronutrients (95-97). Interestingly, diet may not only impact cholesterol metabolism directly, and may also interact with the gut microbiome<sup>(98)</sup>. For example, David et al.<sup>(99)</sup> describe that short-term changes to diet could significantly influence both the microbial community structure and bacterial gene expression. For instance, a 5 d animal-based-diet significantly altered the abundance of twenty-two clusters, elevated faecal bile acid concentration, and increased BSH expression<sup>(99)</sup>. Everard et al.<sup>(100)</sup> describe that high-fat feeding significantly affected twenty genera. This may have been influenced by a decline in the antimicrobial peptide regenerating islet-derived protein three gamma. Interestingly, prebiotic treatment increased regenerating islet-derived protein three gamma about



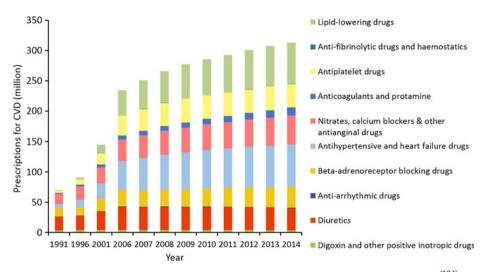


Fig. 6. (Colour online) UK prescription drugs for CVD. Data from British Heart Foundation (134).

6-fold within the colon. Targeting the digestive microbiota with pre- and probiotics is a potential avenue for the treatment of diseases including hypercholesterolaemia.

# Targeting the gut microbiome

There is emerging evidence indicating the important role for the gut microbiome in regulating many biological systems, including cholesterol metabolism<sup>(101,102)</sup>. Ageing is associated with the alteration of the gut microbiome. These changes differ between individuals and populations, as diet, lifestyle, host health and antibiotic use likely play a role in composition<sup>(103,104)</sup>. However, it has been observed Lactobacillus and Bifidobacterium species diversity and total count decline with age<sup>(8,56)</sup>. Supplementation with probiotics may have the ability to partially ameliorate changes associated with ageing, such as immunosenescence, neurodegeneration, carcinogenic transformation and hypercholesterolaemia (105,106).

With regard to hypercholesterolaemia treatment, supplementation with probiotics has been associated with a decline in TC and LDL-cholesterol<sup>(9)</sup>. A meta-analysis of probiotic administration studies concluded probiotic administration resulted in a mean 6.40 mg/dl (0.17 mm/l) reduction in TC and 4.90 mg/dl (0.13 mm/l) reduction in LDL-cholesterol. Additionally, probiotics were observed to have beneficial effects on TC and LDL-cholesterol for individuals with high, borderline high and normal cholesterol levels<sup>(9)</sup>. Probiotics have also been an effective treatment strategy for individuals with underlying conditions. For example, Bernini et al. (107) observed ingestion of 80 ml probiotic milk containing  $3.4 \times 10^8$  colony-forming units/ml Bifidobacterium lactis HN019 for 45 d, significantly lowered TC from 209 to 194 mg/dl (5.40 and 5.02 mm/l), and LDL from 128.5 to 111 mg/dl (3.32– 2.87 mm/l) in patients with metabolic syndrome, in addition to lowering BMI. Moreover, the proinflammatory cytokines TNF-α and IL-6 were significantly reduced with probiotic treatment.

There are several proposed mechanisms to explain the lipid-lowering ability of probiotics<sup>(108)</sup>. For example, some probiotics produce the bile acid deconjugating enzyme BSH, which may increase bile acid excretion, and up-regulate the conversion of cholesterol to bile acids to replace those lost (109). It has also been observed that probiotics increase SCFA, which inhibit the hepatic rate-limiting enzyme in cholesterol synthesis, HMG CoA reductase<sup>(108)</sup>. Furthermore, there may be a reduction in cholesterol absorption due to the assimilation and possible incorporation of cholesterol into the bacterial cell membrane<sup>(110)</sup>. Additionally, some bacteria may act by destabilising cholesterol micelles resulting in the coprecipitation of cholesterol with deconjugated bile acids(110)

# Applying systems biology to our understanding of cholesterol metabolism and ageing

It is apparent from our discussion of cholesterol metabolism, that it is an exceptionally complex system with a multitude of interacting mechanisms<sup>(111,112)</sup>. Many of these mechanisms interact via positive and negative feedback regulators, the dynamics of which is not trivial. Our understanding of this complexity is further compounded by ageing which alters the behaviour of these regulatory mechanisms, thus modifying the overall dynamics of whole-body cholesterol metabolism. Moreover, changes to extrinsic factors such as diet exert a significant influence on the overall behaviour of this system. Thus, it is imperative if we are to gain a more in-depth appreciation of cholesterol metabolism and its interaction with ageing, that we investigate this system in a more integrated manner. The systems biology paradigm contrasts with the more traditional reductionist approach, commonly used in nutrition research, and offers a more integrated way to study this multifaceted system<sup>(14,113,114)</sup>. A fundamental element of this paradigm shift is the close coupling of computational modelling with experimental





