

Epidemiological and serological investigation of a waterborne *Campylobacter jejuni* outbreak in a Danish town

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SUMMARY

Following an unusually heavy rainfall in June 2009, a community-wide outbreak of *Campylobacter* gastroenteritis occurred in a small Danish town. The outbreak investigation consisted of (1) a cohort study using an e-questionnaire of disease determinants, (2) microbiological study of stool samples, (3) serological study of blood samples from cases and asymptomatic members of case households, and (4) environmental analyses of the water distribution system. The questionnaire study identified 163 cases (respondent attack rate 16%). Results showed a significant dose-response relationship between consumption of tap water and risk of gastroenteritis. *Campylobacter jejuni* belonging to two related *flaA* types were isolated from stool samples. Serum antibody levels against *Campylobacter* were significantly higher in cases than in asymptomatic persons. Water samples were positive for coliform bacteria, and the likely mode of contamination was found to be surface water leaking into the drinking-water system. This geographically constrained outbreak presented an ideal opportunity to study the serological response in persons involved in a *Campylobacter* outbreak. The serology indicated that asymptomatic persons from the same household may have been exposed, during the outbreak period, to *Campylobacter* at doses that did not elicit symptoms or alternatively had been exposed to *Campylobacter* at a time prior to the outbreak, resulting in residual immunity and thus absence of clinical signs.

Key words: *Campylobacter*, cohort study, outbreak, serology, waterborne.

INTRODUCTION

Campylobacter spp. are a common cause of acute human bacterial enteritis. Although generally perceived as foodborne infections, primarily from poultry meat

and raw milk, other routes of transmission, including environmental exposures and animal contact, are recognized [1]. Outbreaks of *Campylobacter* gastroenteritis due to contamination of municipal drinking-water supply systems, in some cases affecting thousands of people, have been reported from several countries, including Denmark, Finland, Norway, and Sweden [2–7].

As for other intestinal infections, the true incidence of *Campylobacter* infections is believed to be several times higher than the reported incidence, because

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most cases do not seek medical attention due to short illness duration or mild or absent symptoms [8]. It has been proposed that the number of such cases can be estimated using serological tests, thus obtaining a better measure of the force of transmission in a population (i.e. 'seroincidence'). In clinical medicine, serology is used for confirming recent *Campylobacter* infections in the differential diagnosis of patients with suspected Guillain-Barré syndrome or reactive arthritis [9–12]. The serum antibody response to acute *Campylobacter* infection consists of a sharp rise in immunoglobulin G (IgG), IgA and IgM as measured in an enzyme-linked immunosorbent assay (ELISA), followed by a rapid decrease of IgM and IgA over a period of ~6 weeks [13, 14]. An elevation of at least two of these antibody classes is often considered highly suggestive of recent infection.

In an outbreak situation, serology may be useful to more accurately determine who is infected and who is not, thus providing a better estimate of the true number of affected persons. By avoiding misclassification of asymptotically infected persons (or persons not seeking medical consultation) as non-cases, serology may also improve the power of analytical epidemiological studies to identify the source of an outbreak.

In the week beginning 15 June 2009, general practitioners (GPs) in the town of Tune, ~30 km south of Copenhagen, noticed an unusual increase in patients with acute gastroenteritis. *Campylobacter* spp. were detected in stool cultures from some patients. Preliminary inquiries showed that affected persons apparently lived dispersed over the whole town and that they had not attended a common event. No increase of gastroenteritis cases was observed in neighbouring towns. On 11–12 June the area had been affected by an exceptionally heavy rainfall, causing a backflow of water from the common drainage system for rainwater and sewage into the streets. The suspicion of a waterborne gastroenteritis outbreak was raised, an outbreak investigation began and a tap-water boiling advice was issued on 20 June. In the following, we present the outbreak analysis, including a serological follow-up study of persons affected by this waterborne *Campylobacter* outbreak.

