



Maternal pea fiber supplementation to a high calorie diet in obese pregnancies protects male offspring from metabolic dysfunction in adulthood

Original Article

Cite this article: Andreani GA, Mahmood S, Patel MS, and Rideout TC. (2023) Maternal pea fiber supplementation to a high calorie diet in obese pregnancies protects male offspring from metabolic dysfunction in adulthood. *Journal of Developmental Origins of Health and Disease* **14**: 711–718. doi: [10.1017/S2040174423000399](https://doi.org/10.1017/S2040174423000399)

Received: 16 June 2023
Revised: 28 September 2023
Accepted: 20 November 2023
First published online: 18 January 2024

Keywords:

Maternal; fiber; obesity; pregnancy; dyslipidemia

Corresponding author:

T. C. Rideout; Email: rideout@buffalo.edu

Gabriella A. Andreani¹, Saleh Mahmood¹, Mulchand S. Patel² and Todd C. Rideout¹

¹Departments of Exercise and Nutrition Sciences, School of Public Health and Health Professions, Buffalo, NY, USA and ²Biochemistry, Jacobs School of Medicine and Biomedical Sciences, University at Buffalo, Buffalo, NY, USA

Abstract

We investigated the influence of maternal yellow-pea fiber supplementation in obese pregnancies on offspring metabolic health in adulthood. Sixty newly-weaned female Sprague-Dawley rats were randomized to either a low-calorie control diet (CON) or high calorie obesogenic diet (HC) for 6-weeks. Obese animals were then fed either the HC diet alone or the HC diet supplemented with yellow-pea fiber (HC + FBR) for an additional 4-weeks prior to breeding and throughout gestation and lactation. On postnatal day (PND) 21, 1 male and 1 female offspring from each dam were weaned onto the CON diet until adulthood (PND 120) for metabolic phenotyping. Adult male, but not female, HC offspring demonstrated increased body weight and feed intake vs CON offspring, however no protection was offered by maternal FBR supplementation. HC male and female adult offspring demonstrated increased serum glucose and insulin resistance (HOMA-IR) compared with CON offspring. Maternal FBR supplementation improved glycemic control in male, but not female offspring. Compared with CON offspring, male offspring from HC dams demonstrated marked dyslipidemia (higher serum cholesterol, increased number of TG-rich lipoproteins, and smaller LDL particles) which was largely normalized in offspring from HC + FBR mothers. Male offspring born to obese mothers (HC) had higher hepatic TG, which tended to be lowered ($p = 0.07$) by maternal FBR supplementation.

Supplementation of a maternal high calorie diet with yellow-pea fiber in pre-pregnancy and throughout gestation and lactation protects male offspring from metabolic dysfunction in the absence of any change in body weight status in adulthood.

Introduction

The prevalence of obesity has doubled in the last 50 years, with a third of the global population being considered either overweight or obese.¹ Obesity is a high priority health concern as it is associated with reduced glycemic control and dyslipidemia including altered distribution of lipoprotein particle number and size. Further, individuals with obesity are more likely to develop type 2 diabetes, cardiovascular disease, and some types of cancers.²

The pathogenesis of obesity is complex and multi-factorial, involving genetic predisposition and lifestyle factors including limited exercise and poor nutrition.³ In addition, the more recent rise in pediatric obesity suggests that maternal health status during pregnancy may influence fetal metabolic exposures *in utero* or in early postnatal life to promote longer-term obesity and metabolic disease risk.⁴ Thus, the high incidence of maternal overweight and obesity in women of childbearing age is of particular concern⁵ as this environment has been shown to program metabolic maladaptation in offspring, increasing the risk of childhood obesity and metabolic dysfunction.^{6,7}

There is currently an unmet and urgent need to identify maternal interventions for obesity management that improves maternal pregnancy health and reduces the transgenerational impact of maternal obesity in offspring. In non-pregnancy populations, current recommended treatment approaches for obesity are primarily centered around lifestyle changes where patients are encouraged to improve their diet (i.e. focus on nutrient quality, lower-energy foods, etc.) and exercise regularly, though medication and surgery may be recommended in cases where behavioral modification is not effective.⁸ However, obesity management in women may require special considerations, particularly during pregnancy.⁹ As weight loss and the use of anti-obesity pharmacotherapies are not recommended during pregnancy, the importance of achieving a healthy body weight and improving obesity-associated metabolic issues prior to pregnancy are encouraged to support both maternal and future offspring health.^{8,9} However, a recent

© The Author(s), 2024. Published by Cambridge University Press in association with The International Society for Developmental Origins of Health and Disease (DOHaD). This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted re-use, distribution and reproduction, provided the original article is properly cited.



meta-analysis reported that maternal diet and lifestyle interventions among overweight and obese pregnant women had little effect on the risk of early childhood obesity,¹⁰ re-emphasizing the importance of lifestyle interventions that are initiated before pregnancy and continue throughout the pregnancy and early postpartum periods.

One potentially feasible dietary approach to help achieve a healthy body weight is increased consumption of dietary fiber. As plant-based carbohydrates that are not digested or absorbed in the small intestine, dietary fibers may assist in body weight regulation by reducing dietary energy density, increasing satiety, and promoting a healthy gut microbiome.^{11,12} In non-pregnant adults, high fiber dietary intake has been shown to improve weight loss often without the need for calorie-restriction,^{13,14} decrease waist circumference,^{15,16} and improve a range of obesity-associated metabolic perturbations including dyslipidemia¹⁷ and reduced glycemic control.¹⁸ Despite these findings, there has been limited research to assess the health benefits of maternal fiber consumption in obese pregnancies on the metabolic health of offspring. Therefore, using a diet-induced obese Sprague-Dawley rat model, we aimed to examine the protective effects of maternal yellow-pea fiber supplementation on offspring metabolic function as adults.