work<sup>(115–118)</sup>. Our group has used computational systems biology to investigate cholesterol metabolism and ageing<sup>(119)</sup>. Firstly, we constructed a whole-body model of cholesterol metabolism<sup>(120,121)</sup>. The model is defined by the key components that characterise whole-body cholesterol balance, namely cholesterol ingestion, excretion and synthesis together with LDLr turnover and reverse cholesterol transport. Using in silico experiments we explored the impact of ageing on these fundamental elements of cholesterol metabolism. We investigated the influence of ageing on cholesterol absorption and it was found that for every 10 % increment in the rate of cholesterol absorption, there was a concomitant increase of 12.5 mg/dl in LDL-cholesterol. The model also revealed that increasing cholesterol absorption from 50 to 80 % by age 65 years, resulted in a 34 mg/dl increase in plasma LDL-cholesterol. In addition, the model found that reducing the number of hepatic LDLr had a significant impact on the system. An increase of 116 mg/dl in LDL-cholesterol was observed by 65 years in response to a reduction of 50 % in their numbers; thus emphasising LDLr maintenance as a key component in maintaining cholesterol metabolism during ageing. This model is coded in the Systems Biology Markup Language and archived in the BioModels Database, a repository for models encased in Systems Biology Markup Language exchange framework (http://www.ebi.ac.uk/biomodelsmain/BIOMD000000434). This means the model is straightforward to update and adapt, a feature exploited by other groups working in this area<sup>(122)</sup>. We recently implemented significant updates to the model described earlier for the following reasons<sup>(123)</sup>. The model was lacking several key mechanisms which recent experimental evidence has stressed are central to the regulation of cholesterol metabolism. Briefly, these processes included plasma membrane receptors, and in vivo and intestinal microfloral enzymes. Therefore, it was deemed cogent that these ninety-six additional mechanisms were incorporated into the original model<sup>(5)</sup>. We found that our updated model behaved as a hypo-responder to excessive cholesterol feeding. Moreover, the model was utilised to investigate the effects of ageing coupled with three different CETP genotypes. Ageing in the presence of a genotype conferring low CETP activity resulted in a 0.6 % decrease in LDL-cholesterol after 1000 h. In comparison, ageing with a genotype indicative of high CETP activity, provoked a 1.6 % increase in LDL-cholesterol levels (Fig. 7). Thus, our new model consolidates experimental findings, which emphasise the significance of CETP genotypes in healthspan<sup>(123)</sup>.

# Complementing computational models with simulated digestive tracts

To complement our *in silico* studies of cholesterol metabolism, we aim to develop a simulator of the human digestive tract. The digestive track simulator will be used to refine and inform our computational model. One example is the simulator of the human intestinal microbial ecosystem reactor, which represents the length

of the human digestive system, with several closed reactors used to represent the differing conditions (microbial ecosystem, pH, enzymes, etc.) of each section of the digestive tract<sup>(124)</sup>. The flexible nature of the simulator of the human intestinal microbial ecosystem reactor means that it can be easily adapted. This may include additional reactors to more accurately represent the human digestive tract<sup>(125,126)</sup>. Simulated digestive tracts, such as the simulator of the human intestinal microbial ecosystem reactor provide a suitable test environment to supplement experimental data on potential therapeutic strategies. For example, De Smet et al. (127) determined administration of Lactobacillus to pigs for 4 weeks decreased TC and LDL-cholesterol by 15 and 24 %. respectively. Additionally, TC and LDL-cholesterol reduced a further 18 and 34 % 3 weeks after treatment. Follow-up experimentation using the simulator of the human intestinal microbial ecosystem reactor was able to provide details of the intestinal bacterial population and overall metabolic activity associated with this treatment<sup>(125)</sup>. Recently, a continuous gut adhesion model was developed using mucin-alginate beads to immobilise bacteria, which may more effectively represent the gut lining and intestinal lumen. This system was also used to examine bacterial colonisation following probiotic treatment(128). The TNO gastrointestinal model can be used to examine compound absorption<sup>(129)</sup>. This model simulates ingestion of food and water, digestive enzyme and bile salt release, peristalsis, body temperature, and transit through the stomach and small intestine (TNO gastrointestinal model-1) and large intestine (TNO gastrointestinal model-2) under computer control. The model can be altered to represent a different species, such as a human or pig, or can represent varying age, such as infant, adult and older people (129). This is particularly significant for the augmentation and refinement of our computational model. It is hoped that utilising both these approaches in tandem will help to significantly deepen our understanding of cholesterol metabolism and its relationship with ageing.