METHODS

Questionnaire survey

A cohort study was initiated using an online questionnaire with questions about signs and symptoms, onset

date, duration of illness and exposure to potential sources of *Campylobacter* infection (e.g. consumption of unboiled tap water, poultry or raw milk, eating ready-to-eat meals from local shops, etc.). All residents of the town were invited to participate in the survey via announcements in the local newspaper, via a link on the municipality website and through informal messages from local key persons. Respondents were asked to provide their complete address (street name and house/apartment number) in order to perform the serological investigation at household level (see below).

Case definitions

A clinical case was defined as a person with diarrhoea (>3 loose stools in 24 h) or abdominal pain with either fever or vomiting, with symptom onset during the period 13–26 June 2009. A confirmed case was defined as a clinical case with a stool culture positive for *Campylobacter* spp. Asymptomatic persons were defined as persons without any gastrointestinal symptoms. Individuals with ambiguous symptoms (i.e. symptoms indicative of gastrointestinal illness but not fulfilling the case definition) and individuals reporting international travel between 8 and 19 June were excluded from the statistical analysis.

Gender and age distribution (in 10-year age groups 0–9, 10–19, 20–29 years, etc.) of clinical cases and asymptomatic persons were compared using the χ^2 test.

Serological study

Among respondents to the questionnaire survey, we selected a sample of individuals who reported drinking tap water between 12 and 20 June and who were either clinical cases or asymptomatic persons living in the same household as a clinical case. These individuals were selected at household level, i.e. asymptomatic persons were only included if they shared a household with a clinical case. To include individuals from as wide a geographical area as possible, households from different areas of the town were selected. The selection at household level was made in order to account for the possibility that the water contamination was not homogenous across the town and between households. Within a single household we assumed that both cases and asymptomatic individuals had been drinking water with identical *Campylobacter* concentrations and were thus equally at risk of exposure. After informed

consent, blood samples were collected from participants during home visits or at appointments with local GPs. Blood samples were drawn about 2, 4, 7, and 13 weeks after the presumed first exposure on 12 June.

Blood was centrifuged within 24 h and serum samples were stored at 4 °C until analysis. IgA, IgG and IgM against *Campylobacter* spp. was measured by an in-house-developed ELISA at Statens Serum Institut (SSI) as described previously [13].

Ethical permission for the serological study was given by the Committee for Ethics in Science of Copenhagen and Frederiksberg Municipality (reference no. 11-097/02).

In order to assess changes in log-transformed IgG, IgM and IgA optical density (OD) values over time, separate random-effects linear regression models were fitted for both asymptomatic and symptomatic individuals within each antibody class. The outcome variable of these models was log-transformed antibody titre and the explanatory variable was time since assumed exposure (on 12 June 2009) in weeks. The intercept and slope of the models for ill and asymptomatic individuals for each antibody class was compared using *t* tests (using the asymptomatic model as the reference). Statistical analyses were performed using Stata 14 software (StataCorp., USA) and SAS v. 9.4 (SAS Institute, USA).

Microbiological investigations

Patients consulting their GP with diarrhoea and all participants in the serological study were asked to submit two stool samples, unless they had already tested positive for *Campylobacter*. Stool samples were cultured for enteric pathogens including *Campylobacter* by standard methods at the Danish national reference laboratory at SSI. A subset of samples from patients consulting their GP was additionally tested for norovirus, sapovirus and rotavirus.

Isolates of *Campylobacter* were characterized by *flaA* typing according to Meinersmann *et al.* [15].

Environmental investigations

Water samples were repeatedly taken at the waterworks and at multiple points of the water pipe network following the suspected contamination event. Analysis of water samples as well as assessment of the technical condition of the water supply system and scenarios of a possible contamination of the

drinking-water supply with sewage water were undertaken by a private civil engineering company commissioned by the municipality. The presence of bacteria, bacterial markers and faecal markers was analysed using quantitative polymerase chain reaction (qPCR) and plate counts.

RESULTS

Questionnaire survey

In 2009, the town of Tune had a population of 5052 persons (50.5% female) with an average age of 40 years. The questionnaire survey was completed by a total of 1039 individuals, response rate 21%, (61% females, 39% males) ranging from 1 to 86 years (mean age 42 years).