Materials and methods

Animal's diets, and design

The rats used in this experiment were cared for in accordance with the guidelines established by the Institutional Animal Care and Use Committee. All procedures were reviewed and approved by the Animal Care Committee at the University at Buffalo. Sixty newly-weaned postnatal day (PND) 21 female Sprague-Dawley (Charles River, obese prone, CrI:OP-CD) rats were housed in the Laboratory Animal Facility at the University at Buffalo under controlled environmental conditions (12 h light:12 h dark, 20°C, and 50% humidity). All animals had free access to food and water throughout the experiment. During the initial 6-weeks of prepregnancy, rats were randomized to a low-calorie control diet (CON; $n = 10$; total energy 3.8 kcal/g; % energy from fat, 10; protein, 20; and carbohydrate, 70) (Research Diets, D12450K) or a high caloric obesity-inducing diet (HC; $n = 50$; total energy 4.8 kcal/g; % energy from fat, 44; protein, 20; and carbohydrate, 35) (Research Diets, D12451). Following this 6-week period, HC animals demonstrating a body weight $\geq 20\%$ over the CON animals were considered "obese" and were then randomized to either remain on the HC diet ($n = 20$) or provided the HC diet supplemented with yellow-pea fiber (HC + FBR, $n = 15$) (25%, Roquette® Pea Fiber I 50 m; total energy 3.8 kcal/g; % energy from fat, 50; protein, 22; and carbohydrate, 30) for an additional 4-weeks prior to mating (Table 1). Macronutrient and fiber content of the pea fiber supplement was assessed by third party analyses (Table 1) (Anresco Laboratories, San Francisco, Ca, USA). At end of the prepregnancy period (a total of 10 weeks) the rats were bred with CON-fed male breeders to establish a timed pregnancy,¹⁹ confirmed by the presence of a vaginal plug. Maternal body weights and food intake were collected weekly throughout gestation and lactation. Following delivery, litter size and weights were recorded, and the litters were adjusted to 8 pups per dam (4 males and 4 females) within 24 hours of birth. At weaning on PND 21, 6 offspring from each group (3 males and 3 females) were randomly selected for metabolic phenotyping in the non-fasted

Table 1. Experimental diet formulation (% composition)

Ingredient	Experimental Diets ¹		
	CON	HC	HC + FBR
Casein	19.0	23.3	19.0
L-Cystine	0.3	0.3	0.35
Corn Starch	52.1	8.5	9.3
Maltodextrin	14.2	11.7	9.0
Sucrose	0.4	20.6	9.69
Cellulose, BW200	4.7	5.8	0.00
Yellow Pea Fiber Isolate	–	–	25.0
Soybean Oil	2.4	2.9	2.9
Lard	1.9	20.7	18.5
Mineral Mix	4.7	5.8	5.8
Vitamin Mix	0.1	0.1	0.1
Choline Bitartrate	0.2	0.2	0.2
Energy Contribution			
Total energy (kcal/g)	3.8	4.8	3.8
% energy from fat	10.0	45.2	50.6
% energy from protein	20.1	20.1	22.2
% energy from carbohydrate	69.8	34.7	29.3
Fiber Content (%)			
Insoluble fiber	4.74	5.83	12.11
Soluble	–	–	1.11

¹CON, low-calorie control diet based on D12450 formulation (Research Diets); HC, high calorie obese-inducing diet based on D12451 (Research Diets); HC + FBR, HC diet supplemented with yellow-pea fiber (25%, Roquette® Pea Fiber I 50 m; total energy 3.8 kcal/g; % energy from fat, 50; protein, 22; and carbohydrate, 30; insoluble fiber content, 43.99%; soluble fiber content, 4.45%); HC + FBR diet was formulated to be matched for energy and macronutrient contribution based on proximate nutrient analyses of pea fiber.

state. Following weaning, one male and one female offspring from each litter were weaned onto the CON diet until PND 120. Food intake (*ad libitum*) and body weights were monitored weekly throughout the post-weaning period. On PND 120, adult offspring were anesthetized for terminal non-fasting blood collection by cardiac puncture. The rats used in this experiment were cared for in accordance with the guidelines established by the Institutional Animal Care and Use Committee. All procedures were reviewed and approved by the Animal Care Committee at the University at Buffalo. Livers were quickly excised, weighed, flash frozen in liquid nitrogen, and stored at -80°C for further processing and analyses.

Blood biochemistry

Serum analyses included glucose by colorimetric detection (Invitrogen, EIAGLUC), insulin by ELISA (Millipore, EZRMI-13K), cholesterol (TC, LDL/VLDL-C, and HDL-C) by enzymatic analysis (BioAssay, EHDL-100), and triglyceride (TG) by enzyme assay (Zenbio, STG-1-NC). Direct assessment of lipoprotein subclass distribution, including particle number (nM) and size (nm), for triglyceride-rich lipoproteins (very low-density lipoprotein /chylomicron), low-density lipoprotein (LDL), and high-density lipoprotein (HDL), was conducted by nuclear magnetic resonance spectroscopy (NMR) using automated signal acquisition