# **Conclusions**

We have outlined how the age-related dysregulation of cholesterol metabolism is influenced by many factors. Specifically, (i) an increase in LDL-cholesterol residence time due to reduction in LDLr and concomitant decline in LDL-cholesterol clearance;, (ii) a decline in CYP7AI activity which reduces bile acid synthesis, one pathway for cholesterol removal; (iii) a reduction in digestive bacteria which modify bile acids that reduce reabsorption and promotes excretion; (iv) a potential increase in cholesterol absorption due to an increase in NPC1L1 and decrease in ABCG5/G8, as observed in murine models. Cholesterol metabolism is also influenced by diet and gut microbial dynamics, therefore studying the mechanisms underpinning these complex interactions is challenging. Due to its ability to handle this complexity computational modelling has been used to study the intricacies associated with ageing and cholesterol



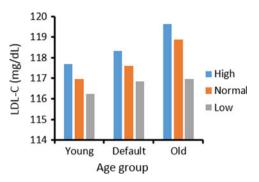


Fig. 7. Model simulation of ageing in the presence of cholesteryl ester transfer protein genotypes. Taken from Morgan et al. (123).

metabolism. Computational modelling is a key component of the systems biology paradigm which nutrition research is beginning to adopt. It is possible nutrition research will benefit significantly from this approach in the coming years. For instance, computational modelling will be needed to deepen our understanding of the role the gut microbiome has to play in host health (130). In tandem with computational modelling, experimental approaches are a key part of systems biology. An important experimental method which dovetails with computational modelling is the use of artificial digestive tracts. These can be used to simulate the activities of the digestive tract under a wide variety of conditions, something which is difficult to achieve in vivo<sup>(131)</sup>. Thus, utilising these methodologies as part of a systems biology framework, provides a means of investigating the dynamics of the gut microbiome together with its interaction with cholesterol metabolism and ageing. In order for this approach to be successful it is imperative for nutritionists to work closely with scientists from the systems biology community. This novel way of conducting nutrition research will in turn facilitate the discovery of nutrientbased strategies to treat hypercholesterolaemia, which so often accompanies the ageing process.

#### Acknowledgement

A. E. M. was funded by a University of Chester PhD scholarship.

## **Financial Support**

This research received no specific grant from any funding agency in the public, commercial or not-for-profit sectors.

# **Conflict of Interest**

None.

# **Authorship**

A. E. M. and M. T. M. A. drafted the manuscript and conceived the idea. K. M. M. provided advice on the nutrition focused sections of the manuscript. N. A. P. provided advice on the microbiology components.

# References

- 1. OFNS (2013) Mortality Assumptions, 2012-based National Population Projections.
- 2. WHO (2014) Aging and life course: facts about aging. http://www.who.int/ageing/about/facts/en/
- 3. Prince MJ, Wu F, Guo Y et al. (2015) The burden of disease in older people and implications for health policy and practice. Lancet 385, 549-562.
- 4. Gould AL, Davies GM, Alemao E et al. (2007) Cholesterol reduction yields clinical benefits: meta-analysis including recent trials. Clin Ther 29, 778-794.
- 5. Morgan AE, Mooney KM, Wilkinson SJ et al. (2016) Cholesterol metabolism: a review of how ageing disrupts the biological mechanisms responsible for its regulation. Ageing Res Rev 27, 108-124.
- 6. Millar JS, Lichtenstein AH, Cuchel M et al. (1995) Impact of age on the metabolism of VLDL, IDL, and LDL apolipoprotein B-100 in men. J Lipid Res 36, 1155-1167.
- 7. Wang DQ-H (2002) Aging per se is an independent risk factor for cholesterol gallstone formation in gallstone susceptible mice. J Lipid Res 43, 1950-1959.
- 8. Hopkins MJ & Macfarlane GT (2002) Changes in predominant bacterial populations in human faeces with age and with Clostridium difficile infection. J Med Microbiol 51, 448-454.
- 9. Guo Z, Liu XM, Zhang QX et al. (2011) Influence of consumption of probiotics on the plasma lipid profile: a metaanalysis of randomised controlled trials. Nutr Metab Cardiovasc Dis 21, 844-850.
- 10. Robinson JG, Smith B, Maheshwari N et al. (2005) Pleiotropic effects of statins: benefit beyond cholesterol reduction?: a meta-regression analysis. J Amer Coll Cardiol 46, 1855-1862.
- 11. Mozaffarian D, Micha R & Wallace S (2010) Effects on coronary heart disease of increasing polyunsaturated fat in place of saturated fat: a systematic review and metaanalysis of randomized controlled trials. PLoS Med 7,
- 12. Lin DS & Connor WE (1980) The long term effects of dietary cholesterol upon the plasma lipids, lipoproteins, cholesterol absorption, and the sterol balance in man: the demonstration of feedback inhibition of cholesterol biosynthesis and increased bile acid excretion. J Lipid Res 21, 1042-1052.
- 13. Beserra BT, Fernandes R, do Rosario VA et al. (2015) A systematic review and meta-analysis of the prebiotics and synbiotics effects on glycaemia, insulin concentrations and lipid parameters in adult patients with overweight or obesity. Clin Nutr 34, 845-858.
- 14. Mc Auley MT, Proctor CJ, Corfe BM et al. (2013) Nutrition research and the impact of computational systems biology. J Comput Sci Syst Biol 6, 271–285.
- 15. Auffray C & Hood L (2012) Editorial: systems biology and personalized medicine – the future is now. Biotechnol J 7, 938-939.