Of the 1039 respondents, 69 (7%) were excluded as they did not provide information on the presence or absence of gastrointestinal symptoms, 17 (2%) were excluded due to ambiguous symptoms as described above and 20 persons (2%) were excluded as they reported international travel between 8 and 19 June. Of the remaining 933 persons, 159 (15.3%) were classified as cases and 774 as non-cases, based on their reported symptoms and onset dates. The 159 cases were distributed in 119 households with one case, nine households with two cases, six households with three cases and one household with four cases. Dates for onset of gastroenteritis ranged from 11 to 30 June with a peak on 18–20 June (Fig. 1).

The only exposure variable showing a statistically significant association with increased risk of illness was consumption of tap water. A significant dose-response relationship was observed with increasing attack rates in persons reporting higher amounts of tap-water consumption (Table 1).

Serological study

A total of 67 individuals provided blood samples for IgG, IgM and IgA measurement of which 35 were clinical cases and 32 were asymptomatic. Twelve (34%) out of 35 clinical cases and 16 (50%) of asymptomatic persons were males which was not significantly different ($P = 0.19$). The age distribution of ill and asymptomatic persons also did not differ significantly.

Campylobacter antibody levels following exposure to contaminated water showed IgG and IgA OD values which overall were 3–4 times higher for cases

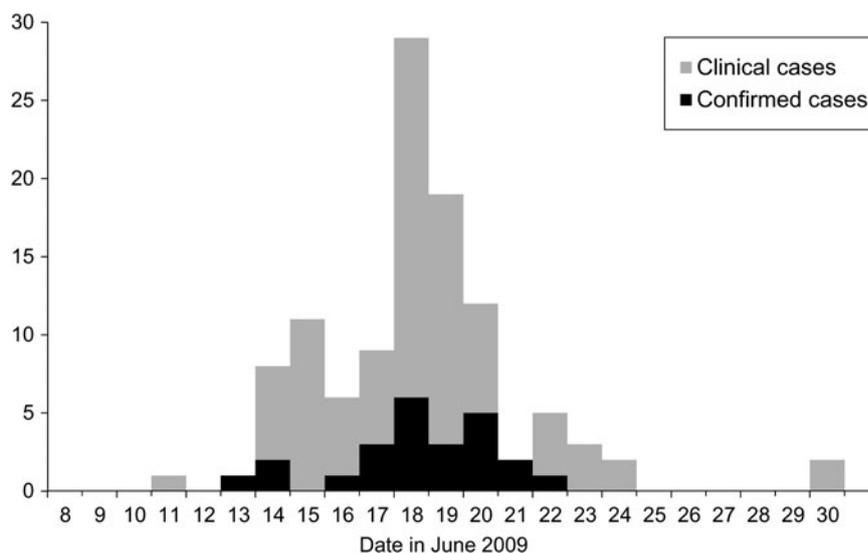


Fig. 1. Persons with acute gastroenteritis by date of symptom onset, waterborne *Campylobacter* outbreak, Denmark, June 2009

Table 1. Attack rate by reported amount of tap-water consumption, waterborne *Campylobacter* outbreak, Denmark, June 2009

Daily tap-water consumption (in glasses of ~200 ml)	Cases (n)	Non-cases (n)	Attack rate (%)	Risk ratio
0–1	5	68	6.9	1 (reference)
2	9	113	7.4	1.08
3	25	131	16.0	2.34
4	34	145	19.0	2.77
≥5	86	317	21.3	3.11

Mantel–Haenszel χ^2 for linear trend = 18.80, $P < 0.001$

compared to asymptomatic persons, while IgM OD values did not differ significantly (Fig. 2, Table 2). Regarding antibody decay profiles, test for similar slopes showed a significantly faster IgG and IgA antibody decay profile in ill compared to asymptomatic individuals while the IgM antibody decay profile did not differ between the two groups (Fig. 2, Table 2).

Microbiological investigations

Stool samples were received from 100 patients consulting their GP with diarrhoea. Of these, 41 were culture-positive for *Campylobacter jejuni*. Cultures for other enteric bacteria were negative. Virological investigations detected sapovirus in two (10%) out of 20 samples

tested, and one patient tested positive for both sapovirus and *C. jejuni*. Stool samples were available for 26 cases enrolled in the serological study and 10 (38%) of these were culture positive for *Campylobacter*. Of the 32 asymptomatic subjects enrolled in the serology study, 12 provided stool samples, of which only one was positive for *Campylobacter*.