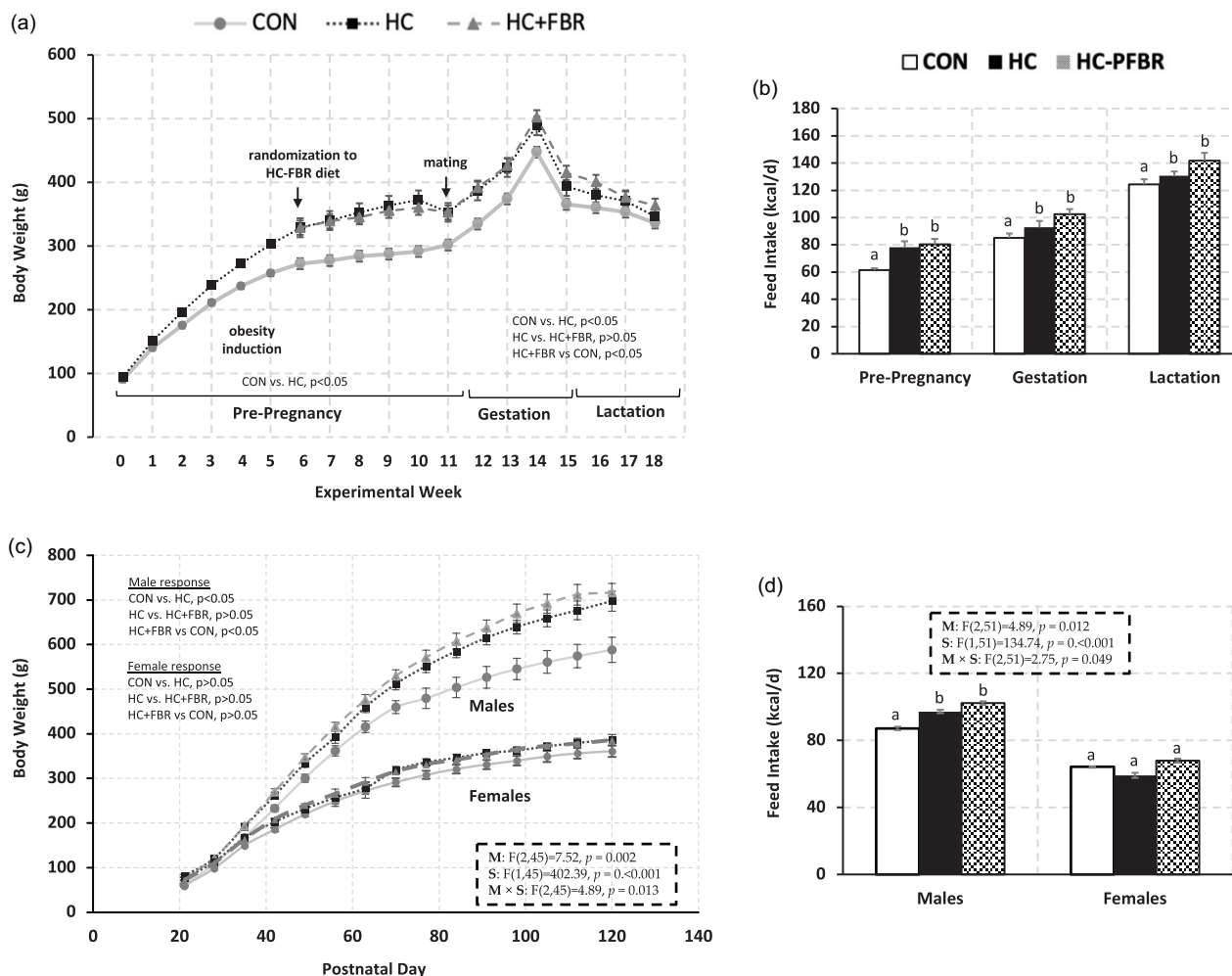


Figure 1. Body weight trajectory and food intake in mothers and offspring. [a] Body weight (g) in pre-pregnancy, gestation, and lactation in mothers consuming a control [CON], high calorie [HC], and HC + yellow-pea fiber [HC + FBR] diet; [b] Maternal caloric intake (kcal/g) in mothers in pre-pregnancy, gestation, and lactation; [c] Postnatal body weight in male and female offspring from CON, HC, and HC + FBR mothers; [d] Average postnatal daily caloric intake (kcal/d) in male and female offspring. Data are means \pm SE; $n = 8-10$ per group; ^ab groups that do not share a superscript are significantly different ($p < 0.05$).

followed by computational analysis and proprietary signal processing algorithms (LabCorp Inc., Raleigh, NC).²⁰ Lipoprotein distribution was assessed in male offspring only, given the lack of treatment effects in serum lipids amongst female offspring. Hepatic TG was extracted from frozen tissue and analyzed with a commercial kit (Zenbio, STG-1-NC) according to previously published work.²¹

Data analyses

All statistical analyses were conducted using SPSS 16 (SPSS Inc., Chicago, IL). Data were checked for normality using the Shapiro-Wilk test. Maternal outcomes were measured with a one-way ANOVA with a least significant difference (LSD) post-hoc test. Litters from each dam were considered as a single observation. Main effects of maternal exposure (CON, HC, and HC + FBR) and sex (male and female from same maternal exposure), and interaction-related effects were analyzed by two-way ANOVA. If a significant main effect or interaction was detected, a one-way ANOVA with an LSD post-hoc test was conducted to assess programming responses. Data are presented as means \pm SE. Differences were considered significant at $p < 0.05$.

Results

Compared with CON, rats consuming the HC diet in pre-pregnancy demonstrated increased ($p < 0.05$) body weight and caloric intake (Fig. 1a, b), which persisted throughout the gestation and lactation phases. Initiation of the HC + FBR diet at week 7 did not ($p > 0.05$) protect against body weight gain or reduce feed intake throughout gestation and lactation compared with the HC group.

Adult male, but not female offspring, from obese mothers (HC) demonstrated increased ($p < 0.05$) body weight and caloric intake (Fig. 1c, d) compared with offspring from CON-fed mothers, however, maternal supplementation of HC + FBR throughout pre-pregnancy, gestation, and lactation did not protect against this response.

Compared to CON, adult male and female offspring from HC-fed mothers showed an increase ($p < 0.05$) in serum glucose and HOMA-IR (Fig. 2a, c). Male HC offspring showed an additional increase ($p < 0.05$) in serum insulin (vs. CON) that was not observed in females (Fig. 2b). Maternal supplementation of pea fiber improved ($p < 0.05$) serum glucose, insulin, and HOMA-IR in adult male offspring compared with HC offspring, but no protective effect was observed in females.

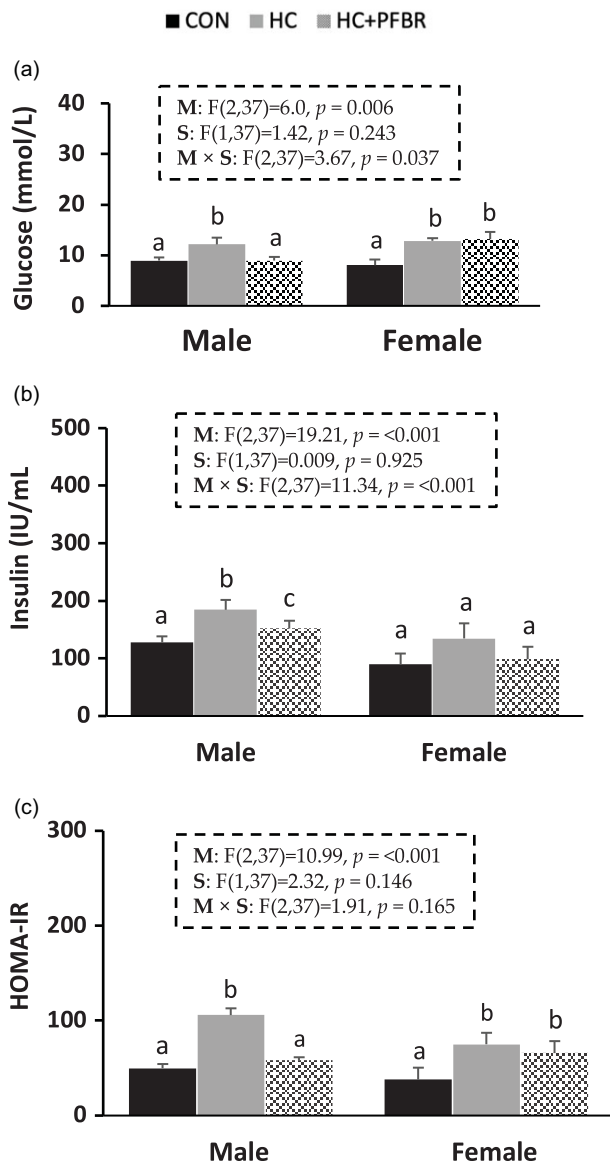


Figure 2. Glycemic control markers in adult male and female offspring from mothers consuming a control [CON], high calorie [HC], and HC + yellow-pea fiber [HC + FBR] diet. [a] Serum glucose (mg/dL); [b] Serum insulin (IU/ml); [c] HOMA-IR. Data are means \pm SE; $n = 8-10$ per group; ^{ab} groups that do not share a superscript are significantly different ($p < 0.05$).