- Kitano H (2002) Computational systems biology. *Nature* 420, 206–210.
- Patti ME & Kahn BB (2004) Nutrient sensor links obesity with diabetes risk. Nat Med 10, 1049–1050.
- 18. de Luca C & Olefsky JM (2008) Inflammation and insulin resistance. *FEBS Lett* **582**, 97–105.
- Gianchandani EP, Brautigan DL & Papin JA (2006) Systems analyses characterize integrated functions of biochemical networks. *Trends Biochem Sci* 31, 284–291
- Lamb RF (2012) Negative feedback loops: nutrient starvation employs a new tr(IKK) to inhibit PI3K. *Mol Cell* 45, 705–706.
- Huang Y, He S, Li JZ et al. (2010) A feed-forward loop amplifies nutritional regulation of PNPLA3. Proc Natl Acad Sci USA 107, 7892–7897.
- Pappu AS, Steiner RD, Connor SL et al. (2002) Feedback inhibition of the cholesterol biosynthetic pathway in patients with Smith-Lemli-Opitz syndrome as demonstrated by urinary mevalonate excretion. J Lipid Res 43, 1661–1669.
- 23. Henderson L, Gregory J, Irving K et al. (2003) The National Diet & Nutrition Survey: Adults Aged 19 to 64 Years. London: Office for National Statistics.
- Iqbal J & Hussain MM (2009) Intestinal lipid absorption. Amer J Physiol Endocrinol Metab 296, E1183–E1194.
- Betters JL & Yu L (2010) NPC1L1 and cholesterol transport. FEBS Lett 584, 2740–2747.
- 26. Ikeda I, Matsuoka R, Hamada T *et al.* (2002) Cholesterol esterase accelerates intestinal cholesterol absorption. *Biochim Biophys Acta Gen Subj* **1571**, 34–44.
- Davis HR Jr, Zhu LJ, Hoos LM et al. (2004) Niemann–Pick C1 like 1 (NPC1L1) is the intestinal phytosterol and cholesterol transporter and a key modulator of whole-body cholesterol homeostasis. J Biol Chem 279, 33586–33592.
- 28. Yu X-H, Qian K, Jiang N *et al.* (2014) ABCG5/ABCG8 in cholesterol excretion and atherosclerosis. *Clin Chim Acta* **428**, 82–88.
- Chang T-Y, Li B-L, Chang CCY et al. (2009) Acyl-coenzyme A: cholesterol acyltransferases. Am J Physiol
   Endocrinol Metab 297, E1–E9.
- 30. Atzel A & Wetterau JR (1993) Mechanism of microsomal triglyceride transfer protein catalyzed lipid transport. *Biochemistry* **32**, 10444–10450.
- Van Dyck F, Braem CV, Chen Z et al. (2007) Loss of the PlagL2 transcription factor affects lacteal uptake of chylomicrons. Cell Metab 6, 406–413.
- Kersten S (2014) Physiological regulation of lipoprotein lipase. *Biochim Biophys Acta* 1841, 919–933.
- Olivecrona G & Beisiegel U (1997) Lipid binding of apolipoprotein CII is required for stimulation of lipoprotein lipase activity against apolipoprotein CII-deficient chylomicrons. Arterioscler Thromb Vasc Biol 17, 1545–1549.
- 34. Cooper AD (1997) Hepatic uptake of chylomicron remnants. *J Lipid Res* 38, 2173–2192.
- 35. Bloch K (1965) The biological synthesis of cholesterol. *Science* **150**, 19–28.
- Renfurm LN, Bandsma RHJ, Verkade HJ et al. (2004) Cholesterol synthesis and de novo Lipogenesis in premature infants determined by mass isotopomer distribution analysis. Pediatr Res 56, 602–607.
- Jones PJ & Schoeller DA (1990) Evidence for diurnal periodicity in human cholesterol synthesis. *J Lipid Res* 31, 667–673.