Campylobacter isolates from 27 cases were characterized by *flaA* sequence analysis. Most isolates belonged to two distinct *flaA* types (13 and nine isolates, respectively) and the remaining five isolates showed <5% sequence difference from these two clusters, showing that the outbreak was caused by two *Campylobacter* strains with different but strongly related *flaA* sequences.

Environmental and technical investigations

The town's drinking water consisted of non-chlorinated groundwater drawn from several boreholes around the town and supplied by the local water plant. Testing of water samples collected on 21 and 26 June and 6, 9 and 10 July from more than 25 points of the water distribution system, including one of the groundwater boreholes, indicated contamination with coliform bacteria (measured by total coliform counts). In addition, following the first positive culture from human stool samples, ~300 water samples were tested specifically for *Campylobacter* by qPCR. These tests were negative.

During 20 June to 30 July, a boiling order was in place for the whole town and the inhabitants were

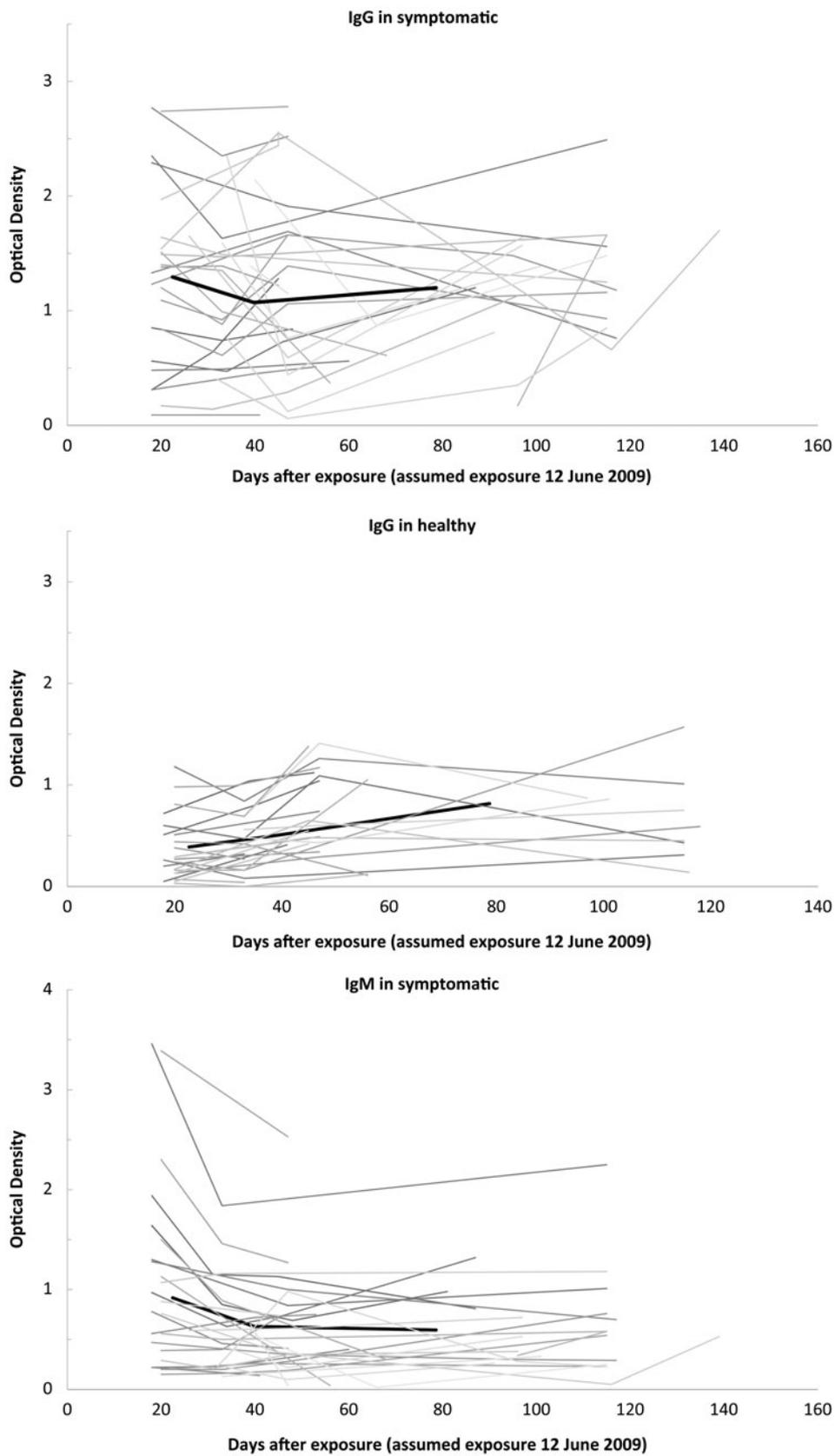


Fig. 2. Serum antibody levels against *Campylobacter* in persons exposed to tap water presumably contaminated with *Campylobacter*, Denmark, June 2009. Optical density (OD) (in arbitrary units). Bold lines indicate the mean OD value.