Compared to CON, adult males from HC-fed mothers had higher ($p < 0.05$) serum total-C and LDL/VLDL-C (Fig. 3a,c), but no difference ($p > 0.05$) in HDL-C or TG (Fig. 3 b, d). Maternal yellow-pea fiber supplementation reduced ($p < 0.05$) serum LDL/VLDL-C (vs. HC) and TG (vs. CON) concentrations in adult male offspring (Fig. 3c, d), but did not influence total-C and HDL-C concentrations. No between group differences in serum lipids were observed in females.

Males from HC mothers demonstrated an altered lipoprotein distribution pattern with increased ($p < 0.05$) number of TG-rich lipoproteins particles and smaller LDL particles compared with offspring from CON mothers (Table 2). Maternal supplementation of yellow-pea fiber normalized LDL size to CON levels and reduced ($p < 0.05$) the size of TG-rich lipoproteins lower than both the CON and HC groups.

Compared with CON, maternal obesity programed an increase ($p < 0.05$) in hepatic TG in adult male offspring and maternal yellow-pea fiber supplementation tended ($p = 0.07$) to protect against this response, however, it was not significant (Fig. 4). No between group differences in liver TG were observed in female offspring.

Discussion

In this study, maternal diet-induced obesity programed hyperphagia, increased body weight, reduced glycemic control, and dyslipidemia, including altered lipoprotein distribution and increased hepatic fat accumulation, in adult male offspring compared with offspring from lean mothers. Although maternal yellow-pea fiber supplementation throughout pre-pregnancy, pregnancy, and lactation did not influence maternal obesity or offspring body weight status, several metabolic health benefits were evident in adult male, but not female, offspring including improved glycemic control and dyslipidemia.

We observed that adult male offspring from obese mothers had higher body weight and food intake than offspring from CON mothers, confirming that maternal obesity can increase the risk of obesity in offspring, at least in males. This sex-specific response has been observed in some,^{22,23} but not all,^{24,25} previous rodent model studies investigating the transgenerational impact of maternal obesity. Previous work in rodent models suggests this hyperphagic/obesogenic response may be mediated, at least partially, by altered expression of central appetite reward systems including preferential enhancement of orexigenic (NPY/AgRP) versus anorexigenic neural peptide signaling in the hypothalamus.^{26,27}

Our findings do not support a protective effect of maternal dietary fiber supplementation on gestational weight gain^{28,29} or offspring adulthood obesity^{29,30} that has been reported in other previous rodent model studies. Hallam *et al.*²⁸ reported that maternal consumption of a high prebiotic fiber diet (21.6% w/w mix of oligofructose and inulin) in a Wistar rat model throughout pregnancy and lactation reduced dam weight gain during pregnancy and lowered body fat mass in female, but not male, offspring compared with a high protein diet. A previous human study also supports a negative association between maternal dietary fiber intake and adiposity in female offspring in the first two years of life.³¹ Unfortunately, as we assessed obesity by weekly body weight measures only, we are not able to evaluate potential changes in body composition between treatment groups. However, a previous study in mice lends support for the possibility that maternal fiber supplementation throughout pregnancy and lactation could influence body weight by selectively influencing organ weight (intestinal and muscle mass) in male offspring (Desbuauds *et al.* 2012). These contradictory sex-specific findings are likely due to many experimental design factors that influence the outcomes of rodent model studies assessing the transgenerational impact of maternal obesity,³² including the source and amount of dietary fiber. The limited amount of work that has been conducted with maternal dietary fiber intervention during pregnancy in rodents has been conducted with a variety of fiber sources including galacto-oligosaccharides,³³ fructooligosaccharides,²⁹ inulin,^{30,34} and oligofructose and inulin.²⁸ Dietary fibers are known to elicit differential metabolic responses based on their physiochemical properties (i.e., viscosity, fermentability, soluble to insoluble ratio, etc.),^{35,36} thus conflicting findings between studies may be expected. For instance, the pea fiber supplement used in the present study had a total fiber content of 48%, composed of

Table 2. Lipoprotein distribution in adult male offspring

Outcome	CON	HC	HC + FBR
Total particle number			
Triglyceride-rich lipoproteins (nmol/L)	2.5 ± 0.3 ^a	4.4 ± 0.3 ^b	2.6 ± 1.0 ^{ab}
Low-density lipoprotein (nmol/L)	671.6 ± 74.6 ^a	693.7 ± 68.0 ^a	734.4 ± 77.1 ^a
High-density lipoprotein (µmol/L)	4.6 ± 0.3 ^a	4.4 ± 0.6 ^a	4.4 ± 0.4 ^a
Lipoprotein size			
Triglyceride-rich lipoproteins (nm)	92.9 ± 1.3 ^a	90.7 ± 2.1 ^a	79.8 ± 5.7 ^b
Low-density lipoprotein (nm)	21.5 ± 0.1 ^a	20.5 ± 0.2 ^b	21.9 ± 0.2 ^a
High-density lipoprotein (nm)	10.9 ± 0.1 ^a	11.0 ± 0.1 ^a	10.9 ± 0.2 ^a

CON, low-calorie control diet; HC, high-calorie obese-inducing diet; HC + FBR, HC high calorie diet with supplemental yellow-pea fiber (25%).
^{a,b,c}Treatment groups that do not share a superscript are significantly different ($p < 0.05$). Data are means ± SE, $n = 8-10$ mothers per group.