- 38. Parker TS, McNamara DJ, Brown C *et al.* (1982) Mevalonic acid in human plasma: relationship of concentration and circadian rhythm to cholesterol synthesis rates in man. *Proc Natl Acad Sci USA* **79**, 3037–3041.
- 39. Sirtori CR (2014) The pharmacology of statins. *Pharmacol Res* **88**, 3–11.
- 40. Hoshino T, Chiba A & Abe N (2012) Lanosterol biosynthesis: the critical role of the methyl-29 group of 2,3-oxidosqualene for the correct folding of this substrate and for the construction of the five-membered D ring. *Chemistry* 18, 13108–13116.
- Luu W, Hart-Smith G, Sharpe LJ et al. (2015) The terminal enzymes of cholesterol synthesis, DHCR24 and DHCR7, interact physically and functionally. J Lipid Res 56, 888–897.
- 42. Risley JM (2002) Cholesterol biosynthesis: lanosterol to cholesterol. *J Chem Educ* **79**, 377.
- 43. Havel RJ (1984) The formation of LDL: mechanisms and regulation. *J Lipid Res* **25**, 1570–1576.
- 44. Spady DK, Turley SD & Dietschy JM (1985) Receptorindependent low density lipoprotein transport in the rat *in vivo*. Quantitation, characterization, and metabolic consequences. *J Clin Invest* **76**, 1113–1122.
- Veniant MM, Zlot CH, Walzem RL et al. (1998) Lipoprotein clearance mechanisms in LDL receptordeficient 'Apo-B48-only' and 'Apo-B100-only' mice. J Clin Invest 102, 1559–1568.
- 46. Shen L, Peng H, Peng R *et al.* (2015) Inhibition of soluble epoxide hydrolase in mice promotes reverse cholesterol transport and regression of atherosclerosis. *Atherosclerosis* **239**, 557–565.
- 47. He Y, Zhang L, Li Z *et al.* (2015) RIP140 triggers foamcell formation by repressing ABCA1/G1 expression and cholesterol efflux via liver X receptor. *FEBS Lett* **589**, 455–460.
- Ji A, Meyer JM, Cai L et al. (2011) Scavenger receptor SR-BI in macrophage lipid metabolism. Atherosclerosis 217, 106–112.
- Gillotte KL, Davidson WS, Lund-Katz S et al. (1998) Removal of cellular cholesterol by pre-beta-HDL involves plasma membrane microsolubilization. J Lipid Res 39, 1918–1928.
- Sorci-Thomas M, Babiak J & Rudel LL (1990) Lecithincholesterol acyltransferase (LCAT) catalyzes transacylation of intact cholesteryl esters. Evidence for the partial reversal of the forward LCAT reaction. *J Biol Chem* 265, 2665–2670.
- 51. Zhang M, Charles R, Tong H *et al.* (2015) HDL surface lipids mediate CETP binding as revealed by electron microscopy and molecular dynamics simulation. *Sci Rep* 5, 8741.
- 52. Brown MS & Goldstein JL (1984) How LDL receptors influence cholesterol and atherosclerosis. *Sci Am* **251**, 58–66
- 53. Repa JJ, Berge KE, Pomajzl C *et al.* (2002) Regulation of ATP-binding cassette sterol transporters ABCG5 and ABCG8 by the liver X receptors α and β. *J Biol Chem* **277**, 18793–18800.
- Aldini R, Montagnani M, Roda A et al. (1996) Intestinal absorption of bile acids in the rabbit: different transport rates in jejunum and ileum. Gastroenterology 110, 459– 468.
- Batta AK, Salen G, Rapole KR et al. (1999) Highly simplified method for gas-liquid chromatographic quantitation of bile acids and sterols in human stool. J Lipid Res 40, 1148–1154.