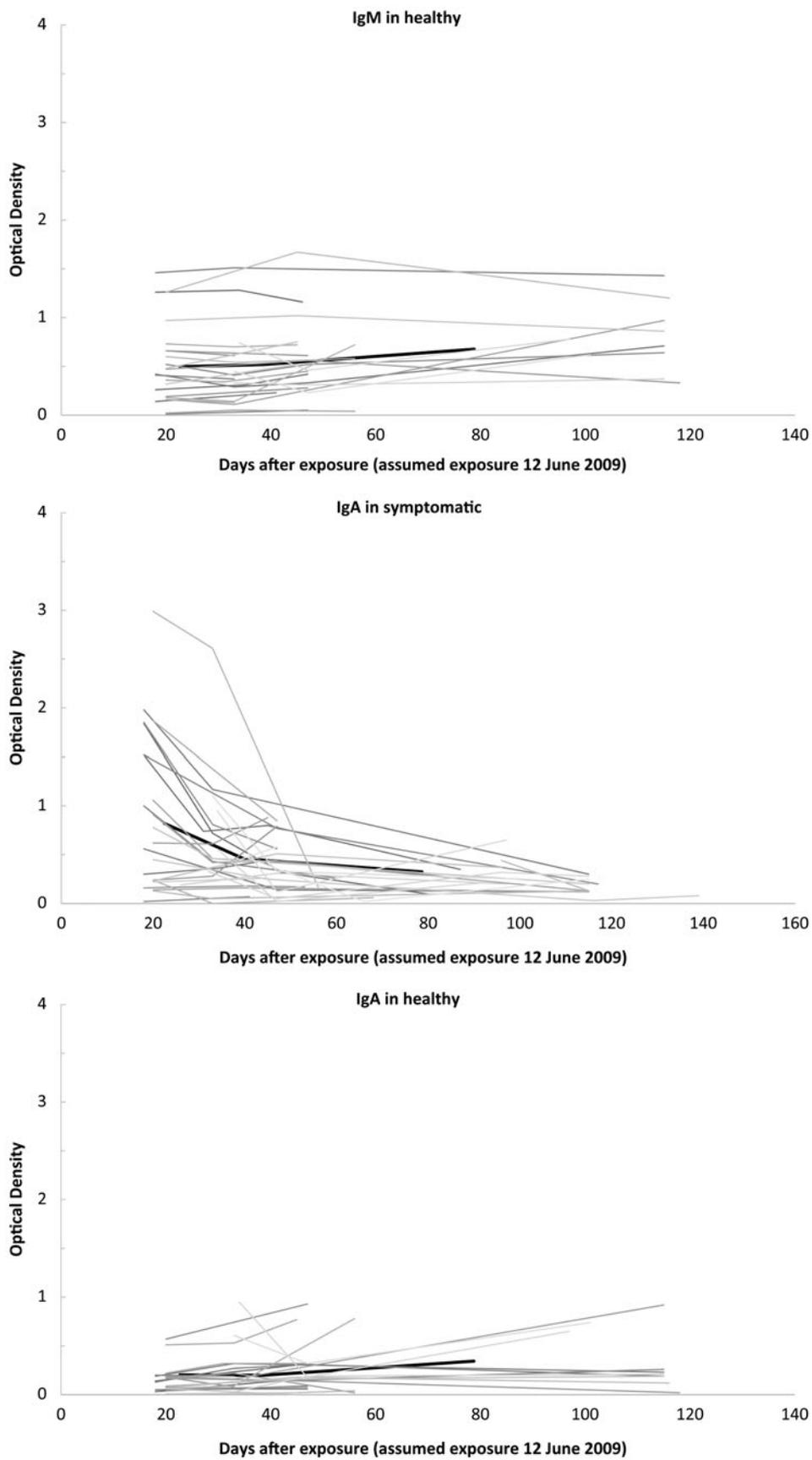


Fig. 2 (cont.)

Table 2. Modelling *Campylobacter* antibody decay profiles in persons exposed to tap water presumably contaminated with *Campylobacter*, Denmark, June 2009

Antibody class	Variable	Clinical form	Coefficient	95% CI	Regression <i>P</i> value	Comparison between ill and asymptomatic (<i>P</i> value) [‡]
IgG	Intercept	Ill	0.73	0.52–1.04	0.08	<0.0001
		Asymptomatic	0.20	0.13–0.32	<0.0001	
	Week	Ill	1.01	0.92–1.11	0.76	0.03
		Asymptomatic	1.24	1.06–1.45	0.008	
IgA	Intercept	Ill	0.38	0.24–0.57	<0.0001	<0.0001
		Asymptomatic	0.08	0.05–1.30	<0.0001	
	Week	Ill	0.80	0.69–0.92	0.003	0.001
IgM	Intercept	Ill	1.30	1.02–1.65	0.04	
		Asymptomatic	0.51	0.37–0.71	0.0002	0.08
	Week	Ill	0.32	0.21–0.48	<0.0001	
		Asymptomatic	0.94	0.84–1.05	0.25	0.09
		Asymptomatic	1.13	0.94–1.37	0.19	

CI, Confidence interval.

[‡] Comparing the values for intercept and slope between ill and asymptomatic persons within each antibody class (with asymptomatic as the reference).

supplied with drinking water from tank trucks. The technical assessment revealed a faulty installation adjacent to one of the boreholes which may have allowed a backflow of sewage water into the gravel surrounding the borehole, when the sewage system was congested due to the heavy rainfall on 12 June.

DISCUSSION

Here we present results from an investigation of a *C. jejuni* outbreak in a confined setting, with a particular focus on antibody development in cases and asymptomatic members of case households. The outbreak was most likely caused by drinking water as shown by several lines of evidence: First, the only exposure found to be associated with gastroenteritis in the cohort