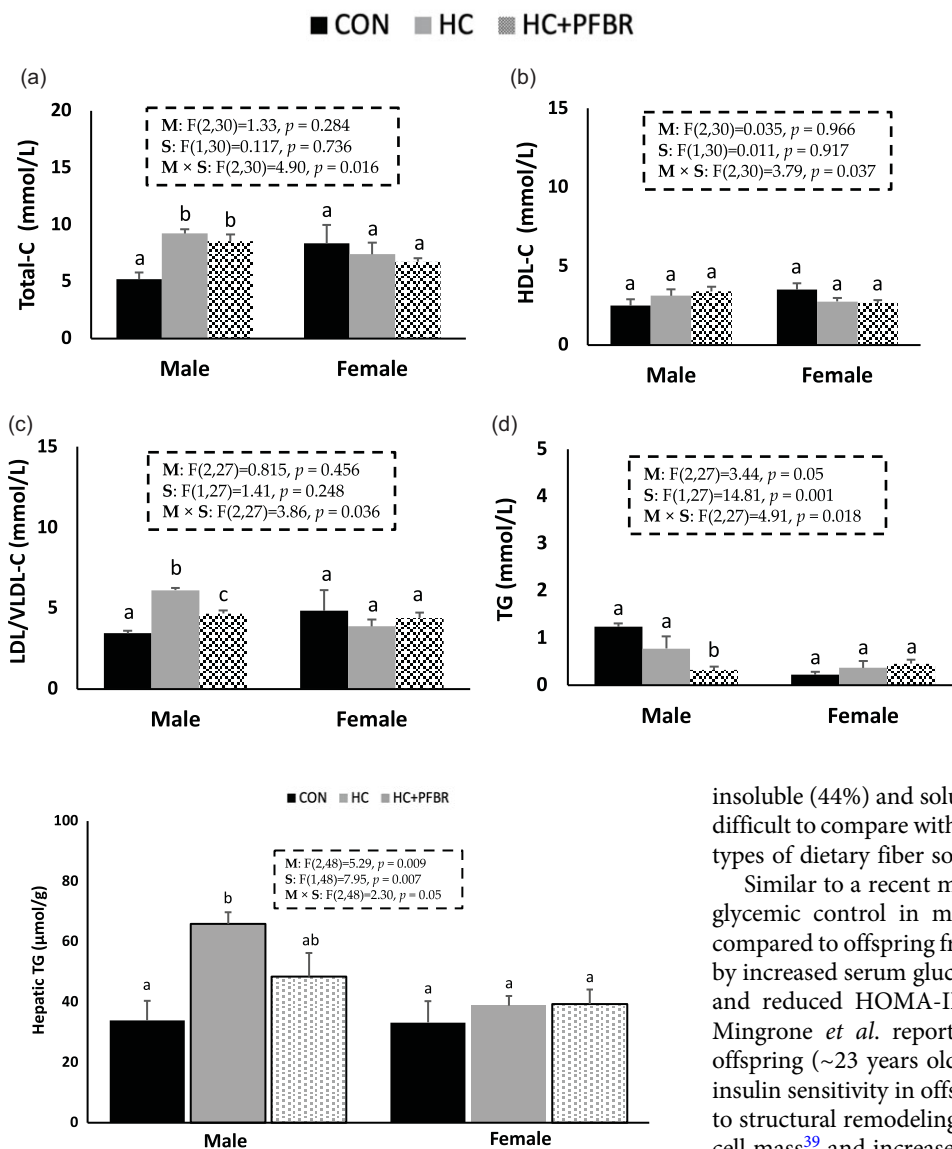


Figure 3. Lipid profile in adult male and female offspring from mothers consuming a control [CON], high calorie [HC], and HC + yellow-pea fiber [HC + FBR] diet including [a] Serum total cholesterol (Total-C) (mg/dL); [b] Serum HDL-C (mg/dL); [c] Serum LDL/VLDL-C (mg/dL); and [d] Serum triglycerides (TG) (mg/dL) in male and female offspring. Data are means ± SE; $n = 8-10$ per group; ^{ab} groups that do not share a superscript are significantly different ($p < 0.05$).

Figure 4. Hepatic triglycerides (µmol/g) in adult male and female offspring from mothers consuming a control [CON], high calorie [HC], and HC + yellow-pea fiber [HC + FBR] diet. Data are means ± SE; $n = 8-10$ per group; ^{ab} groups that do not share a superscript are significantly different ($p < 0.05$).

insoluble (44%) and soluble (4.5%) fiber fractions, likely making it difficult to compare with other studies using different amounts and types of dietary fiber sources.

Similar to a recent mouse model study,³⁷ we observed reduced glycemic control in males and females from HC-fed mothers compared to offspring from lean (CON) mothers, as demonstrated by increased serum glucose, increased serum insulin (males only), and reduced HOMA-IR. Further, in a previous human study, Mingrone *et al.* reported reduced glucose clearance in female offspring (~23 years old) from obese vs lean mothers.³⁸ Reduced insulin sensitivity in offspring from obese pregnancies may be due to structural remodeling of pancreatic islets, including reduced β -cell mass³⁹ and increased α -cells,^{40,41} and decreased expression of genes involved in insulin secretion.^{42,43}

Dietary pea fiber intake has been reported to improve glycemic control in diet-induced obese male rats, an effect which may be

related to changes in the gut microbiome community^{44,45} as well as increased expression of genes involved in gut epithelial function and short chain fatty acid production.⁴⁵ To our knowledge, maternal pea fiber intake has not been previously assessed in pregnancy, however, other types of dietary fibers have shown promise as a maternal intervention to prevent the programming of metabolic dysfunction in offspring. In a diet-induced Golden Syrian hamster model, Mohammed *et al.* 2022 reported that maternal dietary supplementation of fructooligosaccharides from preconception throughout lactation was effective at improving glycemic control in offspring from obese pregnancies.²⁹ Similarly, compared with offspring from low fiber-fed mothers, mouse offspring born to mothers consuming a high fiber inulin-supplemented diet during pregnancy reduced blood glucose and conferred resistance to adult-onset obesity by beneficially shifting maternal gut microbiota composition.³⁰

Consistent with other studies in similar obese rat models,^{46,47} we observed a range of blood lipid abnormalities in adult male offspring from obese vs. lean mothers including higher serum concentration of total-C and LDL/VLDL-C. In addition to changes in the serum cholesterol profile, maternal obesity altered lipoprotein distribution in male offspring with evidence of a higher concentration of TG-rich lipoproteins and smaller LDL particles. The mechanism(s) underlying these responses are not completely understood but are likely associated with the reduction in insulin sensitivity observed in HC offspring compared with offspring from lean (CON) mothers. Insulin influences both postprandial and fasting lipid metabolism by regulating the activity of peripheral lipases and lipid transfer proteins, lipid uptake and efflux transporters, intracellular lipid synthesis and oxidation pathways, and the production and secretion of lipoproteins.^{48,49} Consequently, the blood lipid and lipoprotein profile in type 2 diabetes is exemplified by elevated TG and LDL-C concentrations, large TG-rich VLDL particles, and small LDL particles that are prone to oxidation.^{50–52} Although we are not aware of previous studies that have assessed potential perturbations in lipoprotein distribution in offspring from obese pregnancies, obese mothers have been reported to have an increase in small, dense LDL particles in the third trimester,⁵³ indicating the potential fetal exposure to an atherogenic lipoprotein profile in an obese gestational environment. The observed lipoprotein profile in HC offspring may signal potential longer-term disease risk complications as LDL particle number has been reported to be more strongly associated with CVD than LDL-C alone.^{54,55} Further, several reports suggest that offspring from obese pregnancies have increased risk for cardiovascular events,^{56,57} however, the specific contribution of dyslipidemia to CVD incidence in offspring from obese pregnancies is not known.