- 56. Gérard P (2014) Metabolism of cholesterol and bile acids by the gut microbiota. Pathogens 3, 14-24.
- 57. Oner O, Aslim B & Aydas SB (2014) Mechanisms of cholesterol-lowering effects of lactobacilli and bifidobacteria strains as potential probiotics with their bsh gene analysis. J Mol Microbiol Biotechnol 24, 12-18.
- 58. Ericsson S, Eriksson M, Vitols S et al. (1991) Influence of age on the metabolism of plasma low density lipoproteins in healthy males. J Clin Investig 87, 591-596.
- 59. Abbott RD, Garrison RJ, Wilson PW et al. (1983) Joint distribution of lipoprotein cholesterol classes. The Framingham study. Arteriosclerosis 3, 260–272.
- 60. Bosner MS, Lange LG, Stenson WF et al. (1999) Percent cholesterol absorption in normal women and men quantified with dual stable isotopic tracers and negative ion mass spectrometry. J Lipid Res 40, 302-308.
- 61. Duan LP, Wang HH, Ohashi A et al. (2006) Role of intestinal sterol transporters Abcg5, Abcg8, and Npc111 in cholesterol absorption in mice: gender and age effects. Am J Physiol Gastrointest Liver Physiol 290, G269–276.
- 62. Bertolotti M, Gabbi C, Anzivino C et al. (2007) Agerelated changes in bile acid synthesis and hepatic nuclear receptor expression. Eur J Clin Invest 37, 501-508.
- 63. Bertolotti M, Abate N, Bertolotti S et al. (1993) Effect of aging on cholesterol 7 alpha-hydroxylation in humans. J Lipid Res 34, 1001–1007.
- 64. Einarsson K, Nilsell K, Leijd B et al. (1985) Influence of age on secretion of cholesterol and synthesis of bile acids by the liver. N Engl J Med 313, 277-282.
- 65. Tanaka H, Doesburg K, Iwasaki T et al. (1999) Screening of lactic acid bacteria for bile salt hydrolase activity. J Dairy Sci 82, 2530-2535.
- 66. Woodmansey EJ, McMurdo MET, Macfarlane GT et al. (2004) Comparison of compositions and metabolic activities of fecal microbiotas in young adults and in antibiotic-treated and non-antibiotic-treated elderly subjects. Appl Environ Microbiol 70, 6113–6122.
- 67. Lagace TA, Curtis DE, Garuti R et al. (2006) Secreted PCSK9 decreases the number of LDL receptors in hepatocytes and in livers of parabiotic mice. J Clin Investig 116, 2995-3005.
- 68. Cui Q, Ju X, Yang T et al. (2010) Serum PCSK9 is associated with multiple metabolic factors in a large Han Chinese population. Atherosclerosis 213, 632–636.
- 69. Herron KL, Vega-Lopez S, Conde K et al. (2003) Men classified as hypo- or hyperresponders to dietary cholesterol feeding exhibit differences in lipoprotein metabolism. J Nutr 133, 1036-1042.
- 70. Quintao E, Grundy SM & Ahrens EH Jr (1971) Effects of dietary cholesterol on the regulation of total body cholesterol in man. J Lipid Res 12, 233–247.
- 71. Edington J, Geekie M, Carter R et al. (1987) Effect of dietary cholesterol on plasma cholesterol concentration in subjects following reduced fat, high fibre diet. Br Med J (Clin Res Ed) 294, 333-336.
- 72. Mustad VA, Etherton TD, Cooper AD et al. (1997) Reducing saturated fat intake is associated with increased levels of LDL receptors on mononuclear cells in healthy men and women. J Lipid Res 38, 459-468.
- 73. Glatz JFC & Katan MB (1993) Dietary saturated fatty acids increase cholesterol synthesis and fecal steroid excretion in healthy men and women. Eur J Clin Invest 23, 648–655.
- 74. Jones PJ, Lichtenstein AH, Schaefer EJ et al. (1994) Effect of dietary fat selection on plasma cholesterol synthesis in older, moderately hypercholesterolemic humans. Arterioscler Thromb Vasc Biol 14, 542-548.

- 75. Jansen S, López-Miranda J, Castro P et al. (2000) Low-fat and high-monounsaturated fatty acid diets decrease plasma cholesterol ester transfer protein concentrations in young, healthy, normolipemic men, Am J Clin Nutr **72**, 36–41.
- 76. Berard AM, Dabadie H, Palos-Pinto A et al. (2004) Reduction of dietary saturated fatty acids correlates with increased plasma lecithin cholesterol acyltransferase activity in humans. Eur J Clin Nutr 58, 881-887.
- 77. Sjogren M & Blennow K (2005) The link between cholesterol and Alzheimer's disease. World J Biol Psychiatry 6,
- 78. Kuzu OF, Noory MA & Robertson GP (2016) The role of cholesterol in cancer. Cancer Res 76, 2063-2070.
- 79. Sharrett AR, Ballantyne CM, Coady SA et al. (2001) Coronary heart disease prediction from lipoprotein cholesterol levels, triglycerides, lipoprotein(a), apolipoproteins A-I and B, and HDL density subfractions: the atherosclerosis risk in communities (ARIC) study. Circulation 104, 1108-1113
- 80. Lewington S, Whitlock G, Clarke R et al. (2007) Blood cholesterol and vascular mortality by age, sex, and blood pressure: a meta-analysis of individual data from 61 prospective studies with 55 000 vascular deaths. Lancet 370, 1829-1839.
- 81. Ravnskov U, Diamond DM, Hama R et al. (2016) Lack of an association or an inverse association between lowdensity-lipoprotein cholesterol and mortality in the elderly: a systematic review. BMJ Open http://bmjopen. bmj.com/content/6/6/e010401.full
- 82. Lv Y-B, Yin Z-X, Chei C-L et al. (2015) Low-density lipoprotein cholesterol was inversely associated with 3-year all-cause mortality among Chinese oldest old: data from the Chinese longitudinal healthy longevity survev. Atherosclerosis 239, 137-142.
- 83. Takata Y, Ansai T, Soh I et al. (2014) Serum total cholesterol concentration and 10-year mortality in an 85-yearold population. Clin Intervent Aging 9, 293-300.
- Cannon CP, Blazing MA, Giugliano RP et al. (2015) Ezetimibe added to statin therapy after acute coronary syndromes. N Engl J Med 372, 2387-2397.
- 85. Ras RT, Geleijnse JM & Trautwein EA (2014) LDL-cholesterol-lowering effect of plant sterols and stanols across different dose ranges: a meta-analysis of randomised controlled studies. Br J Nutr 112, 214-219.
- 86. Weng TC, Yang YHK, Lin SJ et al. (2010) A systematic review and meta-analysis on the therapeutic equivalence of statins. J Clin Pharm Ther 35, 139-151.
- 87. Law MR, Wald NJ & Rudnicka AR (2003) Quantifying effect of statins on low density lipoprotein cholesterol, ischaemic heart disease, and stroke: systematic review and meta-analysis. BMJ 326, 1423.
- 88. Pedersen TR, Faergeman O, Kastelein JP et al. (2005) High-dose atorvastatin vs usual-dose simvastatin for secondary prevention after myocardial infarction: the ideal study: a randomized controlled trial. JAMA 294, 2437-2445.
- 89. Cannon CP, Steinberg BA, Murphy SA et al. (2006) Meta-analysis of cardiovascular outcomes trials comparing intensive versus moderate statin therapy. J Am Coll Cardiol 48, 438-445.
- 90. Ridker PM, Cannon CP, Morrow D et al. (2005) C-reactive protein levels and outcomes after statin therapy. N Engl J Med 352, 20–28.
- 91. Berry EM, Eisenberg S, Haratz D et al. (1991) Effects of diets rich in monounsaturated fatty acids on plasma