study was drinking tap water, with a marked dose-response relationship between amount of tap water consumed and the risk of gastroenteritis. Second, an exceptionally heavy rainfall occurred a few days before the outbreak, leading to a drinking-water contamination caused by congestion of the combined rainwater drainage and sewage system. A technical investigation of the water system established a likely scenario as to how sewage-contaminated rainwater could have seeped into one of the groundwater boreholes. Third, detection of coliform bacteria in drinking-water samples confirmed a contamination, even though *Campylobacter* could not be detected,

possibly because the necessary large volume water samples were taken too late after the contamination.

Waterborne outbreaks in Denmark are rare [16], as indeed are *Campylobacter* outbreaks in general [17]. The infrequent occurrence of waterborne outbreaks in Denmark compared to neighbouring countries has been explained by the fact that drinking water in Denmark is almost exclusively provided as groundwater [16]. The only two other known major waterborne *Campylobacter* outbreaks in the country occurred in 1995–1996 [2] and in 2010 [7]. Both were traced back to local water distribution systems following point-source contamination events with single clones of *C. jejuni* leading to widespread illnesses in the local settings. The outbreak described here is in several ways similar, although it differs by being caused by two different clones of *C. jejuni*. Given that the drinking-water system was likely contaminated by surface water leaking in, it is not surprising to find several related clones of *C. jejuni* in this outbreak.