As a maternal intervention, maternal pea fiber supplementation throughout prepregnancy, gestation, and lactation was protective against dyslipidemia induced by maternal obesity by largely normalizing serum LDL/VLDL-C concentration and LDL particle size and by reducing serum TG and the size of TG-rich lipoproteins (vs. both CON and HC). Outside of pregnancy, dietary fiber supplementation, including yellow-pea fiber,⁵⁸ has been shown to have numerous beneficial effects on serum lipid profiles in high fat-fed male animals.⁵⁹ However, few studies have assessed blood lipids in offspring from obese mothers supplemented with dietary fiber during pregnancy. Mohamed *et al.* (2022) reported that daily gavage of mung bean fructooligosaccharides to high fat-fed (60% energy from fat) mothers from preconception to the end of lactation reduced serum triglycerides (6 months of age) and

increased serum HDL concentration (at 6 and 12 months of age) in offspring.²⁹

With higher hepatic TG concentration in HC vs. CON male offspring, our findings support the association between maternal obesity and nonalcoholic fatty liver disease in offspring reported in previous rodent model^{60,61} and human investigations.^{62,63} A similar sex-specific predisposition to nonalcoholic fatty liver disease was reported in F1 male offspring from obese mothers in a Wistar rat model.⁶⁴ Previous work suggests that this connection may be mediated through a variety of mechanisms including reduction in the autophagy-mediated degradation of cytoplasmic lipid droplets,⁶¹ liver transcriptome changes in genes that regulate liver lipid metabolism,⁶⁴ innate immune dysfunction,⁶⁰ and persistent shifts in hepatic DNA methylation and cecal microbiome populations.^{65,66} Maternal yellow-pea fiber consumption offered only partial protection against steatosis in male adult offspring and did not reach statistical significance ($p = 0.07$). In a previous study in male rats, consumption of yellow-pea flour, but not fiber, resulted in a lower hepatic TG concentration.⁴⁴ Additionally, we recently reported that maternal consumption of a yellow-pea protein-supplemented diet throughout pregnancy and lactation protected against hepatic TG accumulation in male offspring.²¹ Hence, the benefits of yellow peas with respect to hepatic steatosis are likely fraction-specific and the greatest potential benefits may be realized through consumption of whole yellow peas or by combining different yellow-pea fractions to obtain a specific desired metabolic effect. Interestingly, a recent longitudinal study in humans using multiple 24-h maternal pregnancy diet recalls and MRI assessment of hepatic fat in children (4–8 years old) reported that higher maternal fiber intake was associated with lower hepatic fat in offspring. This beneficial association was observed for total dietary fiber intake, however, the potential effects of different fiber types (i.e., soluble vs. insoluble) or food fiber source were not assessed.⁶⁷

In summary, our findings suggest that maternal obesity programs a sexual dimorphic obesogenic response in adult male offspring that cannot be rescued through the postnatal consumption of a low-calorie diet. Additionally, offspring from obese pregnancies showed evidence of reduced glycemic control (both males and females) and dyslipidemia (males only). Supplementation of a maternal high calorie diet with yellow-pea fiber in prepregnancy and throughout gestation and lactation protects male offspring from metabolic dysfunction in the absence of any change in body weight status in adulthood.

Acknowledgments. We would like to thank the staff of the Laboratory Animal Facility at the University at Buffalo for their assistance in the care and maintenance of the animals involved in this study.

Financial support. This research was supported by the United States Department of Agriculture, USDA Pulse Crop Health Initiative (58-3060-0-049, to TCR).

Competing interests. None.

References

1. Driscoll AK, Gregory ECW. Increases in prepregnancy obesity: United States, 2016–2019. *NCHS Data Brief*. 2020; 392, 1–8.
2. Waddell IS, Orfila C. Dietary fiber in the prevention of obesity and obesity-related chronic diseases: from epidemiological evidence to potential molecular mechanisms. *Crit Rev Food Sci Nutr*. 2022; 63(27), 1–16. DOI: [10.1080/10408398.2022.2061909](https://doi.org/10.1080/10408398.2022.2061909).