- lipoproteins -- the Jerusalem Nutrition Study: high MUFAs vs high PUFAs. Am J Clin Nutr 53, 899-907.
- 92. Katan MB, Grundy SM, Jones P et al. (2003) Efficacy and safety of plant stanols and sterols in the management of blood cholesterol levels. Mayo Clin Proc 78, 965-978.
- 93. IAS (2013) An Int Atherosclerosis Society Position Paper: Global Recommendations for the Management of Dyslipidemia. Full Report.
- 94. Brown L, Rosner B, Willett WW et al. (1999) Cholesterollowering effects of dietary fiber: a meta-analysis. Am J Clin Nutr 69, 30-42.
- 95. Weickert MO & Pfeiffer AFH (2008) Metabolic effects of dietary fiber consumption and prevention of diabetes. J Nutr 138, 439-442.
- 96. Kristensen M, Jensen MG, Aarestrup J et al. (2012) Flaxseed dietary fibers lower cholesterol and increase fecal fat excretion, but magnitude of effect depend on food type. Nutr Metab 9, 8.
- 97. Slavin J (2013) Fiber and prebiotics: mechanisms and health benefits. Nutrients 5, 1417.
- 98. Gibson GR, Beatty ER, Wang X et al. (1995) Selective stimulation of bifidobacteria in the human colon by oligofructose and inulin. Gastroenterology 108, 975-982.
- 99. David LA, Maurice CF, Carmody RN et al. (2014) Diet rapidly and reproducibly alters the human gut microbiome. Nature 505, 559-563.
- 100. Everard A, Lazarevic V, Gaia N et al. (2014) Microbiome of prebiotic-treated mice reveals novel targets involved in host response during obesity. ISME J 8, 2116–2130.
- 101. Cénit MC, Matzaraki V, Tigchelaar EF et al. (2014) Rapidly expanding knowledge on the role of the gut microbiome in health and disease. Biochim Biophys Acta Mol Basis Dis 1842, 1981–1992.
- 102. Kinross JM, Darzi AW & Nicholson JK (2011) Gut microbiome-host interactions in health and disease. Genome Med 3, 1-12.
- 103. Claesson MJ, Jeffery IB, Conde S et al. (2012) Gut microbiota composition correlates with diet and health in the elderly. Nature 488, 178-184.
- 104. O'Sullivan O, Coakley M, Lakshminarayanan B et al. (2013) Alterations in intestinal microbiota of elderly Irish subjects post-antibiotic therapy. J Antimicrob Chemother 68, 214–221.
- 105. Patel PJ, Singh SK, Panaich S et al. (2014) The aging gut and the role of prebiotics, probiotics, and synbiotics: a review. J Clin Gerontol Geriatr 5, 3-6.
- 106. Moroti C, Souza Magri LF, de Rezende Costa M et al. (2012) Effect of the consumption of a new symbiotic shake on glycemia and cholesterol levels in elderly people with type 2 diabetes mellitus. Lipids Health Dis 11, 1-8.
- 107. Bernini LJ, Simão ANC, Alfieri DF et al. (2016) Beneficial effects of Bifidobacterium lactis on lipid profile and cytokines in patients with metabolic syndrome: a randomized trial. Effects of probiotics on metabolic syndrome. Nutrition 32, 716-719.
- 108. Ishimwe N, Daliri EB, Lee BH et al. (2015) The perspective on cholesterol-lowering mechanisms of probiotics. Mol Nutr Food Res 59, 94-105.
- 109. Tsai C-C, Lin P-P, Hsieh Y-M et al. (2014) Cholesterollowering potentials of lactic acid bacteria based on bile-salt hydrolase activity and effect of potent strains on cholesterol metabolism in vitro and in vivo. Sci World J 2014, 10.
- 110. Brashears MM, Gilliland SE & Buck LM (1998) Bile salt deconjugation and cholesterol removal from media by Lactobacillus casei. J Dairy Sci 81, 2103-2110.