Adding serological analysis to outbreak investigations provides the opportunity to study exposure and immune response over time – although recent evidence suggests that, for *Campylobacter*, seroepidemiological results may sometimes be difficult to interpret. Since the outbreak occurred, we and others have conducted multi-country European seroepidemiological studies for *Salmonella* and *Campylobacter*. This has

allowed us to measure the exposure rate in the population and thus for instance to compare infection levels between countries [18, 19]; something not feasible to do by comparing numbers of registered cases because of the large differences in surveillance systems both within and between European countries. For *Salmonella*, such results have been in line with what might be suspected [20, 21]. For *Campylobacter*, however, a different picture has emerged: the seroincidence is generally very high with much less pronounced variation between countries [22]. The longitudinal serological analysis performed in Denmark does not mirror the quadrupling of registered cases seen throughout the 1990s [23]. This has led to the hypothesis being put forward that Europeans (as indeed probably most populations in the developed world) are frequently (possibly on an almost annual basis) exposed to *Campylobacter* via a number of routes, including environmental. This will lead to some symptomatic, but also many asymptomatic infections, possibly causing a build-up of immunity to infections throughout life. High dose exposure (and likely also exposure to previously unencountered serovariants) will, however, still lead to symptomatic infection [22]. In other words, a simple correlation (a conversion factor) between seroincidence and registered cases of clinical illness does not exist.

In this situation, interpreting the results of our outbreak serostudy is not straightforward. In the outbreak, illness was shown to be associated with water consumption. In the survey, drinking tap water was very commonly reported for both cases and asymptomatic respondents. Thus, *a priori*, we might expect almost all participants in the study to seroconvert – irrespective of case status. We might even expect most participants to have been seropositive even before water exposure, since the level of *Campylobacter* seropositivity in the population is, as mentioned, generally high (which indeed may explain the findings of measurable *Campylobacter* antibody levels in almost all individuals sampled). However, it seems likely that the bacteria would not have been uniformly suspended in the water. Even in the questionnaire respondents who reported drinking ≥ 1 litre tap water daily, the attack rate was a moderate 22%. A possible explanation for the results of our serological study is that only individuals unfortunate enough to have drunk the (most heavily) contaminated water, receiving a large dose of *Campylobacter*, developed symptoms and were most likely to seroconvert.

It is often hypothesized that, following massive exposure, a large number of individuals become

infected (as documented by high antibody levels) but do not develop symptoms. We could not provide support for this hypothesis. In contrast, our findings indicate that asymptomatic persons have low, but measurable, antibody levels and were most likely asymptomatic either due to low levels of exposure or a combination of the low exposure and build-up of immunity from repeated previous exposure episodes. Further, our findings demonstrate that the use of seroepidemiology in an outbreak situation can provide additional understanding of the dynamics of the outbreak. We suggest that other researchers use outbreaks as ‘natural experiments’ to study antibody response in asymptomatic but exposed individuals, thereby gradually increasing the insight into advantages and limitations of seroepidemiology as a method to monitor bacterial gastrointestinal infections in the community.

In conclusion, we investigated one of the very few known waterborne *Campylobacter* outbreaks in Denmark, establishing source, agent and mode of contamination. Further, we explored the use of serology as a tool for investigating outbreaks or – in another perspective – using outbreaks to study the seroepidemiology of *Campylobacter*, highlighting serology as a potentially valuable method to gain a better understanding of the dynamics of the most frequently occurring bacterial enteric infection in Europe.

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DECLARATION OF INTEREST

None.

REFERENCES

1. Evers EG *et al.* *Campylobacter* source attribution by exposure assessment. *International Journal of Risk Assessment Management* 2008; **8**: 174–190.