3. Chooi YC, Ding C, Magkos F. The epidemiology of obesity. *Metabolism*. 2019; 92, 6–10.
4. Di Gesu CM, Matz LM, Buffington SA. Diet-induced dysbiosis of the maternal gut microbiome in early life programming of neurodevelopmental disorders. *Neurosci Res*. 2021; 168, 3–19.
5. Ogunwale SM, Zera CA, Stanford FC. Obesity management in women of reproductive age. *JAMA*. 2021; 325(5), 433–434.
6. Catalano PM, Shankar K. Obesity and pregnancy: mechanisms of short term and long term adverse consequences for mother and child. *BMJ*. 2017; 356, j1.
7. Howell KR, Powell TL. Effects of maternal obesity on placental function and fetal development. *Reproduction*. 2017; 153(3), R97–R108.
8. Siega-Riz AM, Bodnar LM, Stotland NE, Stang J. The current understanding of gestational weight gain among women with obesity and the need for future research. *NAM Perspect*. 2020; 2020.
9. Johnson VR, Anekwe CV, Washington TB, Chhabria S, Tu L, Stanford FC. A women's health perspective on managing obesity. *Prog Cardiovasc Dis*. 2023; 78, 11–16. DOI: [10.1016/j.pcad.2023.04.007](https://doi.org/10.1016/j.pcad.2023.04.007).
10. Louise J, Poprzeczny AJ, Deussen AR, et al. The effects of dietary and lifestyle interventions among pregnant women with overweight or obesity on early childhood outcomes: an individual participant data meta-analysis from randomised trials. *BMC Med*. 2021; 19(1), 128.
11. Benítez-Páez A, Gómez Del Pulgar EM, Kjølbaek L, et al. Impact of dietary fiber and fat on gut microbiota re-modeling and metabolic health. *Trends Food Sci Tech*. 2016; 57, 201–212.
12. Haenen D, Zhang J, Souza da Silva C, et al. A diet high in resistant starch modulates microbiota composition, SCFA concentrations, and gene expression in pig intestine. *J Nutr*. 2013; 143(3), 274–283.
13. Lambert JE, Parnell JA, Tunnicliffe JM, Han J, Sturzenegger T, Reimer RA. Consuming yellow pea fiber reduces voluntary energy intake and body fat in overweight/obese adults in a 12-week randomized controlled trial. *Clin Nutr*. 2017; 36(1), 126–133.
14. Kim SJ, de Souza RJ, Choo VL, et al. Effects of dietary pulse consumption on body weight: a systematic review and meta-analysis of randomized controlled trials. *Am J Clin Nutr*. 2016; 103(5), 1213–1223.
15. Venn BJ, Perry T, Green TJ, et al. The effect of increasing consumption of pulses and wholegrains in obese people: a randomized controlled trial. *J Am Coll Nutr*. 2010; 29(4), 365–372.
16. Mollard RC, Luhovyy BL, Panahi S, Nunez M, Hanley A, Anderson GH. Regular consumption of pulses for 8 weeks reduces metabolic syndrome risk factors in overweight and obese adults. *Br J Nutr*. 2012; 108(S1), S111–122.
17. Ghavami A, Ziaei R, Talebi S, et al. Soluble fiber supplementation and serum lipid profile: a systematic review and dose-response meta-analysis of randomized controlled trials. *Adv Nutr*. 2023; 14(3), 465–474.
18. Xu B, Fu J, Qiao Y, et al. Higher intake of microbiota-accessible carbohydrates and improved cardiometabolic risk factors: a meta-analysis and umbrella review of dietary management in patients with type 2 diabetes. *Am J Clin Nutr*. 2021; 113(6), 1515–1530.
19. Heyne GW, Plisch EH, Melberg CG, Sandgren EP, Peter JA, Lipinski RJ. A simple and reliable method for early pregnancy detection in inbred mice. *J Am Assoc Lab Anim Sci*. 2015; 54(4), 368–371.
20. Jeyarajah EJ, Cromwell WC, Otvos JD. Lipoprotein particle analysis by nuclear magnetic resonance spectroscopy. *Clin Lab Med*. 2006; 26(4), 847–870.
21. Rideout TC, Andreani GA, Pembroke J, et al. Maternal pea protein intake provides sex-specific protection against dyslipidemia in offspring from obese pregnancies. *Nutrients*. 2023; 15(4), 867.
22. Tajaddini A, Kendig MD, Prates KV, Westbrook RF, Morris MJ. Male rat offspring are more impacted by maternal obesity induced by cafeteria diet than females-additive effect of postweaning diet. *Int J Mol Sci*. 2022; 23(3), 1442.
23. Kulhanek D, Abrahante Llorens JE, Buckley L, Tkac I, Rao R, Paulsen ME. Female and male C57BL/6J offspring exposed to maternal obesogenic diet develop altered hypothalamic energy metabolism in adulthood. *Am J Physiol Endocrinol Metab*. 2022; 323(5), E448–E466.
24. Kirk SL, Samuelsson AM, Argenton M, et al. Maternal obesity induced by diet in rats permanently influences central processes regulating food intake in offspring. *PLoS One*. 2009; 4(6), e5870.
25. Samuelsson AM, Matthews PA, Argenton M, et al. Diet-induced obesity in female mice leads to offspring hyperphagia, adiposity, hypertension, and insulin resistance: a novel murine model of developmental programming. *Hypertension*. 2008; 51(2), 383–392.
26. Desai M, Ferrini MG, Han G, Narwani K, Ross MG. Maternal high fat diet programs male mice offspring hyperphagia and obesity: mechanism of increased appetite neurons via altered neurogenic factors and nutrient sensor AMPK. *Nutrients*. 2020; 12(11), 3326.
27. Desai M, Han G, Ross MG. Programmed hyperphagia in offspring of obese dams: altered expression of hypothalamic nutrient sensors, neurogenic factors and epigenetic modulators. *Appetite*. 2016; 99, 193–199.
28. Hallam MC, Reimer RA. A maternal high-protein diet predisposes female offspring to increased fat mass in adulthood whereas a prebiotic fibre diet decreases fat mass in rats. *Br J Nutr*. 2013; 110(9), 1732–1741.
29. Mohammed S, Qadri SSY, Mir IA, Kondapalli NB, Basak S, Rajkumar H. Fructooligosaccharide ameliorates high-fat induced intrauterine inflammation and improves lipid profile in the hamster offspring. *J Nutr Biochem*. 2022; 101, 108925.
30. Kimura I, Miyamoto J, Ohue-Kitano R, et al. Maternal gut microbiota in pregnancy influences offspring metabolic phenotype in mice. *Science*. 2020; 367(6481).
31. Heard-Lipsmeier ME, Diaz EC, Sims CR, et al. Maternal adiposity is associated with fat mass accretion in female but not male offspring during the First 2 Years of life. *Obesity (Silver Spring)*. 2020; 28(3), 624–630.
32. Pregaica I, Alves A, Nunes S, et al. Diet-induced rodent models of obesity-related metabolic disorders-A guide to a translational perspective. *Obes Rev*. 2020; 21(12), e13081.
33. Desbuard N, Gourbeyre P, Haure-Mirande V, Darmaun D, Champ M, Bodinier M. Impact of perinatal prebiotic consumption on gestating mice and their offspring: a preliminary report. *Br J Nutr*. 2012; 107(9), 1245–1248.
34. Hsu CN, Lin YJ, Hou CY, Tain YL. Maternal administration of probiotic or prebiotic prevents male adult rat offspring against developmental programming of hypertension induced by high fructose consumption in pregnancy and lactation. *Nutrients*. 2018; 10(9), 1229.
35. Fak F, Jakobsdottir G, Kulcinskaja E, et al. The physico-chemical properties of dietary fibre determine metabolic responses, short-chain fatty acid profiles and gut microbiota composition in rats fed low- and high-fat diets. *PLoS One*. 2015; 10(5), e0127252.
36. Weitkunat K, Stuhlmann C, Postel A, et al. Short-chain fatty acids and inulin, but not guar gum, prevent diet-induced obesity and insulin resistance through differential mechanisms in mice. *Sci Rep*. 2017; 7(1), 6109.
37. Nicholas LM, Nagao M, Kusinski LC, Fernandez-Twinn DS, Eliasson L, Ozanne SE. Exposure to maternal obesity programs sex differences in pancreatic islets of the offspring in mice. *Diabetologia*. 2020; 63(2), 324–337.
38. Mingrone G, Manco M, Mora ME, et al. Influence of maternal obesity on insulin sensitivity and secretion in offspring. *Diabetes Care*. 2008; 31(9), 1872–1876.
39. Bringhenti I, Ornellas F, Martins MA, Mandarim-de-Lacerda CA, Aguila MB. Early hepatic insult in the offspring of obese maternal mice. *Nutr Res*. 2015; 35(2), 136–145.
40. Gregorio BM, Souza-Mello V, Mandarim-de-Lacerda CA, Aguila MB. Maternal high-fat diet is associated with altered pancreatic remodelling in mice offspring. *Eur J Nutr*. 2013; 52(2), 759–769.
41. Bringhenti I, Ornellas F, Mandarim-de-Lacerda CA, Aguila MB. The insulin-signaling pathway of the pancreatic islet is impaired in adult mice offspring of mothers fed a high-fat diet. *Nutrition*. 2016; 32(10), 1138–1143.
42. Yokomizo H, Inoguchi T, Sonoda N, et al. Maternal high-fat diet induces insulin resistance and deterioration of pancreatic beta-cell function in adult offspring with sex differences in mice. *Am J Physiol Endocrinol Metab*. 2014; 306(10), E1163–1175.
43. Han J, Xu J, Epstein PN, Liu YQ. Long-term effect of maternal obesity on pancreatic beta cells of offspring: reduced beta cell adaptation to high