- 111. Mc Auley MT & Mooney KM (2014) Lipid metabolism and hormonal interactions: impact on cardiovascular disease and healthy aging. Expert Rev Endocrinol Metab 9,
- 112. Mooney KM & Mc Auley MT (2015) Cardiovascular disease and healthy ageing. J Integr Cardiol 1, 76-78.
- 113. Mc Auley MT, Choi H, Mooney K et al. (2015) systems biology and synthetic biology: a new epoch for toxicology research. Adv Toxicol 2015, 14.
- 114. Mooney KM, Morgan AE & Mc Auley MT (2016) Aging and computational systems biology. Wiley Interdiscip Rev: Syst Biol Med 8, 123-139.
- 115. Enrique Salcedo-Sora J & Mc Auley MT (2016) A mathematical model of microbial folate biosynthesis and utilisation: implications for antifolate development. Mol Biosyst 12, 923-933.
- 116. Kilner J, Corfe BM, McAuley MT et al. (2016) A deterministic oscillatory model of microtubule growth and shrinkage for differential actions of short chain fatty acids. Mol Biosyst 12, 93-101.
- 117. Mc Auley MT, Kenny RA, Kirkwood TB et al. (2009) A mathematical model of aging-related and cortisol induced hippocampal dysfunction. BMC Neurosci 10, 26.
- 118. Mc Auley MT, Mooney KM, Angell PJ et al. (2015) Mathematical modelling of metabolic regulation in aging. Metabolites 5, 232-251.
- 119. Mc Auley MT & Mooney KM (2015) Computationally modeling lipid metabolism and aging: a mini-review. Comput Struc Biotechnol J 13, 38-46.
- 120. Mc Auley M, Jones J, Wilkinson D et al. (2005) Modelling lipid metabolism to improve healthy ageing. BMC Bioinformatics 6, 1-1.
- 121. Mc Auley MM, Wilkinson DJ, Jones JJ et al. (2012) A whole-body mathematical model of cholesterol metabolism and its age-associated dysregulation. BMC Syst Biol **6**, 130.
- 122. Mishra S, Somvanshi PR & Venkatesh KV (2014) Control of cholesterol homeostasis by entero-hepatic bile transport - the role of feedback mechanisms. RSC Adv 4, 58964-58975.
- 123. Morgan AE, Mooney KM, Wilkinson SJ et al. (2016) Mathematically modelling the dynamics of cholesterol metabolism and ageing. Biosystems 145, 19-32.
- 124. Van de Wiele T, Van den Abbeele P, Ossieur W et al. (2015) The simulator of the human intestinal microbial ecosystem (SHIME®). In The Impact of Food Bio-Actives on Gut Health, pp. 305-317 [K Verhoeckx, P Cotter and I Lopez-Exposito et al. editors] Heidelberg: Springer International Publishing.
- 125. Nollet LJA, Pereira DI & Verstraete W (1999) Effect of a probiotic bile salt hydrolytic Lactobacillus reuteri on the human gastrointestinal microbiota as simulated in the SHIME Reactor System. *Microb Ecol Health Dis* 11, 13–21.
- 126. Molly K, Vande Woestyne M & Verstraete W (1993) Development of a 5-step multi-chamber reactor as a simulation of the human intestinal microbial ecosystem. Appl Microbiol Biotechnol 39, 254-258.
- 127. De Smet I, De Boever P & Verstraete W (1998) Cholesterol lowering in pigs through enhanced bacterial bile salt hydrolase activity. Br J Nutr 79, 185–194.
- 128. Rodes L, Coussa-Charley M, Marinescu D et al. (2013) Design of a novel gut bacterial adhesion model for probiotic applications. Artif Cells Nanomed Biotechnol 41, 116-124.
- 129. Minekus M (2015) The TNO gastro-intestinal model (TIM). In The Impact of Food Bioactives on Health: In



- Vitro and Ex Vivo Models, pp. 37–46 [K Verhoeckx, P Cotter, I López-Expósito, C Kleiveland, T Lea, A Mackie, T Requena, D Swiatecka and H Wichers, editors]. Cham: Springer International Publishing.
- 130. Christley S, Cockrell C & An G (2015) Computational studies of the intestinal host-microbiota interactome. Computation 3, 2.
- 131. de Wiele TV, Boon N, Possemiers S et al. (2004) Prebiotic effects of chicory inulin in the simulator of the human intestinal microbial ecosystem. FEMS Microbiol Ecol **51**, 143–153.
- 132. Clio-Infra (2016) Life expectancy at birth (total). https:// www.clio-infra.eu/datasets/searchresults.
- 133. UNSD (2016) Population by age, sex and urban/rural residence. http://data.un.org/Data.aspx?d=POP&f=table Code%3A22#f\_1.
- 134. Townsend N, Bhatnagar P, Wilkins E et al. (2015) Cardiovascular Disease Statistics, 2015. London: British Heart Foundation.
- 135. Mc Auley MT & Mooney KM (2015) Computational systems biology for aging research. Interdiscip Top Gerontol **40**, 35–48.