2. Engberg J *et al.* Water-borne *Campylobacter jejuni* infection in a Danish town—a 6-week continuous source outbreak. *Clinical Microbiology and Infection* 1998; **4**: 648–656.
3. Kuusi M, *et al.* An outbreak of gastroenteritis from a non-chlorinated community water supply. *Journal of Epidemiology and Community Health* 2004; **58**: 273–277.
4. Kuusi M, *et al.* A large outbreak of campylobacteriosis associated with a municipal water supply in Finland. *Epidemiology and Infection* 2005; **133**: 593–601.
5. Martin S, *et al.* A case-cohort study to investigate concomitant waterborne outbreaks of *Campylobacter* and gastroenteritis in Söderhamn, Sweden, 2002–3. *Journal of Water Health* 2006; **4**: 417–24.
6. Jakopanec I, *et al.* A large waterborne outbreak of campylobacteriosis in Norway: the need to focus on distribution system safety. *BMC Infectious Diseases* 2008; **8**: 128.
7. Gubbels SM, *et al.* A waterborne outbreak with a single clone of *Campylobacter jejuni* in the Danish town of Køge in May 2010. *Scandinavian Journal of Infectious Diseases* 2012; **44**: 586–594.
8. Hardnett FP, *et al.* Epidemiological issues in study design and data analysis related to FoodNet activities. *Clinical Infectious Diseases* 2004; **38**: S121–126.
9. Ang CW, *et al.* Validation of an ELISA for the diagnosis of recent *Campylobacter* infections in Guillain-Barré and reactive arthritis patients. *Clinical Microbiology and Infection* 2007; **13**: 915–922.
10. Tam CC, *et al.* Guillain-Barré syndrome and preceding infection with *Campylobacter*, influenza and Epstein-Barr virus in the general practice research database. *PLoS ONE* 2007; **2**: e344.
11. Kuhn KG, *et al.* Detection of antibodies to *Campylobacter* in humans using enzyme-linked immunosorbent assays: a review of the literature. *Diagnostic Microbiology and Infectious Diseases* 2012; **74**: 113–118.
12. Mortensen NP, *et al.* Sialylation of *Campylobacter jejuni* lipo-oligosaccharides is associated with severe gastroenteritis and reactive arthritis. *Microbes and Infection* 2009; **11**: 988–994.
13. Strid MA, *et al.* Antibody responses to *Campylobacter* infections determined by an enzyme-linked immunosorbent assay: 2-year follow-up study of 210 patients. *Clinical and Diagnostic Laboratory Immunology* 2001; **8**: 314–319.
14. Taylor BV, *et al.* Sensitivity and specificity of serology in determining recent acute *Campylobacter* infection. *Internal Medicine Journal* 2004; **34**: 250–258.
15. Meinersmann RJ, *et al.* Discrimination of *Campylobacter jejuni* isolates by fla gene sequencing. *Journal of Clinical Microbiology* 1997; **35**: 2810–2814.
16. Guzman-Herrador B, *et al.* Waterborne outbreaks in the Nordic countries, 1998 to 2012. *Eurosurveillance* 2015; **20**: 1–10.
17. Statens Serum Institut. EPI-NEWS No. 11, 2016 (<http://www.ssi.dk/English/News/EPI-NEWS/2016/No11-2016.aspx>).
18. Simonsen J, *et al.* Estimation of incidences of infectious diseases based on antibody measurements. *Statistics in Medicine* 2009; **28**: 1882–1895.
19. Mølbak K, *et al.* Seroincidence of human infections with nontyphoid *Salmonella* compared with data from public health surveillance and food animals in 13 European countries. *Clinical Infectious Diseases* 2014; **59**: 1599–1606.
20. Falkenhorst G *et al.* Serological cross-sectional studies on *Salmonella* incidence in eight European countries: no correlation with incidence of reported cases. *BMC Public Health* 2012; **16**: 523.
21. Simonsen J, *et al.* Sero-epidemiology as a tool to study the incidence of *Salmonella* infections in humans. *Epidemiology and Infection* 2008; **136**: 895–902.
22. Teunis PFM, *et al.* *Campylobacter* seroconversion rates in selected countries in the European Union. *Epidemiology and Infection* 2013; **141**: 2051–2057.
23. Emborg HD, *et al.* Was the increase in culture-confirmed *Campylobacter* infections in Denmark during the 1990s a surveillance artefact? *Eurosurveillance* 2015; **20**: pii = 30041.