- glucose and high-fat diet challenges in adult female mouse offspring. *Diabetologia*. 2005; 48(9), 1810–1818.
44. Eslinger AJ, Eller LK, Reimer RA. Yellow pea fiber improves glycemia and reduces clostridium leptum in diet-induced obese rats. *Nutr Res*. 2014; 34(8), 714–722.
 45. Hashemi Z, Foughse J, Im HS, Chan CB, Willing BP. Dietary pea fiber supplementation improves glycemia and induces changes in the composition of gut microbiota, serum short chain fatty acid profile and expression of Mucins in glucose intolerant rats. *Nutrients*. 2017; 9(11), 1236.
 46. Bariani MV, Correa F, Dominguez Rubio AP, *et al.* Maternal obesogenic diet combined with postnatal exposure to high-fat diet induces metabolic alterations in offspring. *J Cell Physiol*. 2020; 235(11), 8260–8269.
 47. Ribaroff GA, Wastnedge E, Drake AJ, Sharpe RM, Chambers TJG. Animal models of maternal high fat diet exposure and effects on metabolism in offspring: a meta-regression analysis. *Obes Rev*. 2017; 18(6), 673–686.
 48. Klop B, Elte JW, Cabezas MC. Dyslipidemia in obesity: mechanisms and potential targets. *Nutrients*. 2013; 5(4), 1218–1240.
 49. Titchenell PM, Lazar MA, Birnbaum MJ. Unraveling the regulation of hepatic metabolism by insulin. *Trends Endocrinol Metab*. 2017; 28(7), 497–505.
 50. Singh AT, Rainwater DL, Haffner SM, *et al.* Effect of diabetes on lipoprotein size. *Arterioscler Thromb Vasc Biol*. 1995; 15(11), 1805–1811.
 51. Dullaart RP, de Vries R, Lefrandt JD. Increased large VLDL and small LDL particles are related to lower bilirubin in Type 2 diabetes mellitus. *Clin Biochem*. 2014; 47(16–17), 170–175.
 52. Woodman RJ, Watts GF, Playford DA, Best JD, Chan DC. Oxidized LDL and small LDL particle size are independently predictive of a selective defect in microcirculatory endothelial function in type 2 diabetes. *Diabetes Obes Metab*. 2005; 7(5), 612–617.
 53. Meyer BJ, Stewart FM, Brown EA, *et al.* Maternal obesity is associated with the formation of small dense LDL and hypoadiponectinemia in the third trimester. *J Clin Endocrinol Metab*. 2013; 98(2), 643–652.
 54. Cromwell WC, Otvos JD, Keyes MJ, *et al.* LDL particle number and risk of future cardiovascular disease in the framingham offspring study - implications for LDL management. *J Clin Lipidol*. 2007; 1(6), 583–592.
 55. Aday AW, Lawler PR, Cook NR, Ridker PM, Mora S, Pradhan AD. Lipoprotein particle profiles, standard lipids, and peripheral artery disease incidence. *Circulation*. 2018; 138(21), 2330–2341.
 56. Reynolds RM, Allan KM, Raja EA, *et al.* Maternal obesity during pregnancy and premature mortality from cardiovascular event in adult offspring: follow-up of 1 323 275 person years. *BMJ*. 2013; 347(aug13 1), f4539–f4539.
 57. Cooper R, Power C. Pregnancy obesity is associated with increased rates of all-cause mortality and cardiovascular hospital admissions in adult offspring. *Evid Based Nurs*. 2014; 17(4), 104–104.
 58. Liu G, Xiao L, Fang T, *et al.* Pea fiber and wheat bran fiber show distinct metabolic profiles in rats as investigated by a 1H NMR-based metabolomic approach. *PLoS One*. 2014; 9(12), e115561.
 59. Han J, Zhang R, Muheyati D, Lv MX, Aikebaier W, Peng BX. The effect of chickpea dietary fiber on lipid metabolism and gut microbiota in high-fat diet-induced hyperlipidemia in rats. *J Med Food*. 2021; 24(2), 124–134.
 60. Mouralidarane A, Soeda J, Visconti-Pugmire C, *et al.* Maternal obesity programs offspring nonalcoholic fatty liver disease by innate immune dysfunction in mice. *Hepatology*. 2013; 58(1), 128–138.
 61. Han S, Zhu F, Huang X, *et al.* Maternal obesity accelerated non-alcoholic fatty liver disease in offspring mice by reducing autophagy. *Exp Ther Med*. 2021; 22(1), 716.
 62. Hagstrom H, Simon TG, Roelstraete B, Stephansson O, Soderling J, Ludvigsson JF. Maternal obesity increases the risk and severity of NAFLD in offspring. *J Hepatol*. 2021; 75(5), 1042–1048.
 63. Zeng J, Shen F, Zou ZY, *et al.* Association of maternal obesity and gestational diabetes mellitus with overweight/obesity and fatty liver risk in offspring. *World J Gastroenterol*. 2022; 28(16), 1681–1691.
 64. Lomas-Soria C, Reyes-Castro LA, Rodriguez-Gonzalez GL, *et al.* Maternal obesity has sex-dependent effects on insulin, glucose and lipid metabolism and the liver transcriptome in young adult rat offspring. *J Physiol*. 2018; 596(19), 4611–4628.
 65. Wankhade UD, Zhong Y, Kang P, *et al.* Enhanced offspring predisposition to steatohepatitis with maternal high-fat diet is associated with epigenetic and microbiome alterations. *PLoS One*. 2017; 12(4), e0175675.
 66. Paul HA, Collins KH, Nicolucci AC, *et al.* Maternal prebiotic supplementation reduces fatty liver development in offspring through altered microbial and metabolomic profiles in rats. *FASEB J*. 2019; 33(4), 5153–5167.
 67. Cohen CC, Perng W, Sauder KA, *et al.* Maternal diet quality during pregnancy and offspring hepatic fat in early childhood: the healthy start study. *J Nutr*. 2023; 153(4), 1122–1132.